

## Supporting Information

### Novel Propargyl-linked Bisubstrate Analogs as Tight-binding

#### Inhibitors for Nicotinamide N-Methyltransferase

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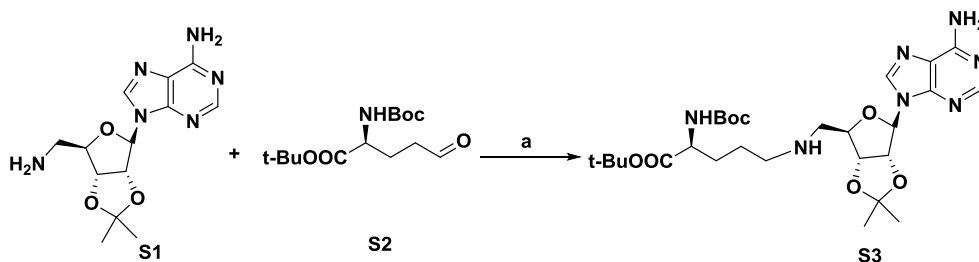
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## General procedures for the synthesis of S3

### Scheme S1. Synthetic route of S3.<sup>a</sup>

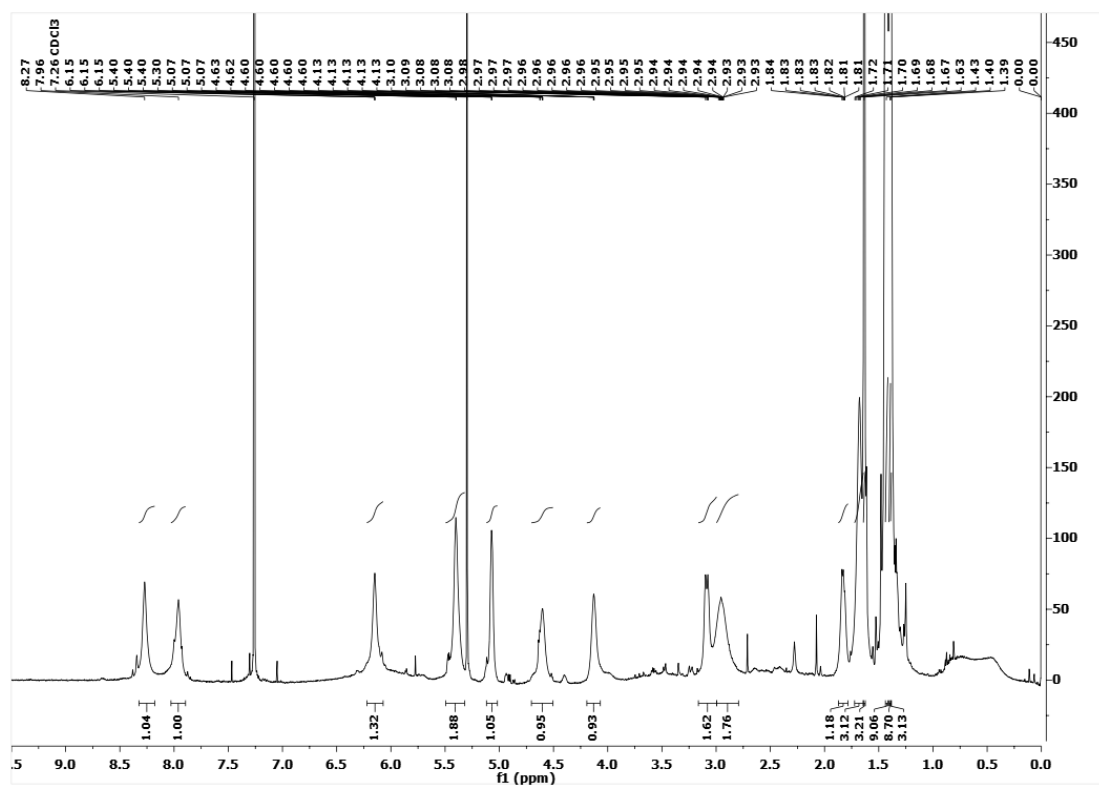
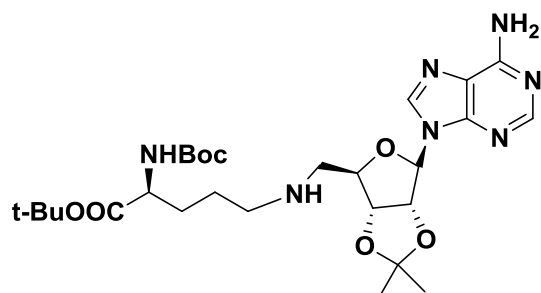


<sup>a</sup>Reagents and conditions: (a) NaBH<sub>3</sub>CN, MeOH, r.t., 64%.

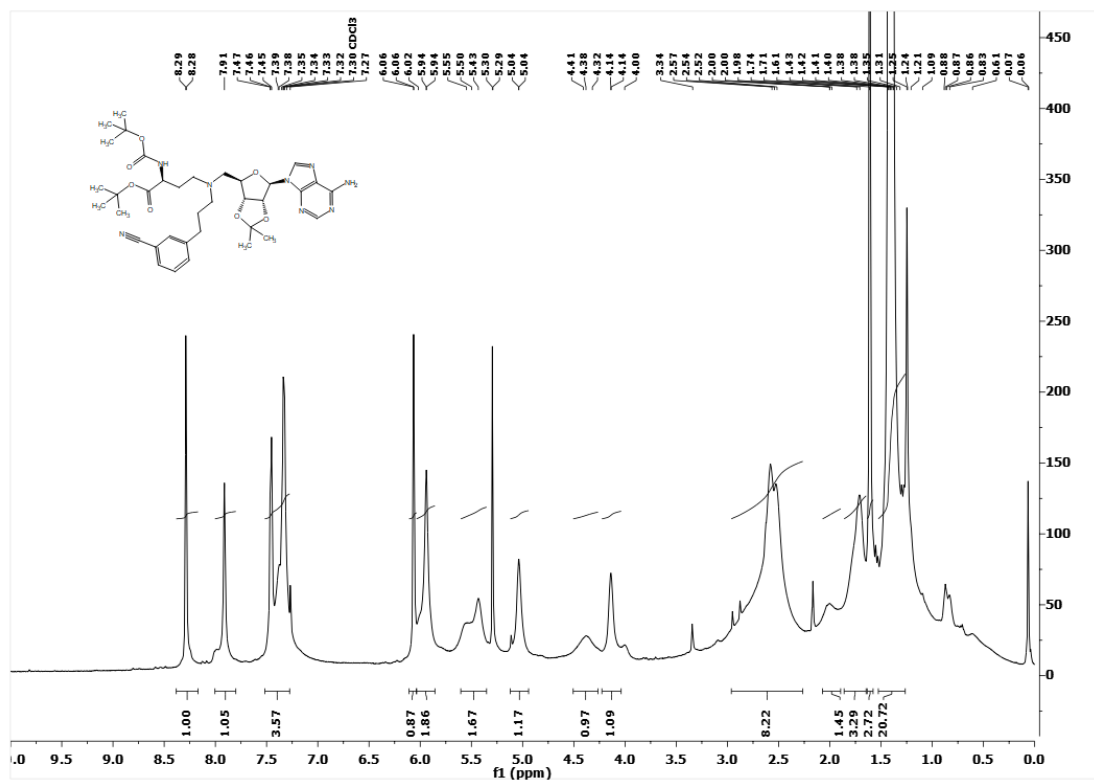
Tert-butyl (S)-5-((((3aR,4R,6S,6aR)-6-((6-amino-9H-purin-9-yl)-2,2-dimethyltetrahydrofuro[3,4-d][1,3]dioxol-4-yl)methyl)amino)-2-((tert-butoxycarbonyl)amino)pentanoate (**S3**). To a solution of 9-((3aR,4S,6R,6aR)-6-(aminomethyl)-2,2-dimethyltetrahydrofuro[3,4-d][1,3]dioxol-4-yl)-9H-purin-6-amine (**S1**)<sup>1</sup> (0.33 g, 1.0 mmol) in methanol, tert-butyl (S)-2-((tert-butoxycarbonyl)amino)-5-oxopentanoate (**S2**)<sup>2</sup> (0.4 g, 1.4 mmol) was added in to the reaction. The mixture stirred for 30min at r.t.. Then NaBH<sub>3</sub>CN (94 mg, 1.5 mmol) was add to the reaction. After 6 h, the solvent was removed in vacuum and the residue was purified by column chromatography (CH<sub>2</sub>Cl<sub>2</sub>:MeOH = 25: 1) to get 0.4 g product (64%). <sup>1</sup>H NMR (500 MHz, Chloroform-*d*) δ 8.27 (s, 1H), 7.96 (s, 1H), 6.22 – 6.07 (m, 1H), 5.50 – 5.32 (m, 2H), 5.12 – 5.02 (m, 1H), 4.70 – 4.50 (m, 1H), 4.19 – 4.07 (m, 1H), 3.16 – 3.00 (m, 2H), 3.00 – 2.79 (m, 2H), 1.87 – 1.78 (m, 1H), 1.72-1.65 (m, 3H), 1.63 (s, 3H), 1.43 (s, 9H), 1.40 (s, 9H), 1.39 (s, 3H).

# NMR spectra of compound S3, 6a-j, 9a-d, and 12a-b

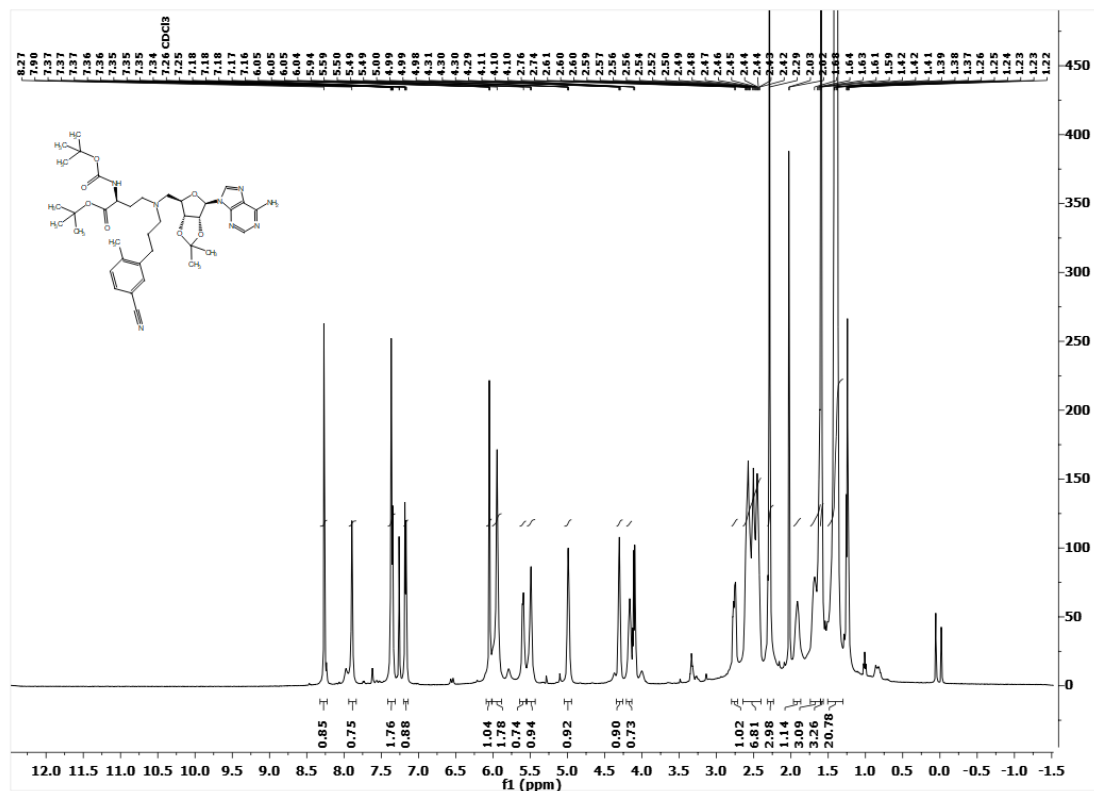
## HNMR of S3



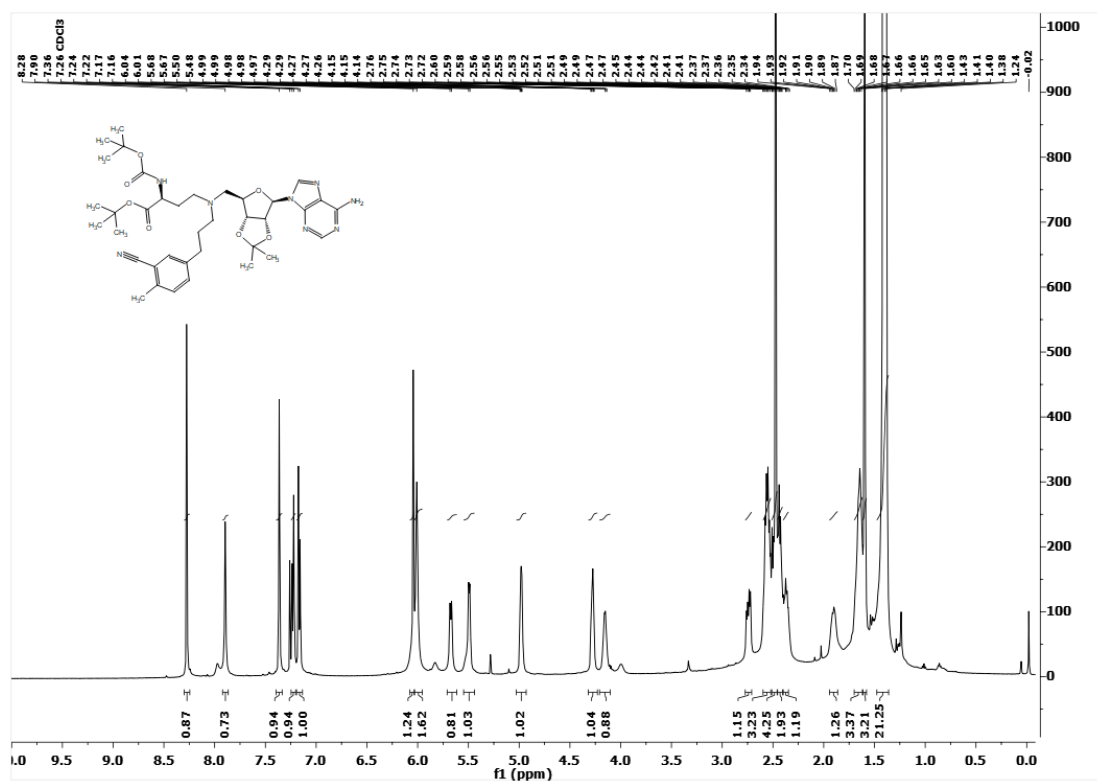
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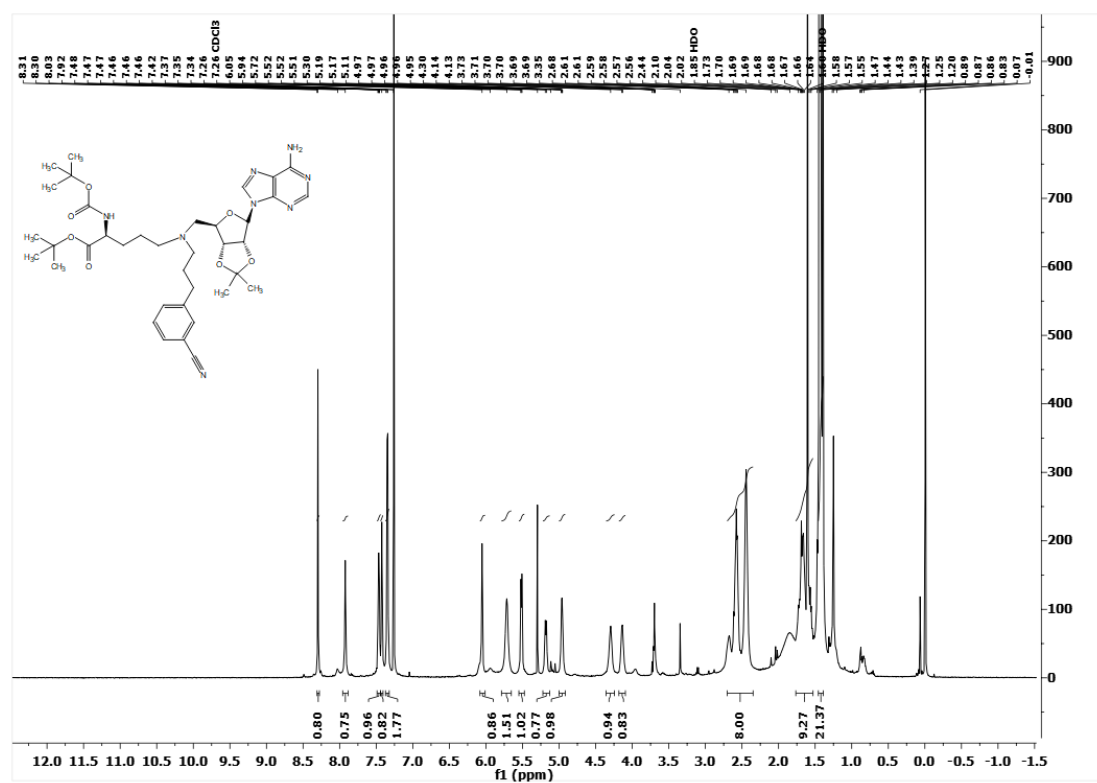
### HNMR of 6b



### HNMR of 6c

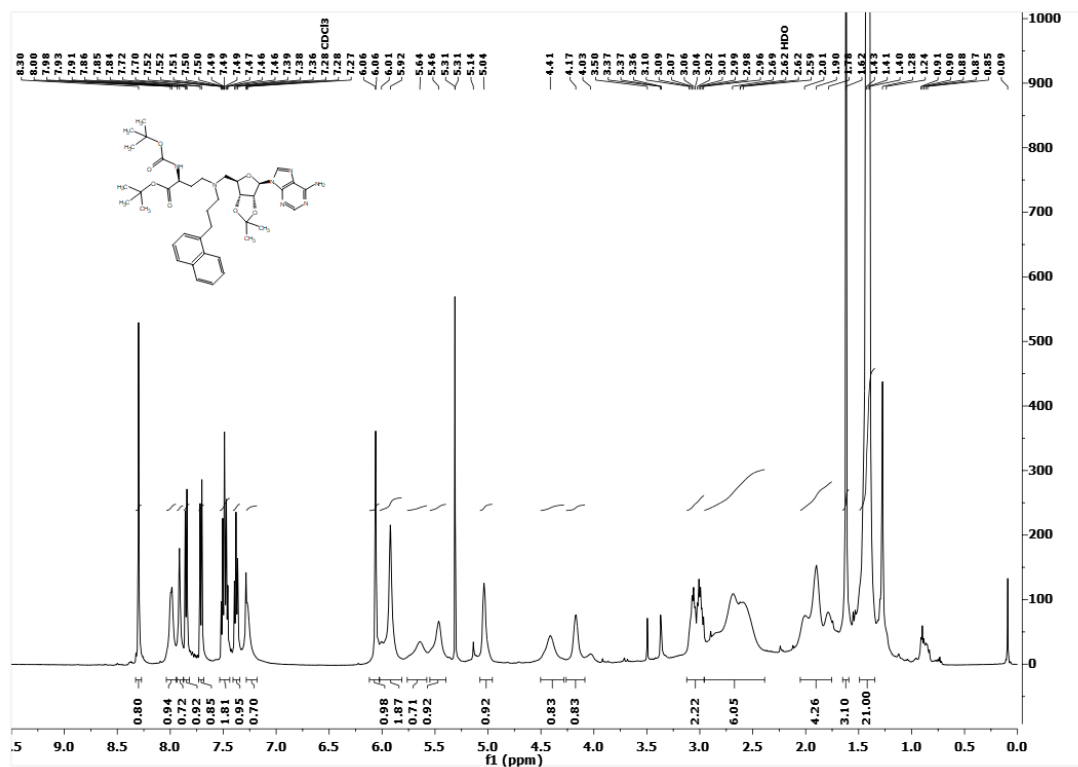


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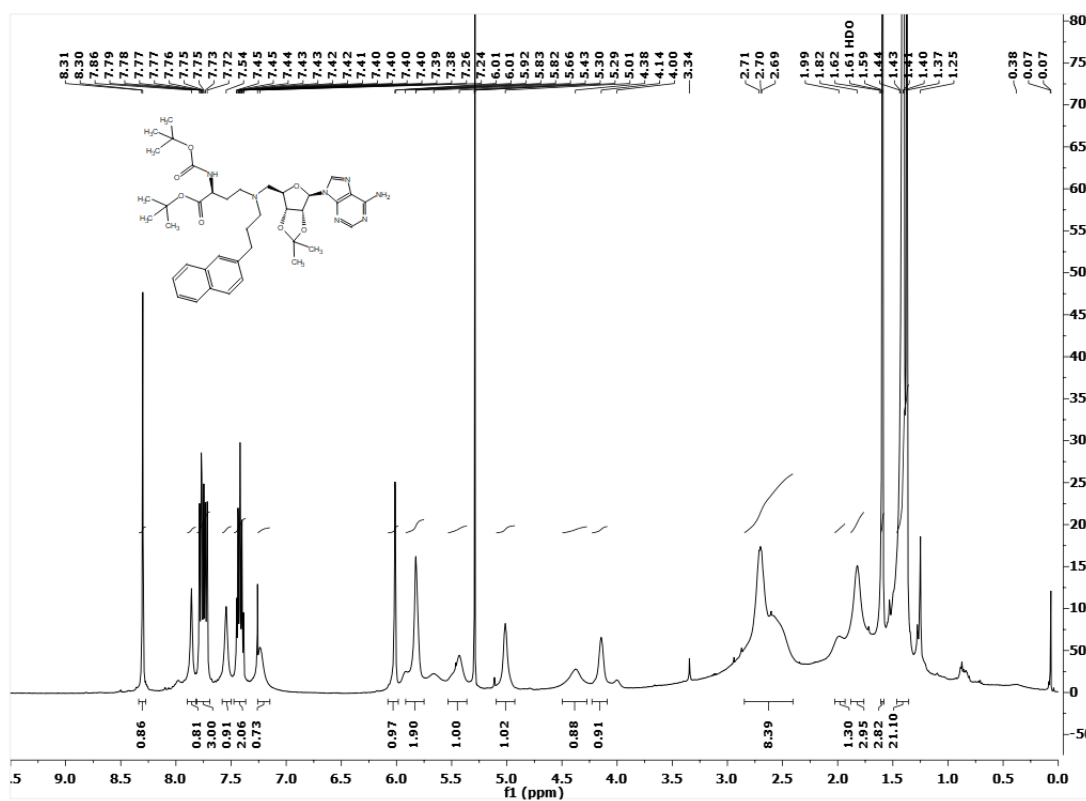




### HNMR of 6g

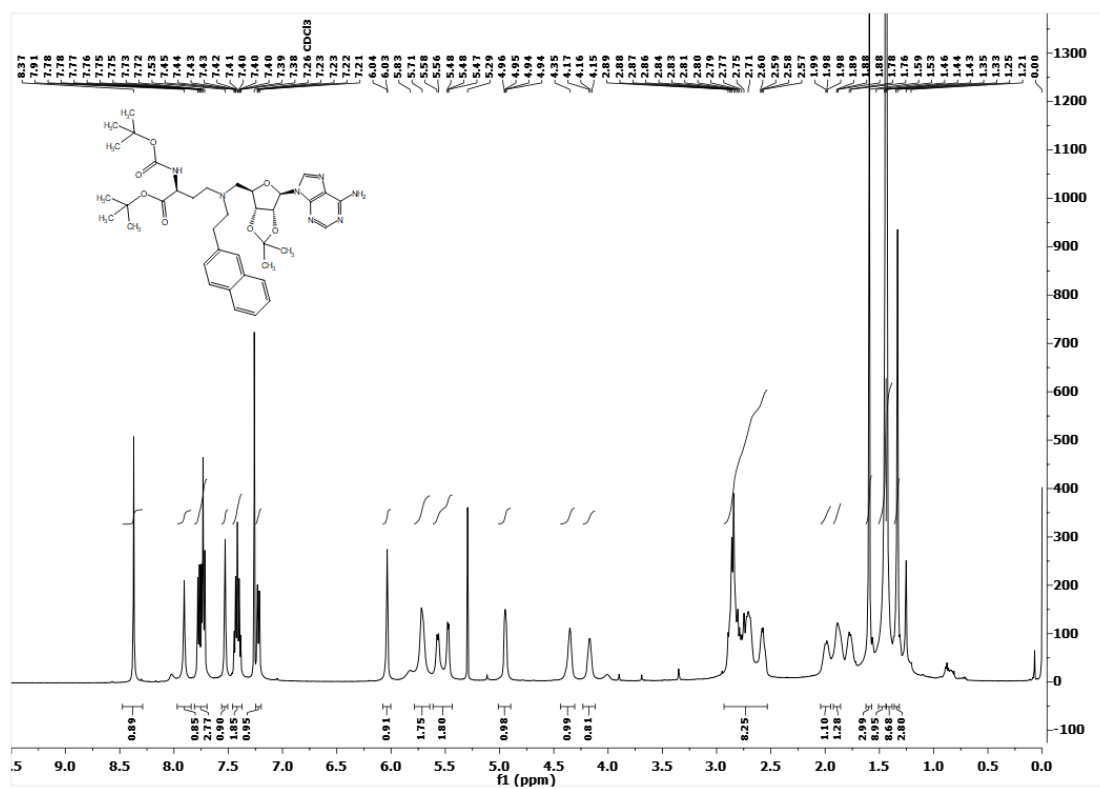


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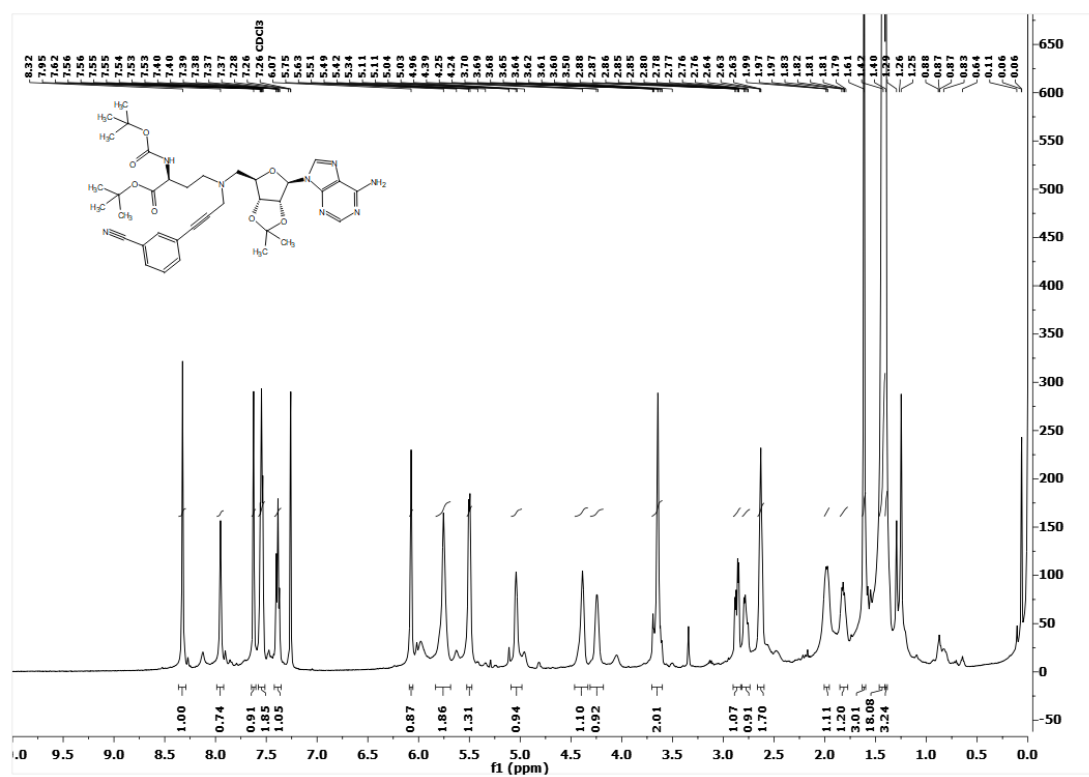




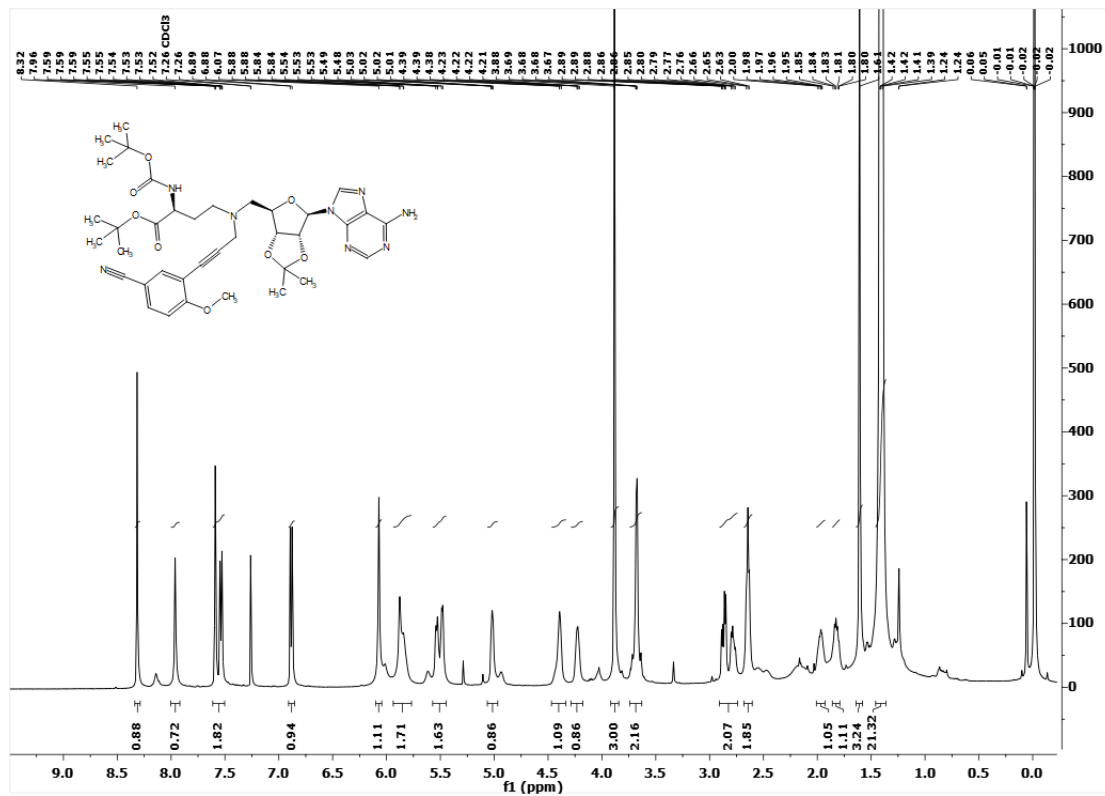
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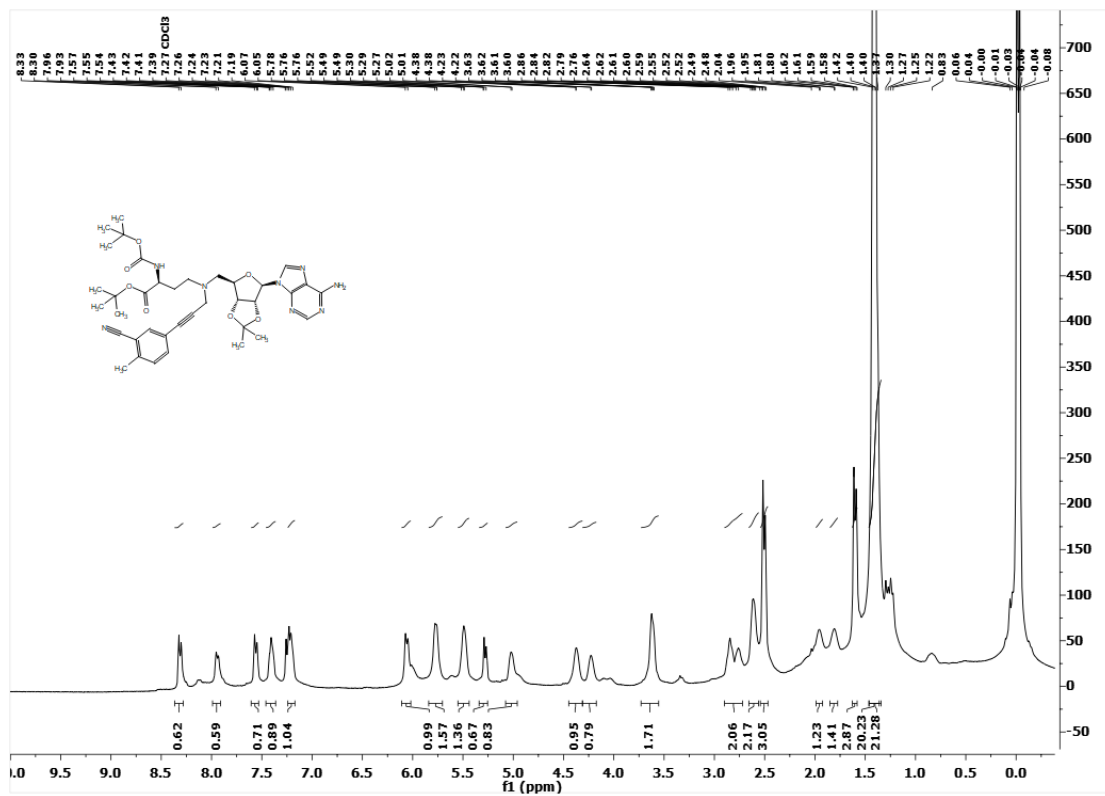
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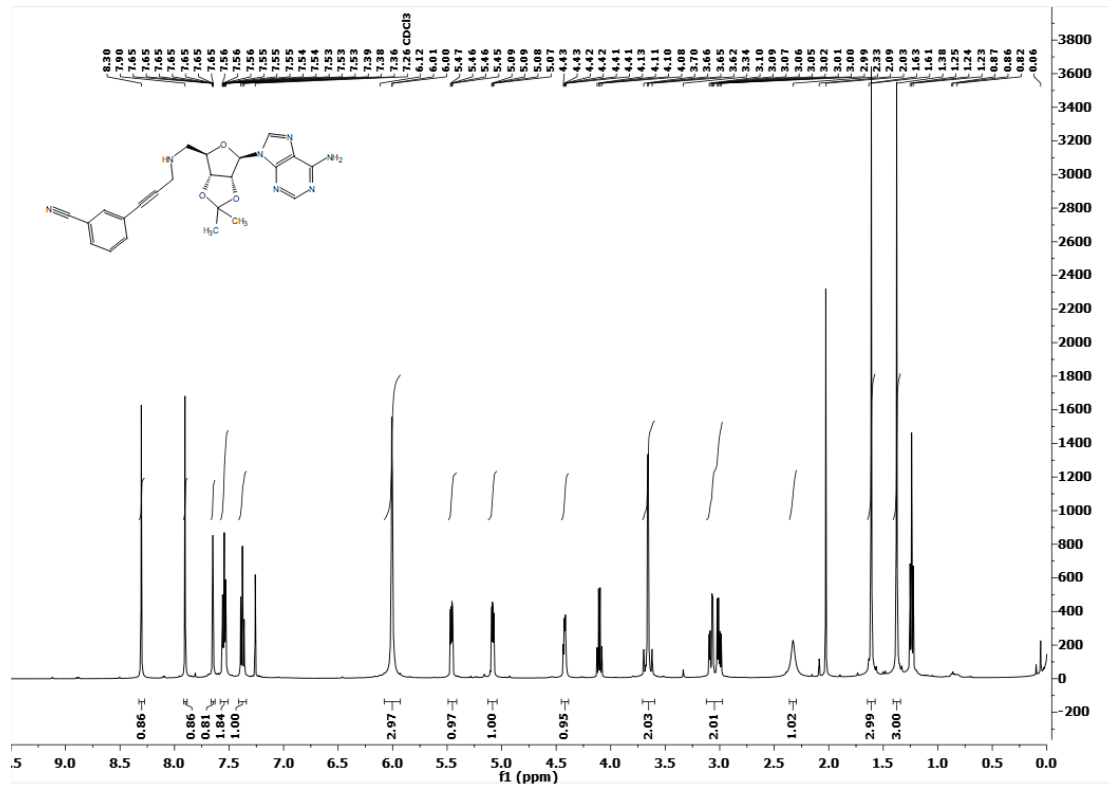
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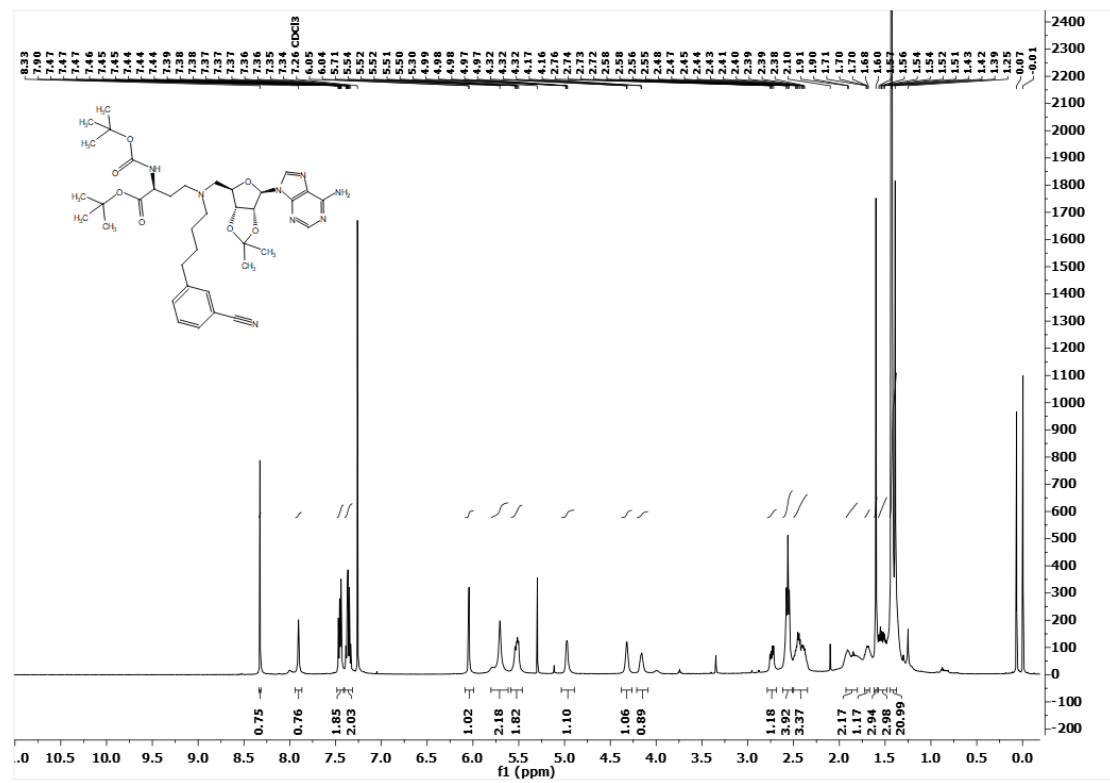
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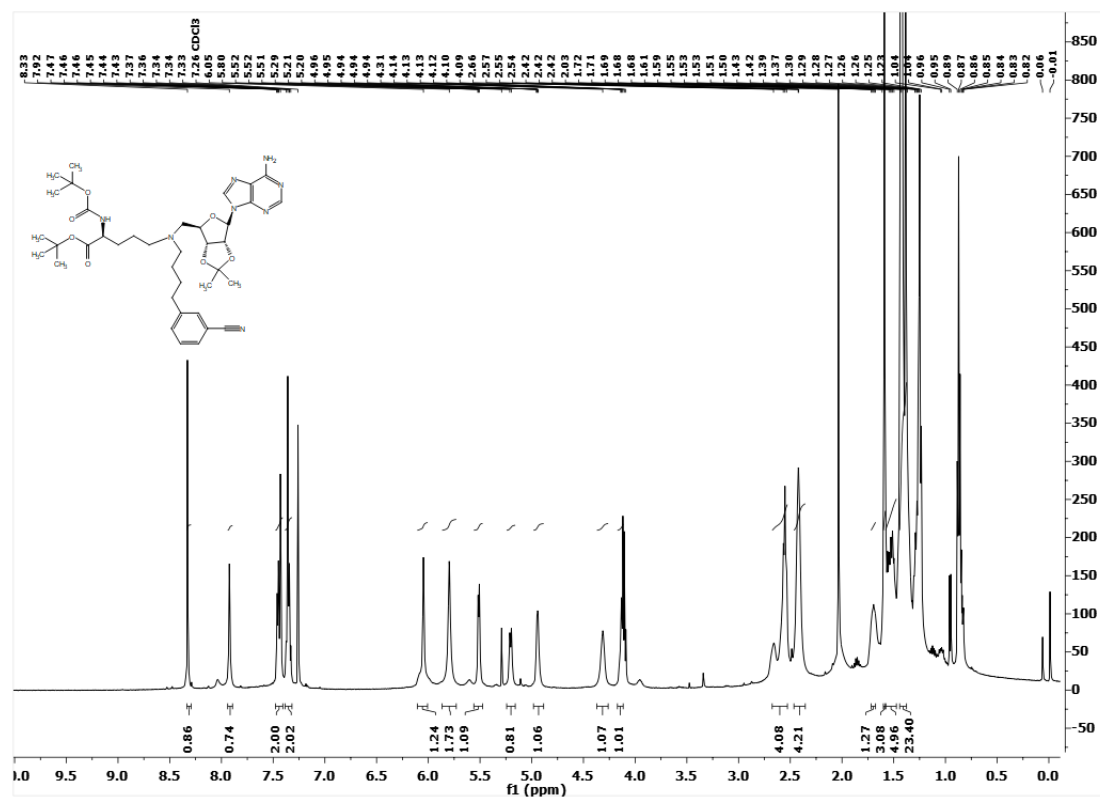
# HNMR of 9d



# HNMR of 12a



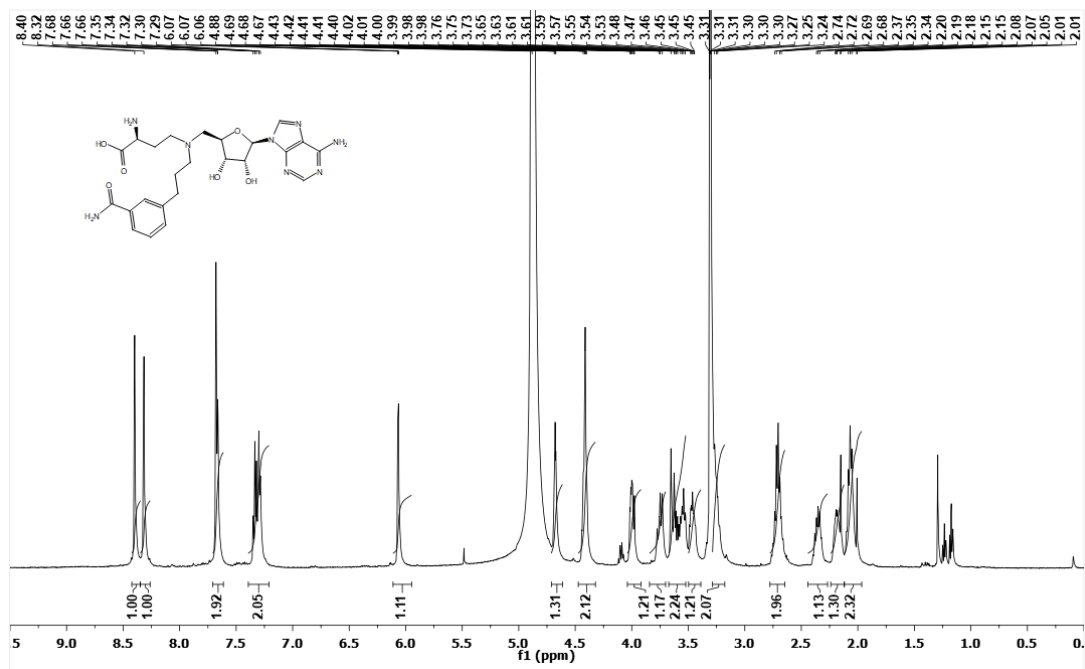
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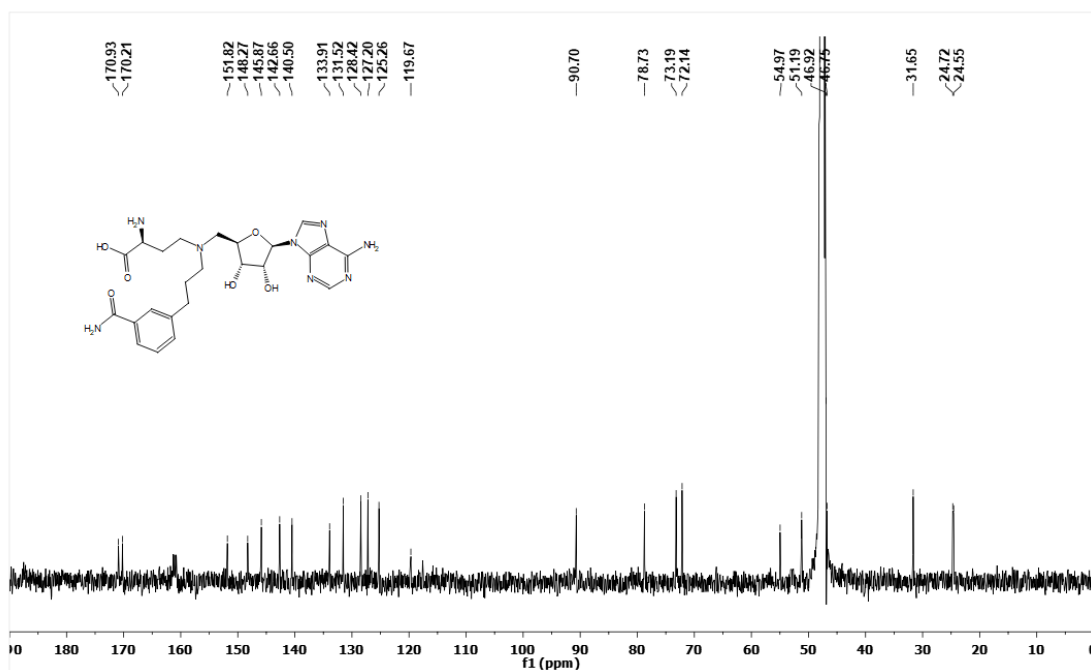
NMR, MALDI-MS and HPLC spectra of compound **1a-j**, **1a\***, **2a-d**, **2a\*** and **3a-b**

NMR, MALDI-MS and HPLC spectra of compound **1a**

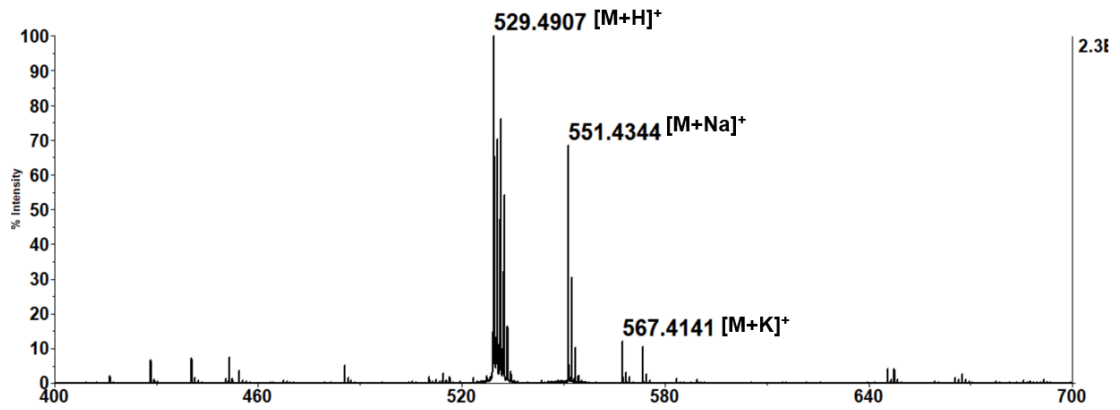
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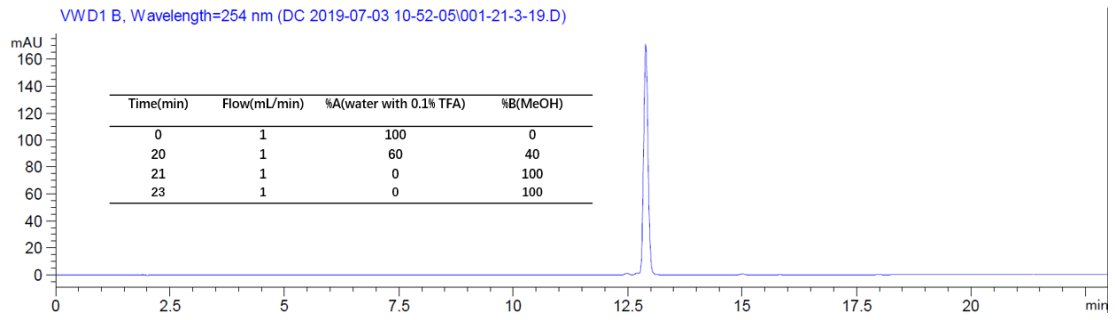
### **CNMR**



## MALDI-MS

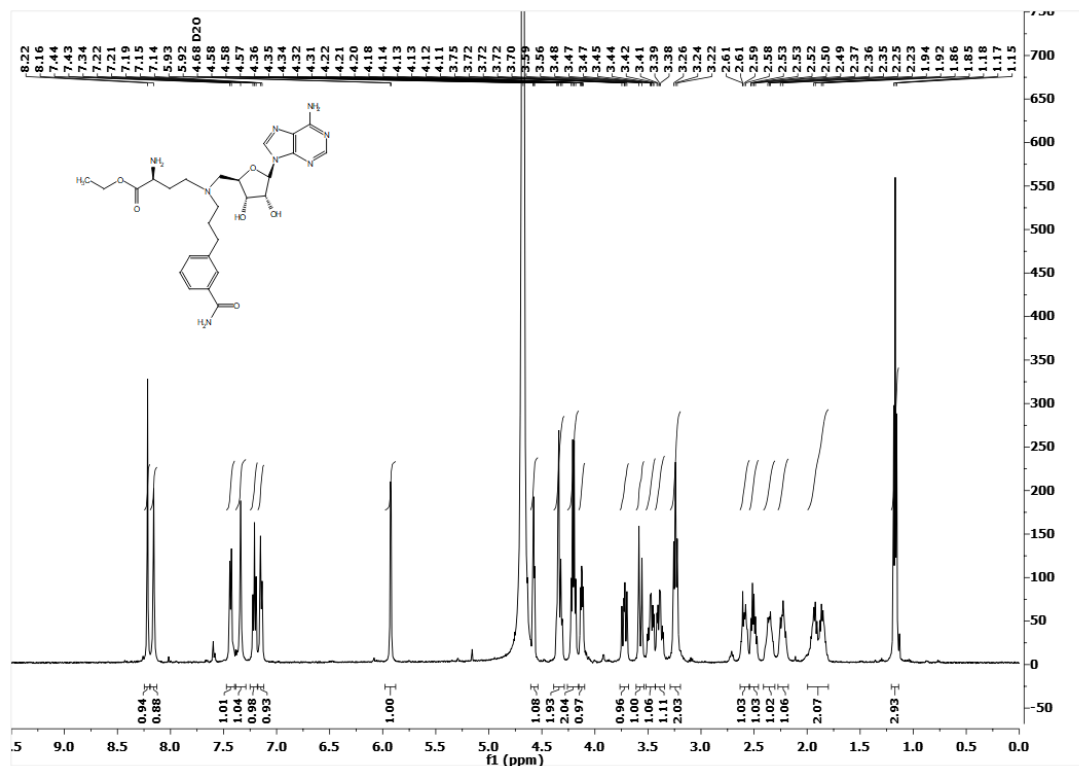


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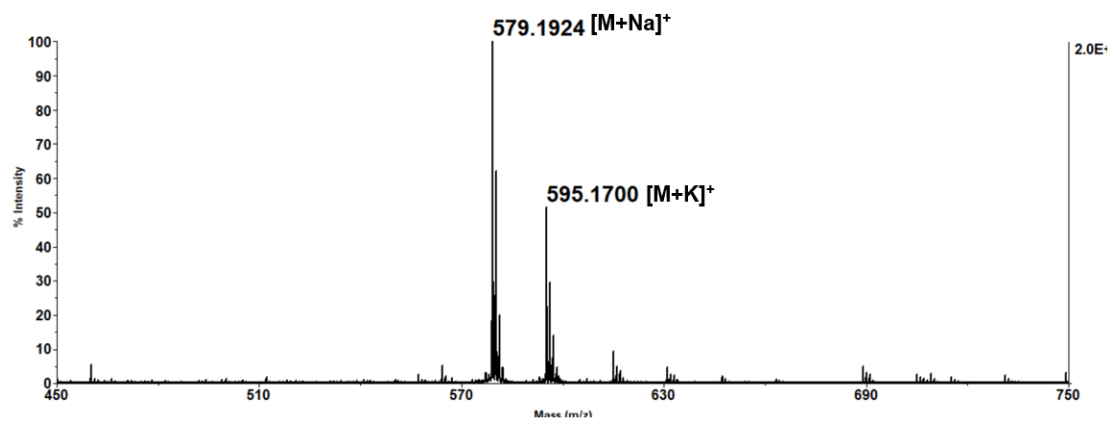


NMR, MALDI-MS and HPLC spectra of compound **1a**\*

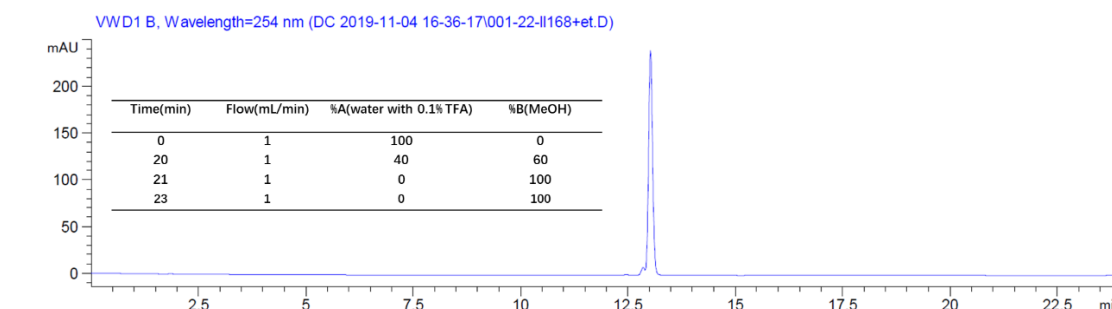
H-NMR



MALDI-MS

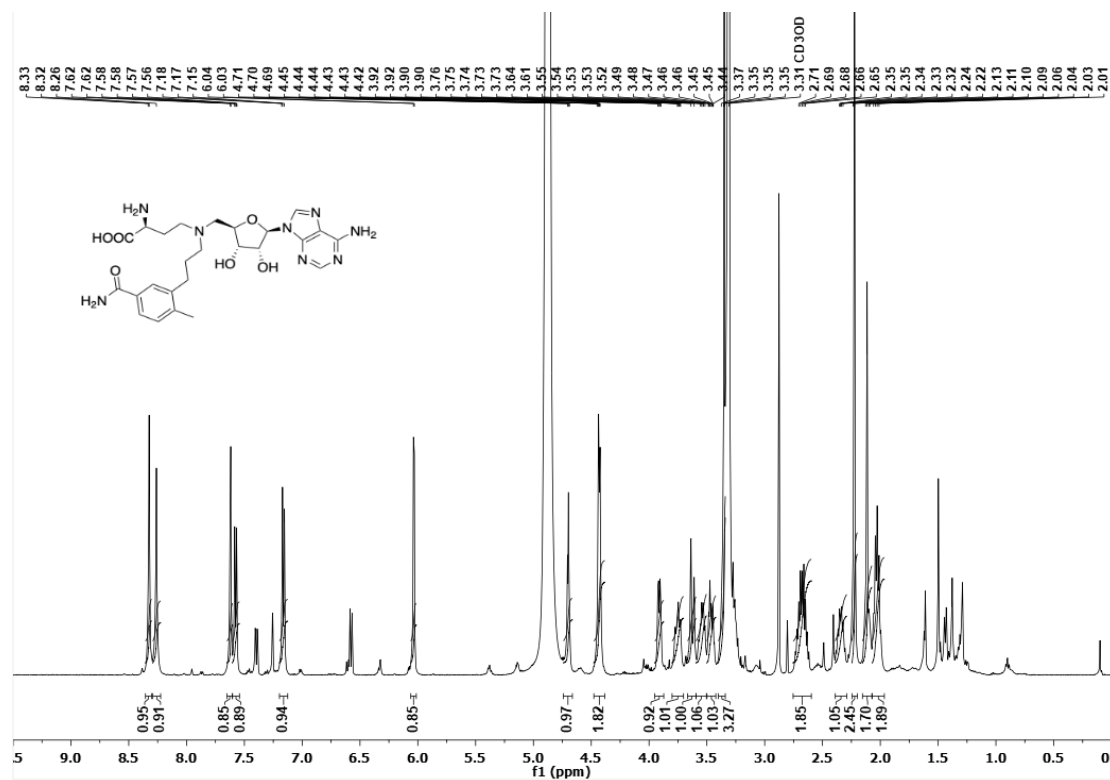


HPLC

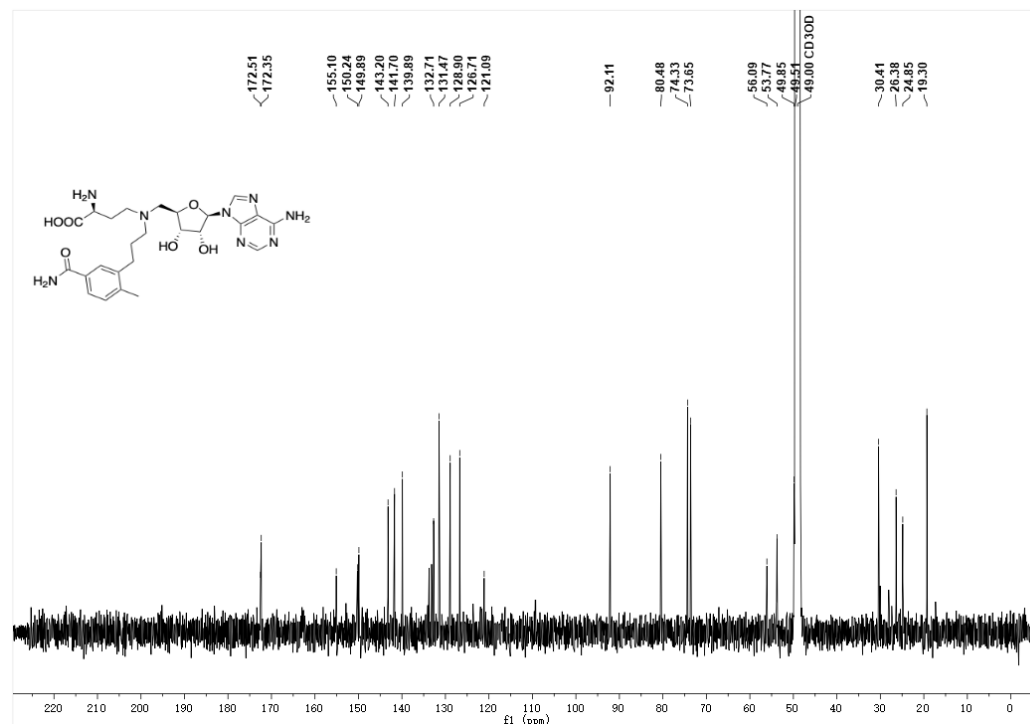


NMR, MALDI-MS and HPLC spectra of compound **1b**

HNMR

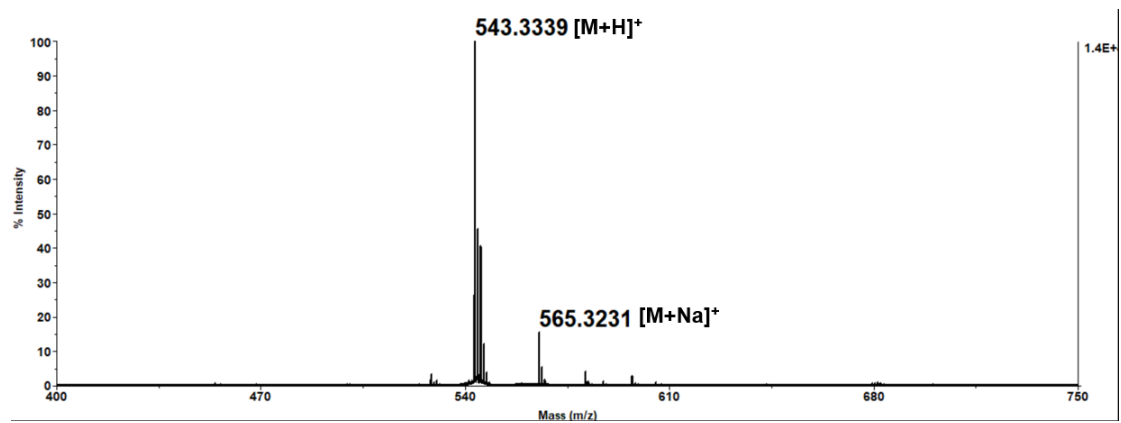


CNMR

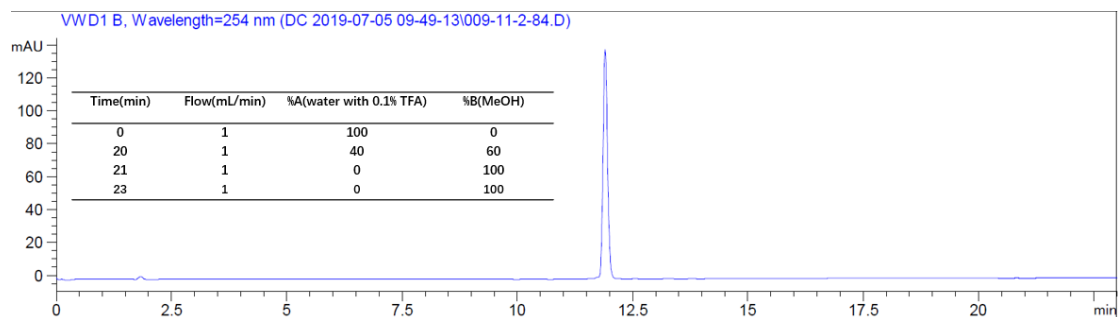




### MALDI-MS

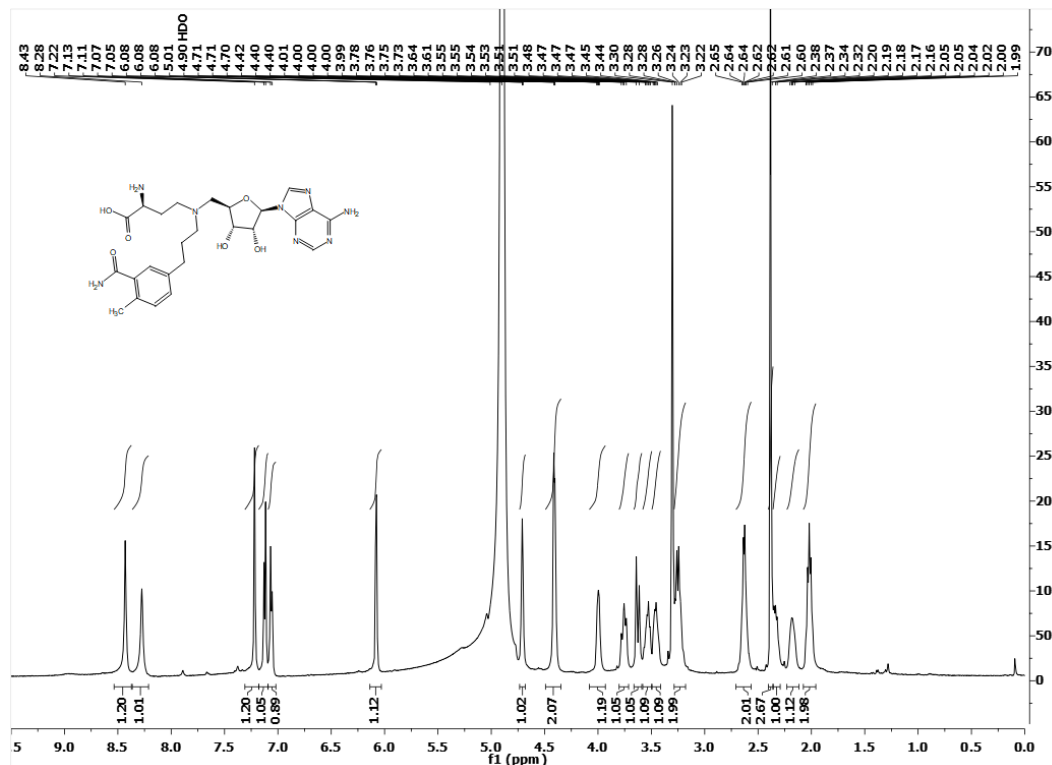


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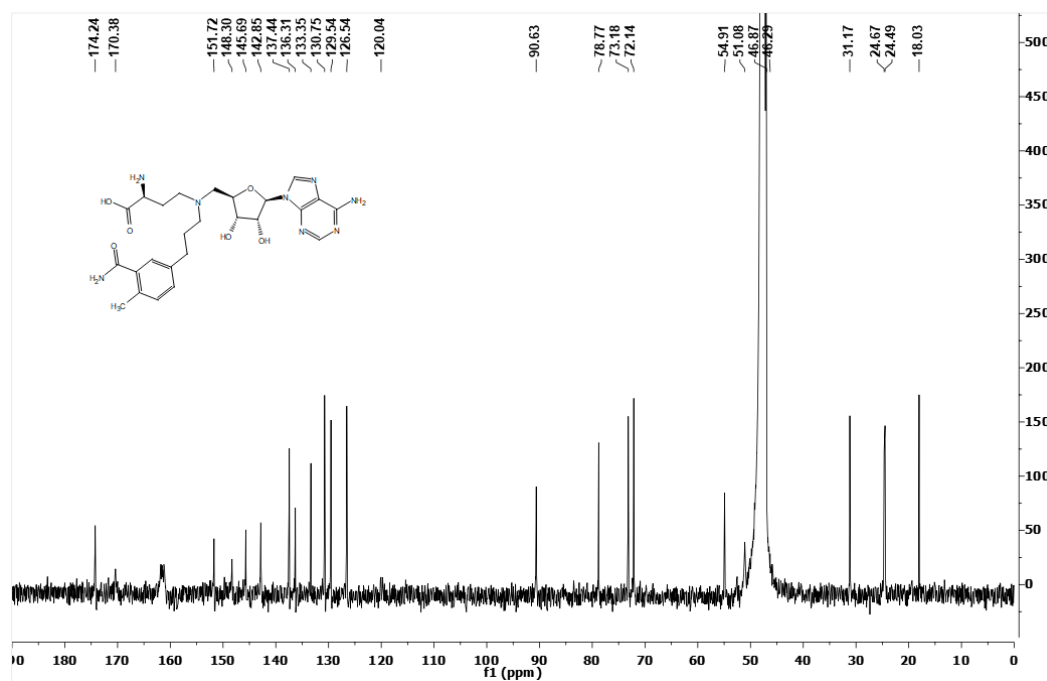


NMR, MALDI-MS and HPLC spectra of compound **1c**

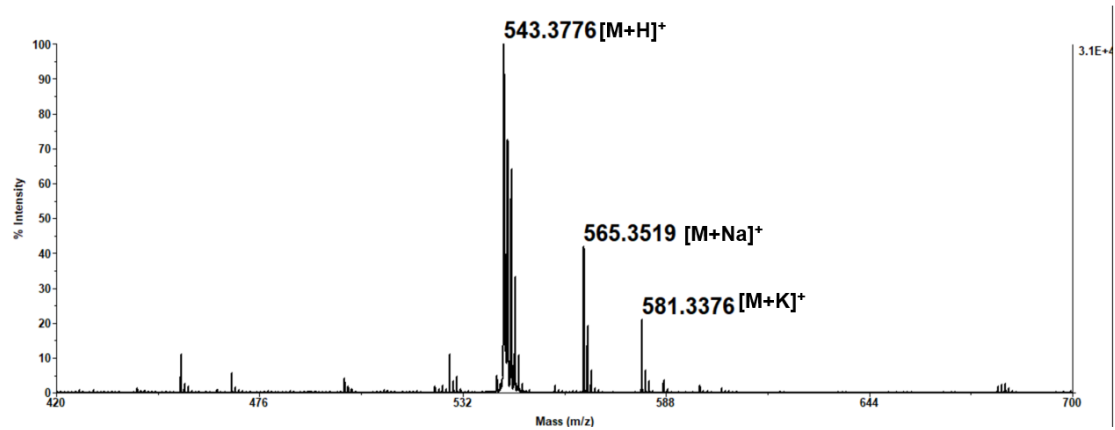
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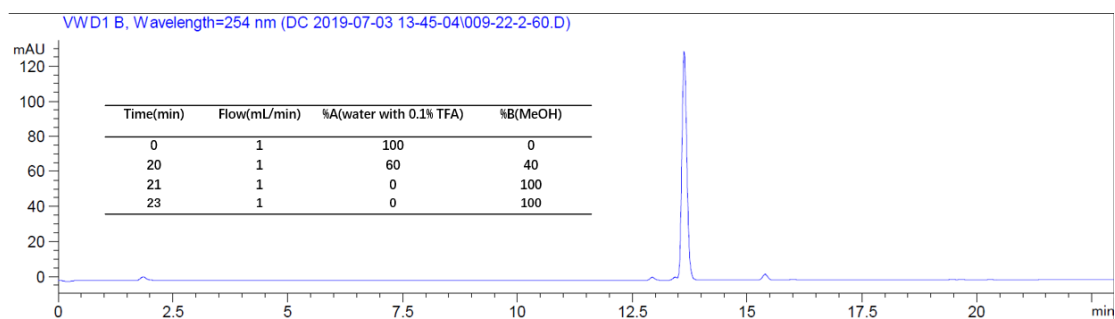
CNMR



## MALDI-MS

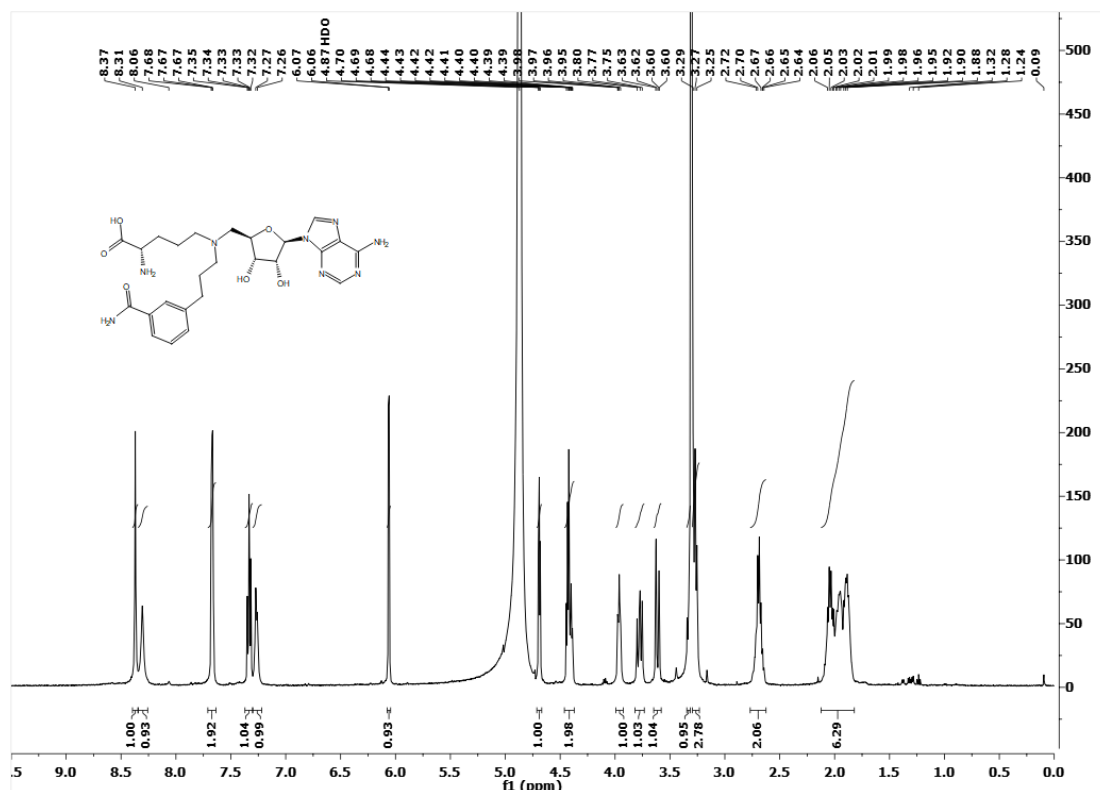


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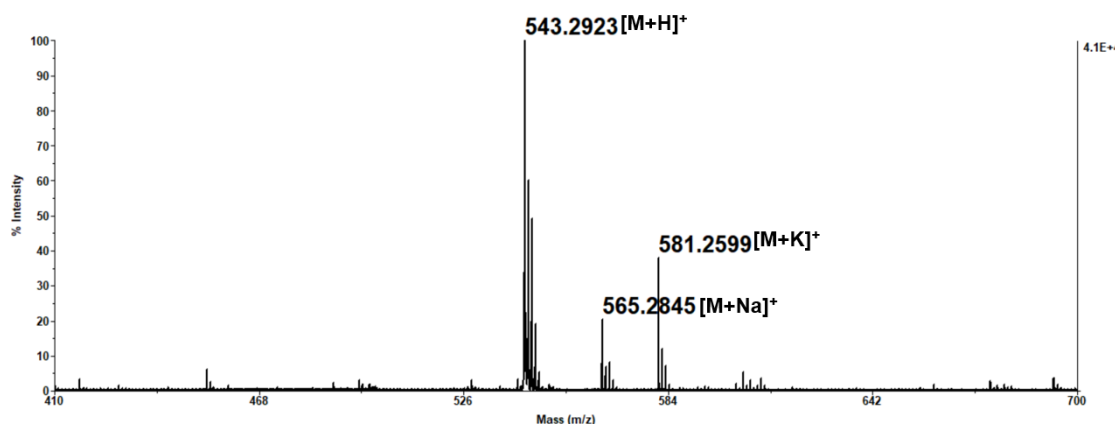


# NMR, MALDI-MS and HPLC spectra of compound 1d

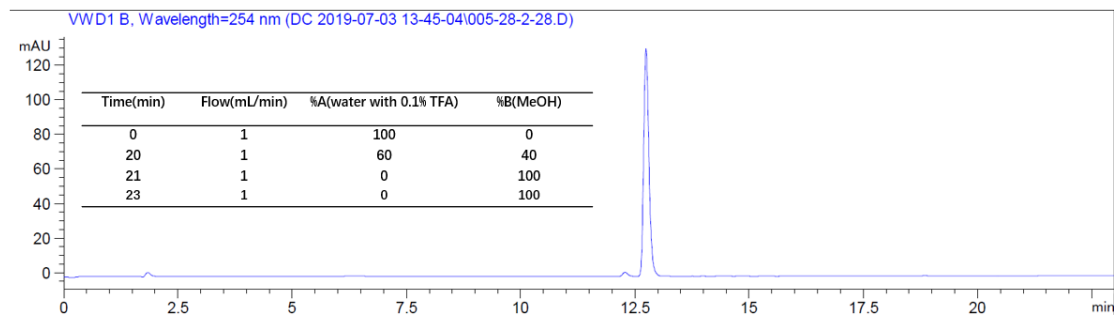
## HNMR



## MALDI-MS

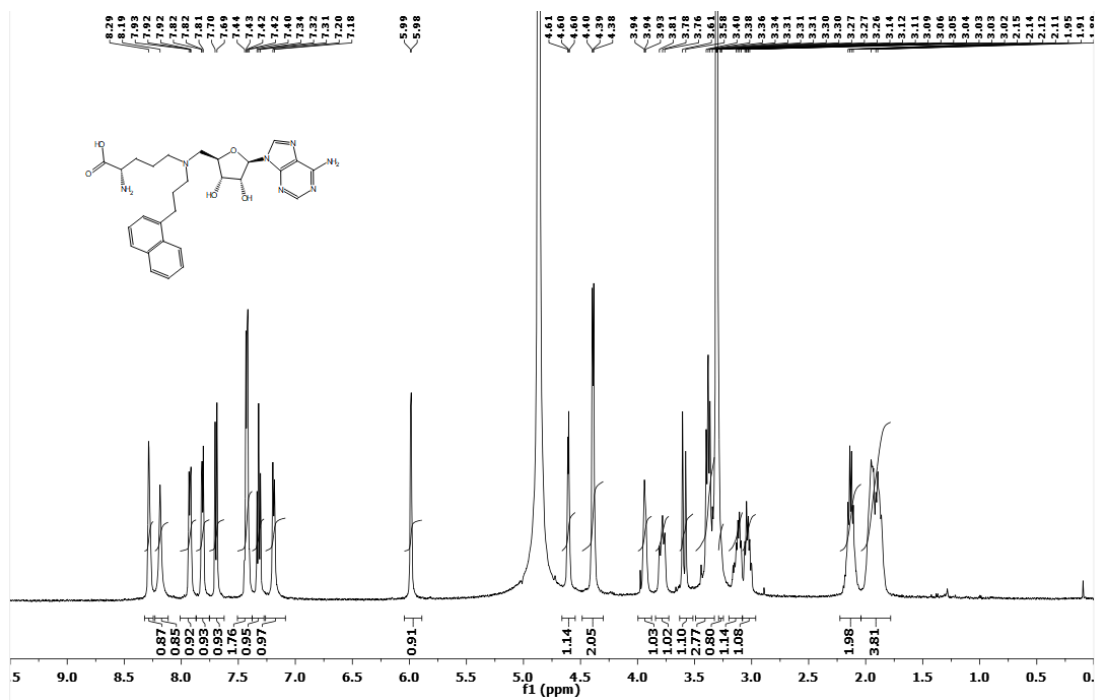


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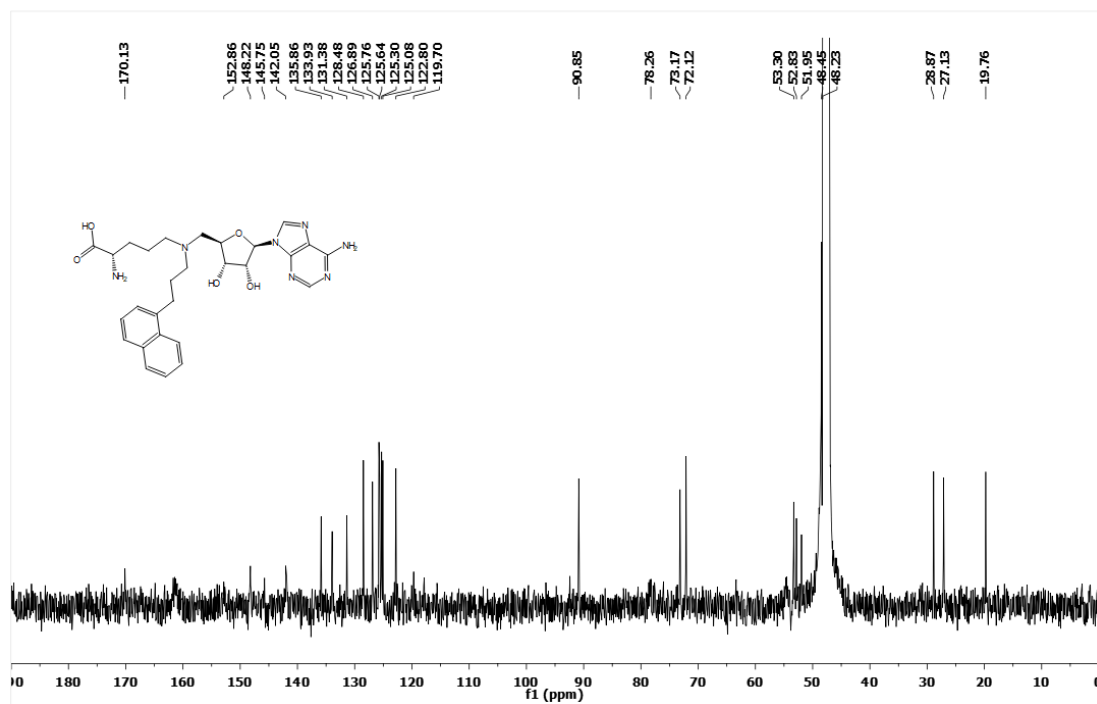


NMR, MALDI-MS and HPLC spectra of compound 1e

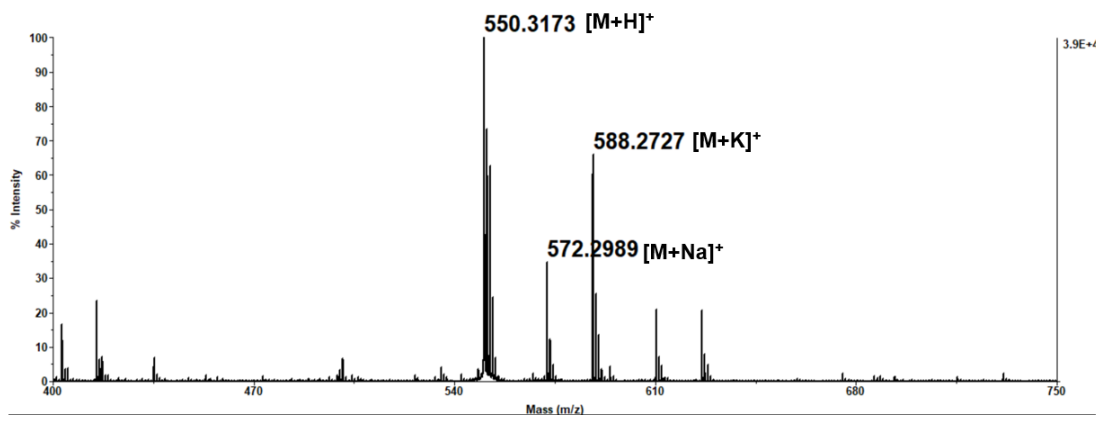
<sup>1</sup>H NMR



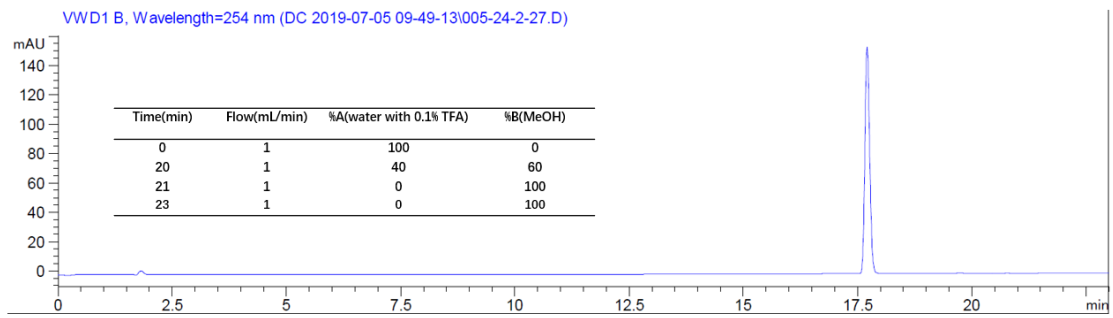
<sup>13</sup>C NMR



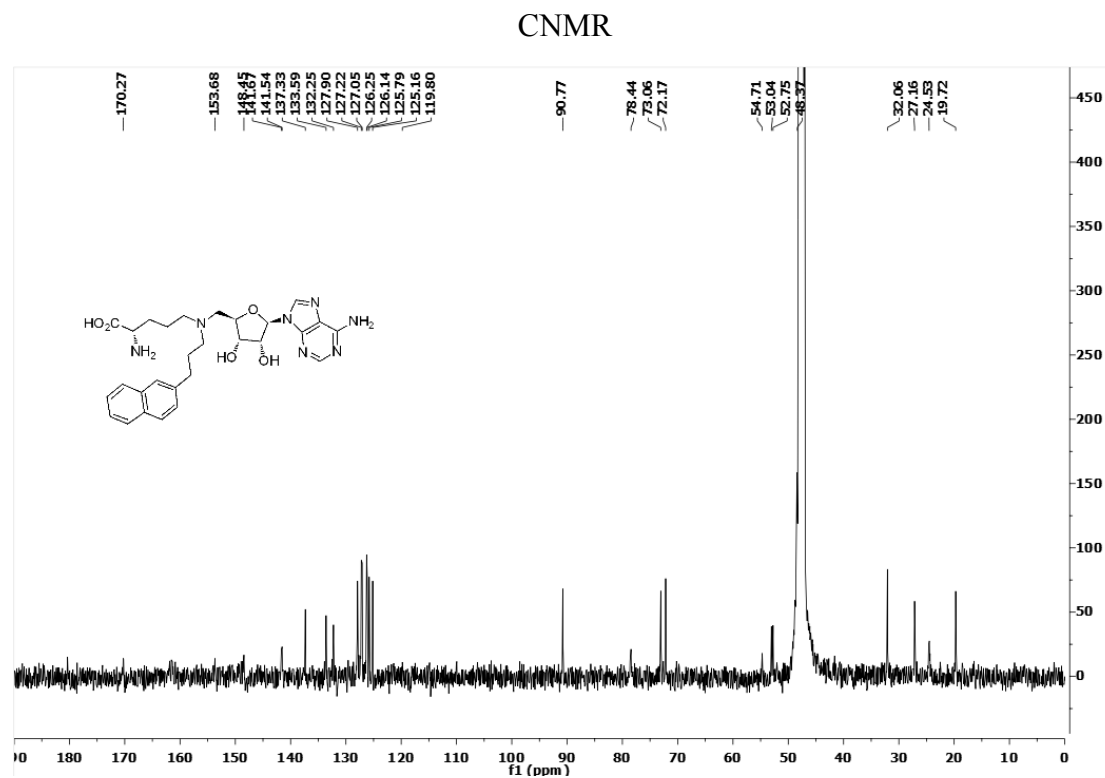
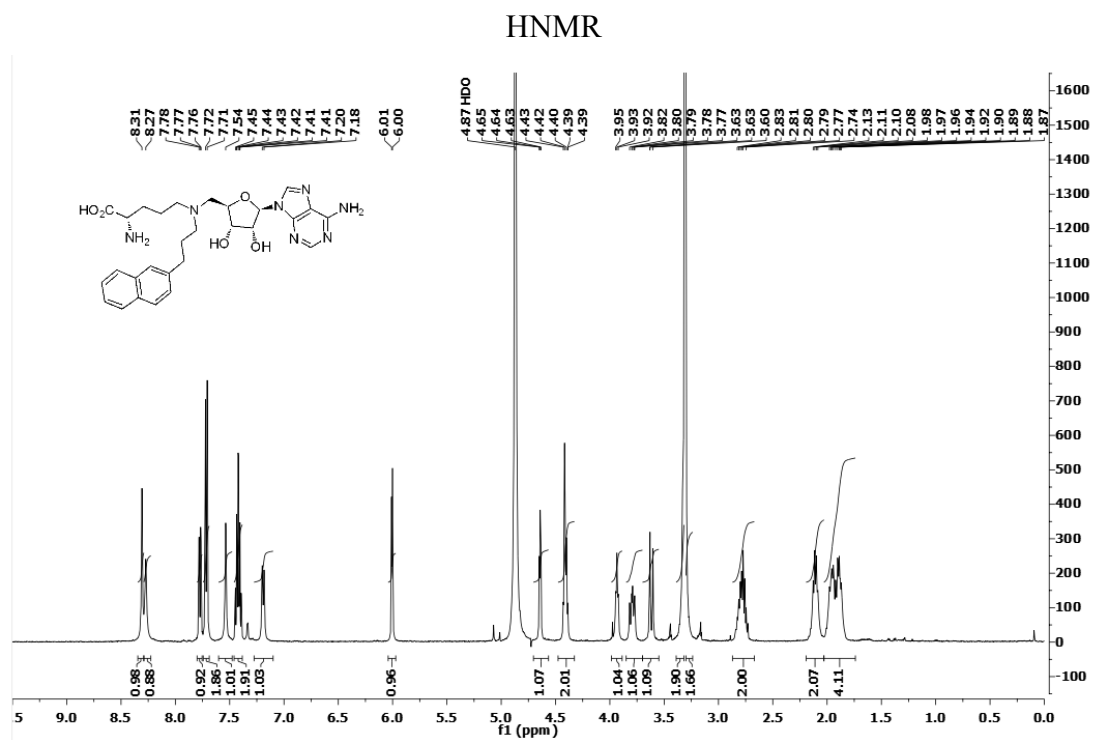
# MALDI-MS



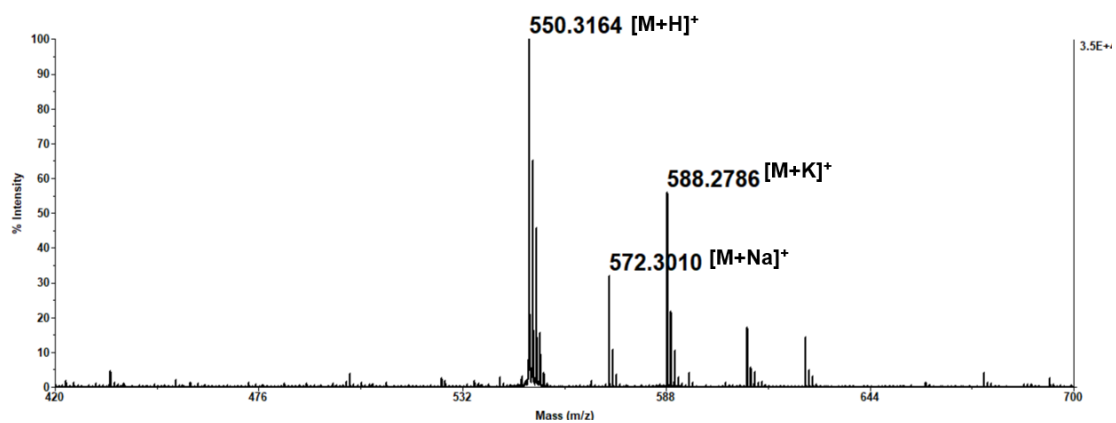
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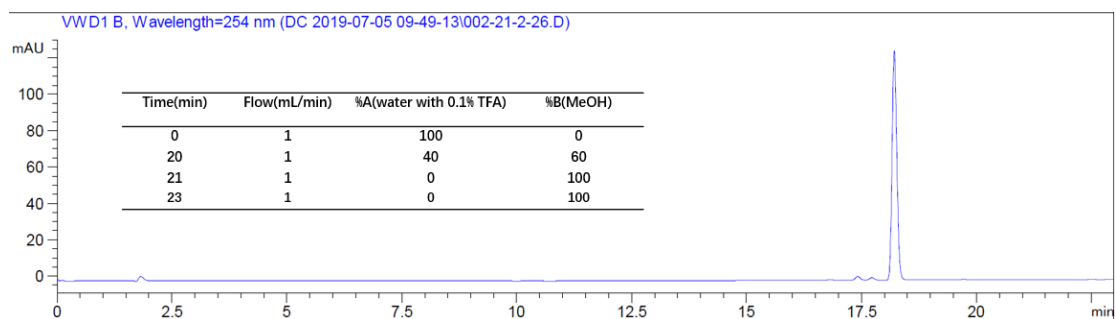
NMR, MALDI-MS and HPLC spectra of compound **1f**



# MALDI-MS



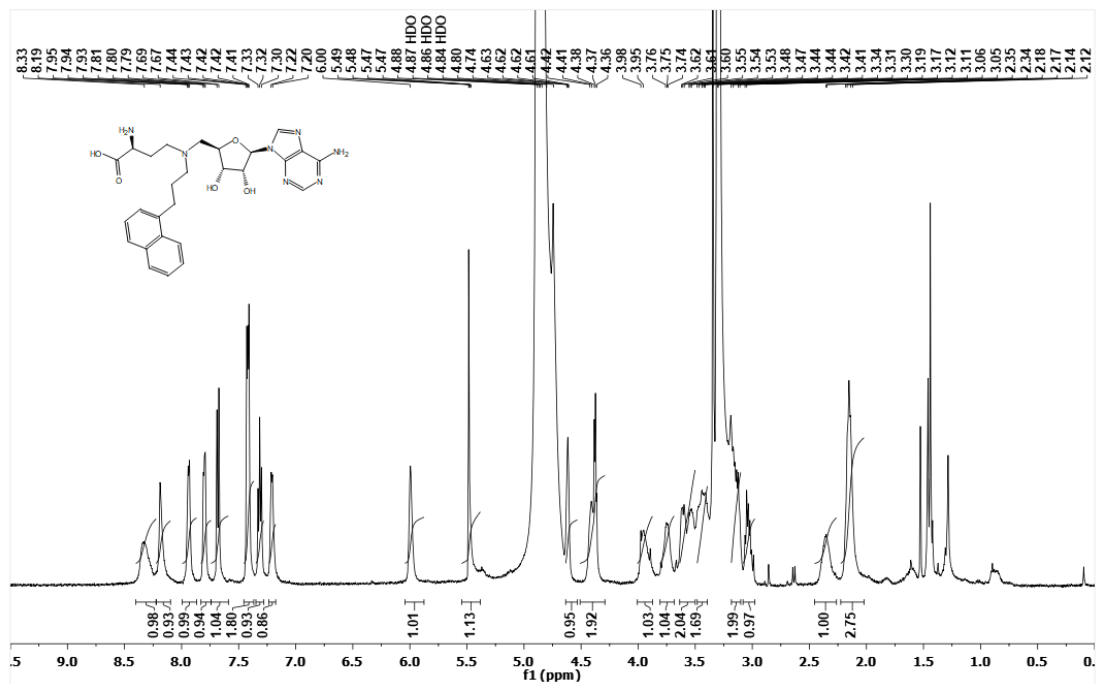
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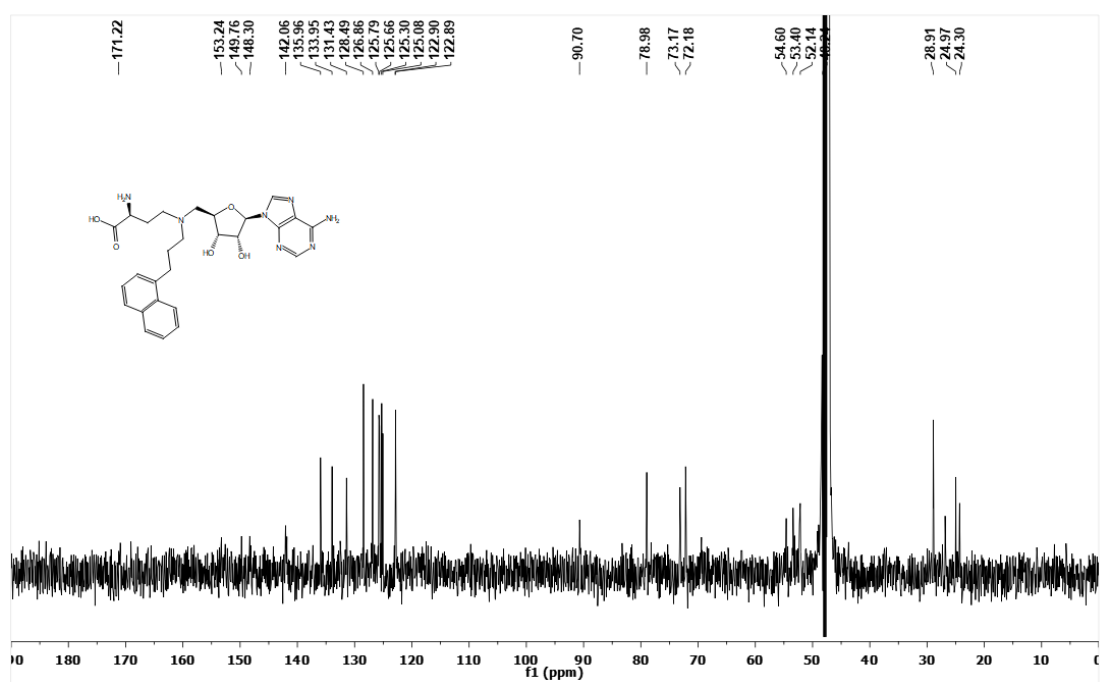


# NMR, MALDI-MS and HPLC spectra of compound **1g**

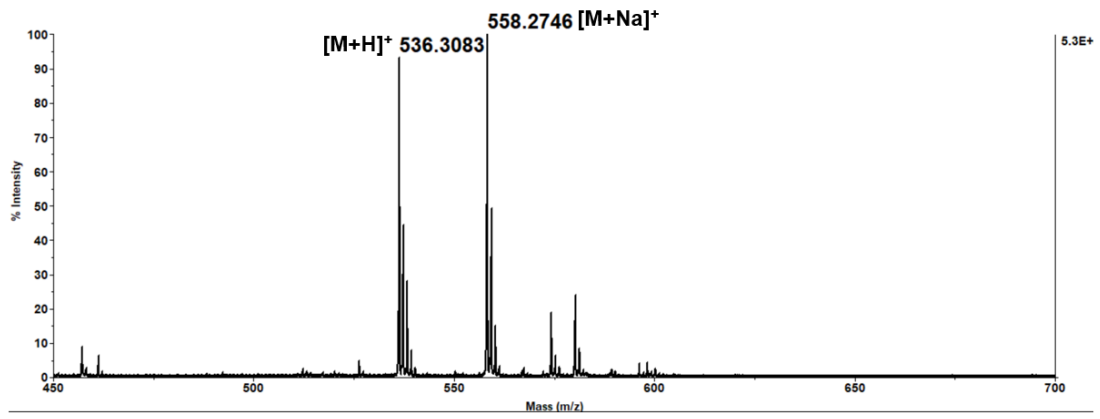
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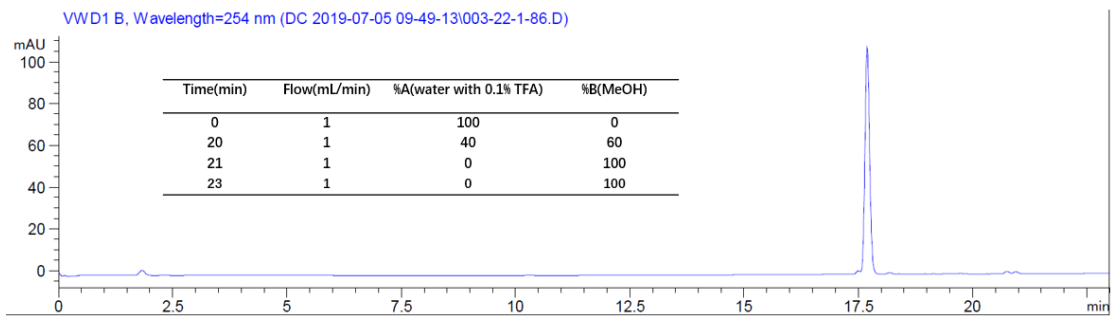
## CNMR



# MALDI-MS

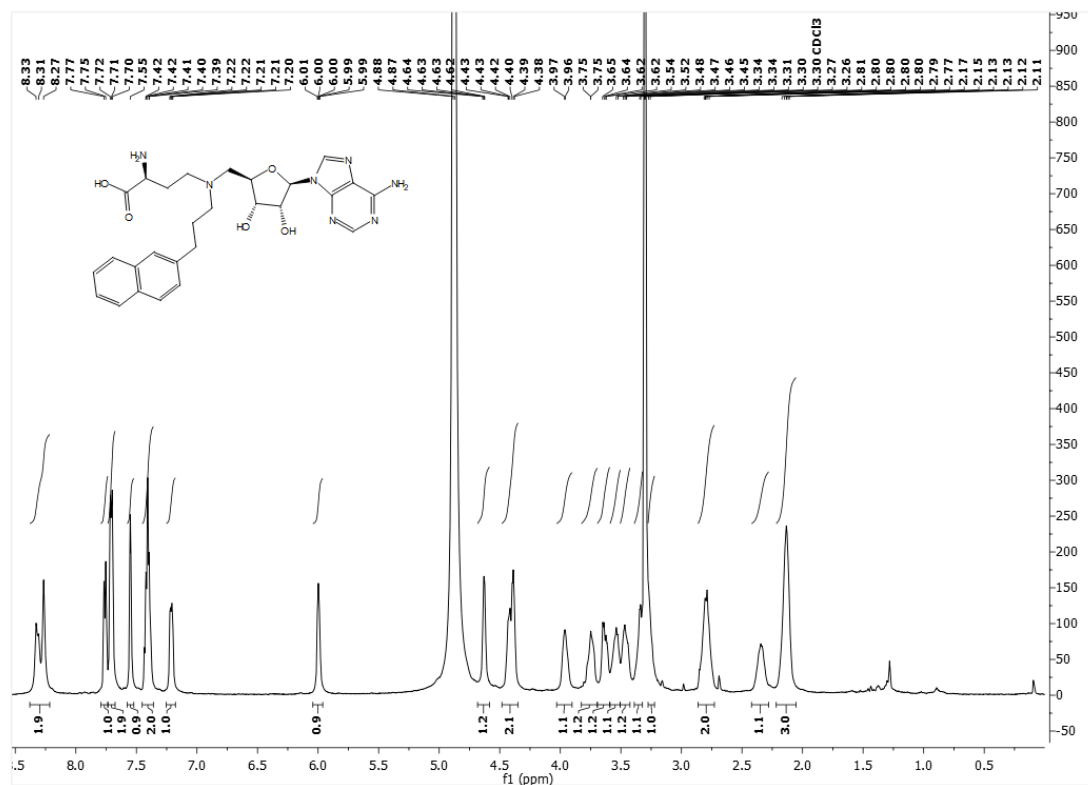


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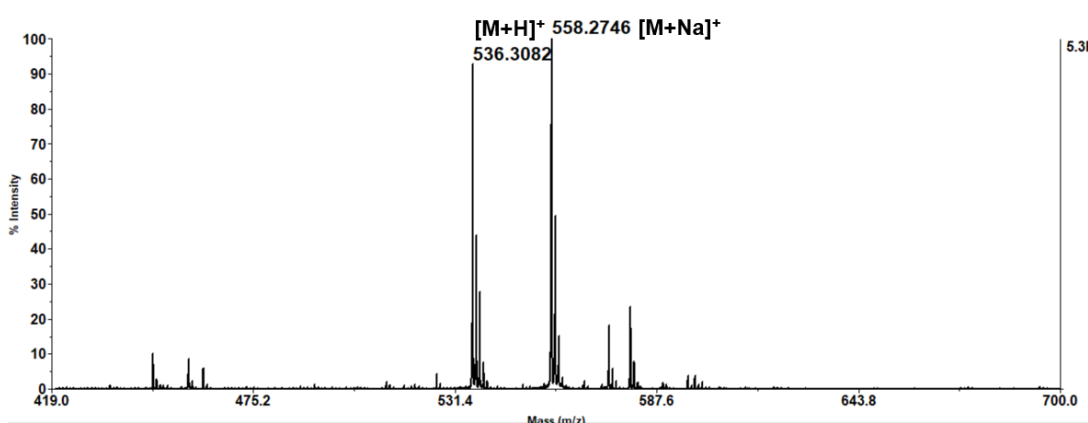


# NMR, MALDI-MS and HPLC spectra of compound 1h

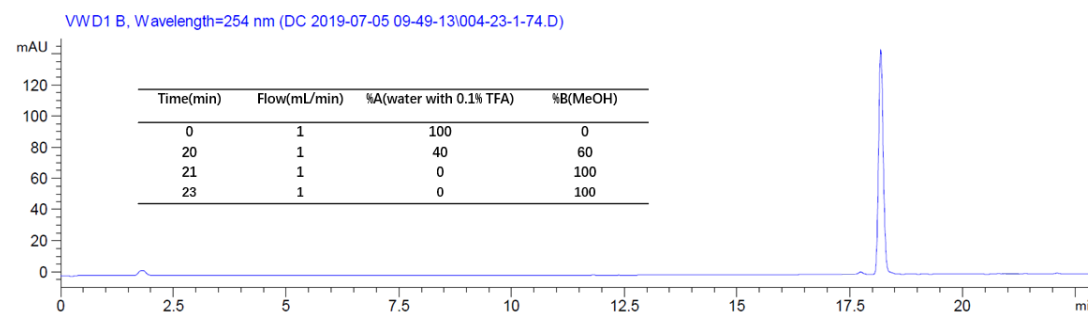
## HNMR



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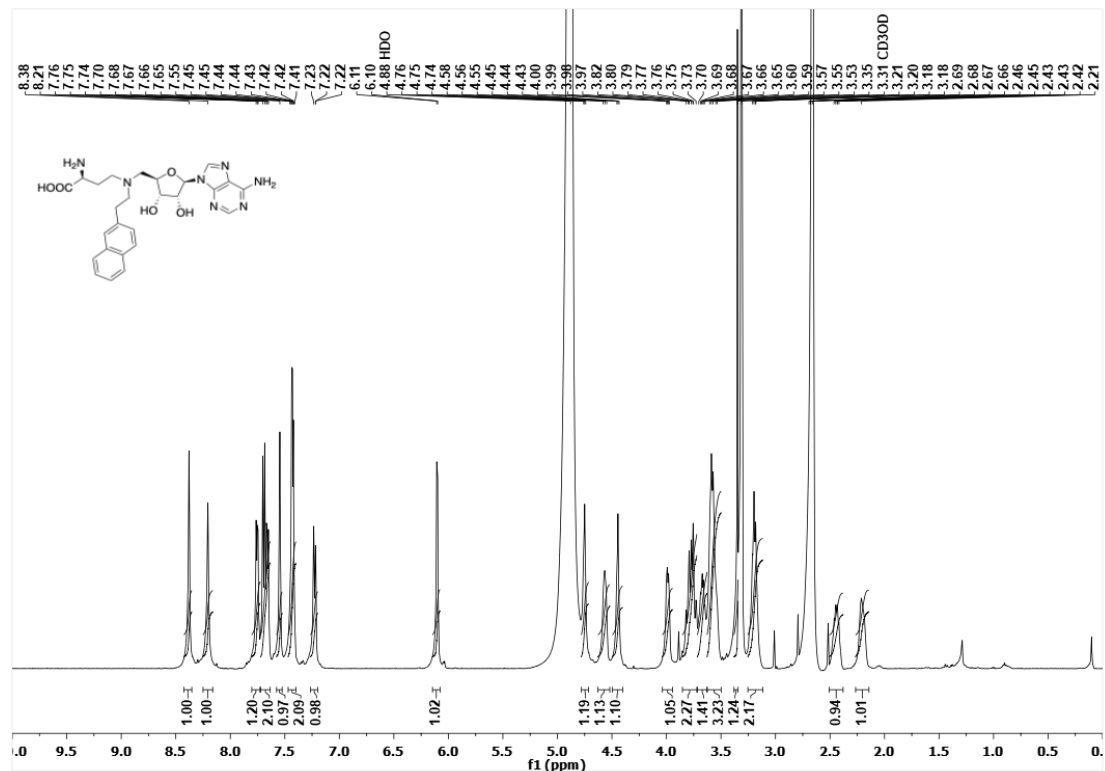


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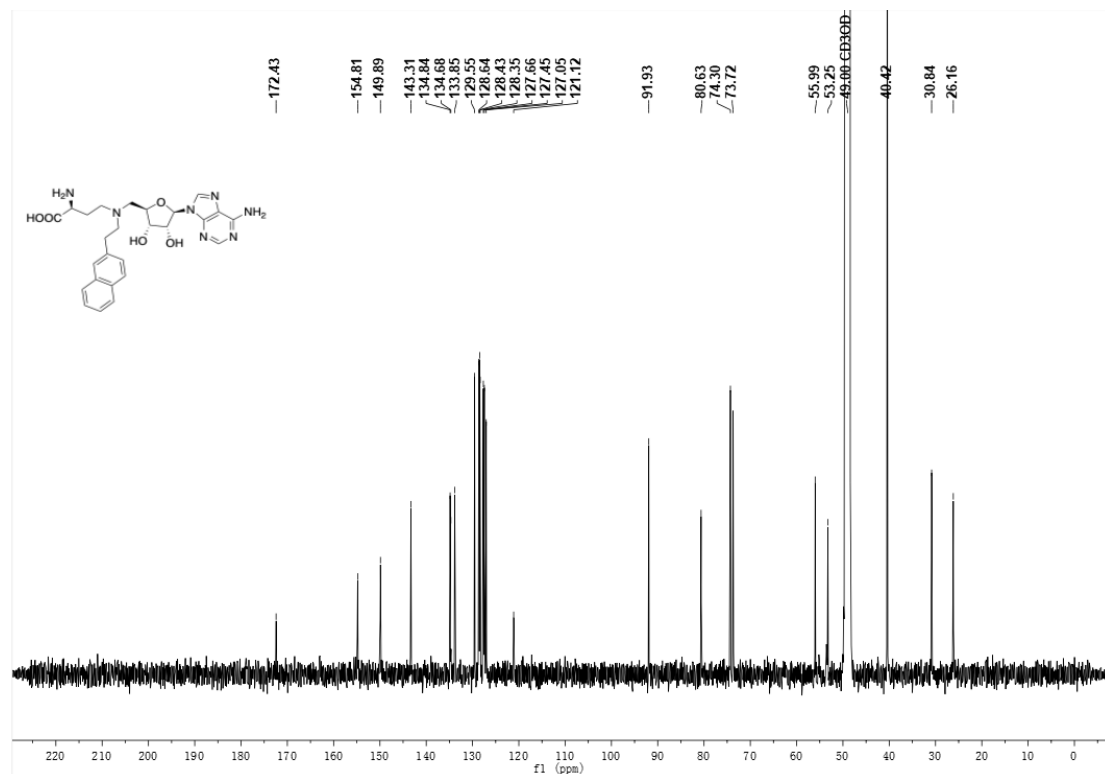


NMR, MALDI-MS and HPLC spectra of compound **1i**

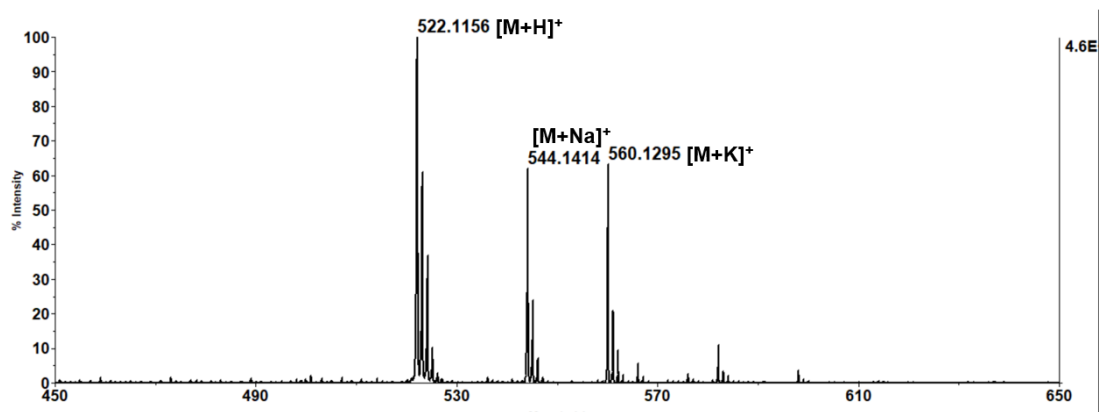
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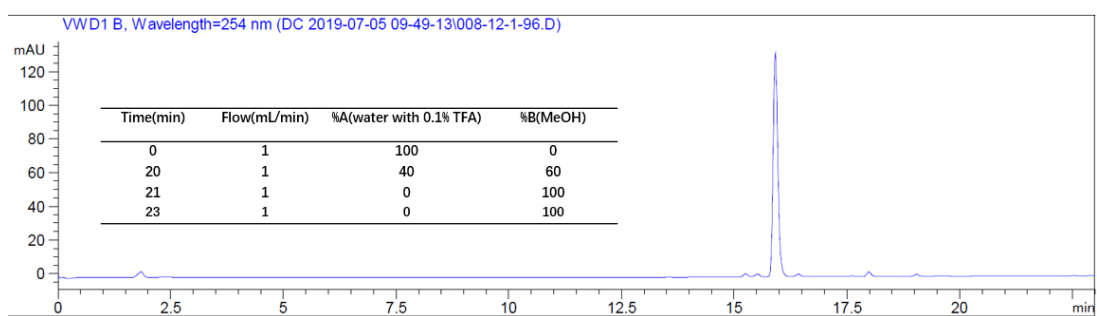
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### MALDI-MS

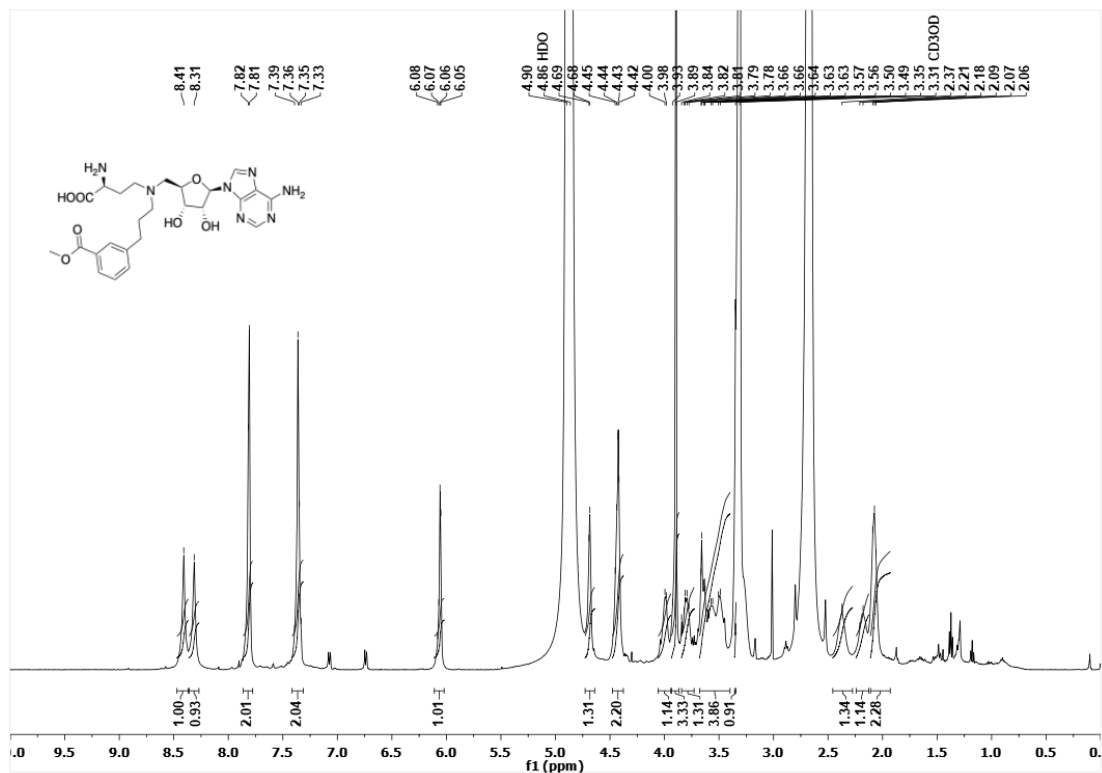


### HPLC

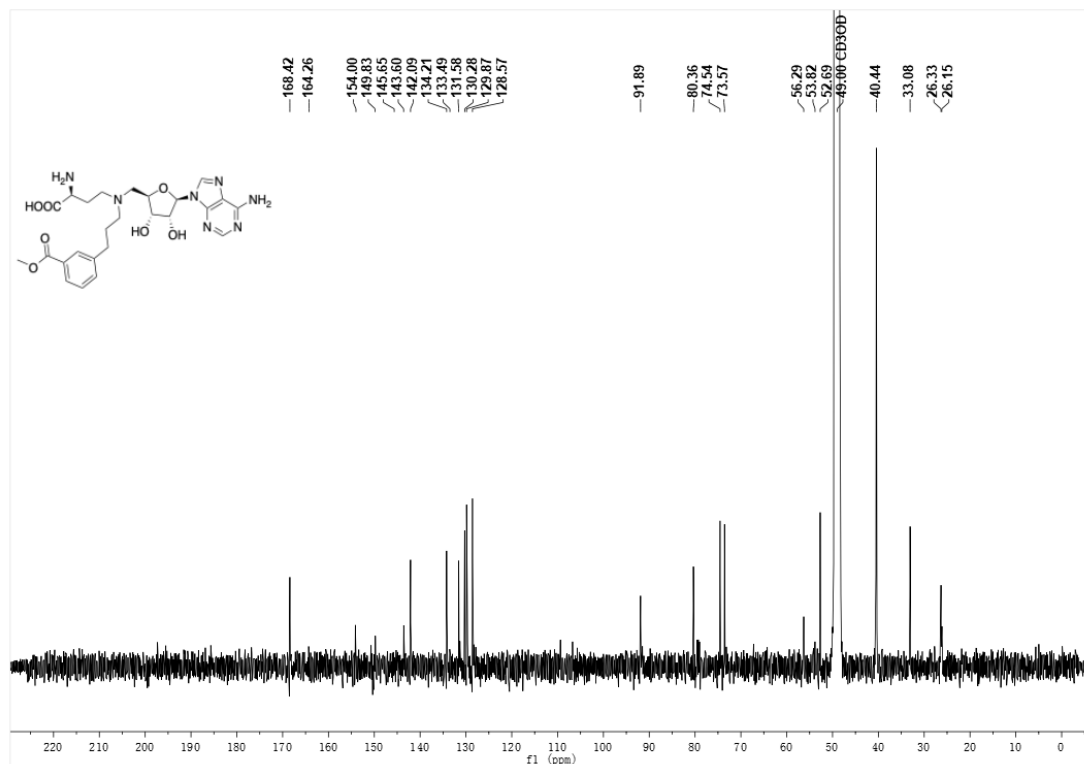


NMR, MALDI-MS and HPLC spectra of compound **1j**

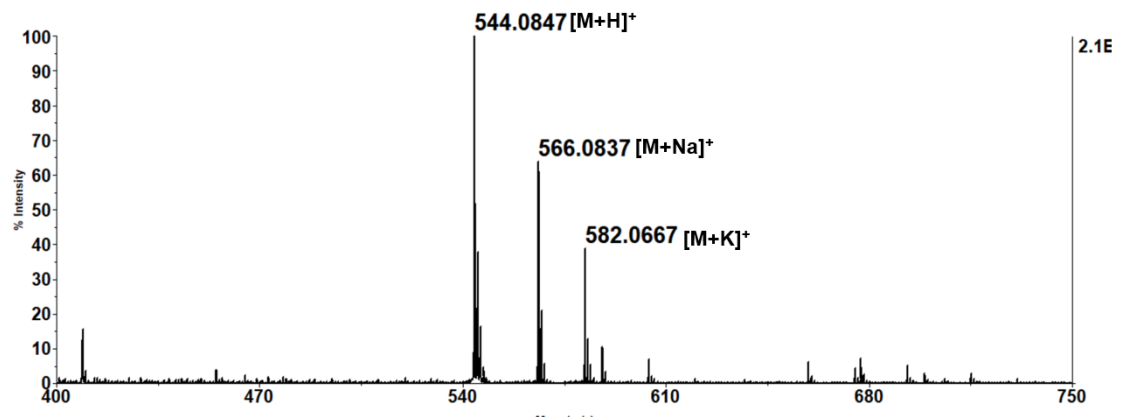
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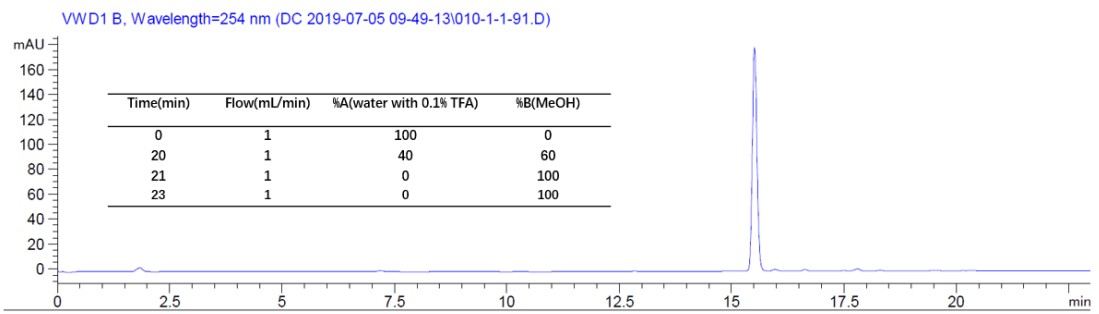
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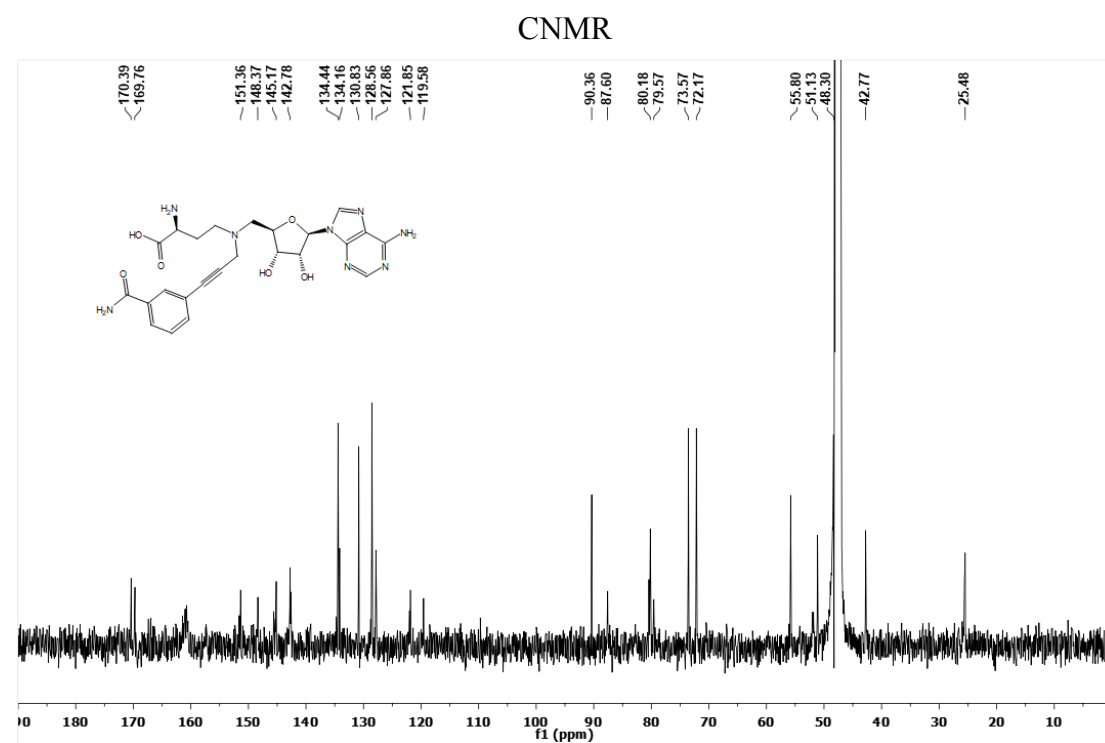
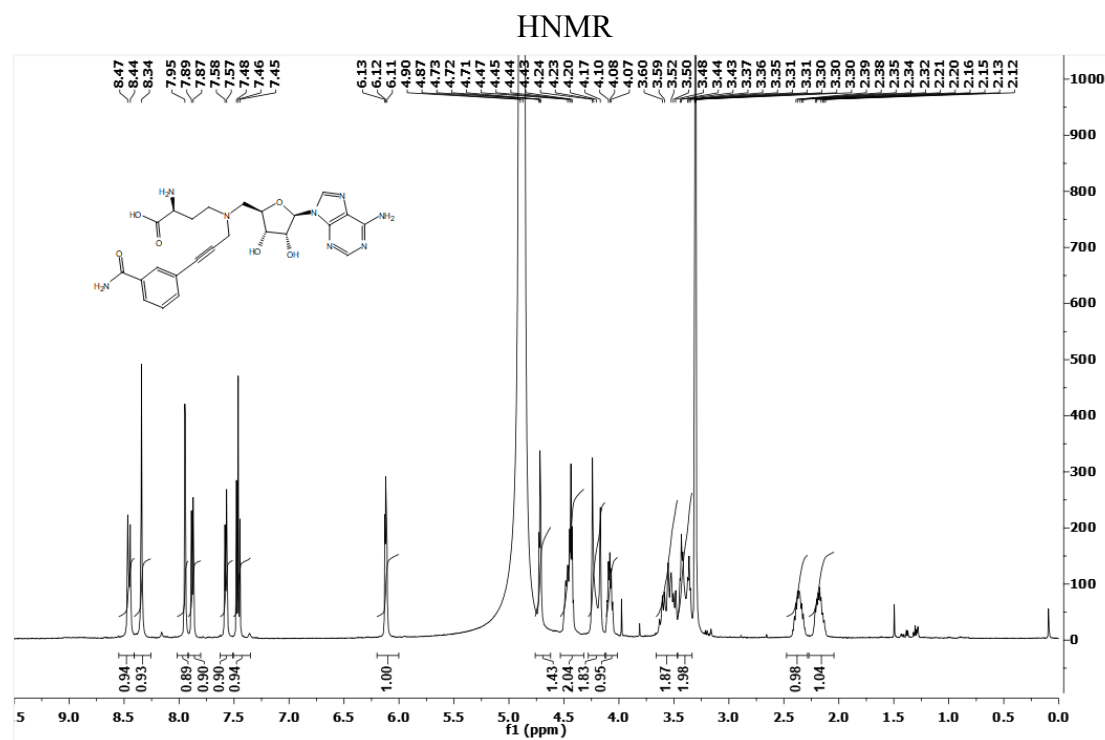
### MALDI-MS



### HPLC

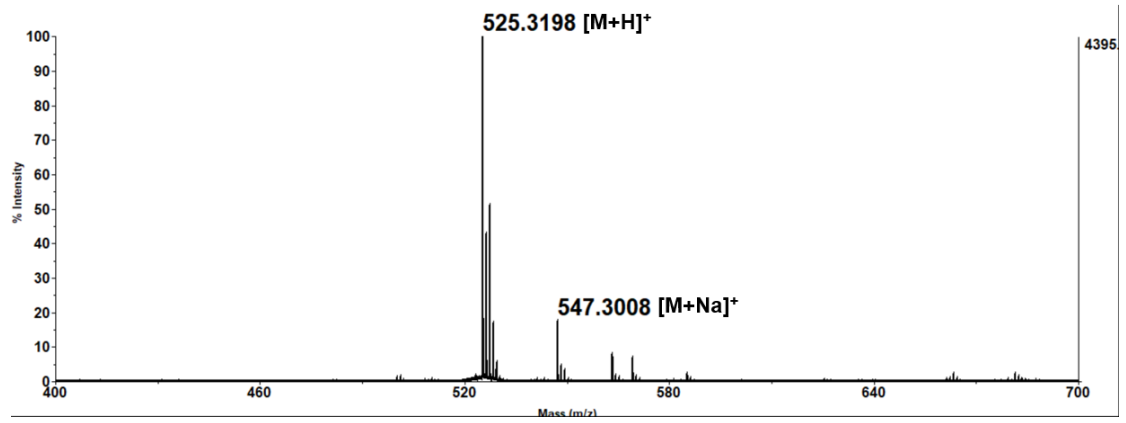


NMR, MALDI-MS and HPLC spectra of compound **2a**

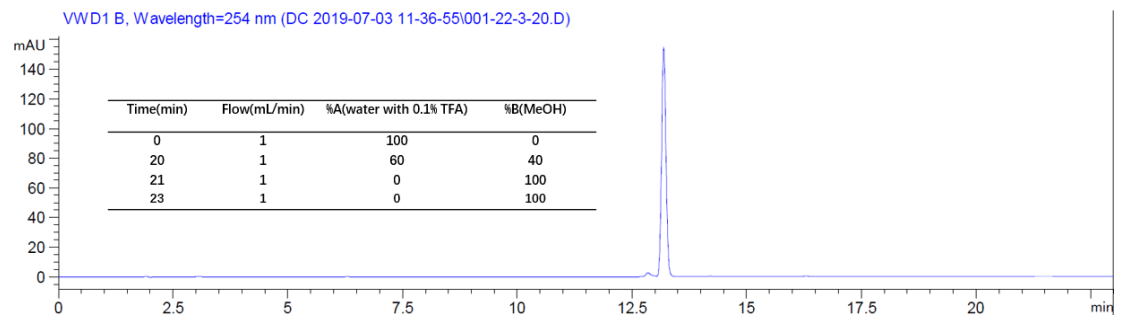




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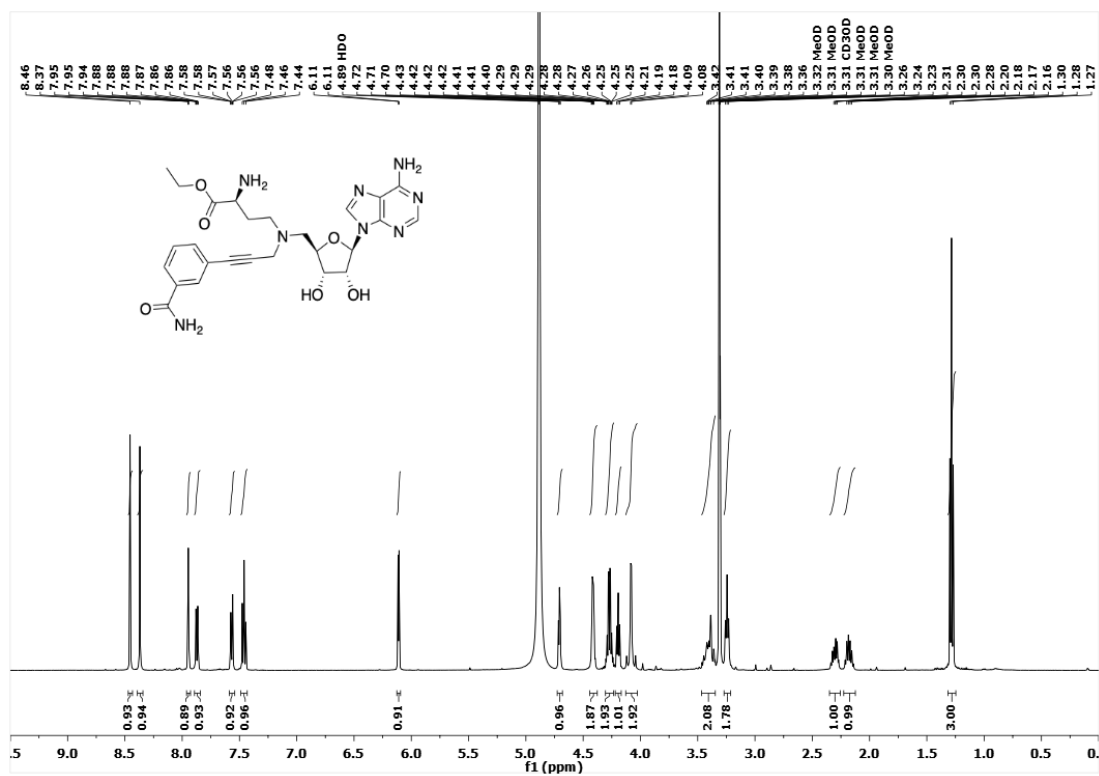


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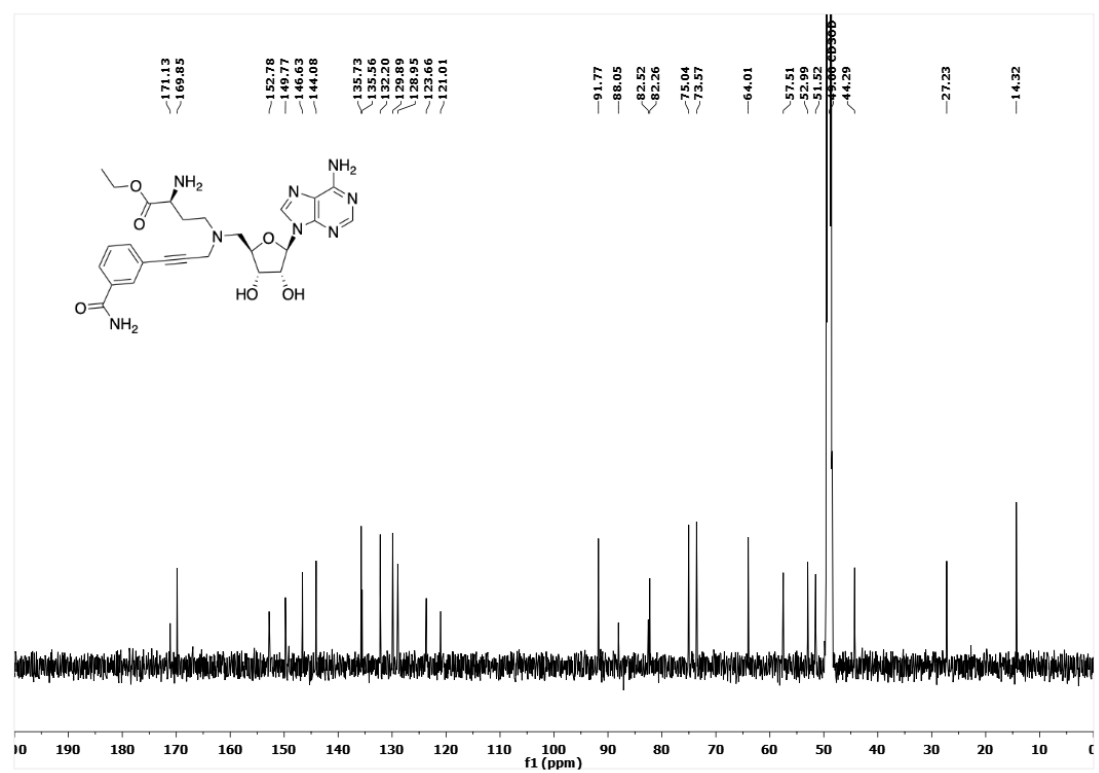


NMR, MALDI-MS and HPLC spectra of compound **2a\***

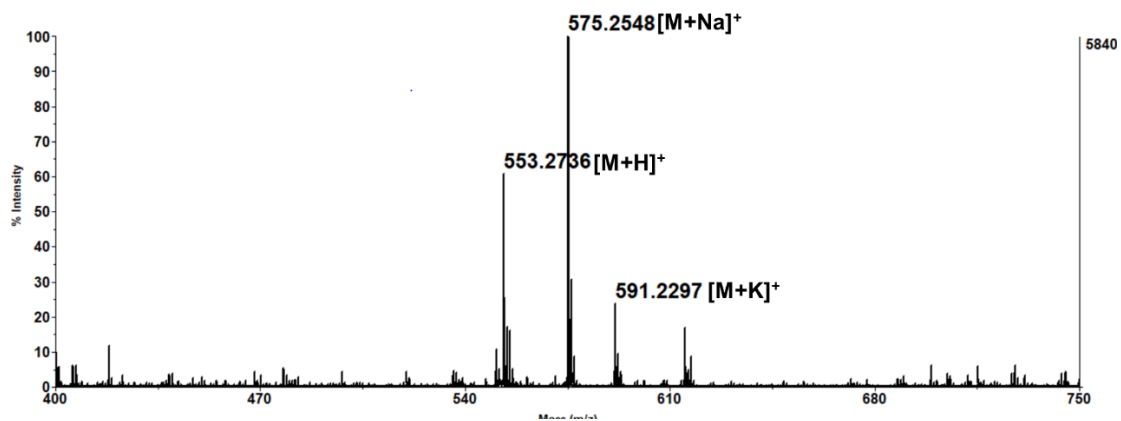
H-NMR



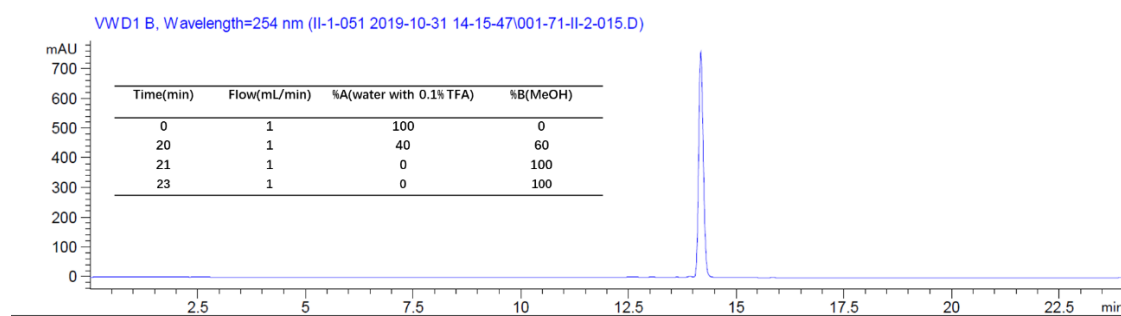
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### MALDI-MS

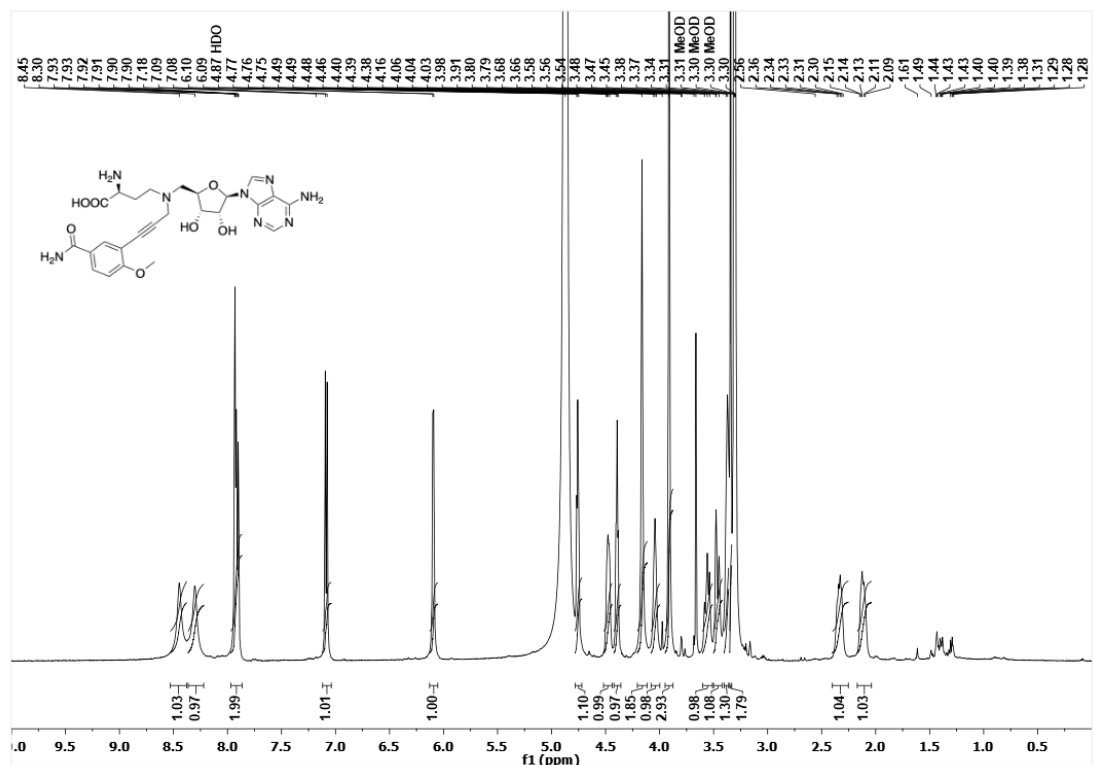


### HPLC

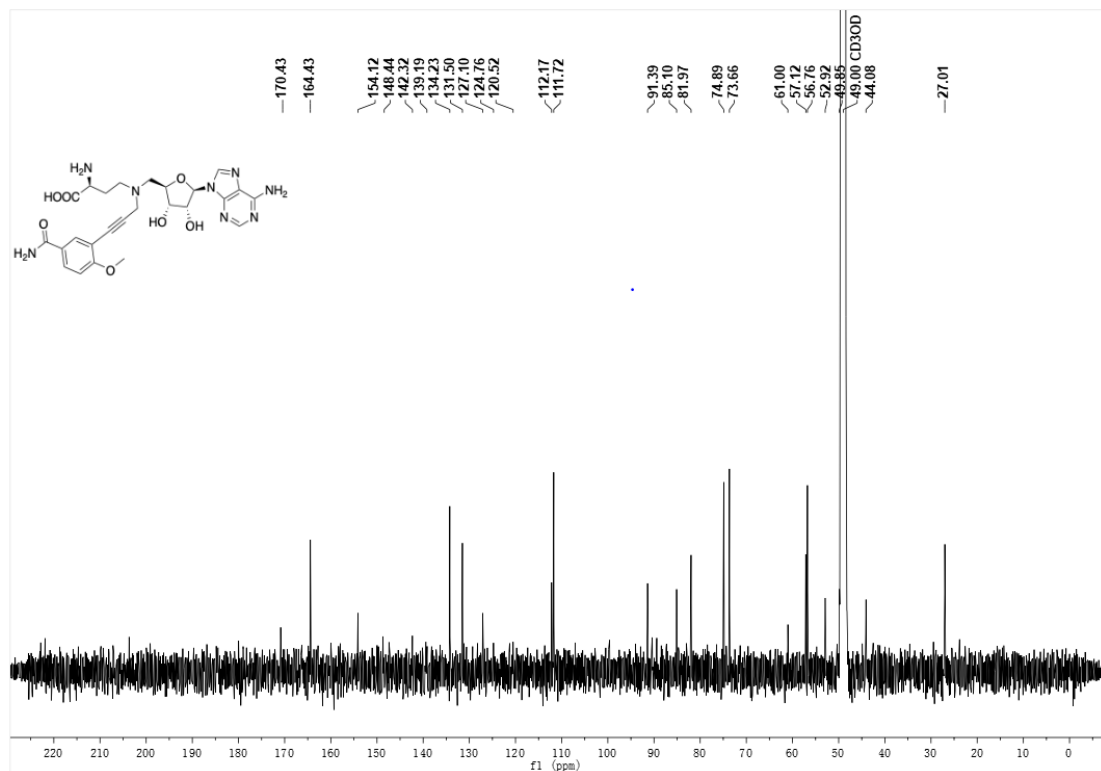


# NMR, MALDI-MS and HPLC spectra of compound 2b

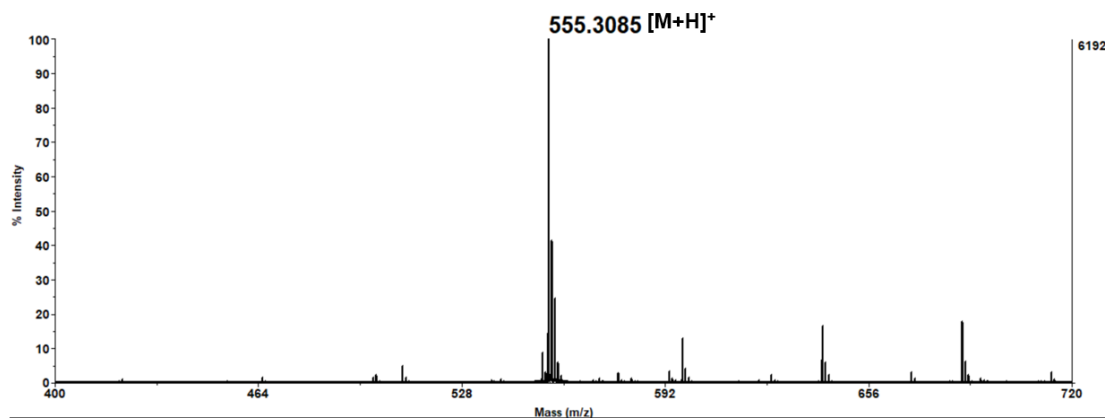
## HNMR



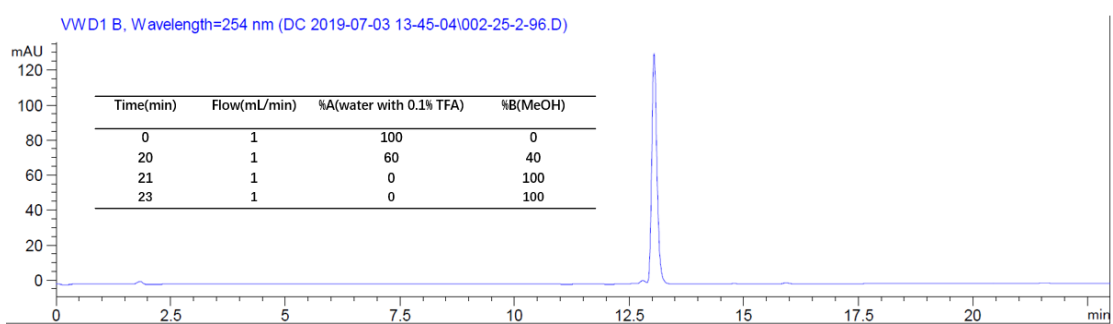
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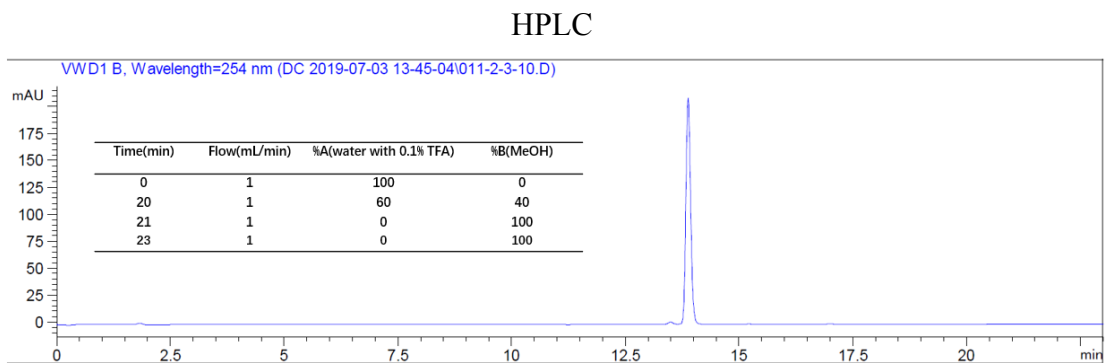
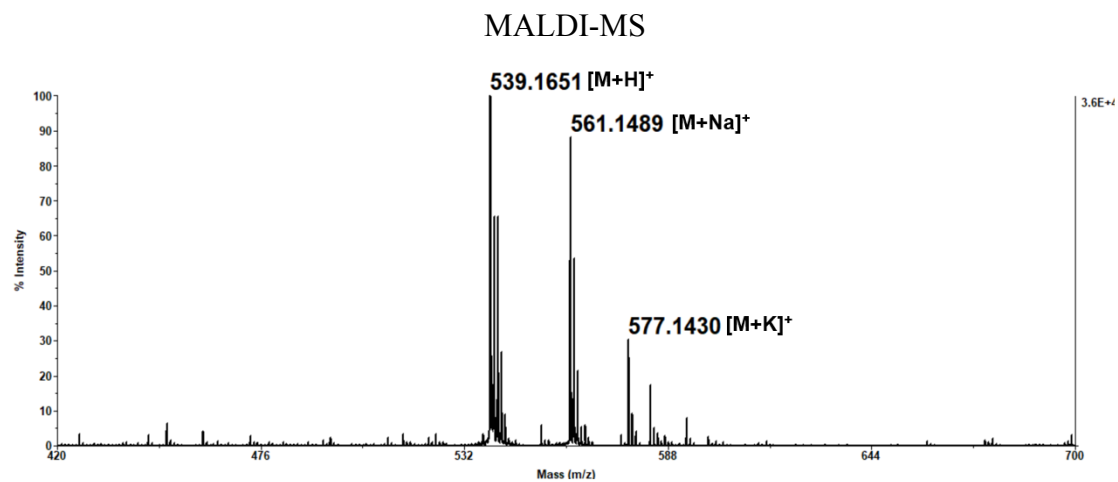
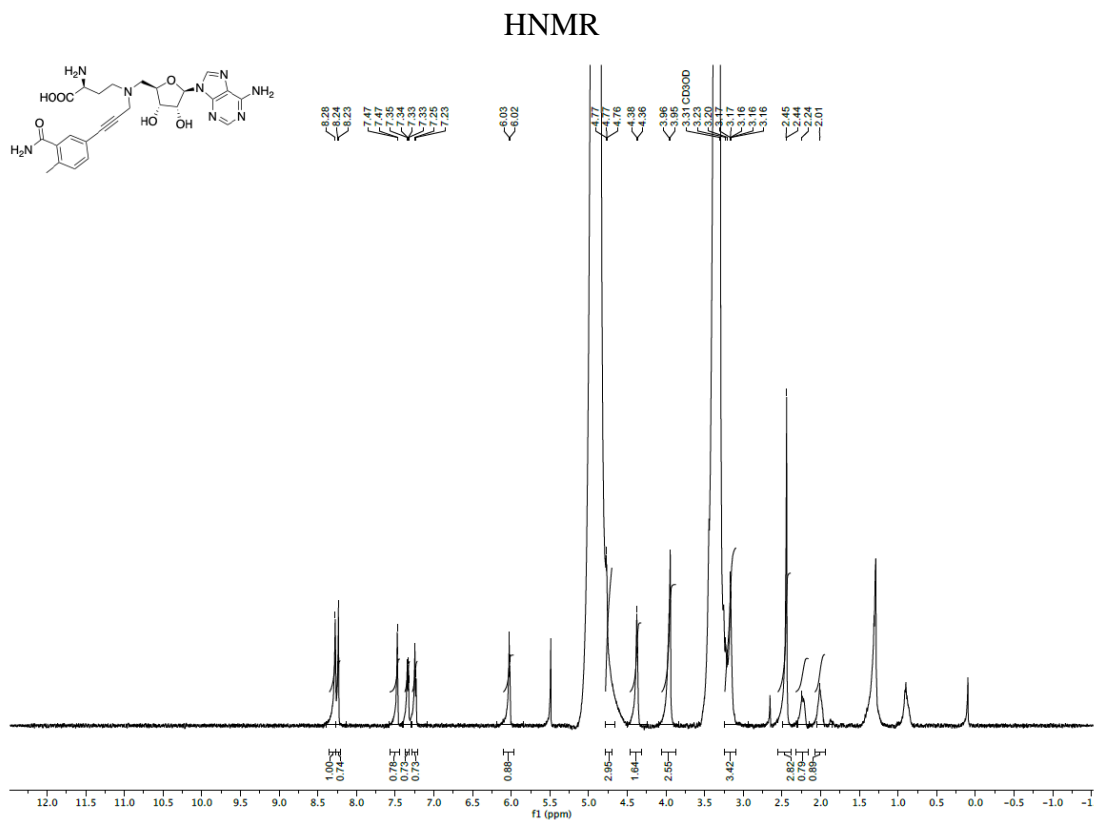
# MALDI-MS



# HPLC

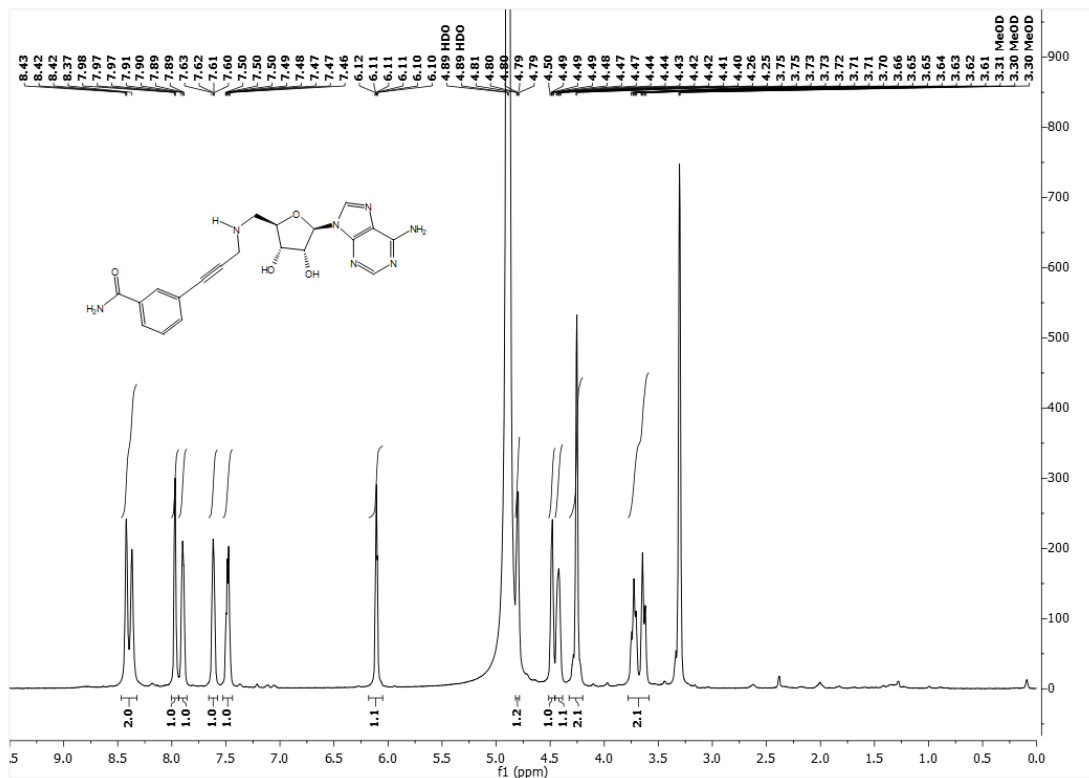


# NMR, MALDI-MS and HPLC spectra of compound 2c

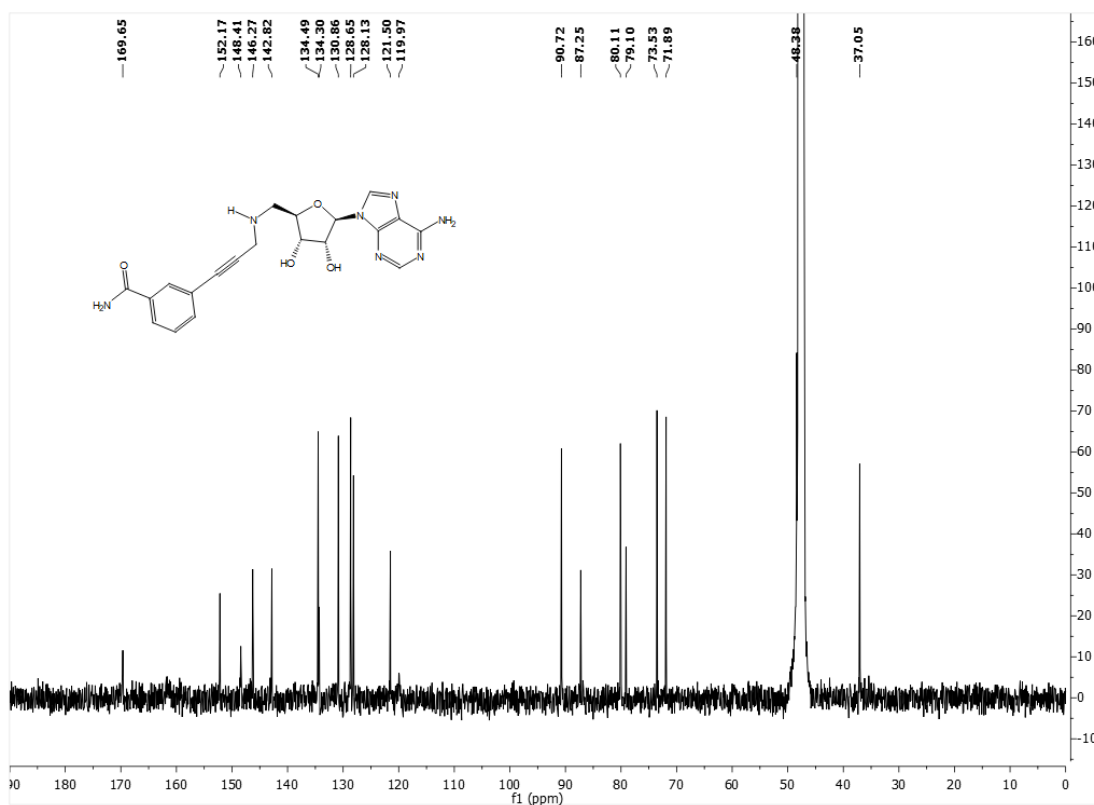


NMR, MALDI-MS and HPLC spectra of compound **2d**

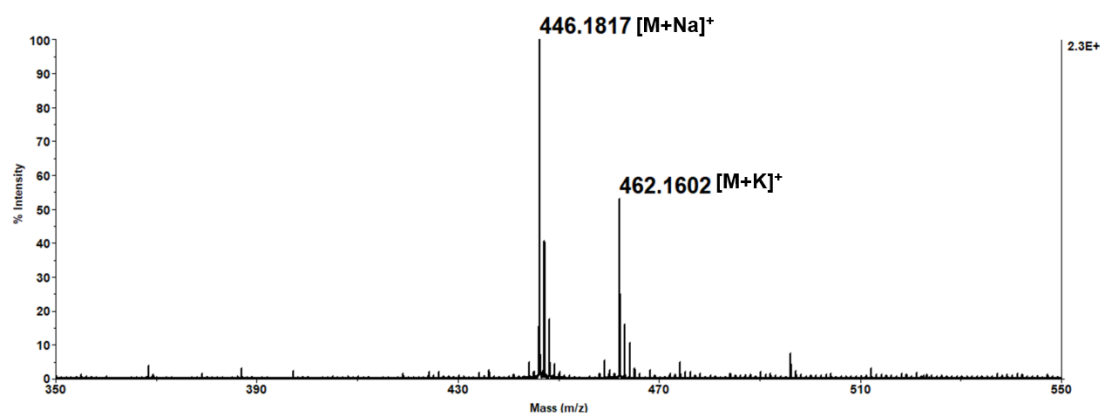
HNMR



CNMR

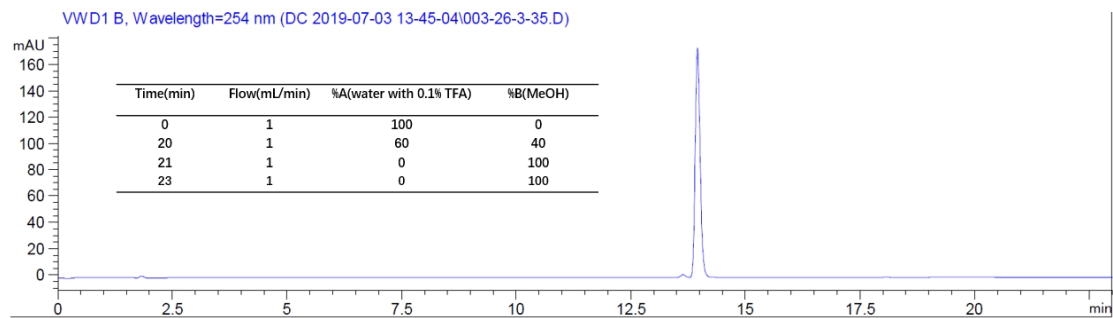


# MALDI-MS



# HPLC

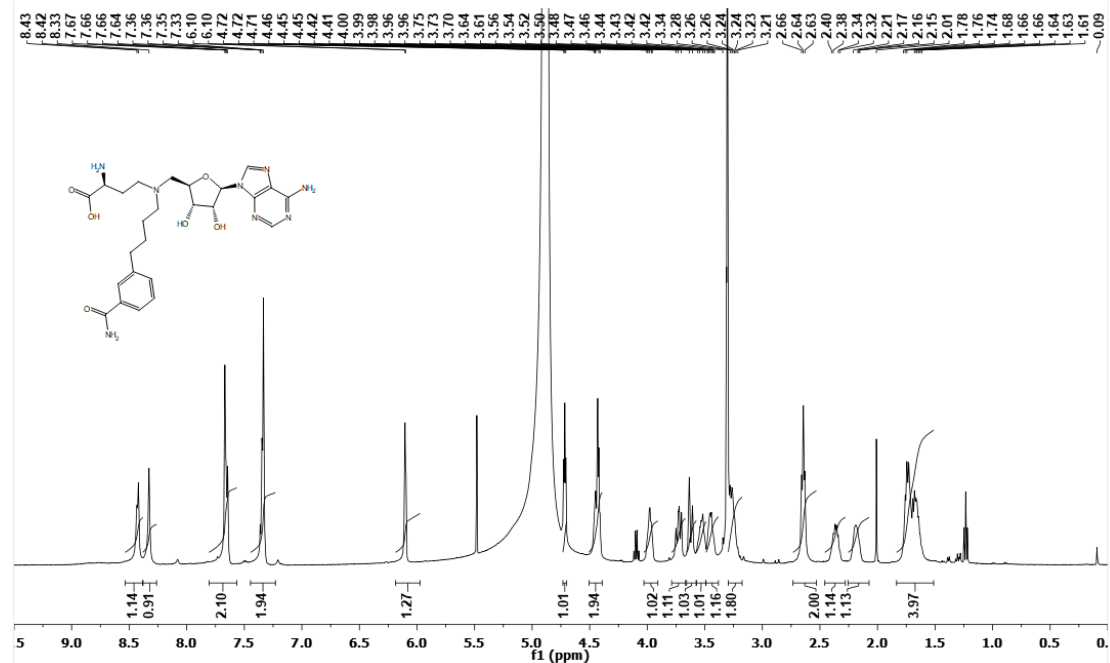
VWD1 B, Wavelength=254 nm (DC 2019-07-03 13-45-041003-26-3-35.D)



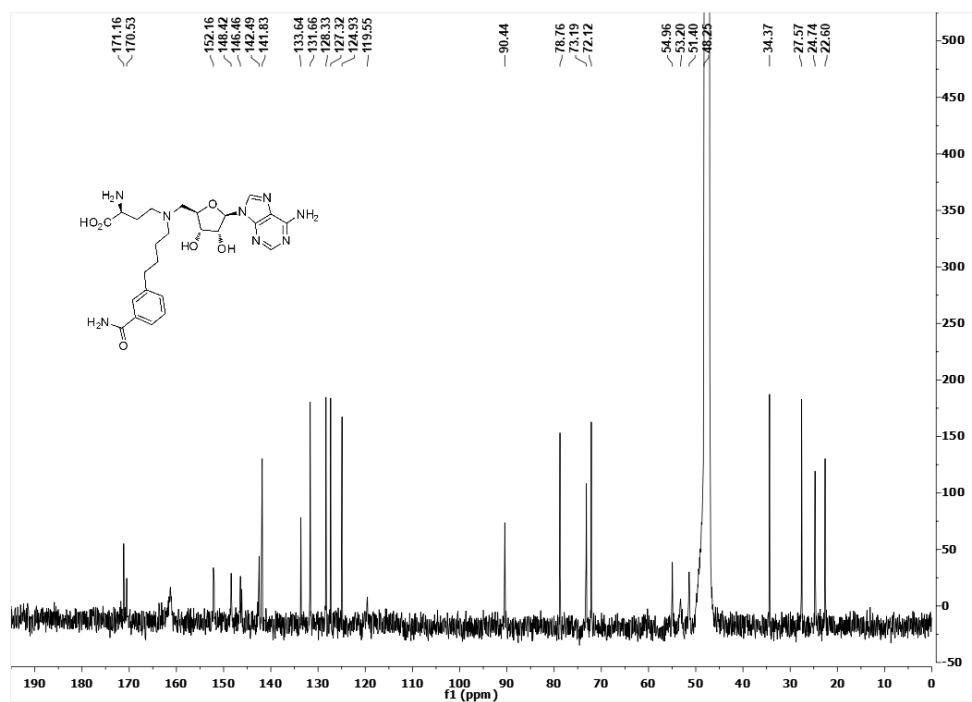


NMR, MALDI-MS and HPLC spectra of compound **3a**

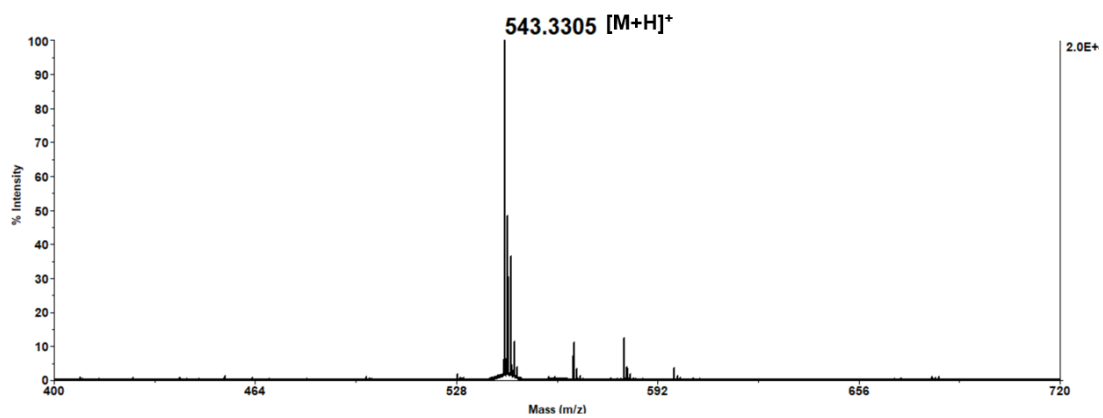
HNMR



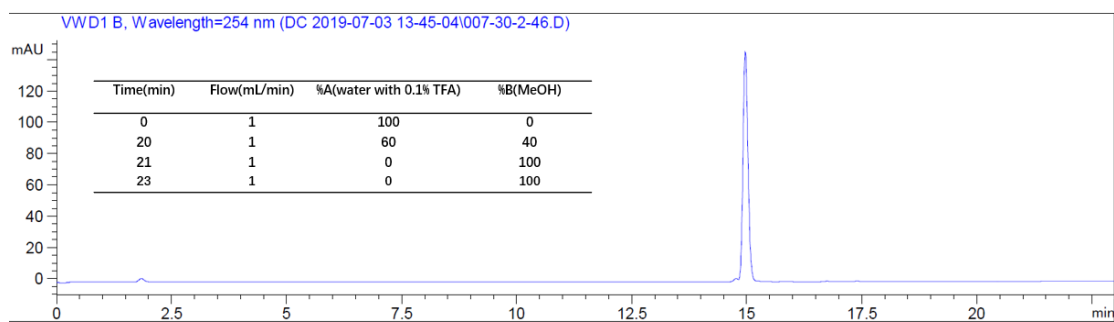
CNMR



# MALDI-MS

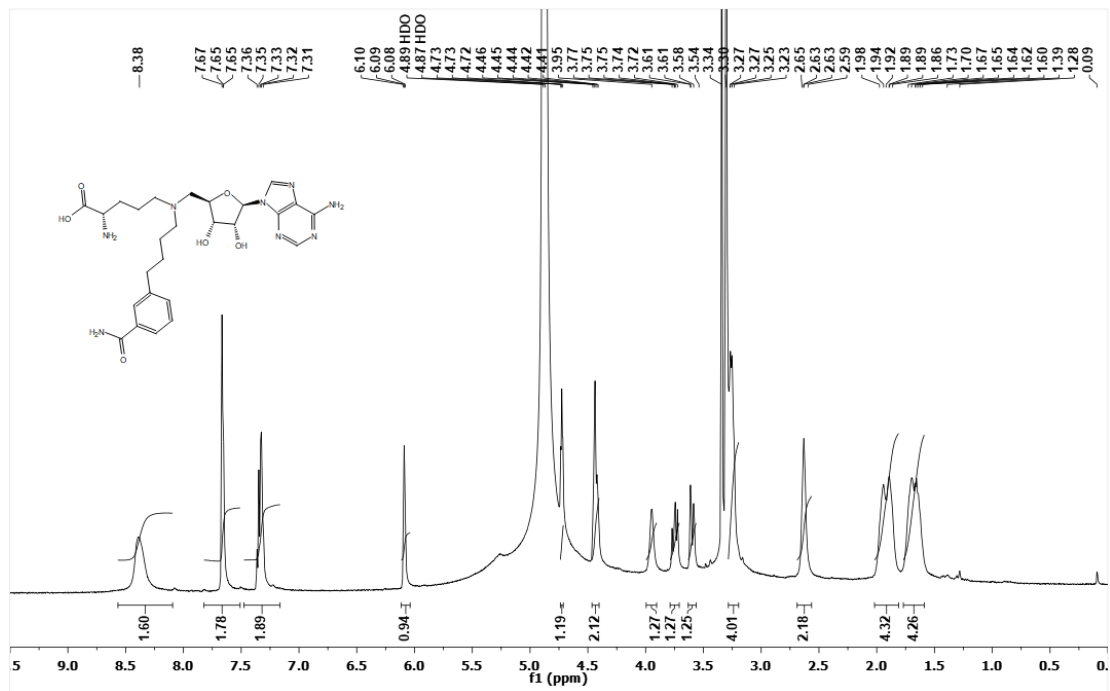


# HPLC

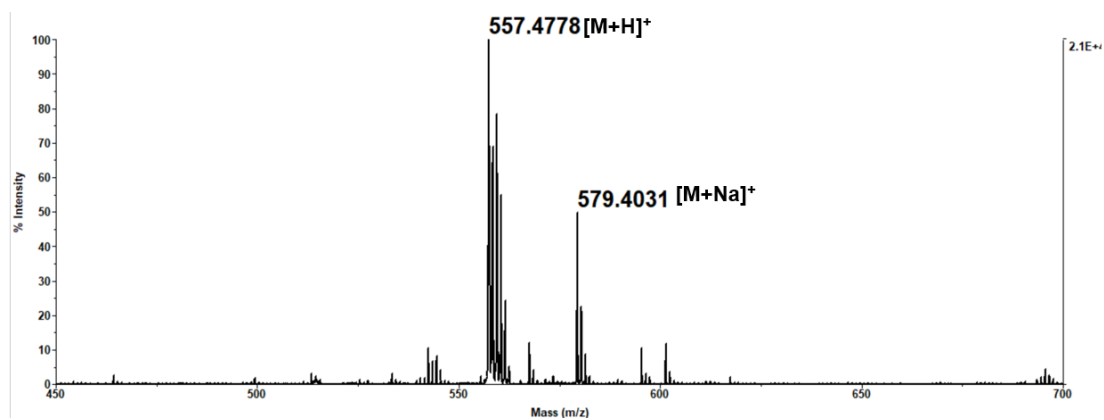


# NMR, MALDI-MS and HPLC spectra of compound 3b

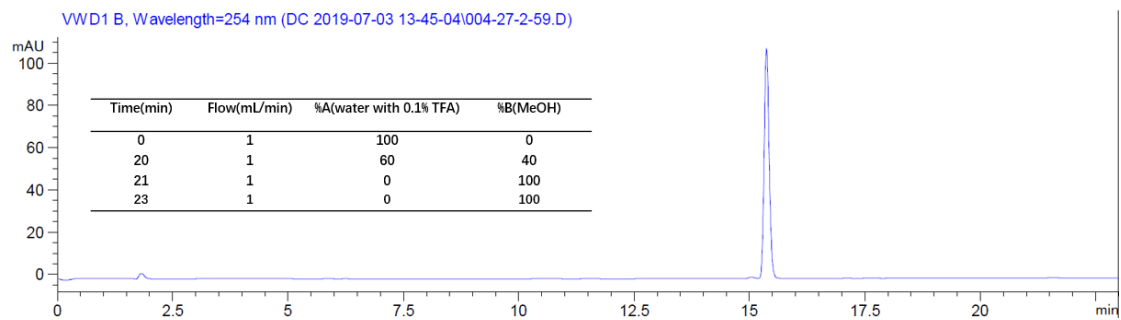
## HNMR

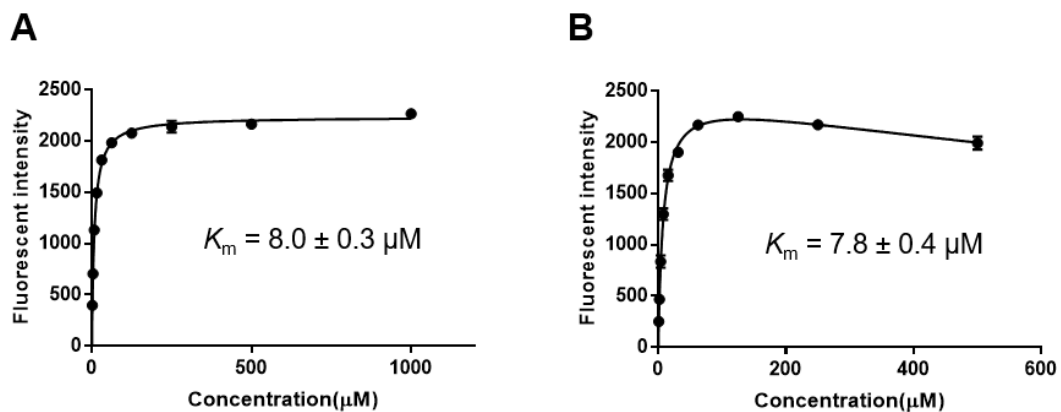


## MALDI-MS

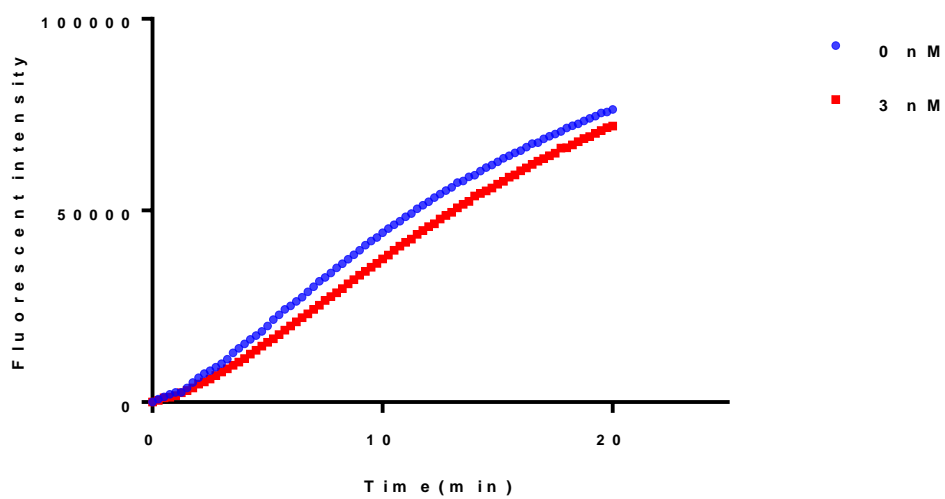


## HPLC

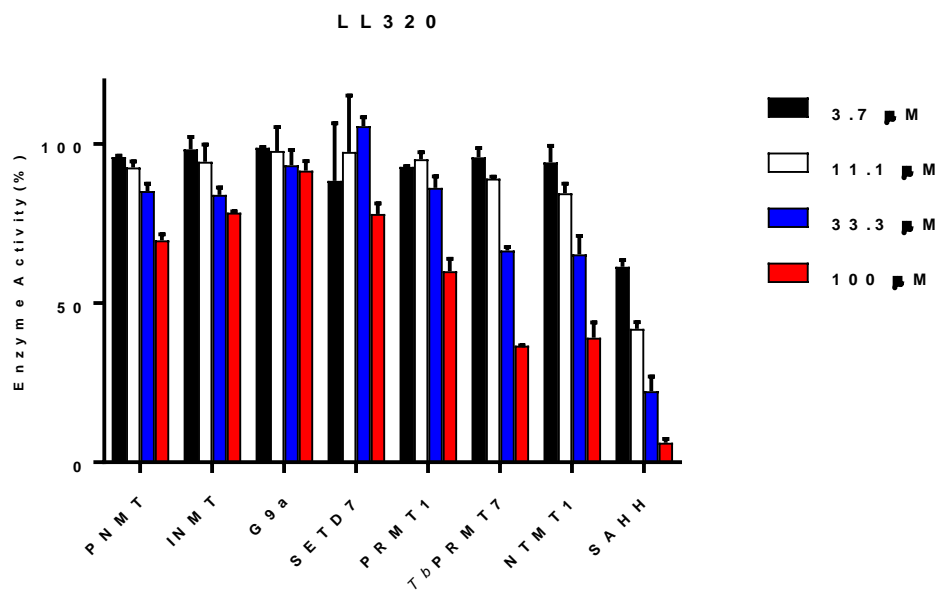




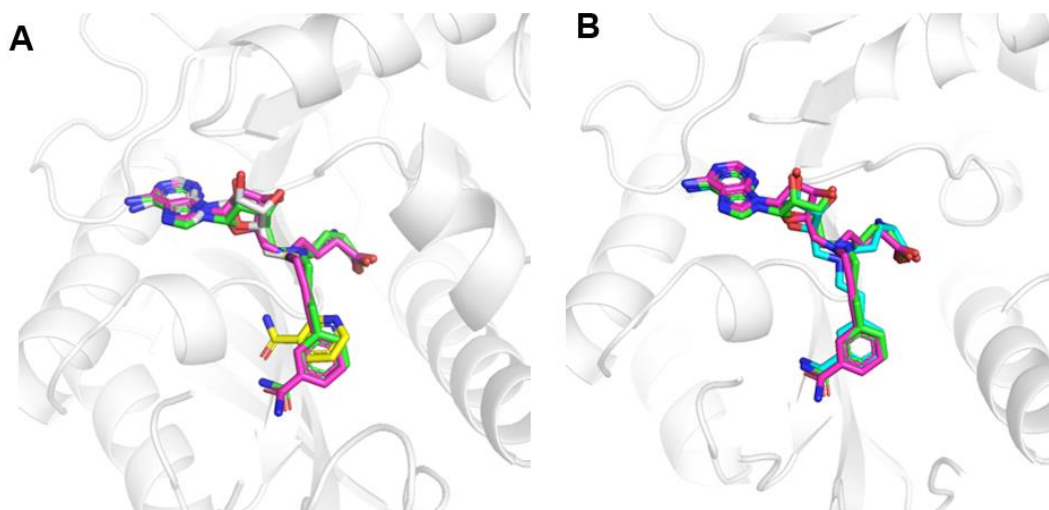
**Supporting Figure S1.** Kinetic parameter determination for NNMT.  $K_m$  values were determined for (A) nicotinamide at fixed SAM concentration (100 μM) and (B) for SAM at fixed concentration of nicotinamide (100 μM). Data were obtained in duplicate.



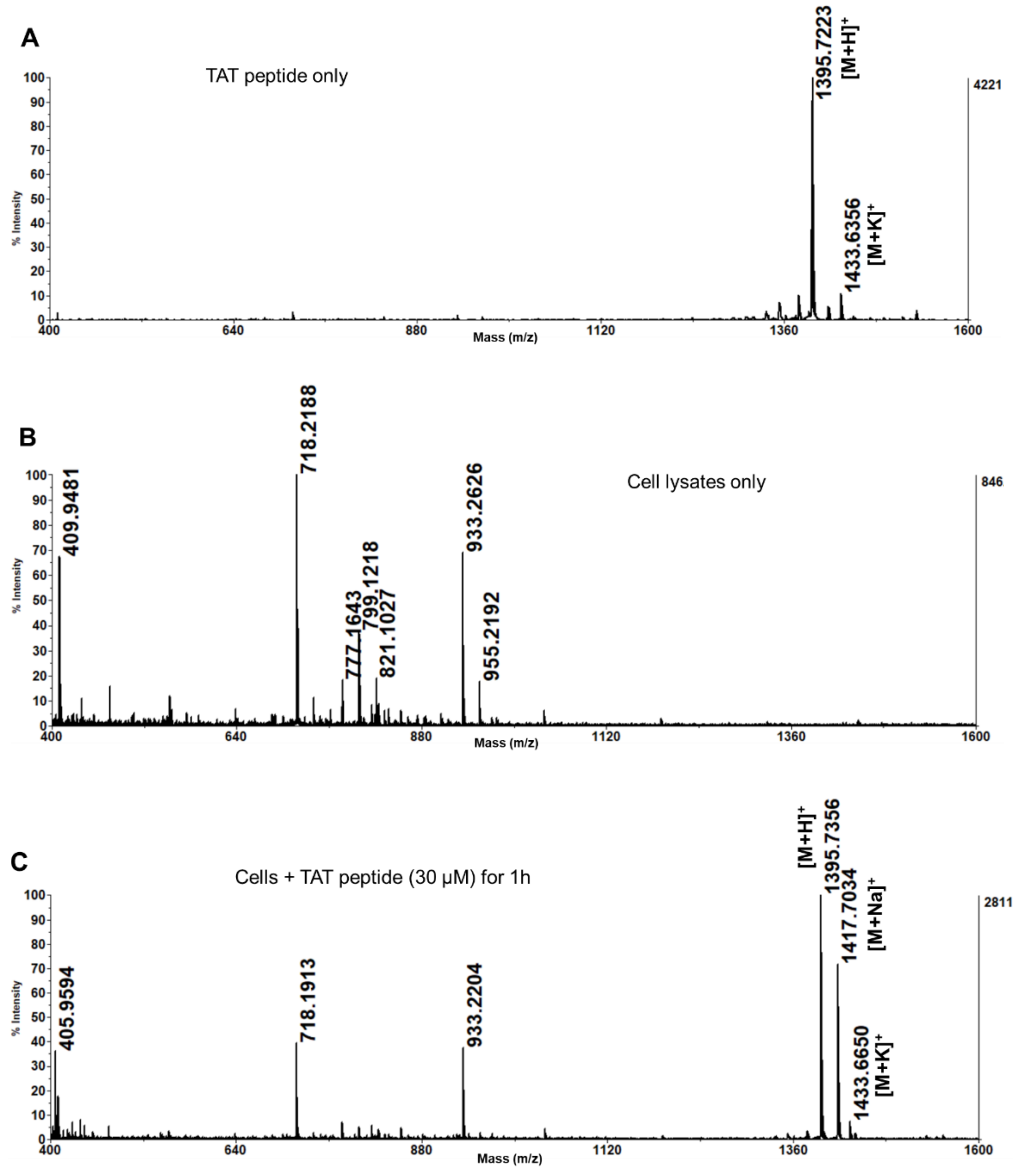
**Supporting Figure S2.** Rapid dilution assay for compound **LL320**



**Supporting Figure S3.** The selectivity study of LL320

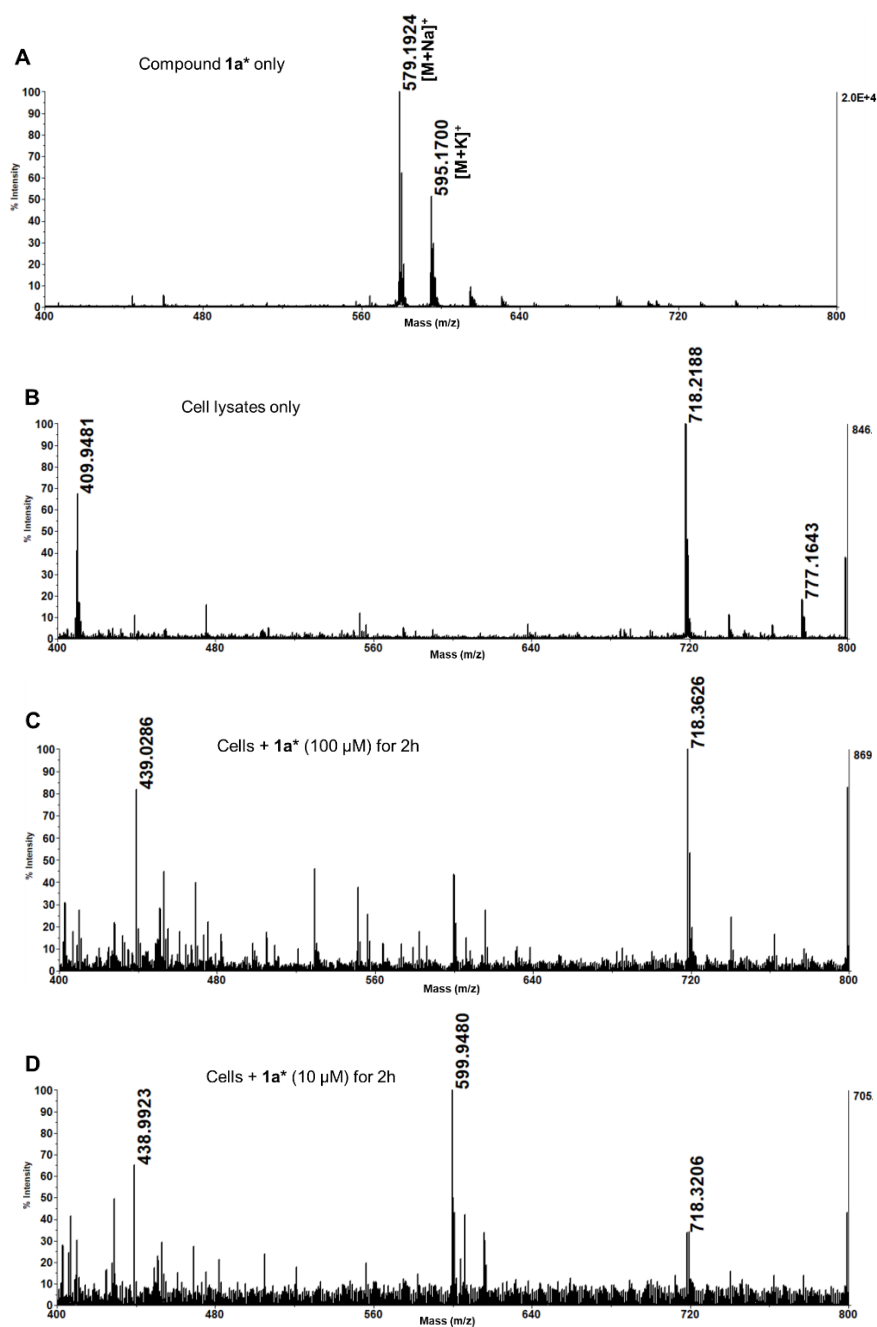


**Supporting Figure S4.** Superimposed co-crystal structures. (A) Overlay co-crystal structures NNMT (gray carton)- LL319 (green stick) (PDB ID: 6PVE), NNMT-LL320 (pink stick) (PDB ID: 6PVS) and NNMT-SAH (gray stick)-nicotinamide (yellow stick) (PDB ID: 3ROD); (B) Overlay co-crystal structures NNMT (gray carton)- LL319 (green stick) (PDB ID: 6PVE), NNMT-LL320 (pink stick) (PDB ID: 6PVS) and NNMT- MS2756 (blue stick) (PDB ID: 6CHH).

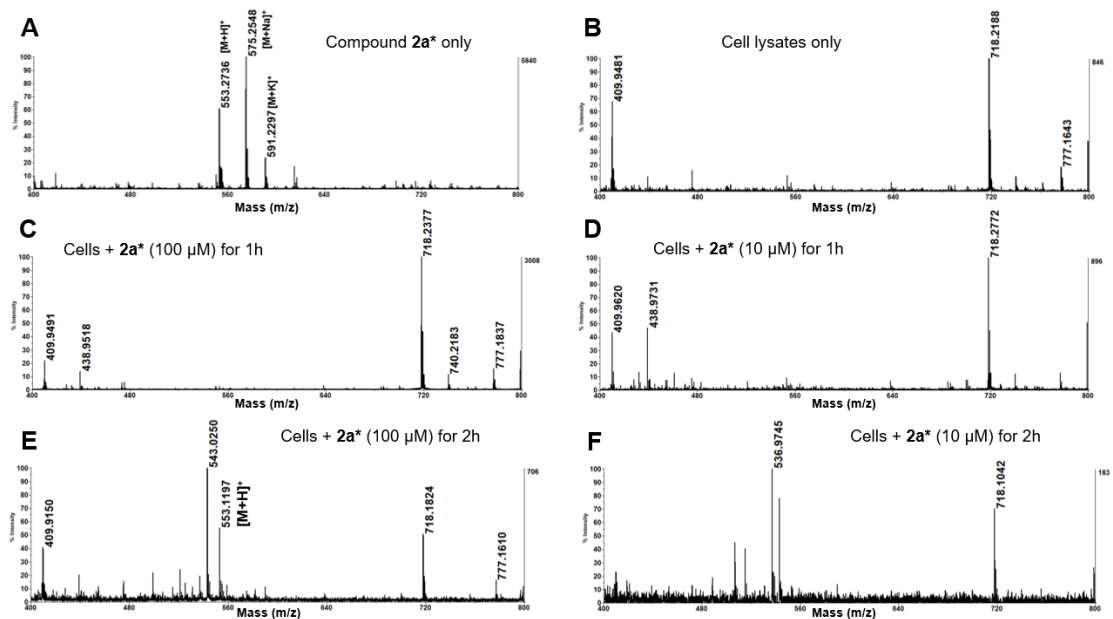


**Supporting Figure S5.** Cell permeability evaluation of TAT peptide as positive control. (A) MALDI-MS for TAT peptide (GRKKRRQRRR-NH<sub>2</sub>) only; (B) MALDI-MS for cell lysates only; (C) MALDI-MS for incubation of 30 μM TAT peptide with HCT116 cells for 1h.





**Supporting Figure S6.** Cell permeability evaluation of **1a\***. (A) MALDI-MS for compound **1a\*** only; (B) MALDI-MS for cell lysates only; (C) and (D) MALDI-MS for incubation of 100  $\mu$ M and 10  $\mu$ M **1a\*** with HCT116 cells for 2h, respectively.  $[\mathbf{1a^*}+\text{H}]^+ = 557.3$ ,  $[\mathbf{1a^*}+\text{Na}]^+ = 579.3$ ,  $[\mathbf{1a^*}+\text{K}]^+ = 595.2$ ,  $[\mathbf{1a^*}\text{-Et}+\text{H}]^+ = 529.3$ ,  $[\mathbf{1a^*}\text{-Et}+\text{Na}]^+ = 551.2$ ,  $[\mathbf{1a^*}\text{-Et}+\text{K}]^+ = 567.2$ .



**Supporting Figure S7.** Cell permeability evaluation of **2a\***. (A) MALDI-MS for compound **2a\*** only; (B) MALDI-MS for cell lysates only; (C) and (D) MALDI-MS for incubation of 100  $\mu\text{M}$  and 10  $\mu\text{M}$  **2a\*** with HCT116 cells for 1h, respectively; (E) and (F) MALDI-MS for incubation of 100  $\mu\text{M}$  and 10  $\mu\text{M}$  **2a\*** with HCT116 cells for 2h, respectively.  $[\mathbf{2a^*}+\text{H}]^+ = 553.3$ ,  $[\mathbf{2a^*}+\text{Na}]^+ = 575.2$ ,  $[\mathbf{2a^*}+\text{K}]^+ = 591.2$ ,  $[\mathbf{2a^*}-\text{Et}+\text{H}]^+ = 525.2$ ,  $[\mathbf{2a^*}-\text{Et}+\text{Na}]^+ = 547.2$ ,  $[\mathbf{2a^*}-\text{Et}+\text{K}]^+ = 563.2$ .

**Supplementary Table S1. Crystallography data and refinement statistics (PDB ID: 6PVE and 6PVS)**

<b>Data Collection</b>	<b>NNMT/LL319</b>	<b>NNMT/LL320</b>
$\lambda$ (Å)	1.0332	1.0332
Space group	P1	P1
a, b, c (Å)	45.90, 62.36, 107.86	46.23, 62.53, 108.36
$\alpha, \beta, \gamma$ (°)	91.78, 97.61, 111.56	82.72, 82.48, 68.34
Resolution (Å)*	50 – 2.30 (2.38 – 2.30)	50.90 – 2.57 (2.71 – 2.57)
Completeness (%)*	98.3 (99.1)	88.4 (94.7)
Redundancy*	2.8 (2.8)	1.8 (1.8)
$R_{\text{sym}}^{\dagger*}$	0.22 (0.75)	0.16 (1.0)
$I / \sigma(I)^*$	7.64 (1.6)	1.9 (0.6)
$CC_{1/2}$	0.95 (0.46)	0.98 (0.40)
<b>Refinement</b>		
Resolution (Å)	2.30	2.57
No. reflections	48089	31186
$R^{\S}/R_{\text{free}}^{\P}$	0.20/0.24	0.23/0.29
<b>r.m.s. deviations</b>		
Bonds (Å)	0.008	0.008
Angles (°)	1.35	1.31
No. Protein atoms	8172	8144
No. Ligand atoms	152	152
No. Waters	500	106
<b>B-factors (Å<sup>2</sup>)</b>		
Wilson B	20.28	42.12
Protein	25.01	44.84
Ligands	16.59	39.83
Waters	26.36	37.69
<b>Ramachandran Analysis<sup>‡</sup></b>		
Favored (%)	97.61	96.08
Allowed (%)	2.20	3.53
Outliers (%)	0.19	0.38
PDB code	6PVE	6PVS

<sup>†</sup>  $R_{\text{sym}} = \sum_{hkl,j} (|I_{hkl} - \langle I_{hkl} \rangle|) / \sum_{hkl,j} I_{hkl}$ , where  $\langle I_{hkl} \rangle$  is the average intensity for a set of j symmetry related reflections and  $I_{hkl}$  is the value of the intensity for a single reflection within a set of symmetry-related reflections.

<sup>§</sup> R factor =  $\sum_{hkl} (||F_o| - |F_c||) / \sum_{hkl} |F_o|$  where  $F_o$  is the observed structure factor amplitude and  $F_c$  is the calculated structure factor amplitude.

<sup>¶</sup>  $R_{\text{free}} = \sum_{hkl,T} (||F_o| - |F_c||) / \sum_{hkl,T} |F_o|$ , where a test set, T (5% of the data), is omitted from the

refinement.

‡ Performed using Molprobit within PHENIX.

\* Indicates statistics for last resolution shell shown in parenthesis.

## Reference

- (1) Zhang, G.; Richardson, S. L.; Mao, Y.; Huang, R. Design, Synthesis, and Kinetic Analysis of Potent Protein N-Terminal Methyltransferase 1 Inhibitors. *Org. Biomol. Chem.* **2015**, *13* (14), 4149–4154.
- (2) Douat, C.; Heitz, A.; Martinez, J.; Fehrentz, J. A. Stereoselective Synthesis of Allyl- and Homoallylglycines. *Tetrahedron Lett.* **2001**, *42* (19), 3319–3321.