## 1 **ELECTRONIC SUPPLEMENTARY MATERIAL**

3 Christie, K. and S.Y. Strauss. (2020). Frequency-dependent fitness and reproductive dynamics<br>4 contribute to habitat segregation in sympatric Jewelflowers. *Proceedings of the Royal Society B*: 4 contribute to habitat segregation in sympatric Jewelflowers. *Proceedings of the Royal Society B:*  5 *Biological Sciences.* DOI/10.1098/rspb.2020.0559.

## 7 **List of Supplementary figures**

9 Figure S1: Seed production and seed viability in experimental crosses using conspecific, mixed, and heterospecific pollen.

12 Figure S2: NMDS ordination of physical site attributes, including soil texture and soil chemistry, of *S. breweri* and 13 *S. hesperidis* sites at McLaughlin.

15 Figure S3: Gravimetric water content of field soils at six adjacent *S. breweri* and *S. hesperidis* sites throughout the growing season.

Figure S4: Germination success in habitat-specific soils in the greenhouse.

Figure S5: Plant growth in a greenhouse competition experiment.

### **List of Supplementary tables**

Table S0: Fixed-effects summary of GLMM predicting seed viability in experimental crosses.

26 Table S1: Summary of physical site attributes, including soil texture and soil chemistry, of *S. breweri* and *S. hesperidis* sites at McLaughlin.

29 Table S2.1: Fixed-effects summary of GLMM predicting germination success at two field sites in 2015.

Table S2.2: Summary of GLM predicting germination success in habitat-specific soils in the greenhouse.

Table S2.3: Fixed-effects summary of GLMM predicting survival in field transplant experiment.

Table S2.4: Fixed-effects summary of GLMM predicting plant growth in field transplant experiment.

Table S2.5: Fixed-effects summary of GLMM predicting fruit number in field transplant experiment.

Table S2.6: Summary of GLM predicting survival in lathhouse soil transplant experiment.

Table S2.7: Summary of GLM predicting plant growth in lathhouse soil transplant experiment.

Table S2.8: Summary of GLM predicting flower number in lathhouse soil transplant experiment.

Table S3.1: Summary of GLM predicting seed production in lathhouse soil transplant.

Table S3.2: Summary of GLM predicting seed viability in lathhouse soil transplant.

Table S3.3: Summary of GLMM predicting seed production in field transplant experiment.

Table S3.4: Summary of GLMM predicting seed viability in field transplant experiment.

Table S4: Model selection table showing best-fitting models explaining seed viability in experimental migrants.

Table S5: Fixed-effects summary of best-fitting GLMM predicting seed viability in experimental migrants.

#### *Experimental pollination*

 To assess the effects of heterospecific pollen transfer on seed viability we performed experimental crosses in the greenhouse using conspecific, heterospecific, and 50:50 pollen mixtures. We made crosses using individuals from six adjacent sites at McLaughlin, the same sites used in the field transplant experiment. We removed mature anthers from paternal plants with forceps, and manually applied pollen using anthers as paintbrushes, to the receptive stigmatic surfaces of bud-emasculated flowers of maternal plants. For the 50:50 pollen mixtures, we first completely saturated one half of the stigmatic surface with one species' pollen, and then completely covered the other half of the stigmatic surface with the other species' pollen in immediate succession. We alternated the order in which we applied pollen, and saturated stigmatic surfaces with pollen, far in excess of the number available ovules. In total, we made 291 crosses, including 97 intraspecific, 83 interspecific, 92 mixed-pollen, and 19 67 control crosses, using multiple maternal ( $n = 24$ ) and paternal ( $n = 38$ ) donors. Of these, 118 successfully 68 set fruit; we used these crosses ( $n = 42$  conspecific pollen,  $n = 42$  mixed pollen,  $n = 34$  heterospecific pollen) to determine if seed viability differed depending on pollen treatment.

 We modeled seed viability (viable or inviable) using a binomial GLMM in R (g*lmer* function from the *lme4* package) with maternal species (*S. breweri, S. hesperidis*), pollen treatment (conspecific, 50:50 mixed, heterospecific), and a maternal species\*pollen treatment interaction as fixed effects, with maternal and paternal individuals as random effects. We tested for differences among pollen treatments with Tukey's HSD tests using the *glht* function from the *multcomp* package in R.

#### *Field soil moisture*

 To determine if soils at sites occupied by *S. breweri* and *S. hesperidis* differed in their water holding capacity, we measured gravimetric water content at six adjacent sites at three time points throughout the growing season (04/17/17, 05/14/17, 06/14/17). At each site, at each time point, we 80 collected 10 soil cores from immediately beneath native plants at randomly selected microsites ( $n = 60$ ) total samples per time point). Cores consisted of the top 15cm of the substrate, the predominant area of root growth for *S. breweri* and *S. hesperidis*. We passed soils through 4mm and then 2mm sieves, transferred 20mL aliquots of field-wet soil into aluminum boats, and dried the soils at 100 C**°** overnight. 84 We used the weights of field-wet soils and dried soils to calculate gravimetric water content ( $\Theta_d$  = [weight of wet soil - weight of dry soil] / weight of dry soil). We tested for differences in water-holding 86 capacity between sites occupied by each species using t-tests (04/17/17 and 06/14/17 measurements) and 87 Mann-Whitney tests (05/14/17 measurements) with Bonferroni correction.

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#### *Germination*

 Two hundred seeds, of a total of 1000 seeds planted in one S*. breweri* site and an adjacent *S. hesperidis* site, germinated in the field in 2015. In addition to assessing germination success in different habitats/soils in the field (see main text), we also conducted a follow-up experiment in the greenhouse, using seeds and soils collected from all six sites represented in the field transplant experiment. Here, we 95 first collected field soil from immediately under native plants, then planted field-collected seeds ( $n = 240$ ) total) into field soils in germination trays, and placed trays under a mist-bench in the greenhouse. The trays received automated mist for five minutes each hour, and ambient springtime light and temperature conditions in the UC Davis greenhouse. We assayed germination weekly for five weeks and counted the total number of germinants. We modeled germination success using a binomial GLM in R with species, 100 soil type, and a species\*soil type interaction as fixed effects.

#### *Lathhouse soil transplant*

 We first collected 76 soil cores from six adjacent *S. breweri* and *S. hesperidis* sites at McLaughlin (n soil cores = 152 total). While keeping the soil structure as undisturbed as possible, we transferred soil cores into cylindrical greenhouse pots (D40 Deepots), and then transplanted greenhouse-grown 106 germinants ( $n = 76$  per species) into site-specific soils. We grew plants in a mesh-covered lathhouse at UC Davis, and used a conservative watering schedule, and ambient light and temperature conditions in the late spring to approximate field conditions. We did not hand pollinate plants, but allowed insect 109 pollinators to freely visit the plants, entering and exiting the lathhouse through a wide mesh netting. We randomized the position of pots and racks every two weeks throughout the experiment. We scored survival and plant height at the end of the growing season as in our field experiment, and used flower number as a proxy for potential fruit production. We assessed patterns of local adaptation to specific soils using GLMs with species, soil type and a species\*soil type interaction as fixed effects.

 Additionally, to test the possibility that seed production and seed viability are associated with intrinsic properties of the soil (and unrelated to other ecological interactions in the field) we also scored 116 seed production and seed viability of all survivors that produced fruit ( $n = 77$ ), using a randomly-selected subset of five fruits per maternal individual. We modeled seed production (Gaussian) and seed viability (binomial) using GLMs in R with species, soil type, and a species\*soil type interaction as fixed effects. 

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**Supplementary Figures**



 Figure S1: Seed viability in experimental crosses using conspecific, mixed (50:50), and heterospecific pollen. The total height of each bar represents the total number of seeds produced; the blue (*S. breweri*) and green (*S. hesperidis*) portions represents the average number of viable seeds, and the light gray 130 portions represent the average number of inviable seeds. Text above each bar indicates median model 131 predictions of seed viability from a GLMM.

133 Seed viability was reduced in both heterospecific ( $p < 0.001$ ) and mixed pollen ( $p < 0.001$ ) treatments compared to conspecific pollinations (Table S0; Fig. S1), and all cross types were significantly different from one another (Tukey's HSD, all p-values < 0.001). *S. breweri* produced 91-99% viable seeds in conspecific crosses (90% confidence intervals from model predictions), 47-87% viable seeds in mixed-pollen crosses, and 2-12% viable seeds in heterospecific crosses. *S. hesperidis* produced 100% viable seeds in conspecific crosses, 30-84% viable seeds in mixed-pollen crosses, and 43-100% viable seeds in heterospecific crosses, however the majority of *S. hesperidis* crosses (including intraspecific crosses) failed in this experiment, so these predictions are based on only 13 successful crosses. Potentially due to this small sample size, we did not observe a significant species\*pollen source interaction (Table S0), however qualitatively, *S. hesperidis* suffered a less severe reduction in seed viability in heterospecific crosses compared to *S. breweri* (Fig. S1). This asymmetric reduction in seed viability following heterospecific pollen transfer is consistent with previous work showing less severe 145 intrinsic postzygotic reproductive isolation at the seed production stage for *S. hesperidis*<sup>57</sup>.





 Figure S2: NMDS ordination of 33 physical site attributes, including soil texture and soil chemistry, for 148 20 *S. breweri* (n = 10) and *S. hesperidis* (n = 10) sites at McLaughlin. Points represent individual sites; ellipses represent 95% confidence intervals for abiotic niche breadth. We found no evidence for abiotic niche differences with respect to the combination of physical site attributes, and soil texture and soil chemistry variables (*adonis2* permutation test, p = 0.49).





 Figure S3: Germination success in site-specific soils in the greenhouse. Points indicate mean model predictions, and error bars show 95% confidence intervals. Similar to findings from the field (Fig. 2A), we found no evidence for a home-soil germination advantage when we planted seeds in site-specific field

158 soils in the greenhouse (species\*soil source interaction,  $p = 0.30$ ). Seeds of both species germinated equally well in both soils.

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Figure S4: Average fitness associated with survival and growth in habitat-specific soils in the lathhouse.

A.) Survival; B.) Plant height at the end of the growing season; C.) Flower production. Points represent

mean model predictions; error bars show 95% confidence intervals; text depicts significance of

species\*soil type interactions.

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 Figure S5: A. Plant growth when each species competed with conspecific and heterospecific competitors in single pots in the greenhouse. We found no differences in growth depending upon whether the 181 competitor was a conspecific or a heterospecific (species\*competitor interaction,  $p = 0.12$ ). B. Plant growth for *S. breweri* (blue) when growing alone, with a conspecific competitor, and with a heterospecific competitor; treatments not sharing a letter are significantly different at the 95% confidence level using a Tukey's HSD test. C. The same data for *S. hesperidis* (green).

 There were no differences in the relative effects of intra- compared to interspecific competition for either species (Fig. S5B; Fig S5C). Interestingly, the smaller *S. hesperidis* (often 50% smaller than *S. breweri*) showed no decrease in height when it competed with its larger congener, when it competed with a conspecific, and when it grew without a competitor (Fig. S5C).

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# 197 **Supplementary Tables**

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Table S0: Seed viability in experimental pollinations in the greenhouse Model (binomial): seed viability $\sim$ maternal species + pollen treatment + maternal species*pollen treatment + $(1 $ maternal individual $) + (1 $ paternal individual $)$				
Factor	Estimate	<b>SE</b>	z value	p-value
maternal species (S. <i>hesperidis</i> )	15.7454	457.9476	0.034	0.973
treatment (mixed pollen)	$-2.5196$	0.2573	$-9.792$	$\leq$ 2E-16
treatment (heterospecific pollen)	$-6.4463$	0.3524	$-18.294$	$\leq$ 2E-16
maternal species $(S. \; h \neq is)$ * treatment (mixed pollen)	$-16.2169$	457.9469	$-0.035$	0.972
maternal species $(S. \; h \text{esperidis})$ * treatment (heterospecific pollen)	$-11.5811$	457.9494	$-0.025$	0.98

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### **Table S3.2: Seed viability in a lathhouse soil transplant experiment**

Model (binomial): seed viability  $\sim$  species + soil type + site + species\*soil type



# **Table S3.3: Seed production in a field transplant experiment**

Model (Gaussian): total seeds per fruit ~ species + habitat + site + species\*habitat + (1|experimental block)





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