

**Figure S1. *Dnase113<sup>-/-</sup>Cd40lg<sup>-/-</sup>* mice do not develop autoimmunity, Related to Figure 1.**

Mice: *Dnase113<sup>-/-</sup>* (red symbols), CD40L (*Cd40lg<sup>-/-</sup>*, grey symbols), *Dnase113<sup>-/-</sup>Cd40lg<sup>-/-</sup>* mice (green symbols) or wild-type controls (WT, open symbols). In all panels, symbols represent individual mice and bars indicate median, wherever applicable.

**(A)** Frequency of CD23<sup>+</sup>CD21<sup>-</sup> follicular (FO) B cells among total CD19<sup>+</sup>B220<sup>+</sup> B cells from 12-mo-old mice as determined by flow cytometry.

**(B)** ELISpot image showing comparative analysis of ImmunoSpots of anti-IgG, anti-dsDNA, and anti-nucleosome antibody-forming cells (AFCs), in the spleen of 12-mo-old mice.

**(C)** Total number of anti-IgG antibody-forming cells (AFCs) in the spleen of 12-mo-old mice as determined by ELISpot (AFC per  $10^6$  total cells).

**(D)** Total IgG in the serum of 12-mo old mice as measured by ELISA.

**(E)** Frequency of CD25<sup>+</sup>FOXP3<sup>+</sup> T-regulatory cells among CD4<sup>+</sup> T cells of 12-mo-old mice as determined by flow cytometry.

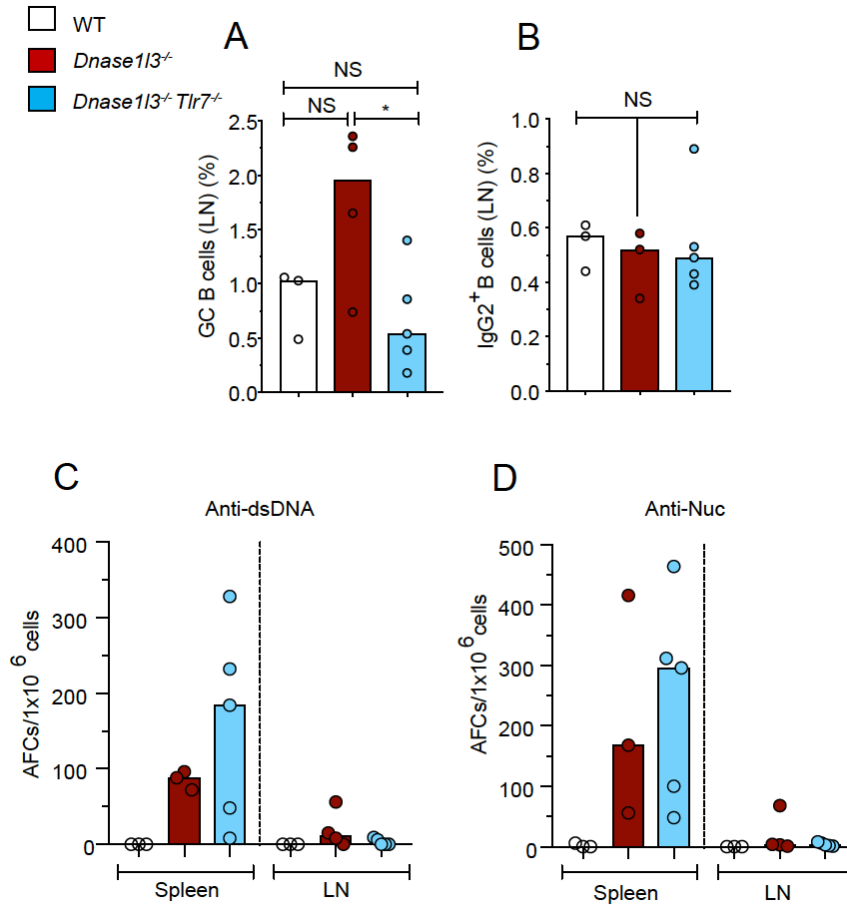
**(F)** Representative immunofluorescence images of spleen sections from 12-mo-old-mice of indicated strains stained for IgD<sup>+</sup> naïve follicular B cells (blue) and GL-7<sup>+</sup> GC B cells (green). Upper panels scale bar 100  $\mu$ m, lower panels 30  $\mu$ m.

**(G)** Fractions of CD38<sup>-</sup> GL-7<sup>+</sup> GC B cells among total CD19<sup>+</sup>B220<sup>+</sup> splenic B cells from 12-mo-old mice, as determined by flow cytometry.

**(H)** Fractions of CXCR5<sup>hi</sup>PD-1<sup>hi</sup> GC-Tfh population among CD4<sup>+</sup> splenic T cells from 12-mo-old mice, as determined by flow cytometry.

**(I)** Spleen weight of mice at 12-months of age.

One-way ANOVA followed by the Tukey multiple-comparison test was used for statistical analysis of more than two groups. Statistical significance: \*  $p \leq 0.05$ , \*\*  $p \leq 0.01$ , and \*\*\*\*  $p \leq 0.0001$ .



**Figure S2. Anti-dsDNA and anti-Nuc AFCs in *Dnase113*<sup>-/-</sup> and *Dnase113*<sup>-/-</sup> *Tlr7*<sup>-/-</sup> mice develop in the spleen and not in LNs, Related to Figure 2.**

Mice: *Dnase113*<sup>-/-</sup> were examined along with *Dnase113*<sup>-/-</sup> *Tlr7*<sup>-/-</sup> mice (blue symbols) and wild-type controls (WT, open symbols). In panels A-D, symbols represent individual mice and bars indicate median.

**(A and B)** Fraction of CD38<sup>+</sup>GL-7<sup>+</sup> GC B cells (A) and IgD<sup>+</sup>IgM<sup>+</sup>IgG2a/2b<sup>+</sup> (switched) cells among CD19<sup>+</sup>B220<sup>+</sup> B cells from inguinal and mesenteric lymph nodes (LNs) of 6-8-mo-old mice as determined by flow cytometry.

**(C and D)** Total number of anti-dsDNA (C) and anti-nucleosome (D) antibody forming cells (AFCs) in the spleen and corresponding inguinal/ mesenteric lymph nodes (LN) of 6-8-mo-old mice determined by ELISpot assay.

One-way ANOVA followed by the Tukey multiple-comparison test was used for statistical comparison of more than two groups. Statistical significance: NS = not significant, and \*  $p \leq 0.05$ .

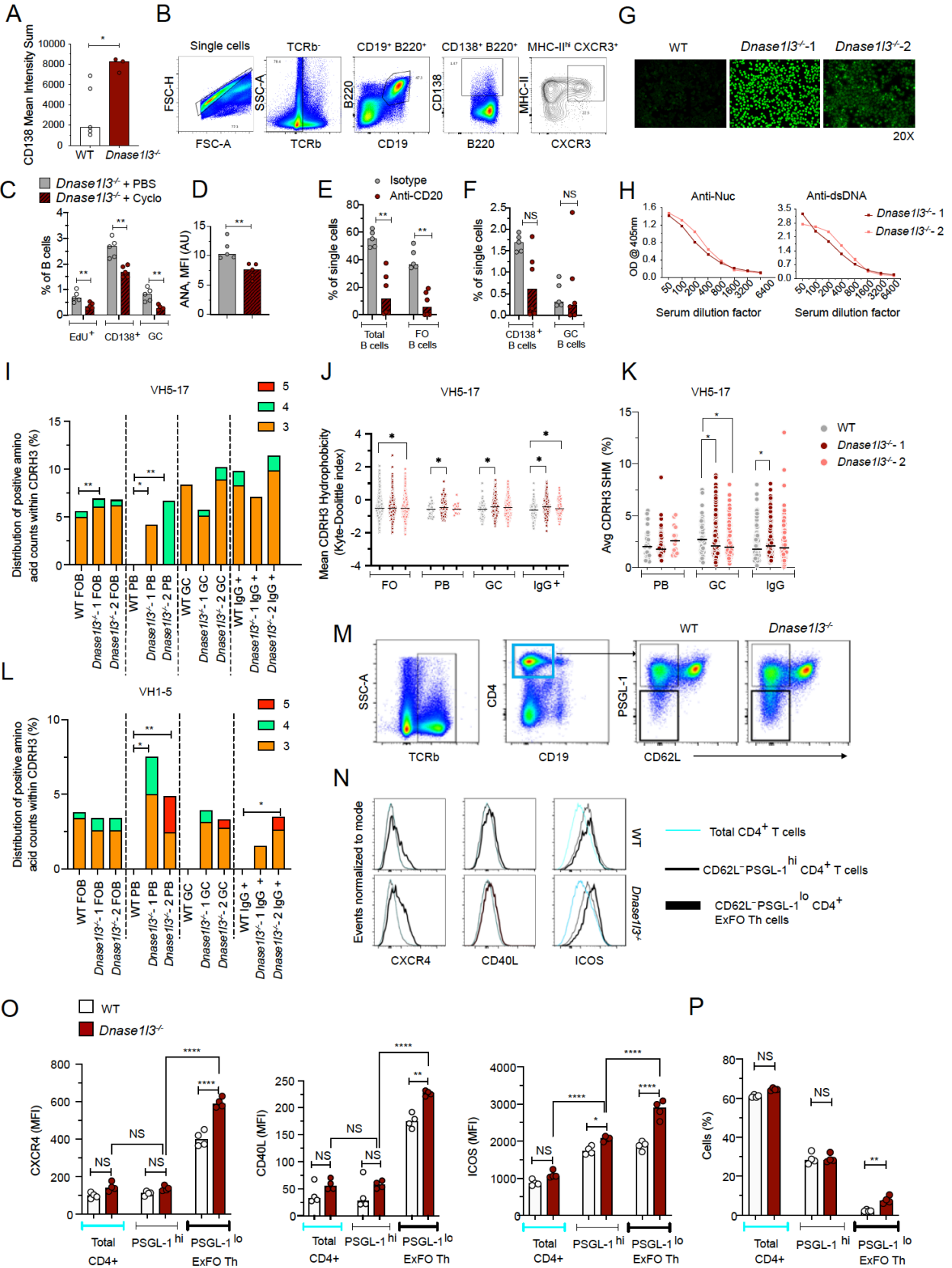


Figure S3. Characteristics of ExFO B cells and ExFO Th cells in WT vs *Dnase1/3<sup>-/-</sup>* mice, Related to Figure 3.

**(A)** Sum of mean intensity of CD138<sup>+</sup> cells in spleen sections of WT (open symbols) vs *Dnase113*<sup>-/-</sup> spleen (red symbols), acquired using confocal microscopy, as in main figure 3, panel A. Symbols represent individual mice and bars indicate median.

**(B)** Representative flow plots show successive gating strategy used for the characterization of splenic TCRβ<sup>+</sup> CD19<sup>+</sup> B220<sup>+/low</sup> CD138<sup>hi</sup> cells with high expression of CXCR3 and MHC-II (ExFO B cells) from a *Dnase113*<sup>-/-</sup> mouse.

**(C)** Frequency of EdU<sup>+</sup> cells, total CD138<sup>+</sup> cells and GL-7<sup>+</sup> GC cells among total B220<sup>+</sup> CD19<sup>+</sup> B cells from the spleens of 6-mo-old *Dnase113*<sup>-/-</sup> mice treated with PBS (grey symbols) or cyclophosphamide (cyclo, red symbols).

**(D)** ANA fluorescence intensity quantified from HEp-2 slides stained with sera from PBS (grey symbols) or cyclo-treated (red symbols) *Dnase113*<sup>-/-</sup> mice, and probed with secondary anti-Igκ antibody to detect ANAs.

**(E, F)** Fraction of TCRβ<sup>+</sup> CD19<sup>+</sup> total B cells and TCRβ<sup>+</sup> CD19<sup>+</sup> CD23<sup>hi</sup> CD21<sup>-</sup> FO B cells (E); B220<sup>+</sup> CD138<sup>+</sup> Plasmablasts and B220<sup>+</sup> CD38<sup>+</sup> GL-7<sup>+</sup> GC B cells (F), among splenic single lymphocytes in IgG2a isotype (open symbols), and anti-CD20 treated (red symbols) *Dnase113*<sup>-/-</sup> mice at 4 weeks after 2 doses of treatment, as determined by flow cytometry. Significance was estimated using nonparametric Mann-Whitney test.

**(G)** Fluorescence microscopy images of HEp-2 slides stained with WT or *Dnase113*<sup>-/-</sup>-1 or *Dnase113*<sup>-/-</sup>-2 mouse sera and probed with secondary anti-Igκ antibody to detect ANAs.

**(H)** Serum anti-Nucleosome (Nuc, left panel) and anti-dsDNA (right panel), IgG reported as OD at 405nm at the indicated serum dilutions, measured by ELISA.

**(I)** Percentage of IGHV5-17 clonotypes in each repertoire that contain 3 (yellow), 4 (green), or 5 (red) positively charged amino acids within the CDR-H3 region of indicated B cell subsets and mice.

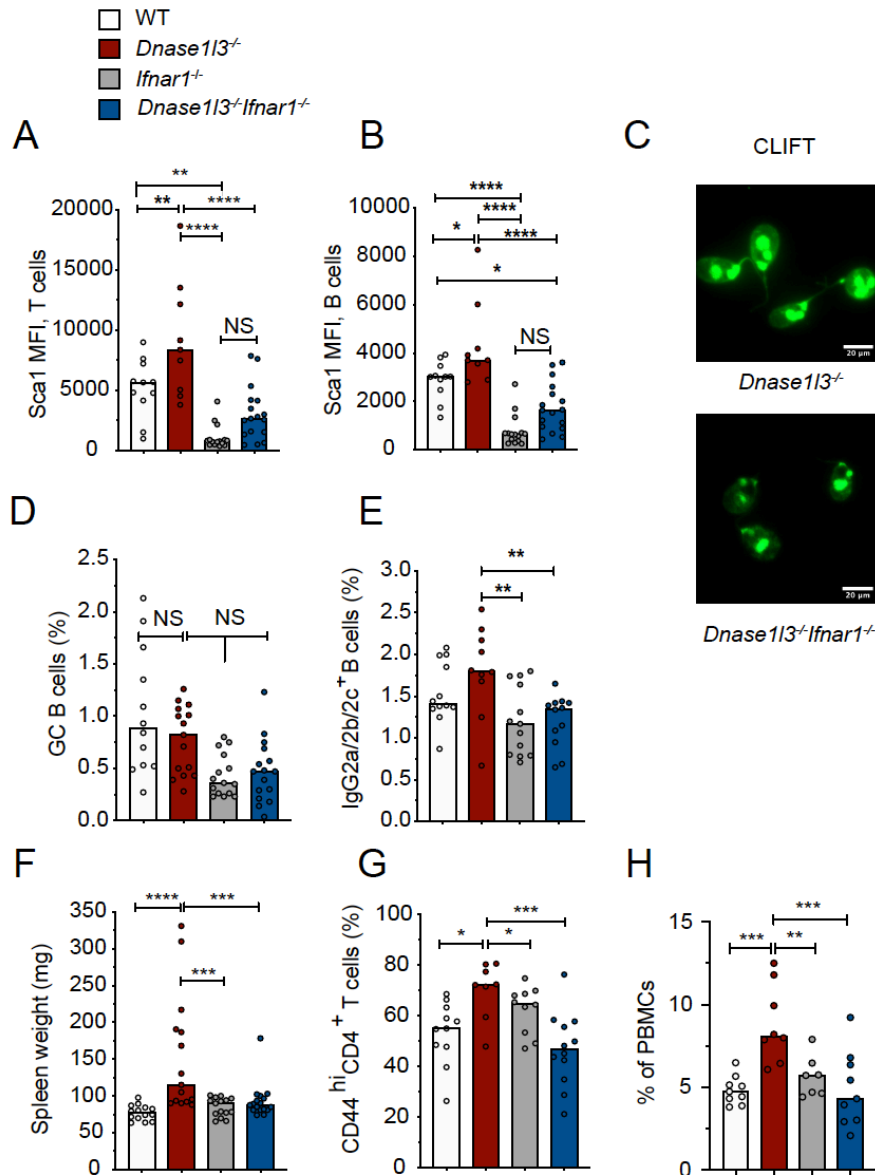
**(J, K)** Mean hydrophobicity (Kyte-Doolittle index), and percent distribution of SHM (somatic hypermutation) of the CDR-H3 region among the clonotypes that use V-gene IGHV5-17. Each point represents one clonotype. Error bars represent the mean and S.D. of all clonotypes in each group.

**(L)** Percentage of IGHV1-5 clonotypes in each repertoire that contain 3, 4, or 5 positively charged amino acids within the CDR-H3 region.

**(M)** Representative flow plots show gating strategy for the characterization of splenic CD19<sup>-</sup> TCRβ<sup>+</sup> CD4<sup>+</sup> cells (blue gate) with low levels of PSGL-1 and CD62L (ExFO Th cells, thick black gate) or high-PSGL-1 and low CD62L expression (thin black gate) from WT or *Dnase113*<sup>-/-</sup> mice.

**(N, O)** Representative histograms show comparative surface expression profile (N) and bar graphs show mean fluorescence intensity (O) of CXCR4, CD40L or ICOS on total CD4<sup>+</sup> T cells (blue line), CD4<sup>+</sup> CD62L<sup>-</sup> PSGL-1<sup>lo</sup> ExFO Th cells (thick black line) and CD4<sup>+</sup> CD62L<sup>-</sup> PSGL-1<sup>hi</sup> T cells (thin black line), from splenocytes of 3-mo-old WT (open symbols) and *Dnase113*<sup>-/-</sup> mice (red symbols).

**(P)** Fraction of CD4<sup>+</sup> cells among total splenocytes and PSGL-1<sup>hi</sup> cells or PSGL-1<sup>lo</sup> cells among CD4<sup>+</sup> T cells from splenocytes of 3-4-mo-old WT (open symbols) or *Dnase113*<sup>-/-</sup> mice (red symbols). Symbols represent individual mice and bars indicate median. Significance was estimated using nonparametric Mann-Whitney test. Statistical significance: \*  $p \leq 0.05$ , \*\*  $p \leq 0.01$ , and \*\*\*\*  $p \leq 0.0001$ .



**Figure S4. Immune cell activation in *Dnase113*<sup>-/-</sup> mice is governed by type-I IFN signaling, Related to Figure 4.**

*Dnase113*<sup>-/-</sup> (red symbols), *Ifnar1*<sup>-/-</sup> (grey symbols) were examined along with *Dnase113*/*Ifnar1* double-deficient mice (*Dnase113*<sup>-/-</sup>*Ifnar1*<sup>-/-</sup>, dark blue symbols) or wild-type controls (WT, open symbols). In panels A-G, symbols represent individual mice and bars indicate median.

**(A and B)** Mean fluorescence intensity of IFN-inducible antigen Sca-1 on splenic TCRβ<sup>+</sup>CD4<sup>+</sup> T cells (A), and CD19<sup>+</sup>B220<sup>+</sup> B cells (B), as measured by flow cytometry in the splenocytes of 12-mo-old mice.

**(C)** Representative images from *Crithidia luciliae* immunofluorescence test (CLIFT). CL substrates were stained with a 1:25 dilution of serum samples from ≥ 4 mice per group. Scale bar: 20 μm.

**(D and E)** Fraction of CD38<sup>+</sup>GL-7<sup>+</sup> GC B cells (D) and IgD<sup>+</sup> IgM<sup>+</sup> IgG2a/2b<sup>+</sup> (switched) B cells (E), among CD19<sup>+</sup> B220<sup>+</sup> splenic B cells in the splenocytes of 12-mo-old mice, as determined by flow cytometry.

**(F)** Spleen weight of 12-mo-old mice.

**(G)** Fraction of CD62L<sup>-</sup>CD44<sup>hi</sup> T cells among splenic CD4<sup>+</sup> T cells in 12-mo-old mice as determined by flow cytometry in the splenocytes of 12-mo-old mice.

**(H)** Fraction of CD11c<sup>+</sup> CD11b<sup>+</sup> Ly6c<sup>-</sup> population among total PBMCs, in 12-mo-old mice as determined by flow cytometry.

One-way ANOVA followed by the Tukey multiple-comparison test was used for statistical comparison of more than two groups. Statistical significance: NS= not significant, \*  $p \leq 0.05$ , \*\*  $p \leq 0.01$ , \*\*\*  $p \leq 0.001$  and \*\*\*\*  $p \leq 0.0001$ .

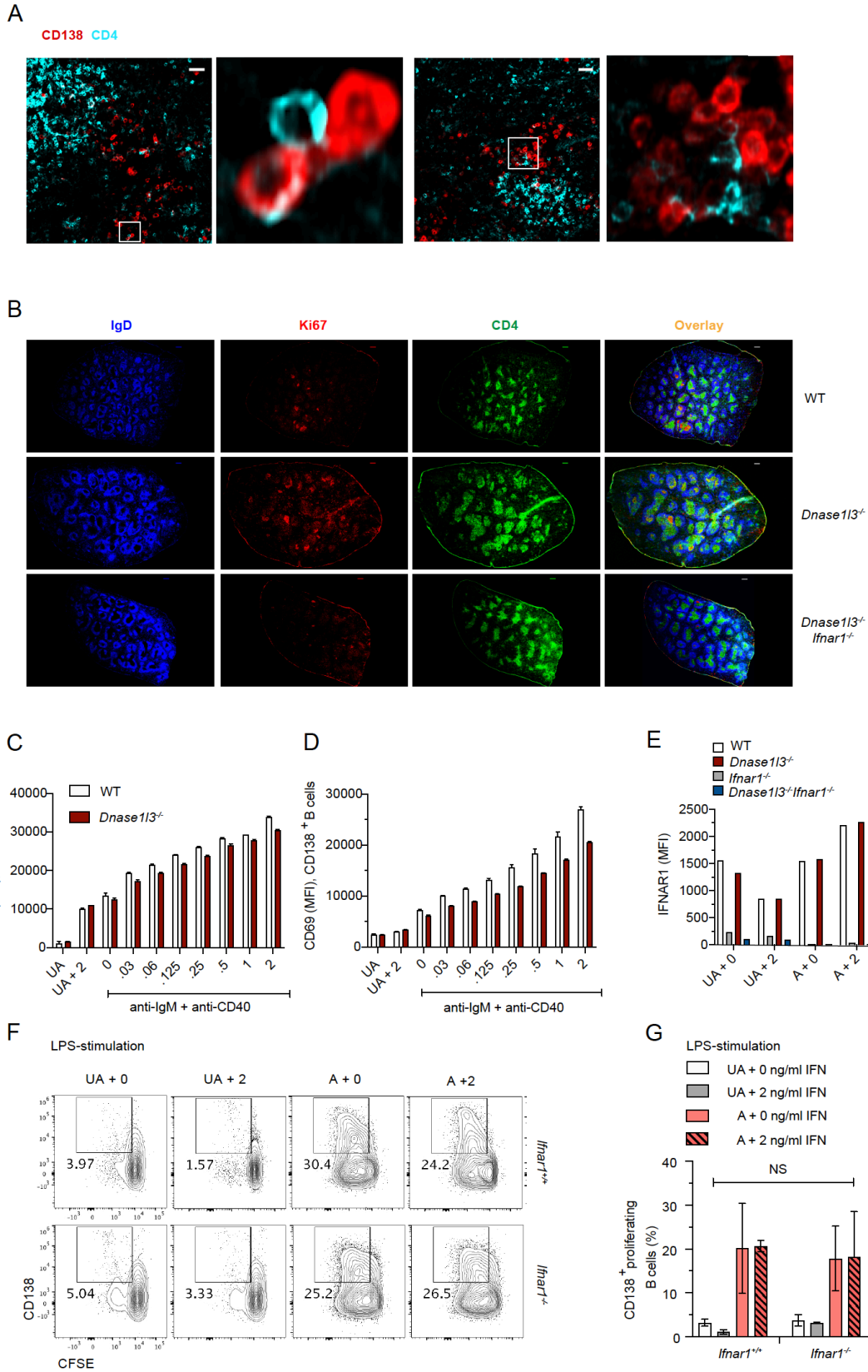


Figure S5. Role of IFN $\alpha$  in the differentiation of B cells into plasmablasts in vitro, Related to Figure 5.

**(A)** Representative confocal images of spleen sections from three, 8-9-mo-old *Dnase113*<sup>-/-</sup> mice showing the distribution of CD4<sup>+</sup> T cells near CD138<sup>+</sup> cell clusters. White insets highlight the close interactions of CD4<sup>+</sup> T cells with CD138<sup>+</sup> cells, as shown in magnified images in adjacent panels. Scale bars, 40μm

**(B)** In situ analysis of proliferating cells in the spleens of WT, *Dnase113*<sup>-/-</sup> and *Dnase113*<sup>-/-</sup>*Ifnar1*<sup>-/-</sup> mice by confocal microscopy. Shown are representative stitched images of spleen sections from at least three >8-mo-old mice of the indicated strains, stained for IgD<sup>+</sup> naïve follicular B cells (blue), Ki67<sup>+</sup> proliferating cells (Red), and CD4<sup>+</sup> T cell zones (green). Right most panels show overlay. Scale bars, 150μm.

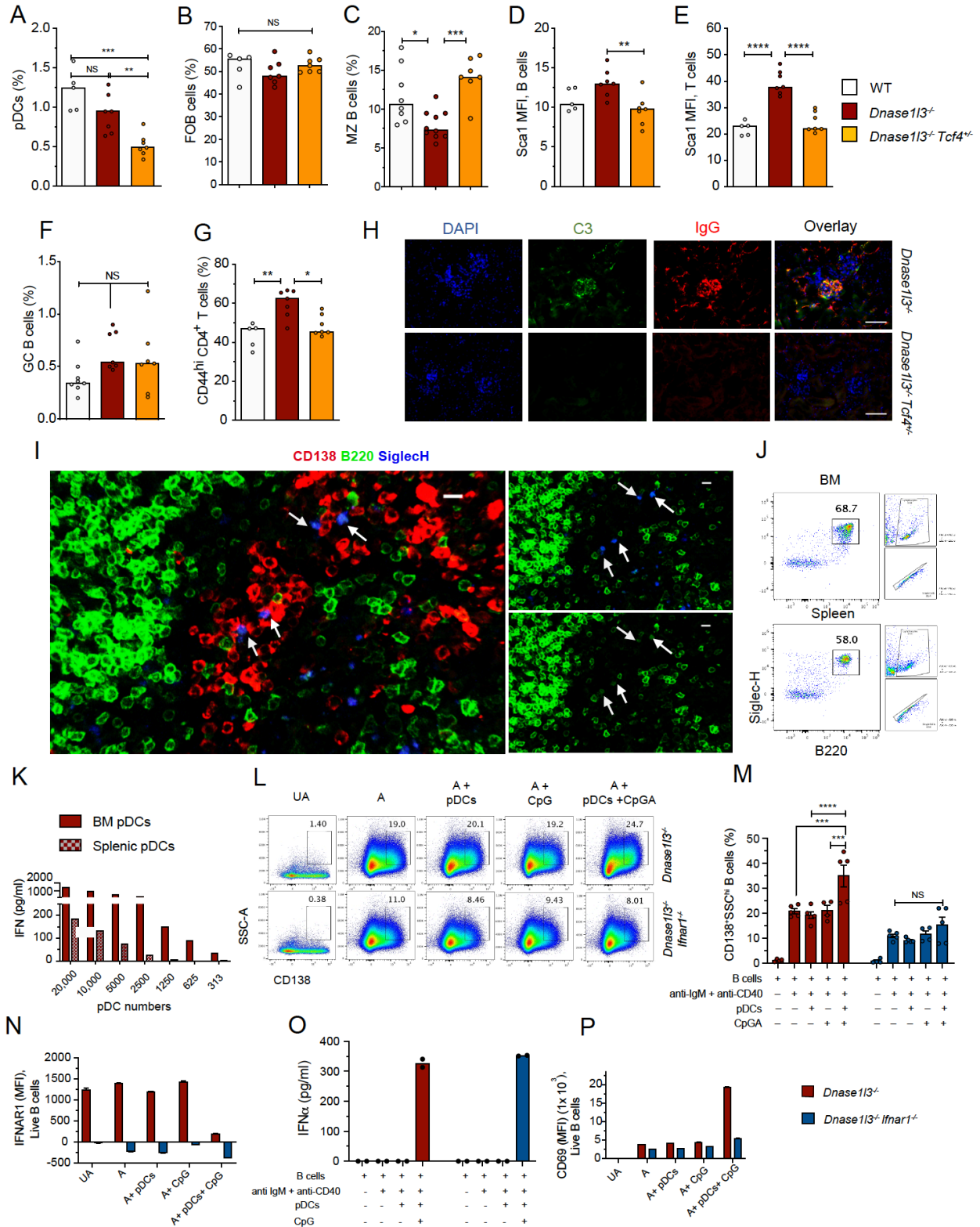
**(C and D)** Analysis of in-vitro activated B cells supplemented with increasing concentration of IFNα. Mean fluorescence intensity of IFN-inducible markers Sca-1 (A) and CD69 (B), on purified B cells from WT (open bars) or *Dnase113*<sup>-/-</sup> (red bars) mice, left unactivated (UA), or cultured with 2 ng/ml IFNα (UA+2) or activated with anti-IgM and anti-CD40 for 72h in the presence increasing concentration of IFNα (0-2 ng/ml), as determined by flow cytometry. Data are representative of two independent experiments with each treatment in duplicate.

**(E)** Mean fluorescence intensity (MFI) of *Ifnar* on B cells from WT (open bars), *Ifnar1*<sup>-/-</sup> (grey bars), *Dnase113*<sup>-/-</sup> (red bars) and *Dnase113*<sup>-/-</sup>*Ifnar1*<sup>-/-</sup> (blue bars) mice, left unactivated (UA) or activated with anti-IgM + anti-CD40 (A) for 72h in the absence (0) or presence of 2 ng/ml of IFNα, measured by flow cytometry. Data are representative of five experiments.

**(F)** Analysis of proliferation and differentiation of naïve B cells upon in-vitro activation with 1 μg/ml LPS in the presence or absence of IFNα. Shown are representative flow plots of purified CFSE labeled unactivated (UA) or LPS activated (A) B cells from IFNAR-sufficient (*Ifnar1*<sup>+/+</sup>) or IFNAR-deficient (*Ifnar1*<sup>-/-</sup>) mice cultured for 72h in the absence (0 ng/ml) or presence of 2 ng/ml of IFNα. The cells in the gate indicate CFSE<sup>lo</sup>CD138<sup>+</sup> proliferating B cells, under the indicated conditions.

**(G)** Quantification of the percentage of CD138<sup>+</sup> proliferating *Ifnar1*<sup>+/+</sup> or *Ifnar1*<sup>-/-</sup> B cells 72h after in-vitro activation with 1 μg/ml LPS in the presence or absence of IFNα. Error bars show mean ± SD. Data are representative of two independent experiments. 2-way ANOVA followed by the Tukey multiple-comparison test was used to compare different treatments within a genotype or 2-way ANOVA followed by Sidak's multiple comparison test was used to compare different treatments between the two genotypes. Statistical significance: NS= not significant.





**Figure S6. IFN $\alpha$  produced by pDCs promotes differentiation of B cells into plasmablasts, Related to Figure 6.** *Dnase113*<sup>-/-</sup> (red symbols) were examined along with *Dnase113*<sup>-/-</sup> mice with monoallelic deficiency of TCF4 (*Dnase113*<sup>-/-</sup>.*Tcf4*<sup>+/-</sup>, orange symbols) and wild-type controls (WT, open symbols). In panels A- G, symbols represent individual mice and bars indicate median.

**(A)** Fraction of CD11c<sup>int</sup>Siglec-H<sup>+</sup> pDCs among total splenocytes in 12-mo-old mice, determined by flow cytometry.

**(B and C)** Fraction of TCR $\beta$ -CD19<sup>+</sup>CD23<sup>hi</sup>CD21<sup>-</sup> FO B cells (B) and TCR $\beta$ -CD19<sup>+</sup> CD23<sup>lo</sup>CD21<sup>hi</sup> MZ B cells (C), among splenic single lymphocytes from 12-mo-old mice as determined by flow cytometry.

**(D and E)** Mean fluorescence intensity of IFN- $\gamma$  inducible antigen Sca-1 on total splenic CD19<sup>+</sup>B220<sup>+</sup> B cells (D) and TCR $\beta$ <sup>+</sup> T cells (E), as measured by flow cytometry.

**(F and G)** Fraction of CD38<sup>+</sup>GL-7<sup>+</sup> GC B cells among CD19<sup>+</sup>B220<sup>+</sup> splenic B cells (F), and fraction of CD62L<sup>-</sup>CD44<sup>hi</sup> effector T cells among splenic CD4<sup>+</sup> T cells (G) in the splenocytes of 12-mo-old mice as determined by flow cytometry. One-way ANOVA followed by the Tukey multiple-comparison test was used for statistical comparison of more than two groups. GraphPad Prism 8 software (La Jolla, CA) was used for all the analyses. Statistical significance: NS= not significant, \*  $p \leq 0.05$ , \*\*  $p \leq 0.01$ , \*\*\*  $p \leq 0.001$  and \*\*\*\*  $p \leq 0.0001$ .

**(H)** Representative immunofluorescence images from kidney sections stained for glomerular immune complexes (IgG, red) and C3 (green). Glomeruli identified as nuclei dense (DAPI<sup>+</sup>, blue) areas in the cortical region of the kidney. Kidneys from at least 3-4, 12-mo-old mice per strain were analyzed. Right most panels show overlay. Scale bars, 50  $\mu$ m.

**(I)** In situ analysis of pDCs in the spleens of *Dnase1/3*<sup>-/-</sup> mice by confocal microscopy. Shown is a representative image of a spleen section from at least three >8-mo-old *Dnase1/3*<sup>-/-</sup> mice stained for CD138<sup>+</sup> plasmablasts (red), B220<sup>+</sup> (green) and Siglec-H<sup>+</sup> (blue) pDCs. Left panel shows the overlay staining with all antibodies while the right side upper and lower panels show B220<sup>lo</sup> and SiglecH<sup>+</sup> pDCs. White arrows indicate the specific pDCs (SiglecH<sup>+</sup> B220<sup>lo</sup>) in close contact with CD138<sup>+</sup> plasmablasts in the red pulp. Scale bar, 20  $\mu$ m.

**(J)** Assessment of Siglec-H<sup>+</sup>B220<sup>lo</sup> pDC purity by flow cytometry after isolation from bone marrow (BM) cells or splenocytes (Spleen) by negative selection using magnetic-activated cell sorting.

**(K)** Purified BM pDCs (red bars) or splenic pDCs (checkered bars) were cultured at the indicated cell numbers along with 1 $\mu$ M CpGA for 18h and the concentration IFN $\alpha$  secreted in respective culture supernatants were measured by ELISA.

**(L-P)** Purified CFSE labeled B cells from *Dnase1/3*<sup>-/-</sup> (red) or *Dnase1/3*<sup>-/-</sup>*Ifnar1*<sup>-/-</sup> (dark blue) mice: left unactivated (UA); activated with anti-IgM+ anti-CD40 (A); activated with anti-IgM+ anti-CD40 in the presence of unstimulated pDCs (A+ pDCs); Or 1 $\mu$ M CpGA (A+ CpGA) or in the presence of pDCs stimulated with 1 $\mu$ M CpGA (A+ pDCs+ CpGA) for 72h.

**(L)** Representative flow plots show gating strategy for blasting SSC<sup>hi</sup> B cells positive for CD138.

**(M)** Quantification of the fraction of CD138<sup>+</sup>SSC<sup>hi</sup> blasting B cells. Error bars show mean  $\pm$  SD from at least four independent experiments. For statistical analysis of grouped data, two-way ANOVA followed by Tukey's multiple-comparison test was used. Statistical significance: NS= not significant, \*\*\* $p \leq 0.001$  and \*\*\*\*  $p \leq 0.0001$ .

**(N)** Representative experiment showing mean fluorescence intensity of *Ifnar1* on B cells by flow cytometry after the indicated treatments.

**(O)** Representative experiment showing concentration of IFN $\alpha$  in the culture supernatants of indicated co-cultures as determined by ELISA.

**(P)** Representative experiment showing mean fluorescence intensity of CD69 on B cells by flow cytometry.

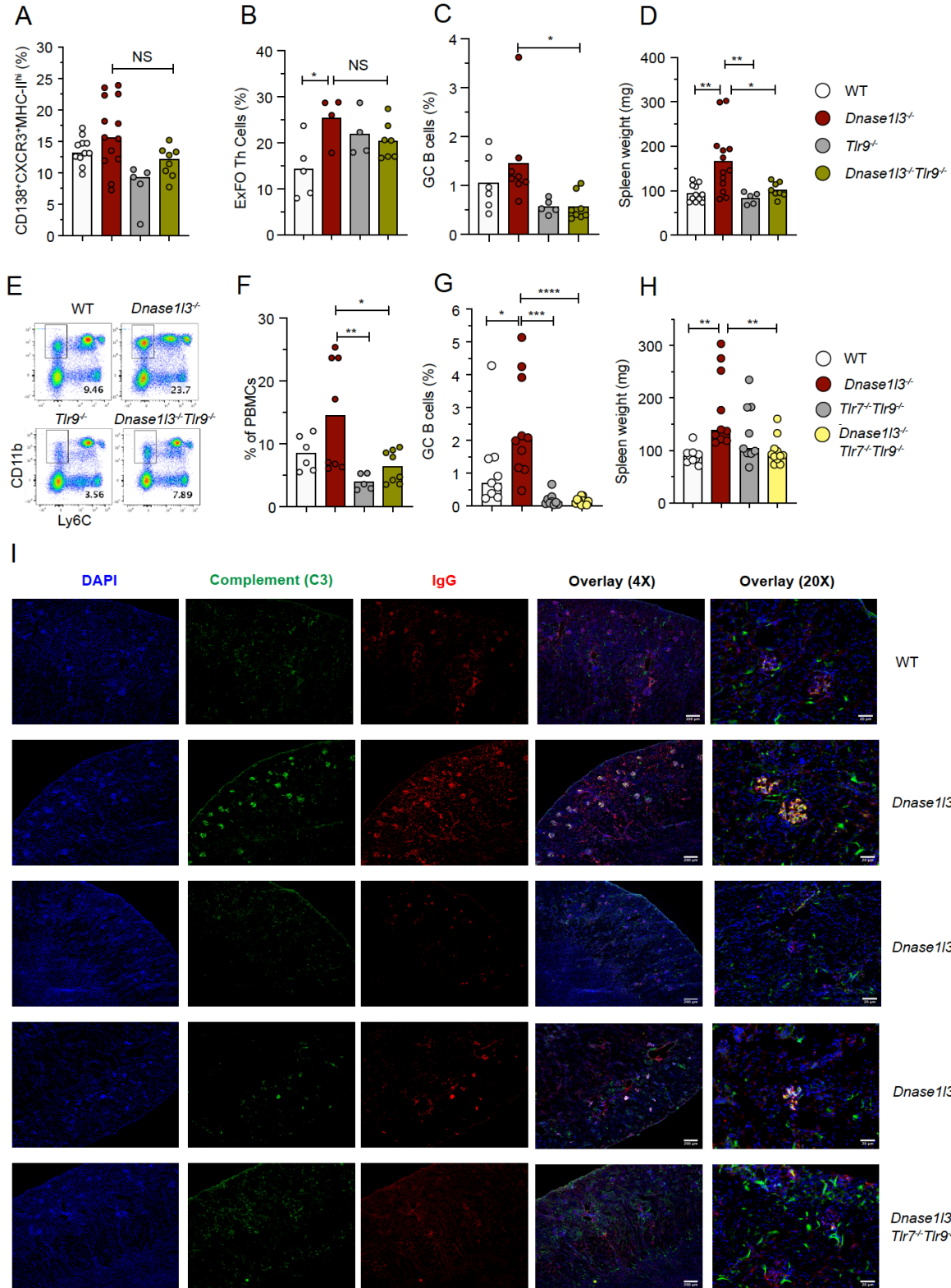


Figure S7. Contribution of TLR9 and TLR7 to autoimmune manifestations of *Dnase113*<sup>-/-</sup> mice, Related to Figure 7.

In panels A-F, 12-mo-old WT control mice (open symbols) *Dnase1l3*<sup>-/-</sup> (red symbols) were examined along with TLR9-deficient mice (*Tlr9*<sup>-/-</sup>, grey symbols) and *Dnase1l3*<sup>-/-</sup> mice with TLR9 deficiency (*Dnase1l3*<sup>-/-</sup>*Tlr9*<sup>-/-</sup>, olive-green symbols).

**(A)** Fraction of CXCR3<sup>+</sup>MHC-II<sup>hi</sup> cells among CD138<sup>+</sup> B cell, as determined by flow cytometry.

**(B)** Fraction of CD62L<sup>+</sup>PSGL-1<sup>lo</sup> cells among CD4<sup>+</sup> T cells, as determined by flow cytometry.

**(C)** Fraction of CD38<sup>+</sup>GL-7<sup>+</sup> cells among CD19<sup>+</sup> B cells, as determined by flow cytometry.

**(D)** Comparative analysis of spleen weights of 12-mo-old mice from the indicated strains.

**(E)** Representative flow plots of PBMCs from 12-mo-old indicated mice with CD11b<sup>+</sup>Ly6c<sup>-</sup> myeloid cells highlighted within the gate.

**(F)** Fractions of CD11b<sup>+</sup>Ly6c<sup>-</sup> population among total PBMCs, as determined by flow cytometry.

**(G-H)** Fraction of CD38<sup>+</sup>GL-7<sup>+</sup> cells among CD19<sup>+</sup> B cells (G) and comparative analysis of spleen weights (H), of 12-mo-old WT control (open symbols), *Dnase1l3*<sup>-/-</sup> (red symbols), dual Tlr7 and Tlr9-deficient (*Tlr7*<sup>-/-</sup>*Tlr9*<sup>-/-</sup>, grey symbols) and *Dnase1l3*, Tlr7 and Tlr9 triple deficient (*Dnase1l3*<sup>-/-</sup>*Tlr7*<sup>-/-</sup>*Tlr9*<sup>-/-</sup>, yellow symbols) mice.

One-way ANOVA followed by the Tukey multiple-comparison test was used for statistical comparison of more than two groups. GraphPad Prism 8 software (La Jolla, CA) was used for all the analyses. Statistical significance: NS= not significant, \*  $p \leq 0.05$ , \*\*  $p \leq 0.01$ , \*\*\*  $p \leq 0.001$  and \*\*\*\*  $p \leq 0.0001$ .

**(I)** Representative immunofluorescence images from kidney sections stained for glomerular immune complexes IgG (red), and C3 (green). Glomeruli identified as nuclei dense (DAPI<sup>+</sup>, blue) areas in the cortical region of the kidney. Kidneys from at least four, 12-mo-old mice per strain were analyzed. Right most panels show overlay at 4X magnification of the cortical region of the kidney (scale bar: 200  $\mu\text{m}$ ), and overlay at 20X magnification highlighting individual glomeruli (scale bars, 20 $\mu\text{m}$ ).