Supporting Information

Cell-Permeable Cyclic Peptidyl Inhibitor against the Keap1-Nrf2 Interaction

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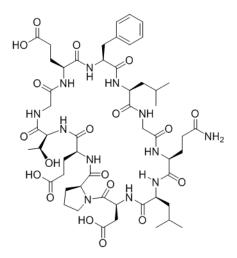
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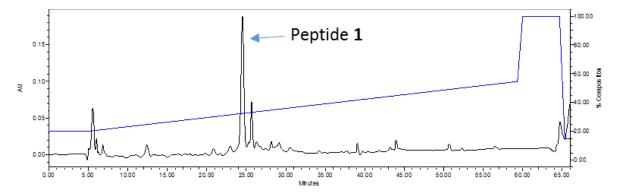
Figure S1	
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Figure S1. Structures, purity (as assessed by reversed-phase analytical HPLC), and HR-MS (MALDI FT-ICR) of peptides used in this work. Note: Some of the NF-labeled peptides eluted as two separate peaks, because the commercially available NF dye is a mixture of 5- and 6-carboxy isomers. The mixtures of two isomers were used in all experiments.

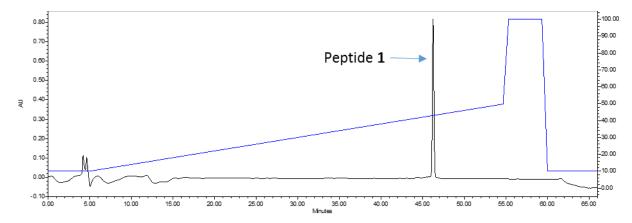
Peptide 1



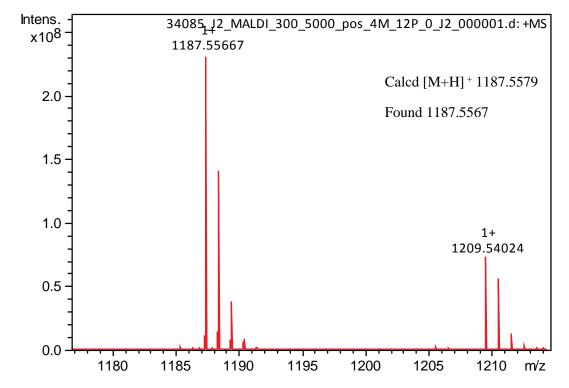
Crude peptide on semi-preparative reversed-phase HPLC (monitored at 214 nm):



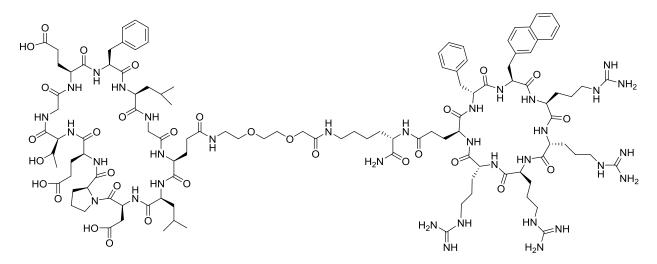
Purity check on analytical reversed-phase HPLC (214 nm):

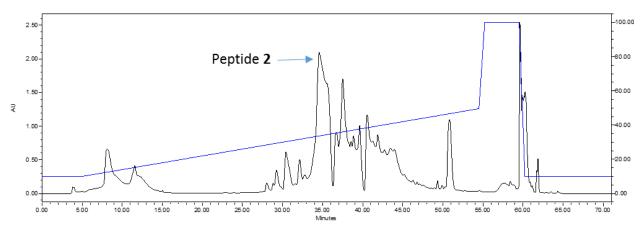






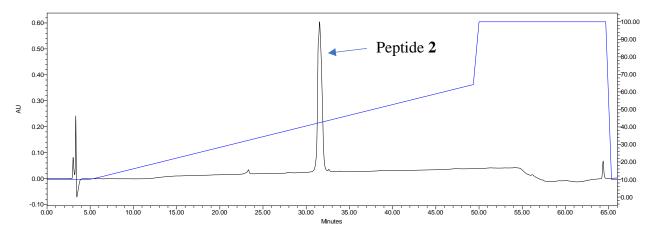
Peptide 2



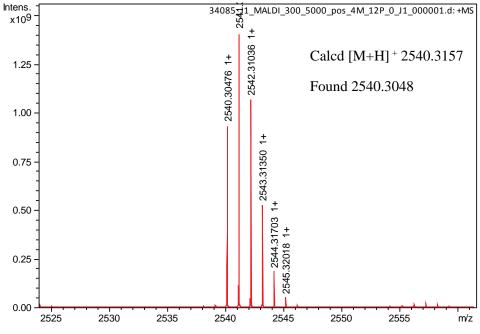


Crude peptide on semi-preparative reversed-phase HPLC (monitored at 214 nm):

Purity check (analytical reversed-phase HPLC at 214 nm):

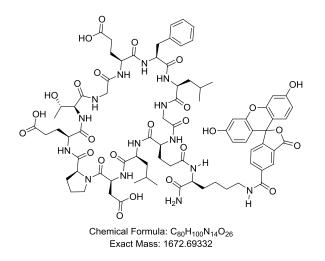




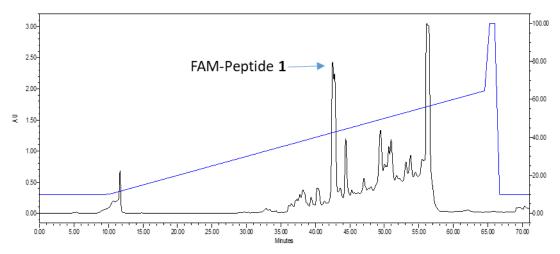


S4

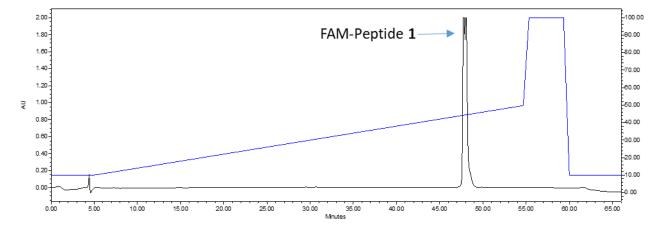
FAM-Peptide 1



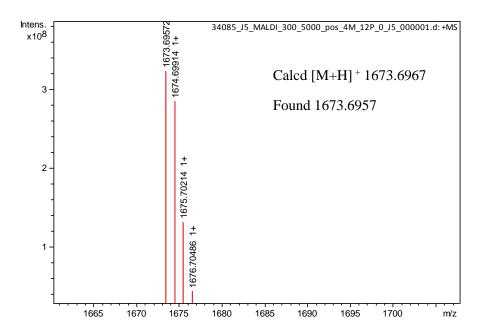
Crude peptide on semi-preparative reversed-phase HPLC (monitored at 214 nm):

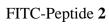


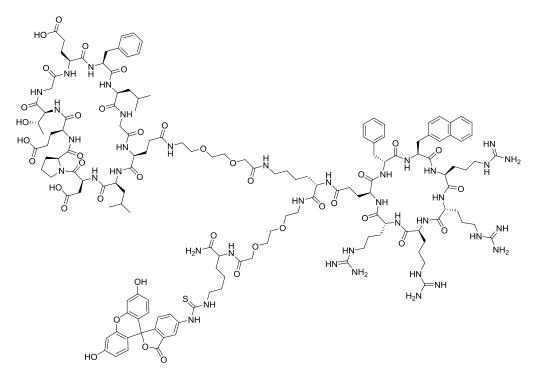
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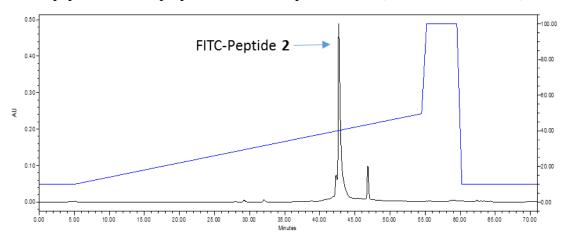






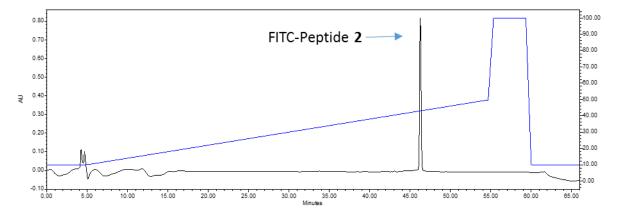




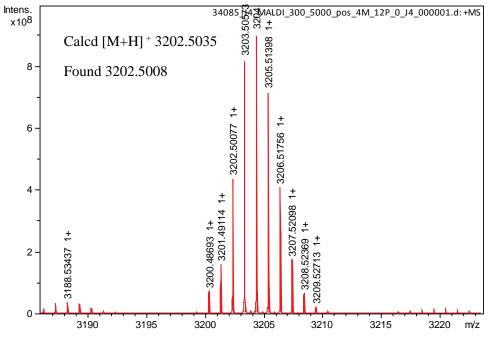


Crude peptide on semi-preparative reversed-phase HPLC (monitored at 214 nm):

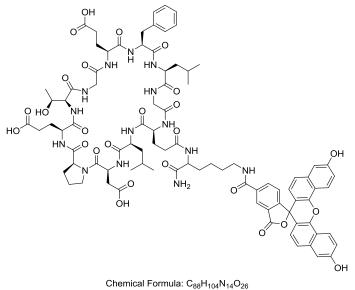
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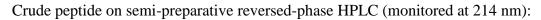


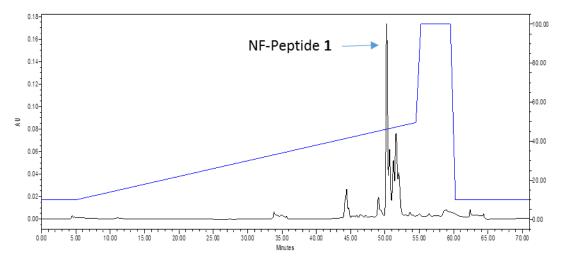


NF-Peptide 1

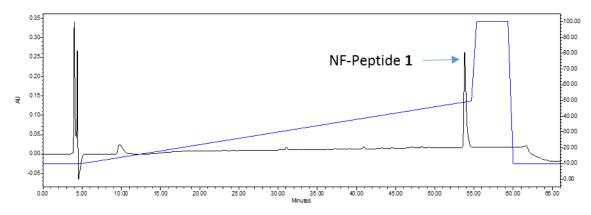


Exact Mass: 1772.72462

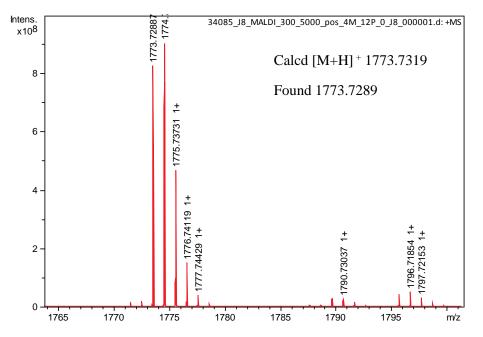


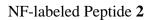


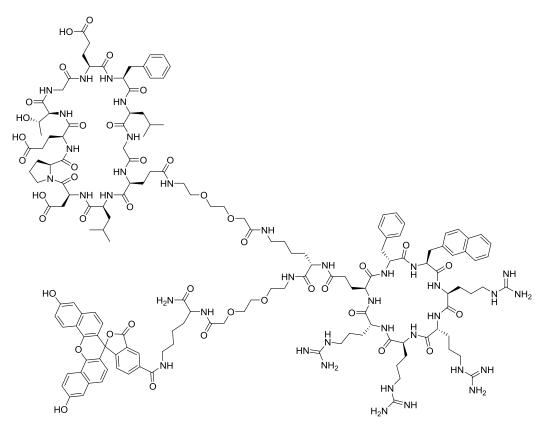
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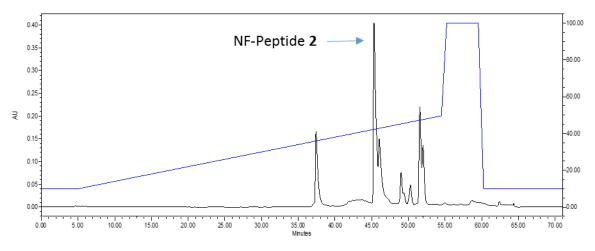






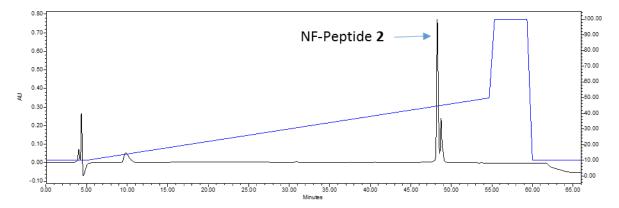


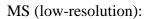


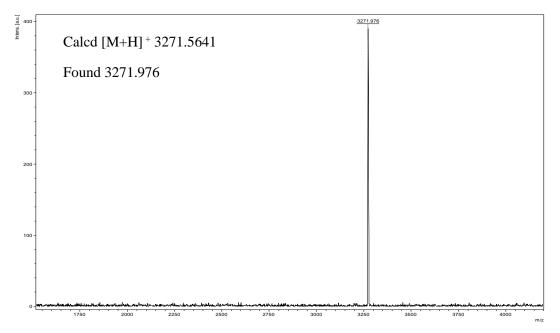


Crude peptide on semi-preparative reversed-phase HPLC (monitored at 214 nm):

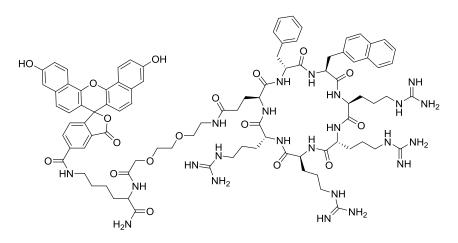
Purity check on analytical reversed-phase HPLC (214 nm):



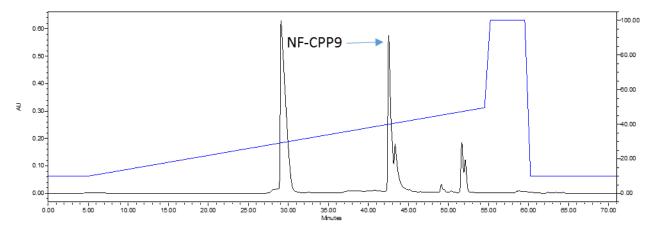




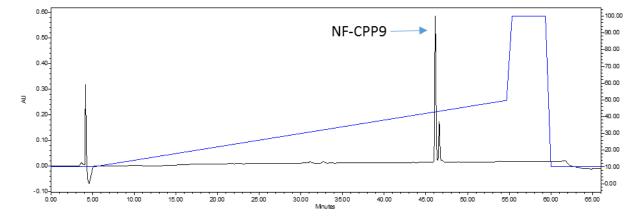
NF-CPP9



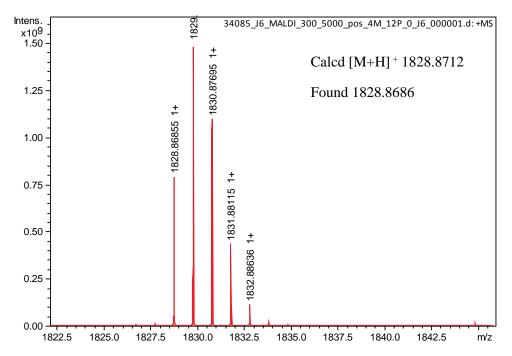
Crude peptide on semi-preparative reversed-phase HPLC (monitored at 214 nm):



Purity check on analytical reversed-phase HPLC (214 nm):







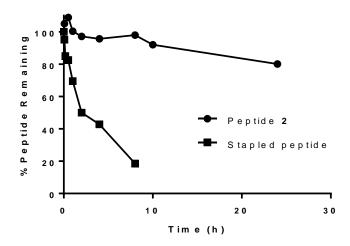


Figure S2. Serum stability of peptide **2** and a stapled peptide, Ac-GGYPED*ILDK*HLQRVIL-(miniPEG)₂-Dap-CPP9, in which an amide bond is formed between the side chains of D* and K*.

Procedure: Human serum was diluted to 25% in sterile DPBS and incubated at 37 °C for 15 min. Peptide was added to the diluted serum to a final concentration of 100 μ M and incubated at 37 °C with gentle mixing. Aliquots of 100 μ L were withdrawn at various time points and mixed with 100 μ L of 15% trichloroacetic acid in MeOH (w/v) and 100 μ L of acetonitrile to quench the reaction. After overnight storage at 4 °C, aliquots were centrifuged at 15000 g for 5 min at 4°C and analyzed by RP-HPLC on a C18 column. The amount of remaining peptide was assessed by integration of the area under the peak in the HPLC chromatogram and comparing it with that at zero-time point.

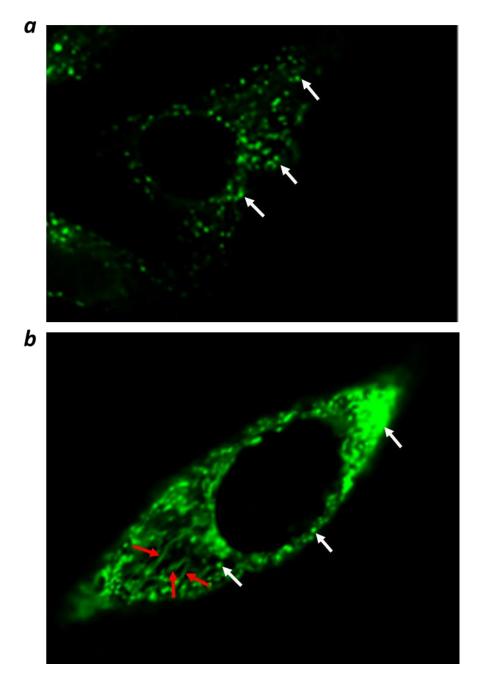


Figure S3. Enlarged confocal microscopy images from Figure 2 (main text) to show the differential intracellular localization of peptides 1 (*a*) and 2 (*b*). Representative round, intensely fluorescent structures (presumably endosomes and lysosomes) are indicated by white arrows, whereas the fibrous structures in (*b*) are marked with red arrows.

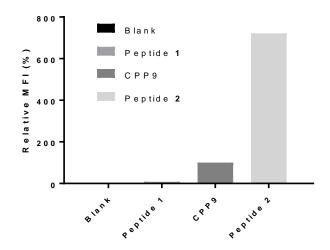


Figure S4. Cytosolic entry efficiencies of peptides 1 and 2. HeLa cells were treated with 5 μ M NF-labeled peptide for 2 h in the presence of 10% FBS, washed, and suspended in a pH 6.5 buffer immediately before flow cytometry analysis. All MFI values are relative to that of CPP9-NF (100%).

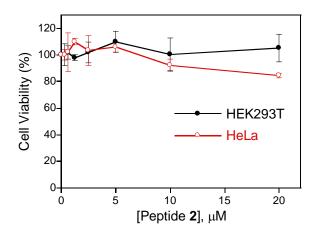


Figure S5. Effect of peptide 2 on the viability of HeLa and HEK293T cells as monitored by the MTT assay.