

Supporting Information for

**Multi-site SUMOylation restrains DNA Polymerase  $\eta$  interactions with DNA damage sites**

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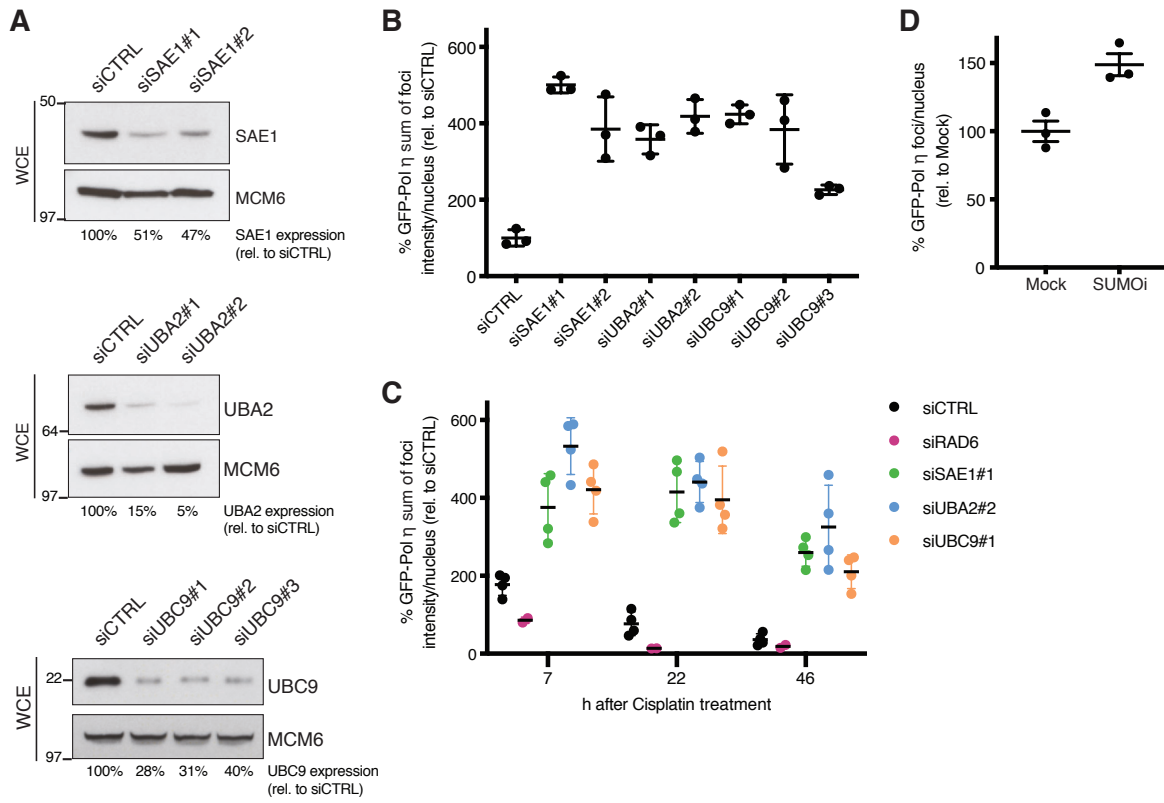
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**Table S1.****Results of high-throughput microscopy screen for regulators of Pol  $\eta$  interaction with DNA damage sites**

U2OS/GFP-Pol  $\eta$  cells were transfected with an siRNA library targeting ubiquitin and UBL signaling factors (1251 genes, two siRNAs per gene). After 72 h, cells were treated with 30  $\mu$ M Cisplatin for 6 h and subsequently grown overnight in fresh media. Cells were then pre-extracted, fixed and stained with DAPI. GFP-Pol  $\eta$  foci counts were determined using QIBC. Table shows modified Z-scores for all siRNAs in the library.

**Table S2.****List of validated Ub/UBL signaling factors whose knockdown leads to enhanced formation of DNA damage-induced Pol  $\eta$  foci****Table S3.****Overview of endogenous Pol  $\eta$  SUMOylation sites identified by mass spectrometry****Figure S1.****SUMO-dependent regulation of Pol  $\eta$  association with DNA damage sites.**

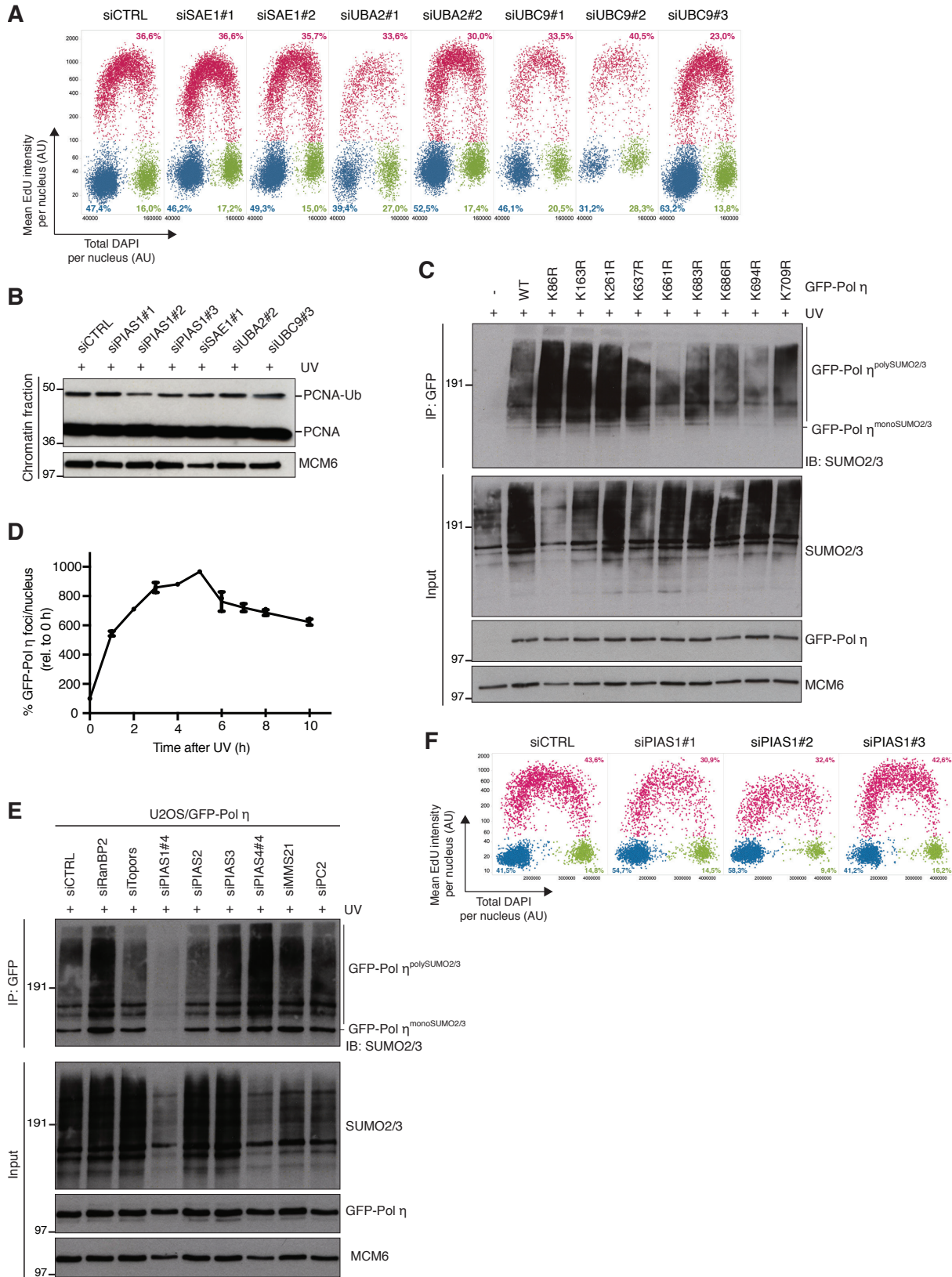
**A.** Immunoblot analysis of whole-cell extracts (WCE) of U2OS cells transfected with non-targeting control (CTRL) siRNA or siRNAs targeting indicated SUMO enzymes. Expression levels of SAE1, UBA2 and UBC9 (normalized to MCM6 expression) were quantified using ImageJ. **B.** Results of validation screen. The sum of GFP-Pol  $\eta$  foci intensity in U2OS/GFP-Pol  $\eta$  cells transfected with indicated siRNAs, exposed to cisplatin for 6 h and fixed 16 h later was quantified using QIBC analysis (mean $\pm$ SD;  $\geq$ 294 cells quantified per condition). **C.** Results of validation screen. The sum of GFP-Pol  $\eta$  foci intensity at different time points after exposure to cisplatin in cells treated as in (B) was quantified using QIBC analysis (mean $\pm$ SD;  $\geq$ 3,000 cells quantified per condition). **D.** U2OS/GFP-Pol  $\eta$  cells were treated or not with SUMO E1 inhibitor (SUMOi) for 30 min, exposed to UV and fixed 6 h later. GFP-Pol  $\eta$  foci count per nucleus was quantified by QIBC analysis (mean $\pm$ SEM;  $n=3$  independent experiments;  $\geq$ 7,482 cells quantified per condition).



**Figure S2.**

**Impact of removing SUMO modification components on cell cycle distribution, PCNA monoubiquitylation and Pol  $\eta$  SUMOylation**

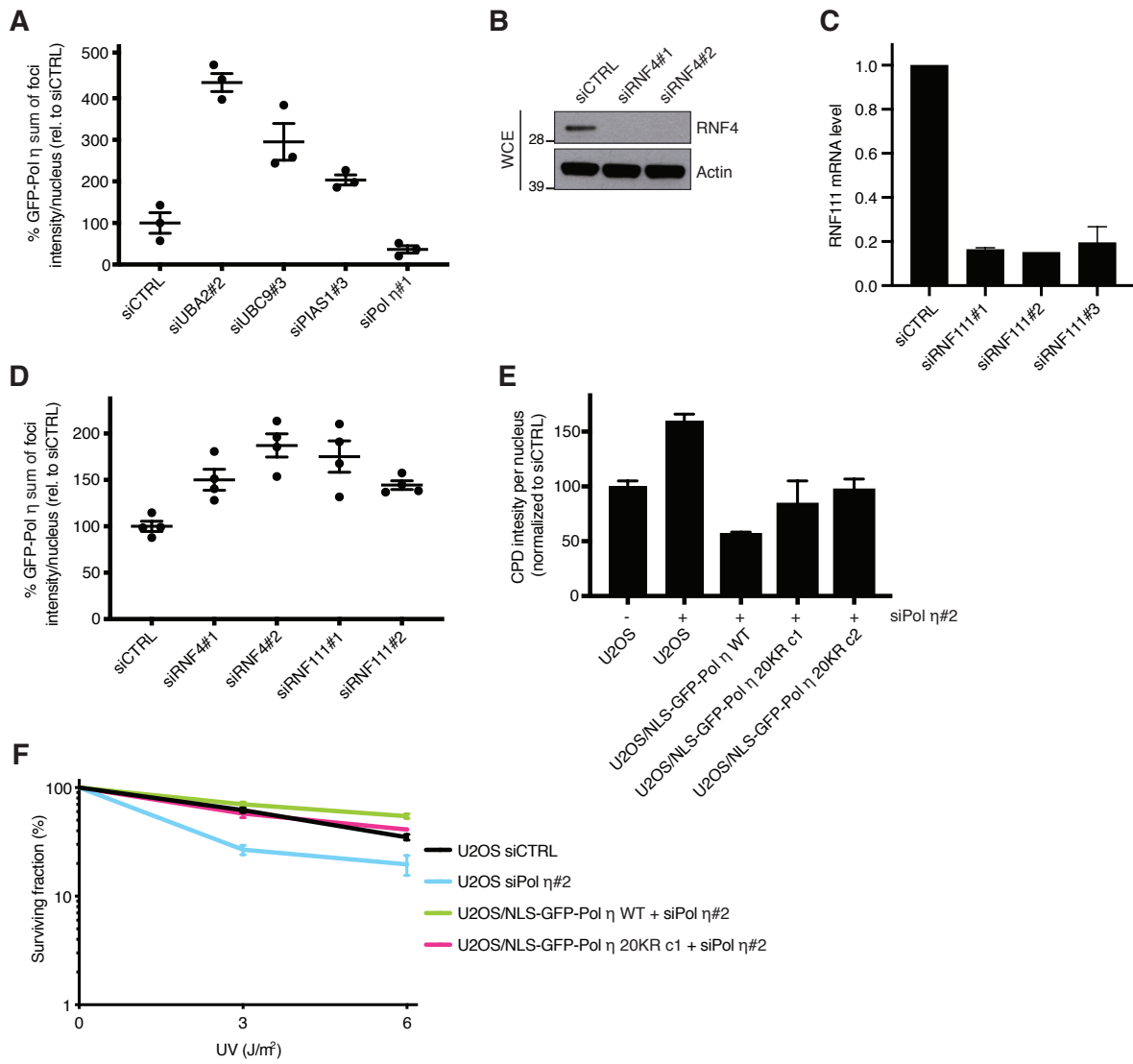
**A.** U2OS cells transfected with indicated siRNAs were pulse-labeled with EdU, fixed and processed for EdU labeling and staining of total DNA with DAPI. Cell cycle profiles were analyzed using QIBC ( $\geq 1869$  cells quantified per condition). Percentage of cells in different cell cycle stages (G1 phase, blue; S phase, pink; G2/M phase, green) is indicated. **B.** Immunoblot analysis of chromatin-enriched fractions of U2OS cells transfected with indicated siRNAs and exposed to UV. **C.** U2OS left untreated or transfected with plasmids encoding the indicated GFP-Pol  $\eta$  alleles and exposed to UV were lysed and subjected to GFP immunoprecipitation (IP) under denaturing conditions, followed by immunoblotting with SUMO2/3, GFP and MCM6 (loading control) antibodies. **D.** Quantification of GFP-Pol  $\eta$  foci formation in U2OS/GFP-Pol  $\eta$  collected at indicated times after exposure to UV (mean $\pm$ SEM;  $n=2$  independent experiments;  $\geq 508$  cells quantified per condition). **E.** U2OS/GFP-Pol  $\eta$  cells transfected with indicated siRNAs and exposed to UV were lysed and subjected to GFP immunoprecipitation (IP) under denaturing conditions, followed by immunoblotting with SUMO2/3, GFP and MCM6 (loading control) antibodies. **F.** Cell cycle profiles of cells transfected with indicated PIAS1 siRNAs or non-targeting control (CTRL) siRNA were determined as in (A) ( $\geq 2936$  cells quantified per condition).



### Figure S3.

#### Impact of impaired SUMO-dependent processing of Pol $\eta$ on its association with DNA damage sites and TLS functionality

**A.** Quantification of the sum of GFP-Pol  $\eta$  foci intensity using QIBC analysis in U2OS/GFP-Pol  $\eta$  cells transfected with indicated siRNAs, exposed to UV and fixed 6 h later (mean $\pm$ SEM;  $n=2$  independent experiments;  $\geq 1991$  cells quantified per condition). **B.** Immunoblot analysis of U2OS cells transfected with non-targeting control (CTRL) or RNF4 siRNAs using indicated antibodies. **C.** qPCR analysis of RNF111 mRNA levels in U2OS cells transfected with indicated siRNAs relative to siCTRL. Primers to GAPDH were used as a normalization control (mean $\pm$ SEM;  $n=2$  independent experiments). **D.** Quantification of the sum of GFP-Pol  $\eta$  foci intensity using QIBC analysis in U2OS/GFP-Pol  $\eta$  cells transfected with indicated siRNAs and treated as in (A) (mean $\pm$ SEM;  $n=4$  independent experiments;  $\geq 1254$  cells quantified per condition). **E.** U2OS cell lines stably expressing GFP-Pol  $\eta$  WT or 20KR mutant (clone 1 (c1) and clone 2 (c2)) were transfected or not with siRNA targeting the 3'UTR of endogenous Pol  $\eta$  (siPol  $\eta$ #2), exposed to UV (5 J/m<sup>2</sup>) and fixed 6 h later. Cells were then stained with antibody specific to cyclobutane pyrimidine dimers (CPDs) under native conditions, where only CPDs in single-stranded DNA (ssDNA) are exposed, and CPD intensity per nucleus was quantified by QIBC analysis (mean $\pm$ SEM;  $n=2$  independent experiments;  $\geq 10,086$  cells quantified per condition). Both GFP-Pol  $\eta$  WT or 20KR are able to rescue the increased occurrence of CPDs in ssDNA resulting from knockdown of endogenous Pol  $\eta$ , indicating that both alleles are proficient for TLS-mediated bypass of these lesions. **F.** Clonogenic survival of U2OS cells and derivative stable U2OS/NLS-GFP-Pol  $\eta$  WT and U2OS/NLS-GFP-Pol  $\eta$  20KR mutant (c1) cell lines transfected with siCTRL or siPol  $\eta$ #2 and exposed to the indicated doses of UV. After UV radiation the medium was supplemented with 0,4 mM caffeine (mean $\pm$ SEM;  $n=2$  experiments).

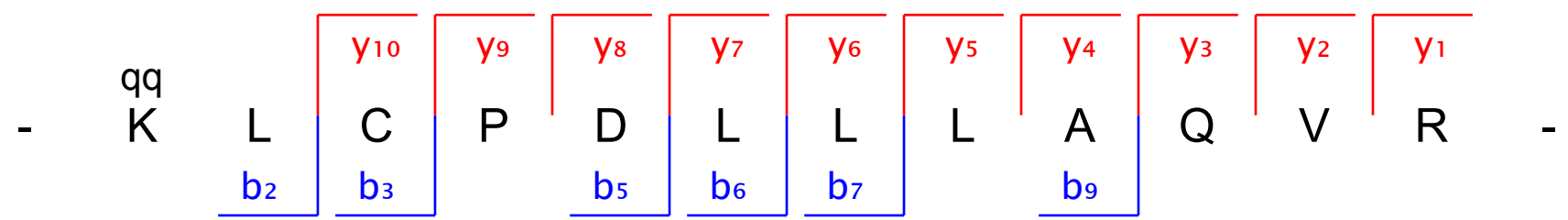
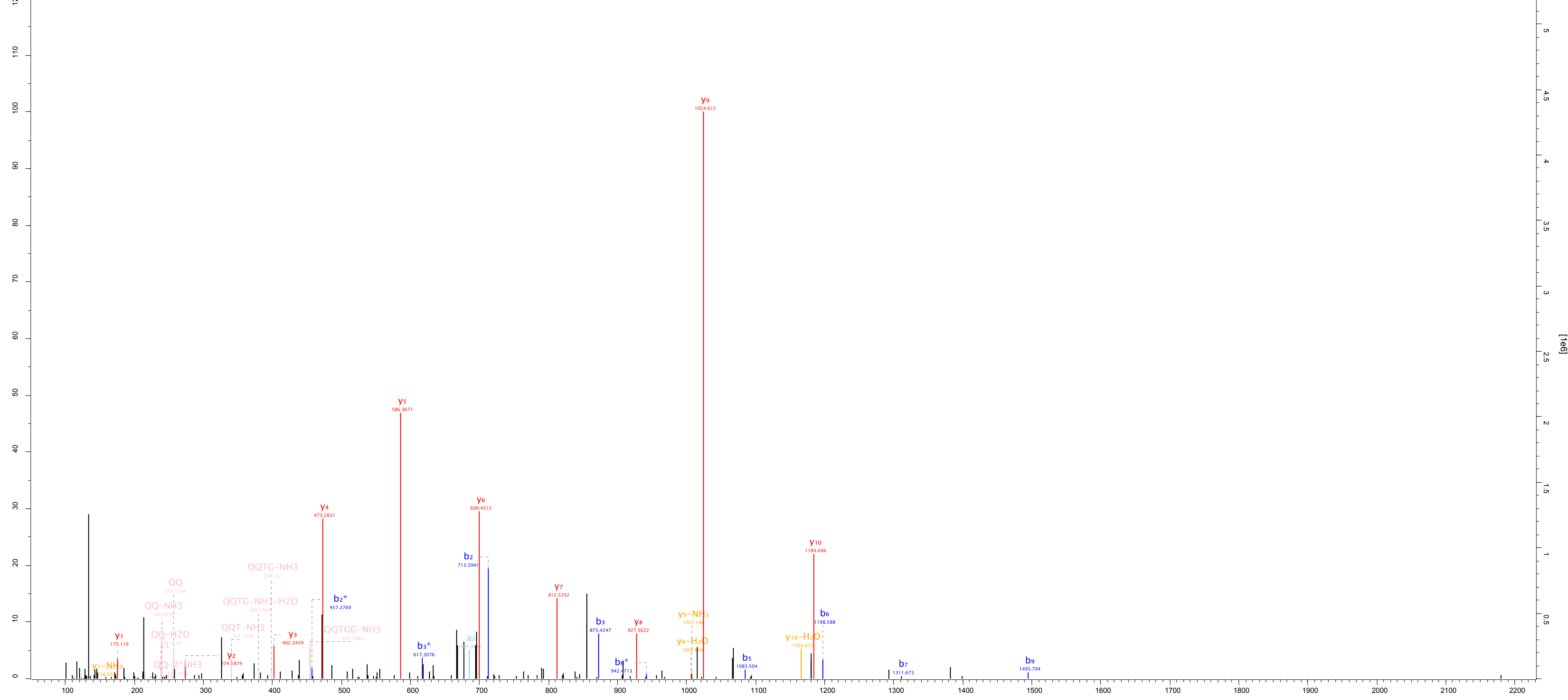


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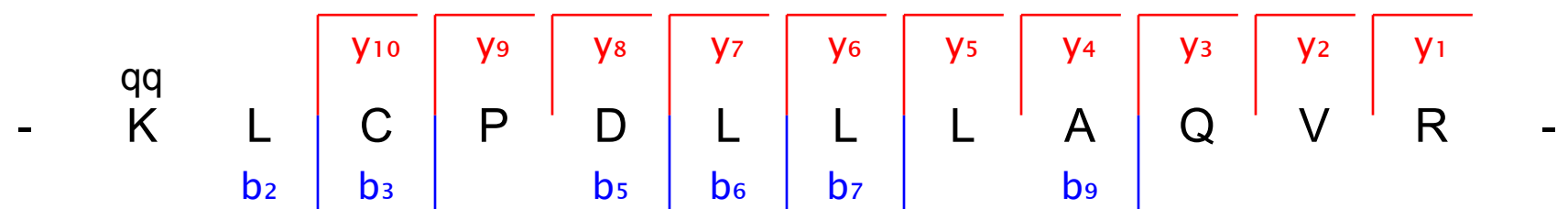
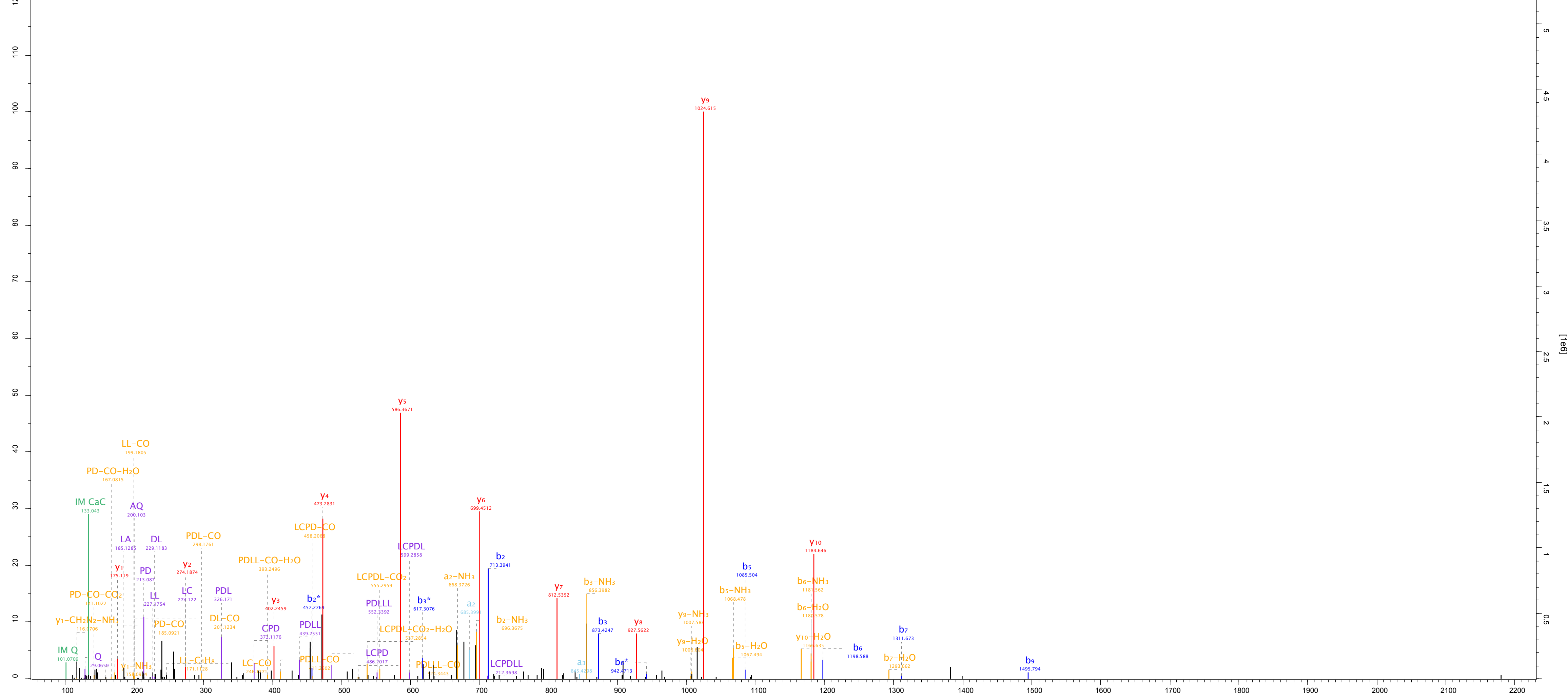
**MS spectra for identified SUMO sites in human Pol  $\eta$**

A collection of 19 fully annotated MS/MS spectra corresponding to all POLH SUMO sites reported in this work. Both “Standard” and “Advanced” annotations via the built-in MaxQuant Viewer software are available for each spectrum. Note that “qq” in the peptide sequence corresponds to attachment of the SUMO C-terminal mass remnant, QQTGG, covalently linked to the lysine residue via its C-terminal glycine residue.

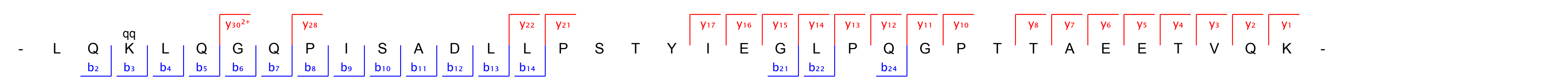
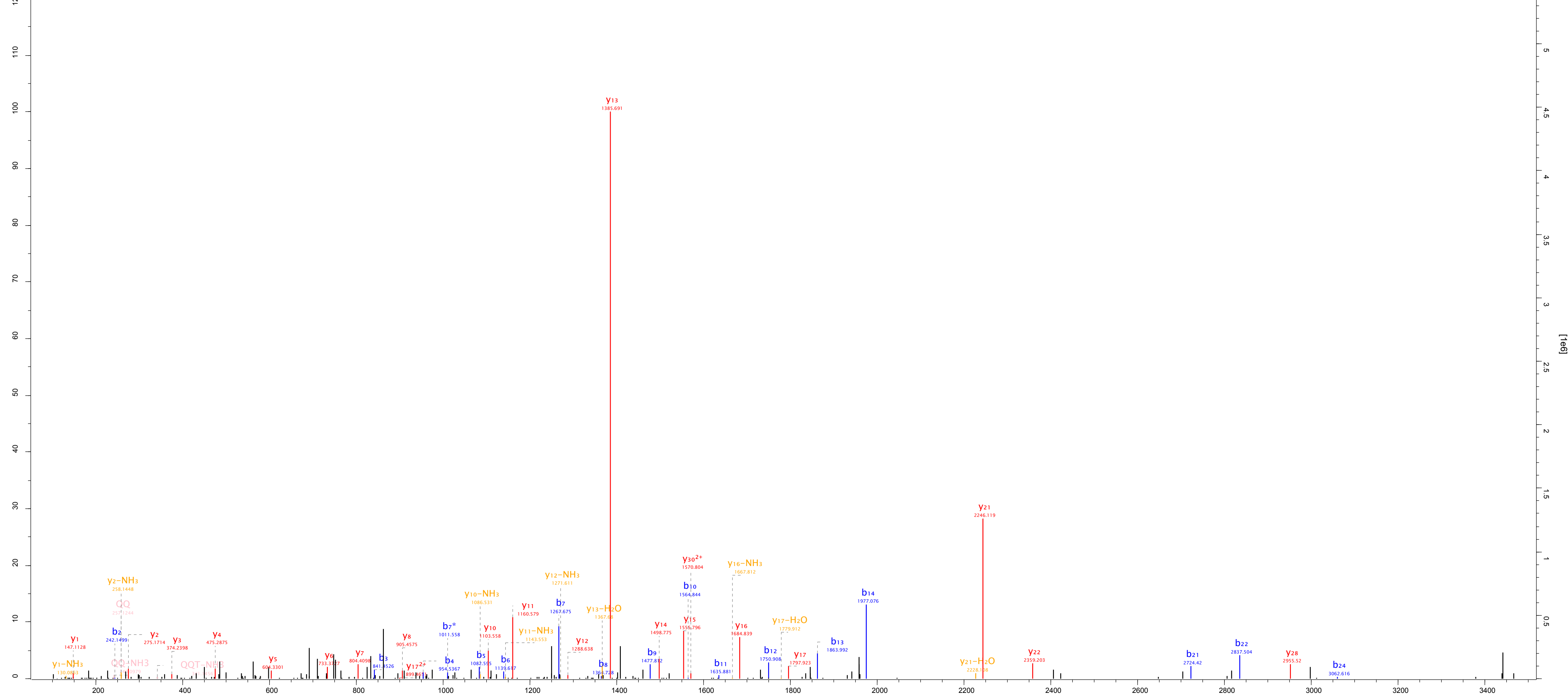




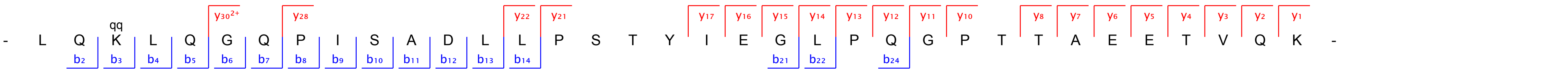
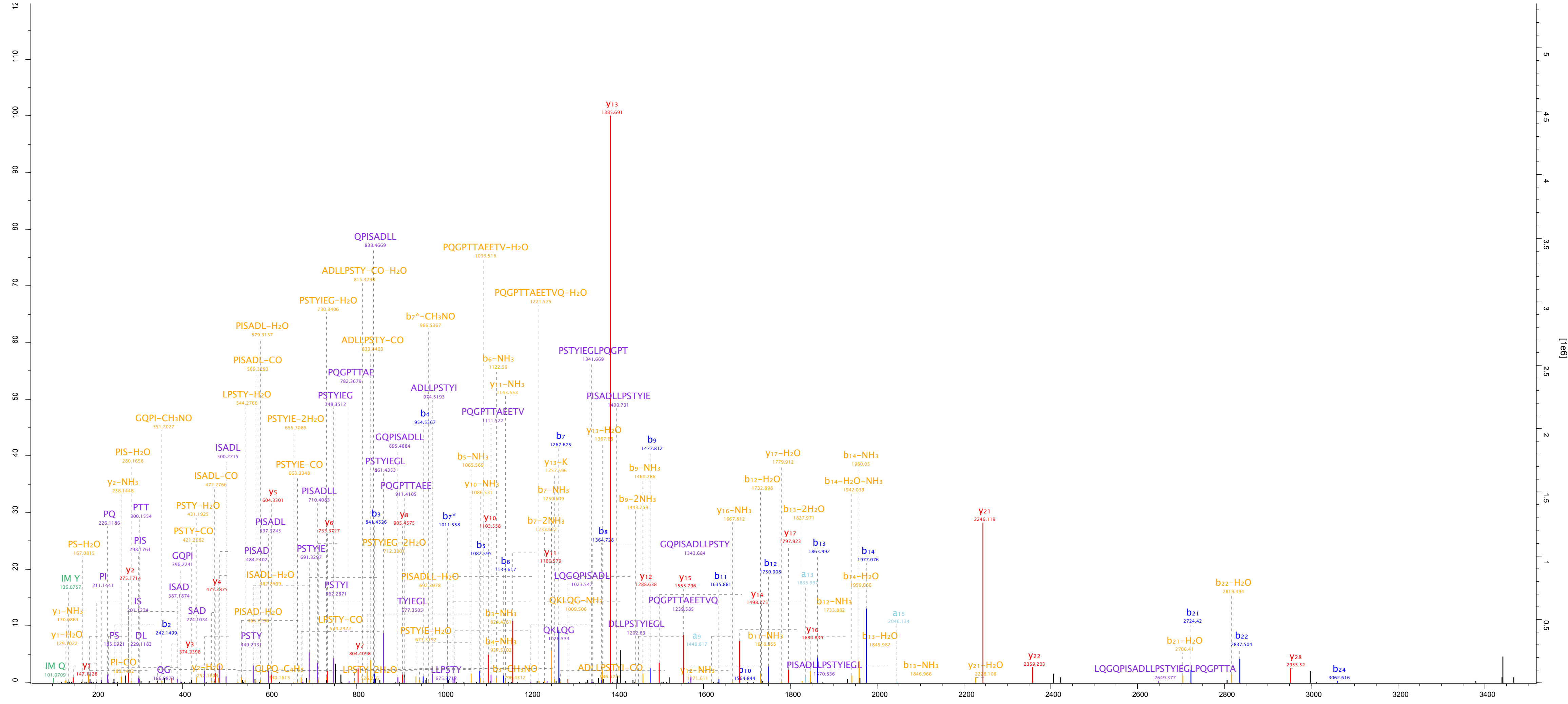
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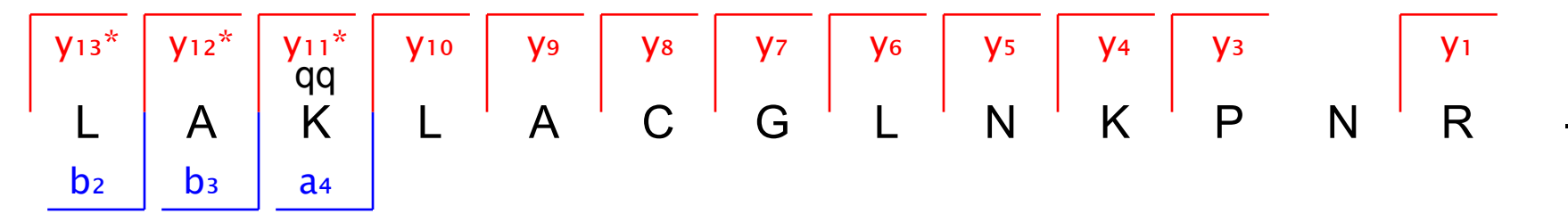
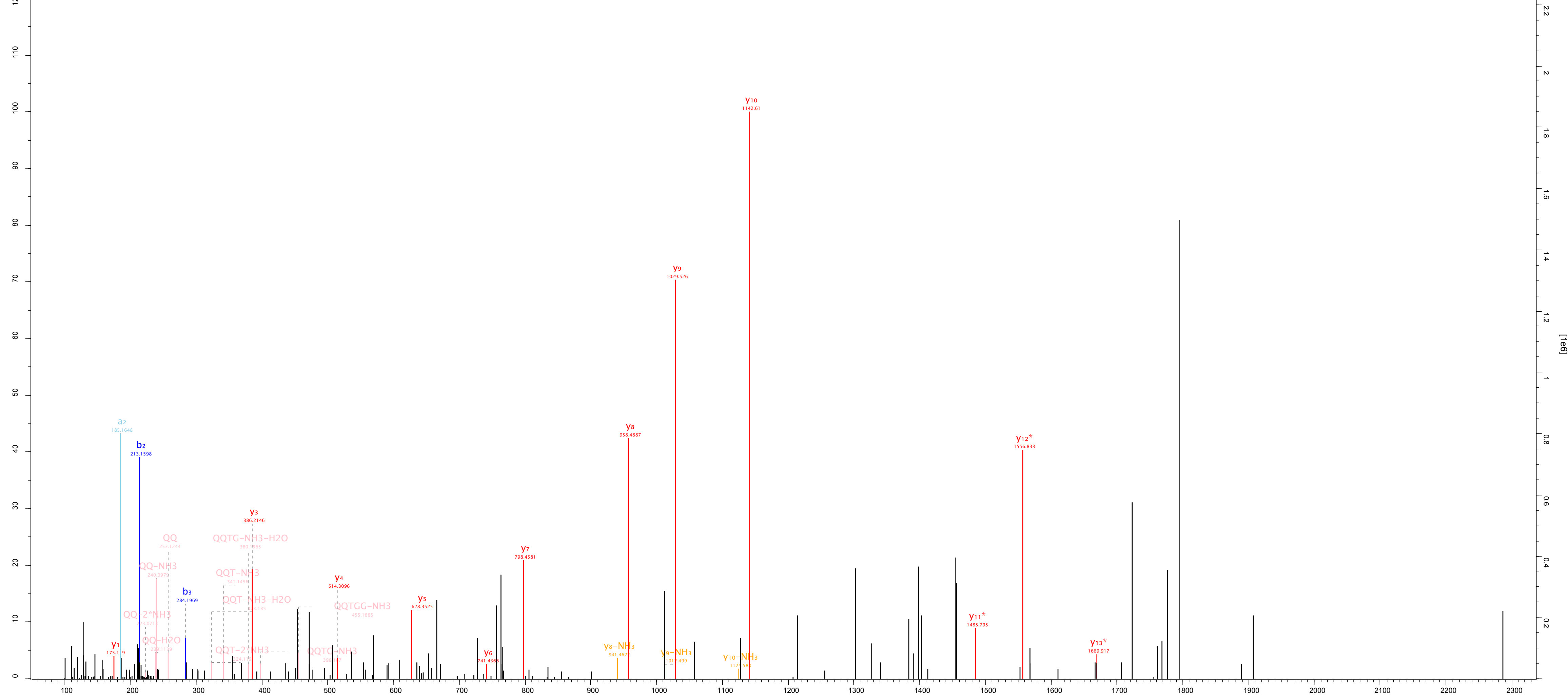
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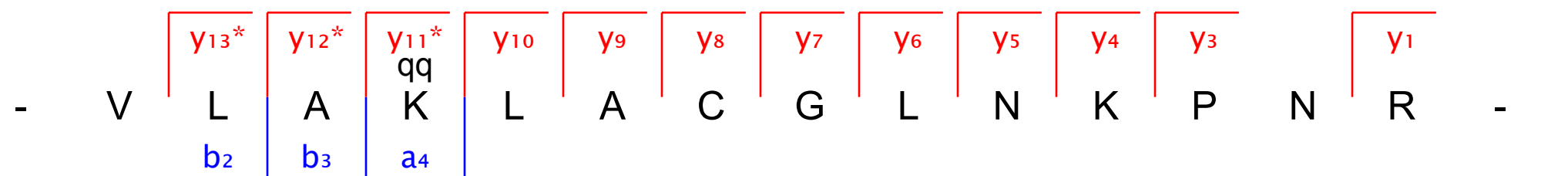
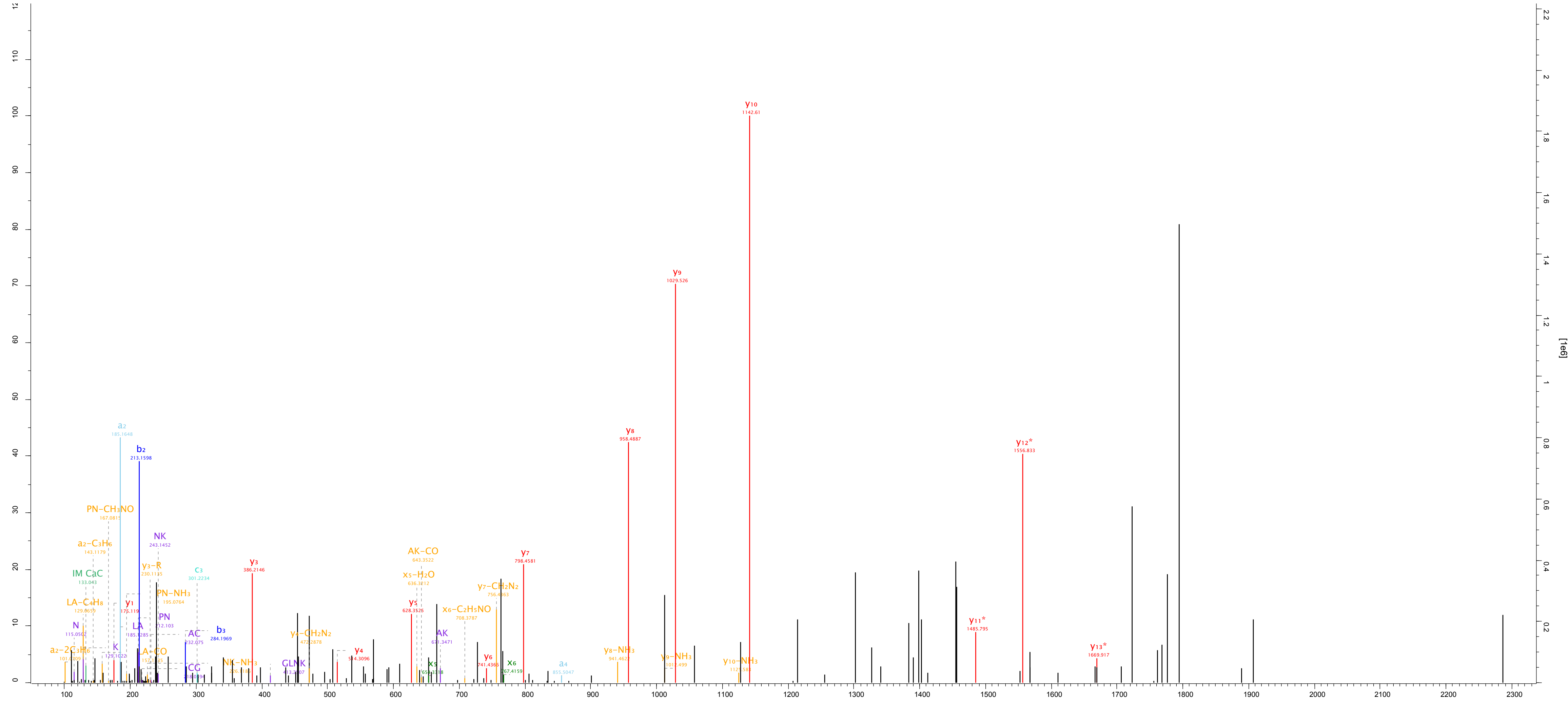
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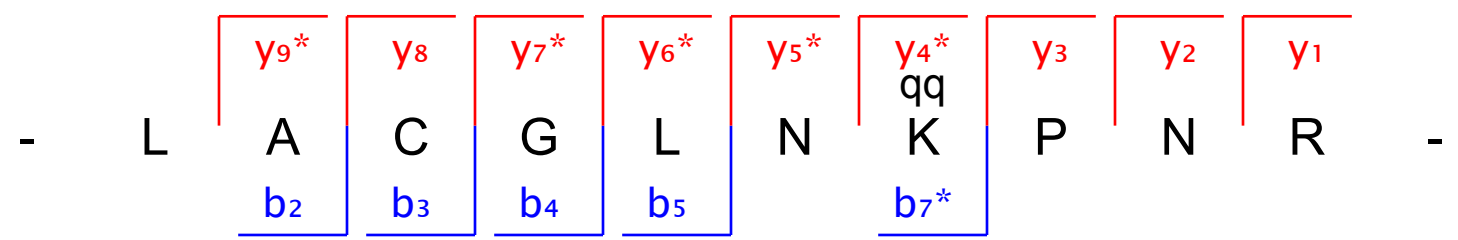
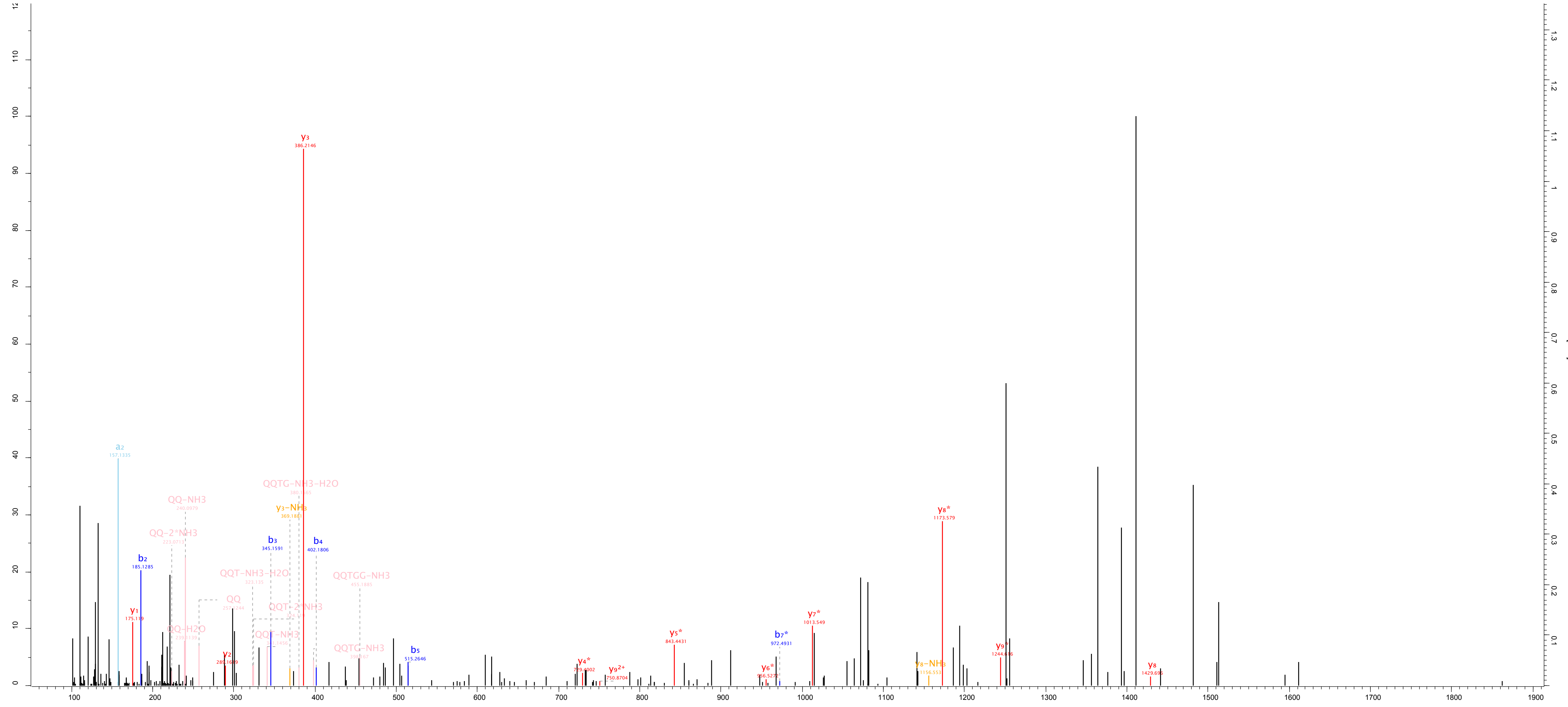
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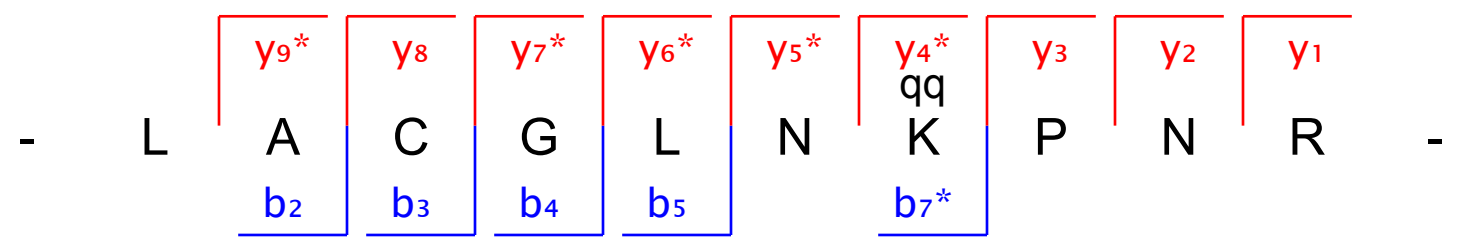
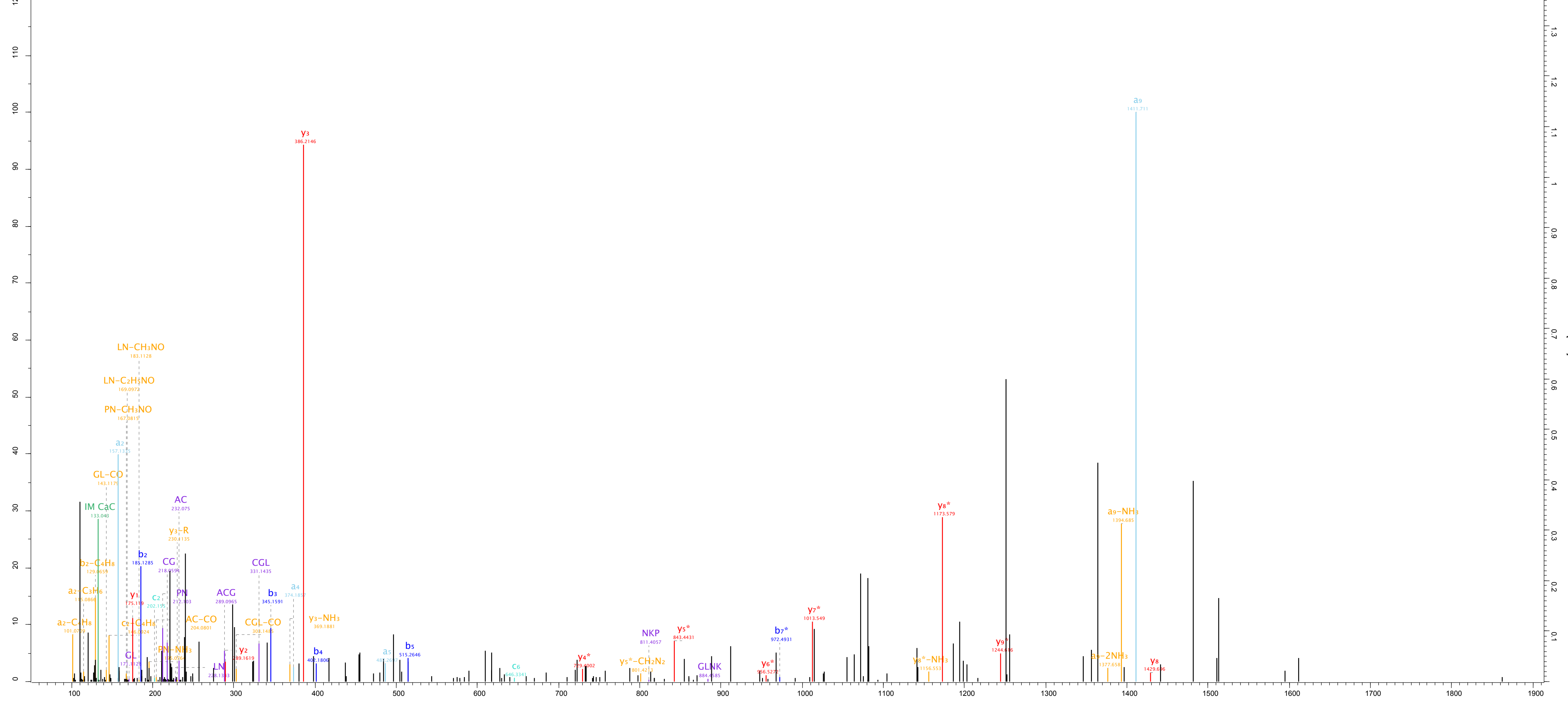
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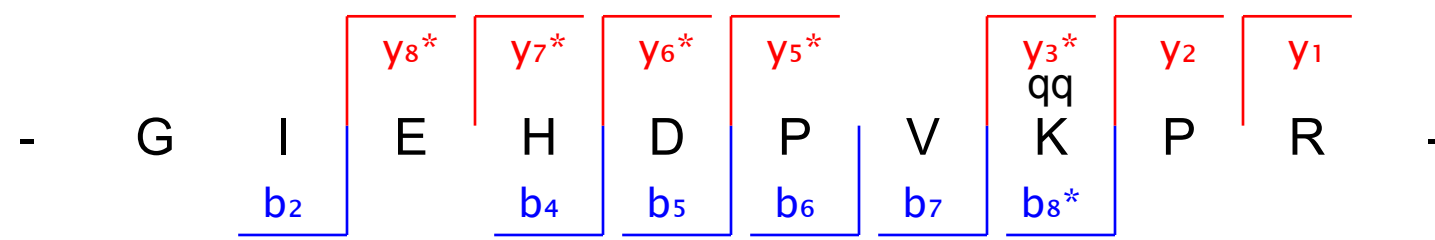
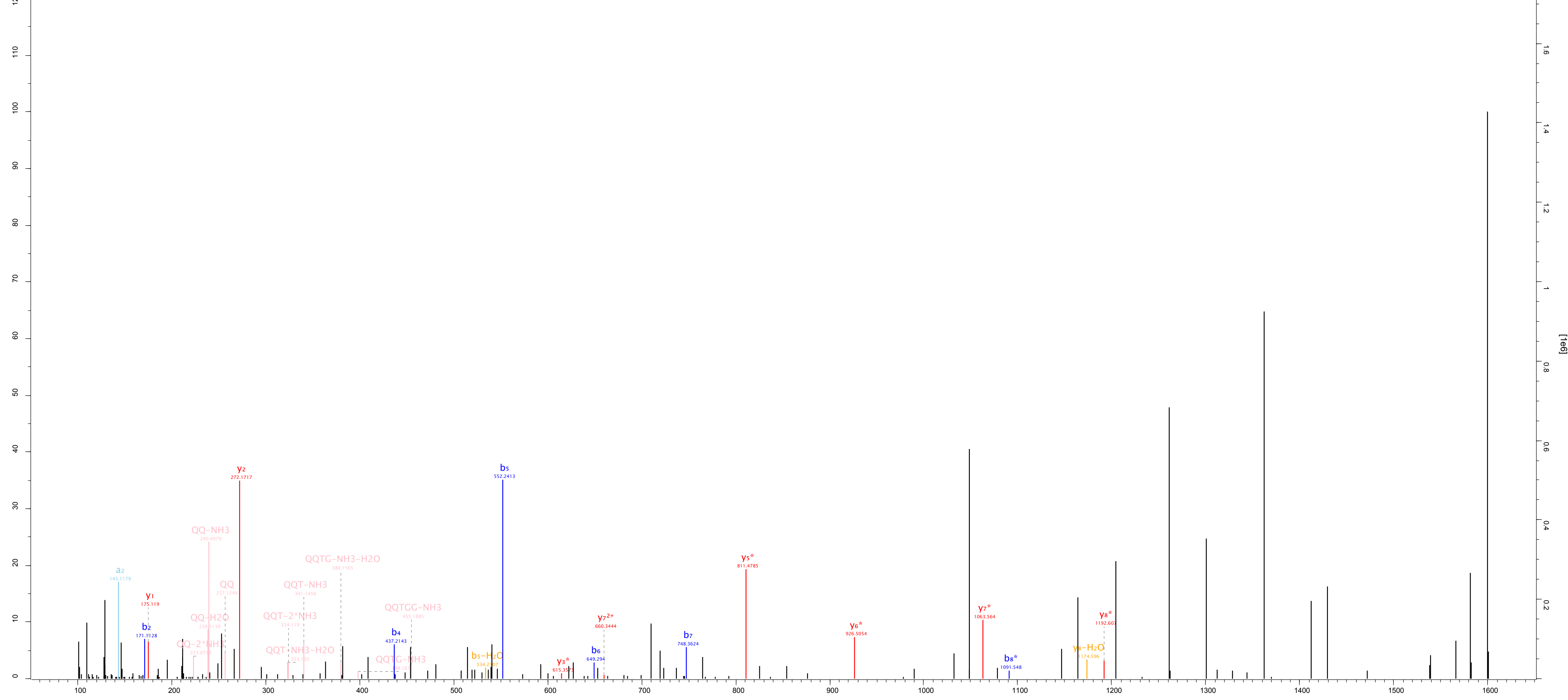


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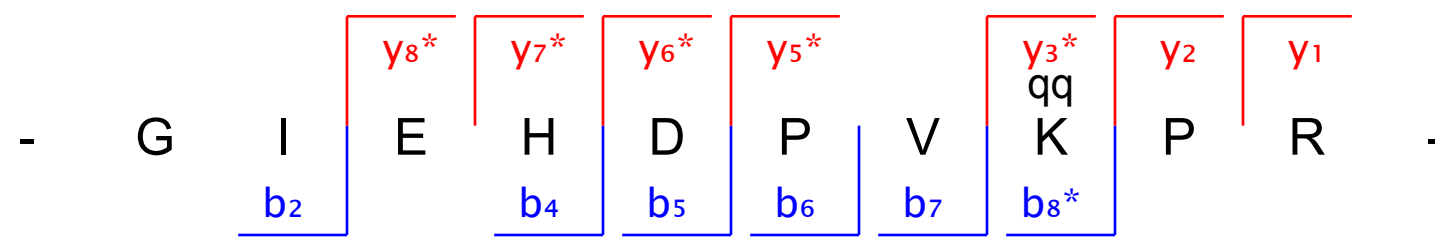


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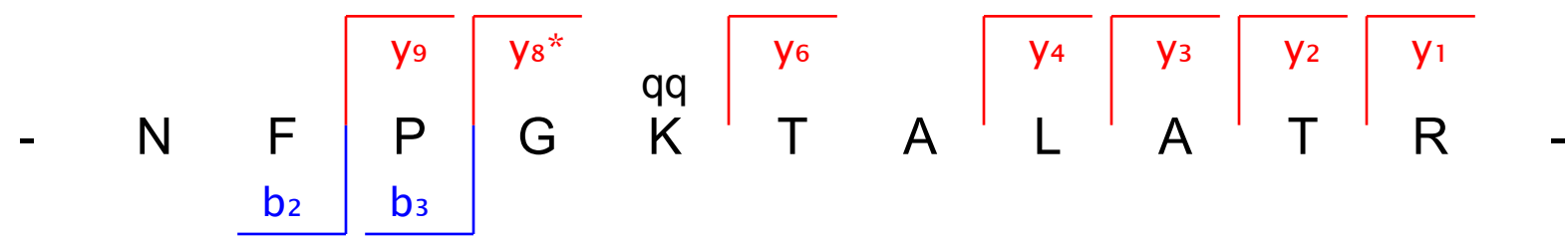
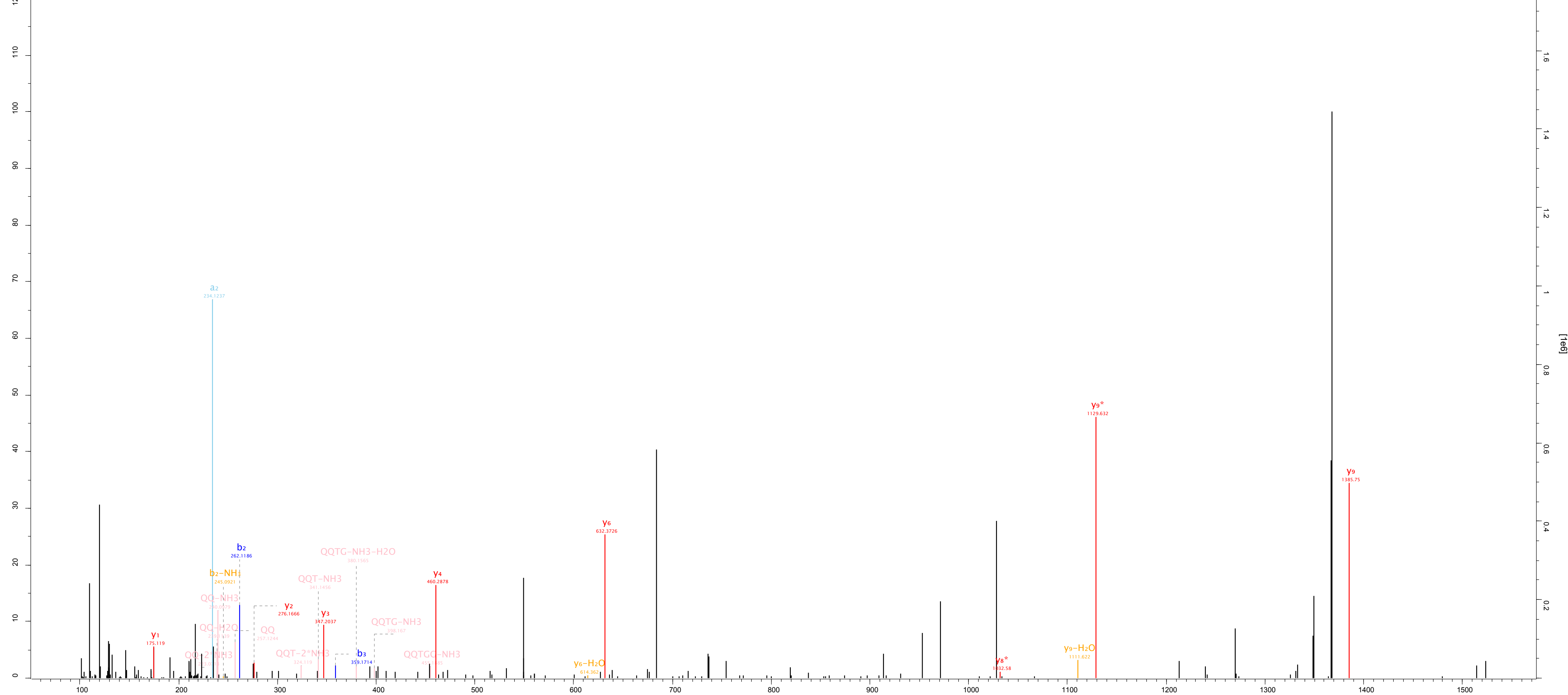




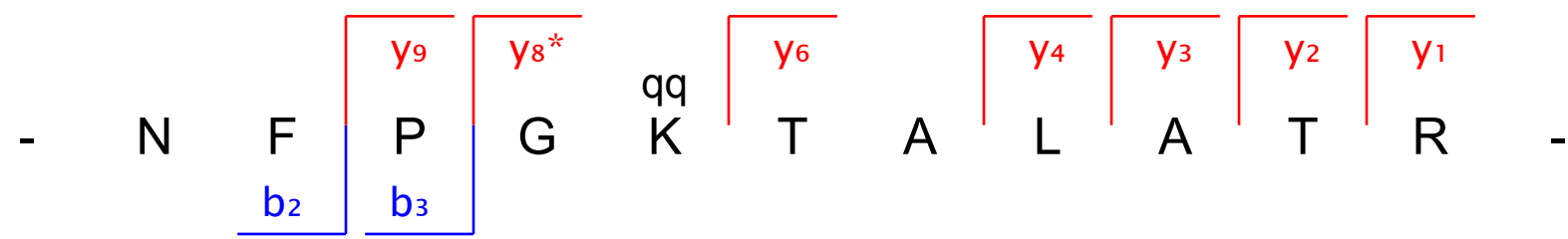
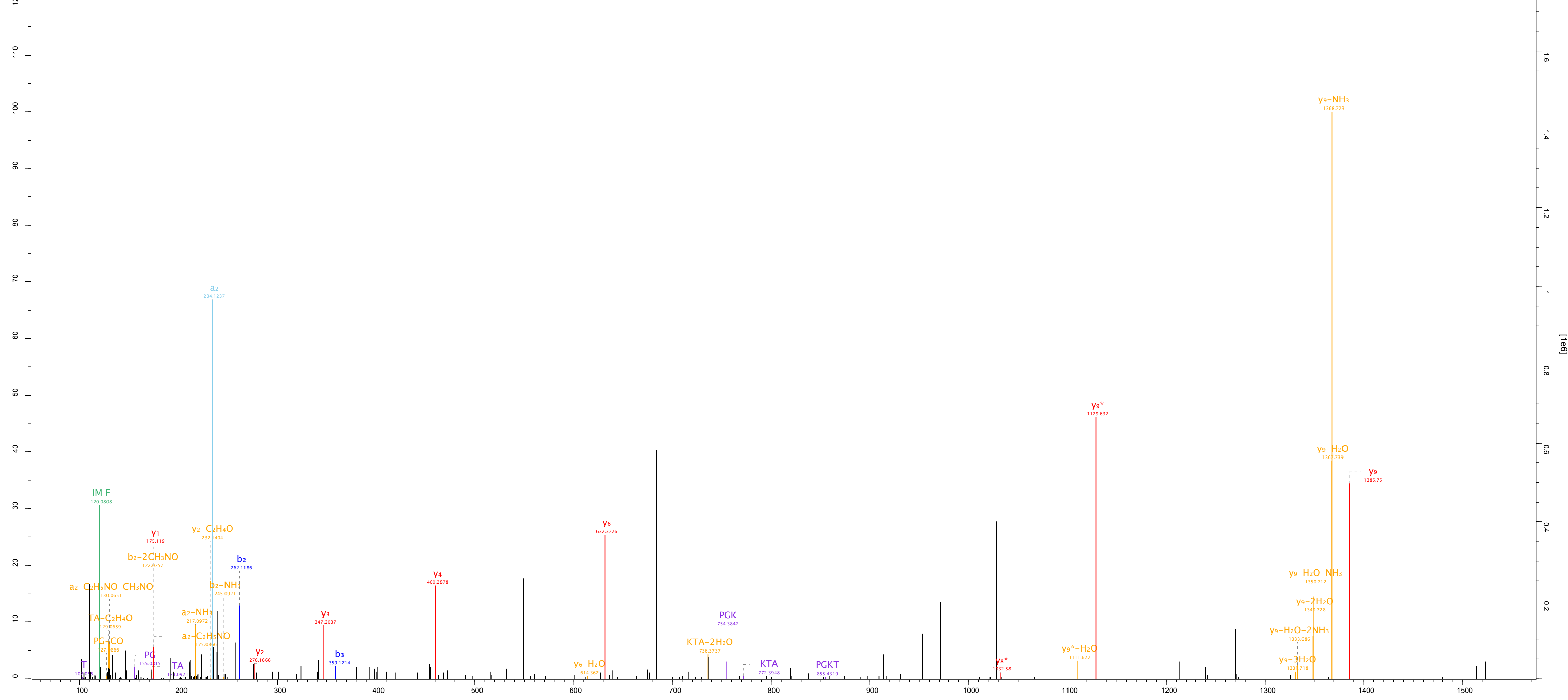
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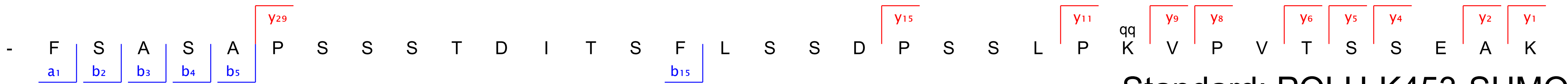
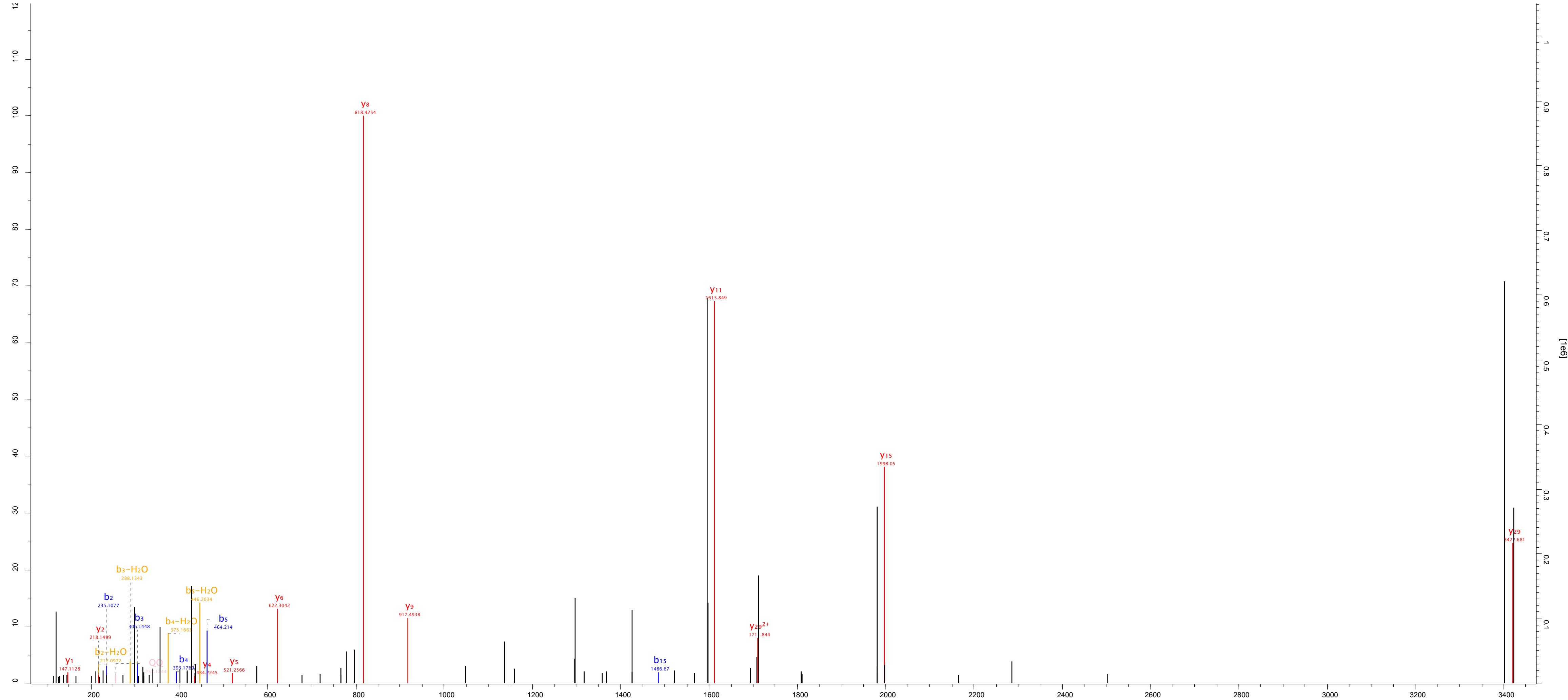
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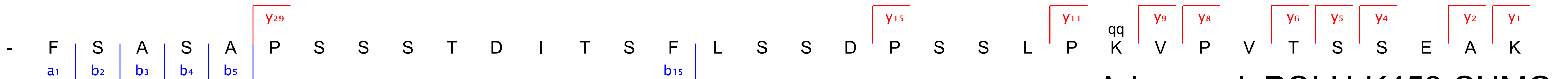
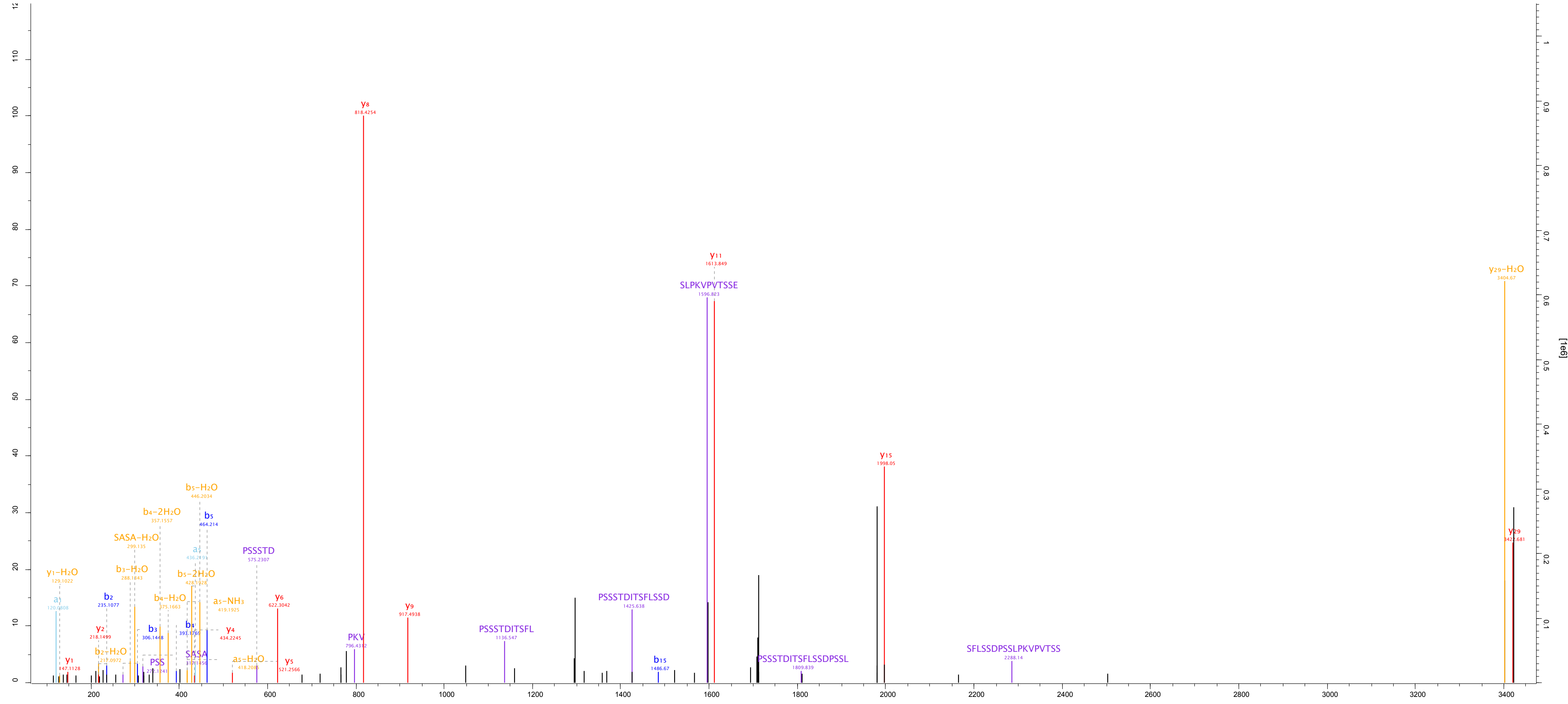
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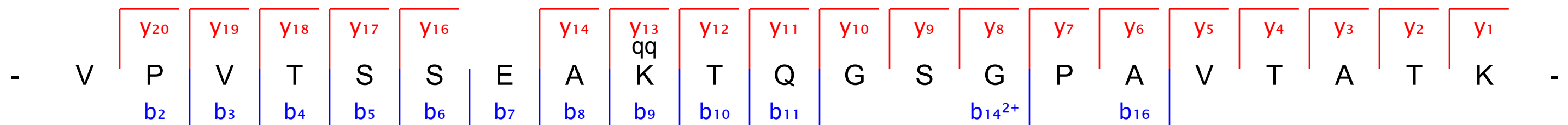
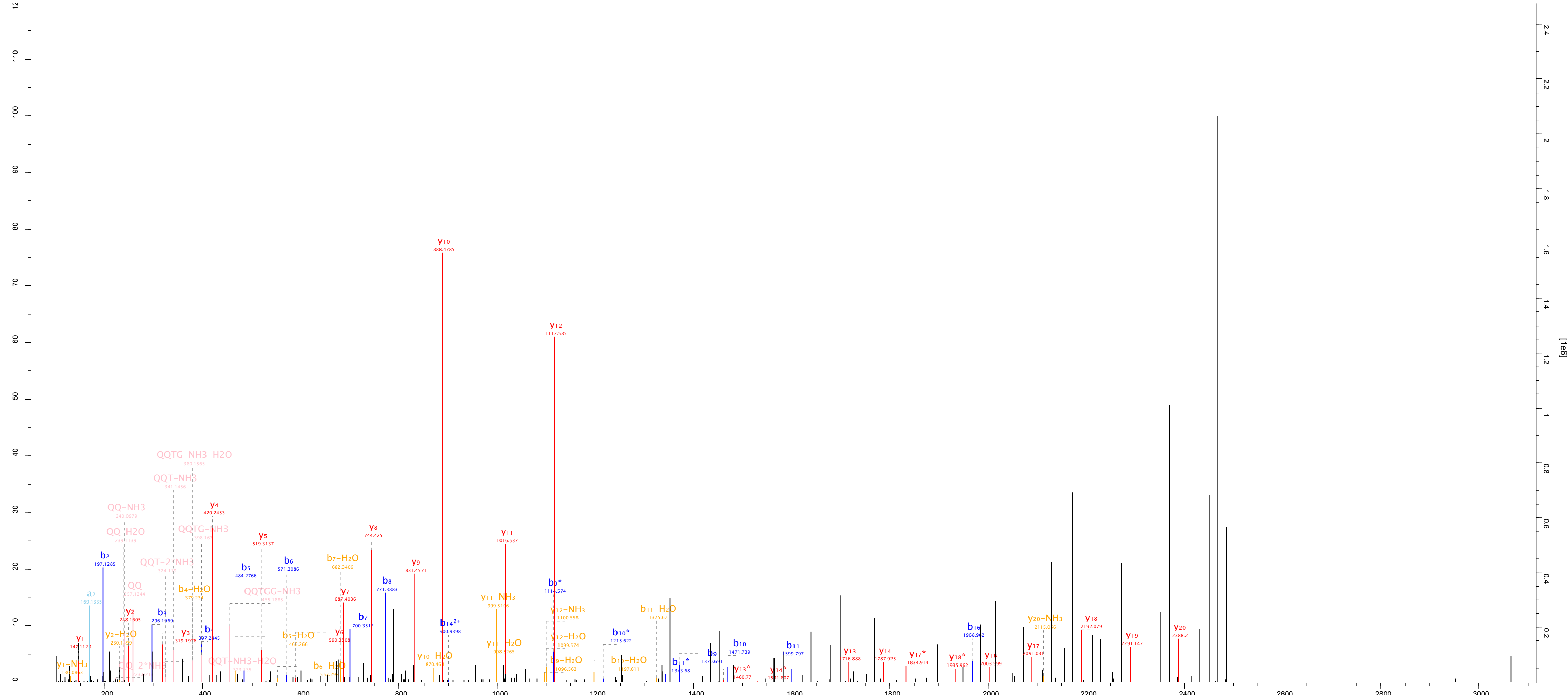
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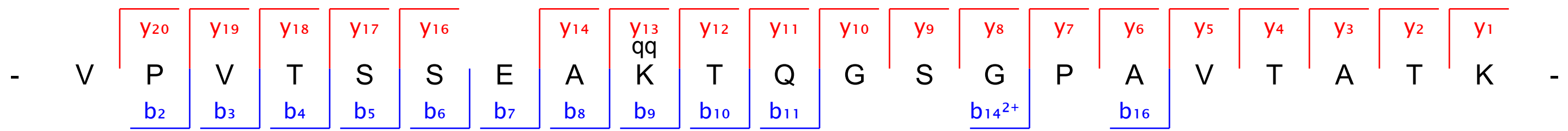
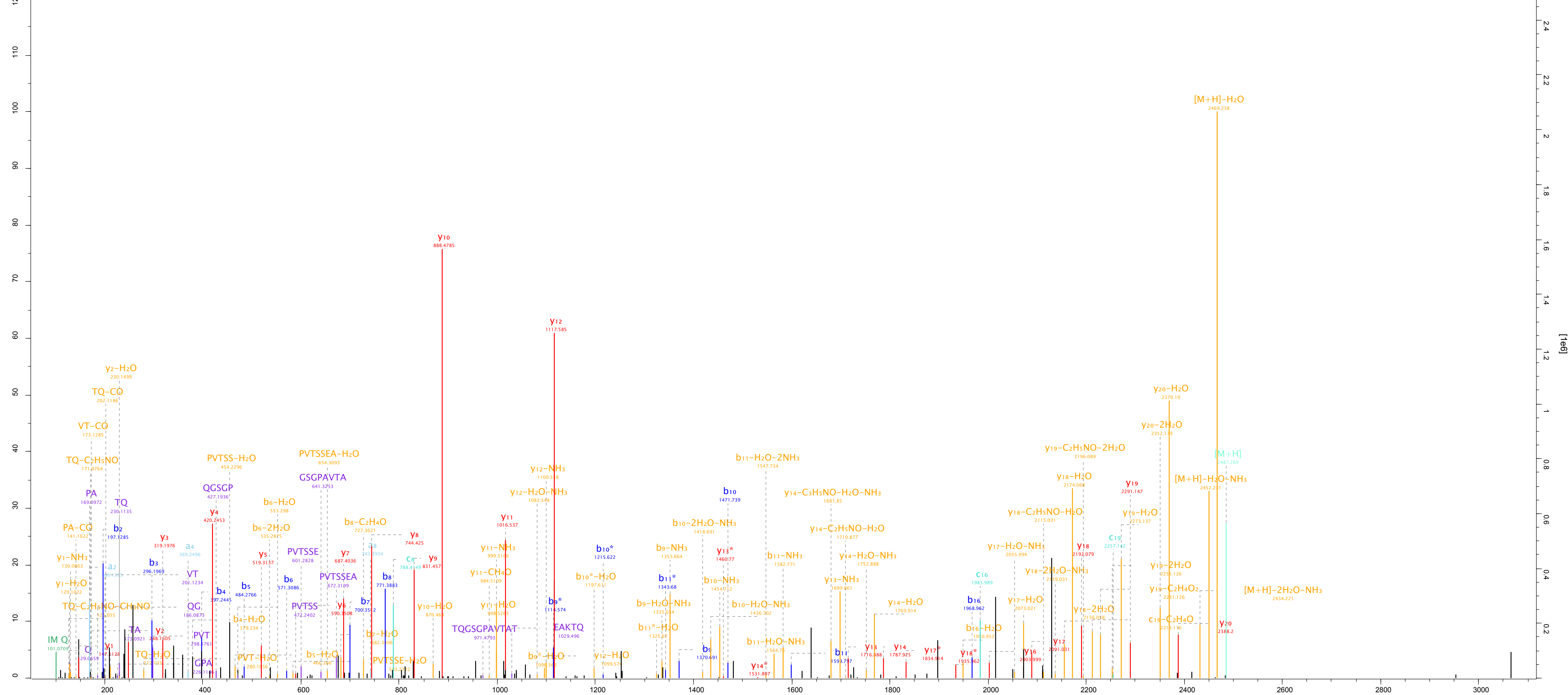
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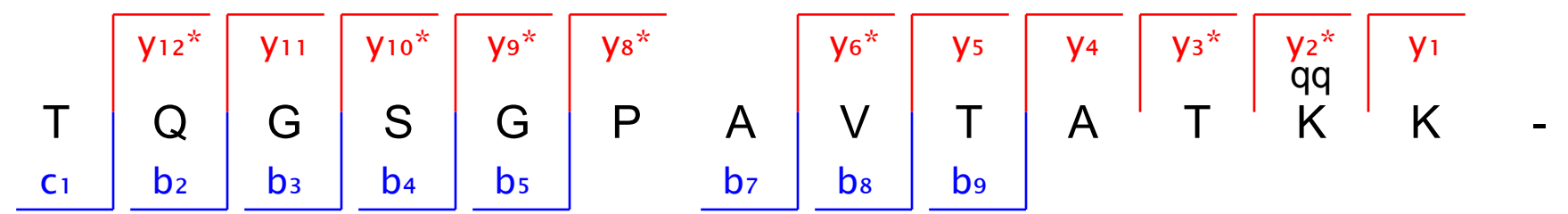
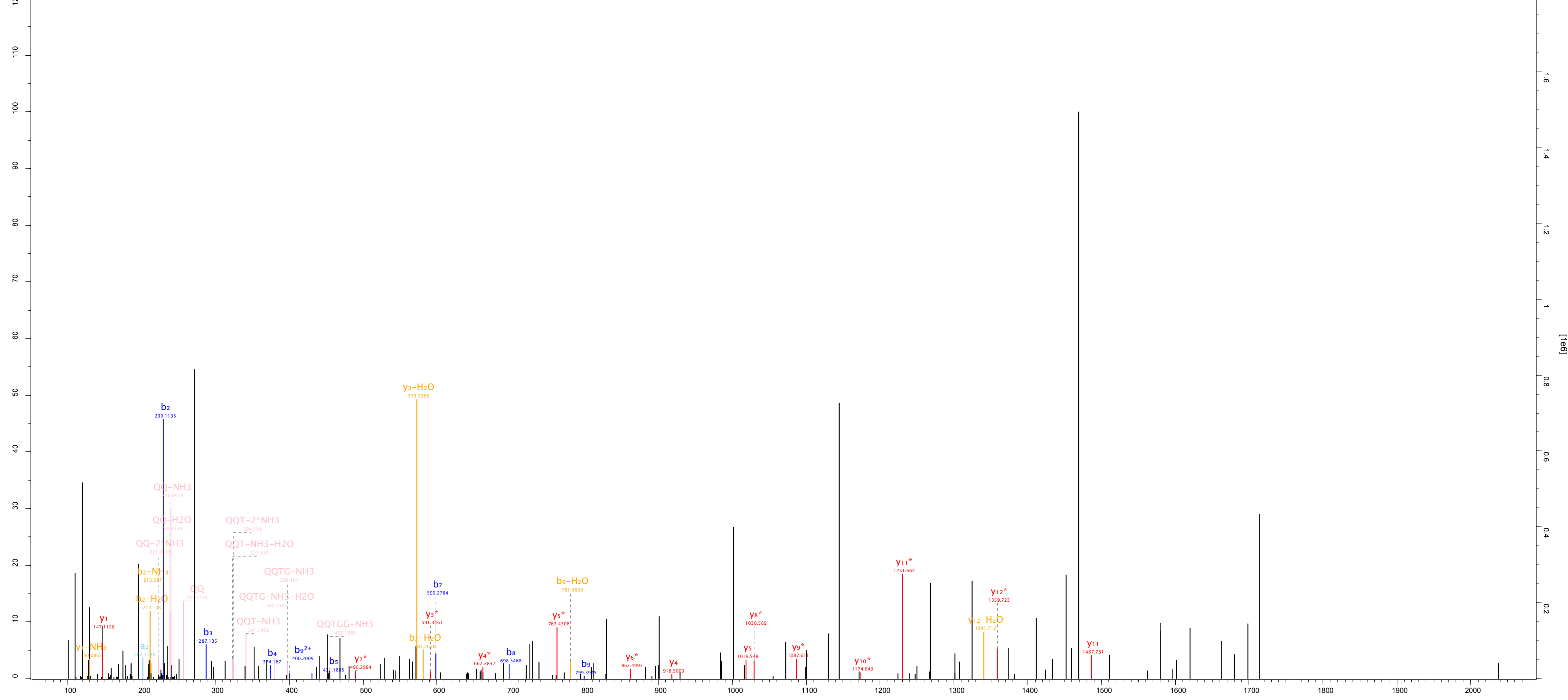


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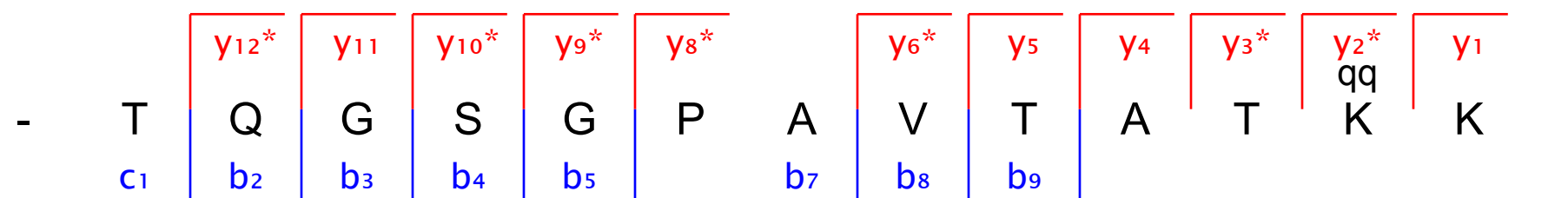
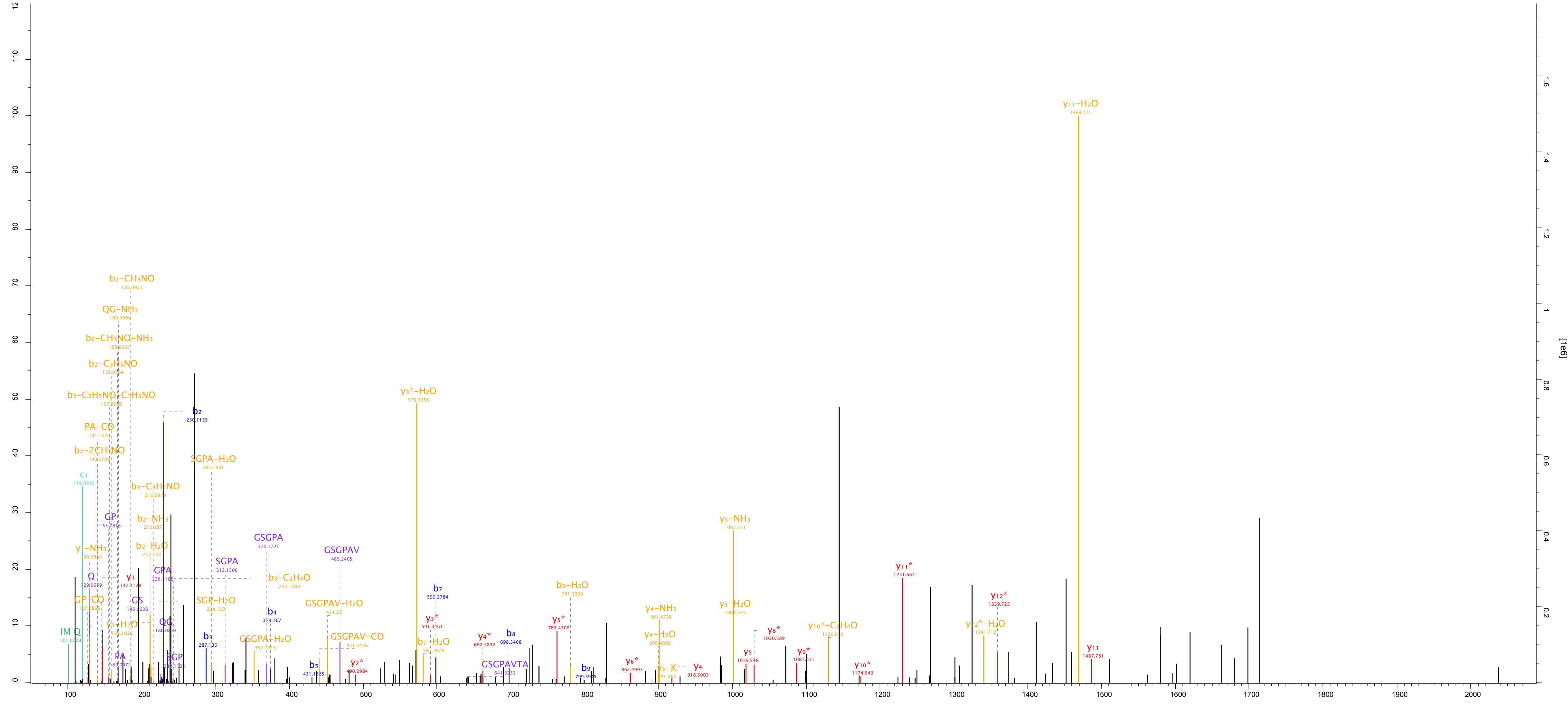


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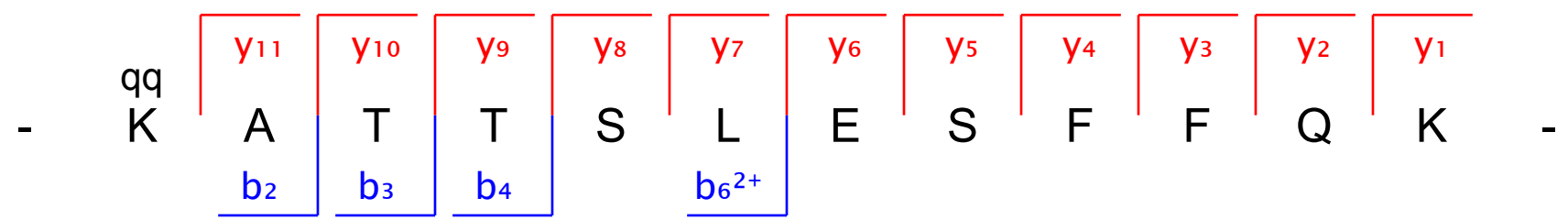
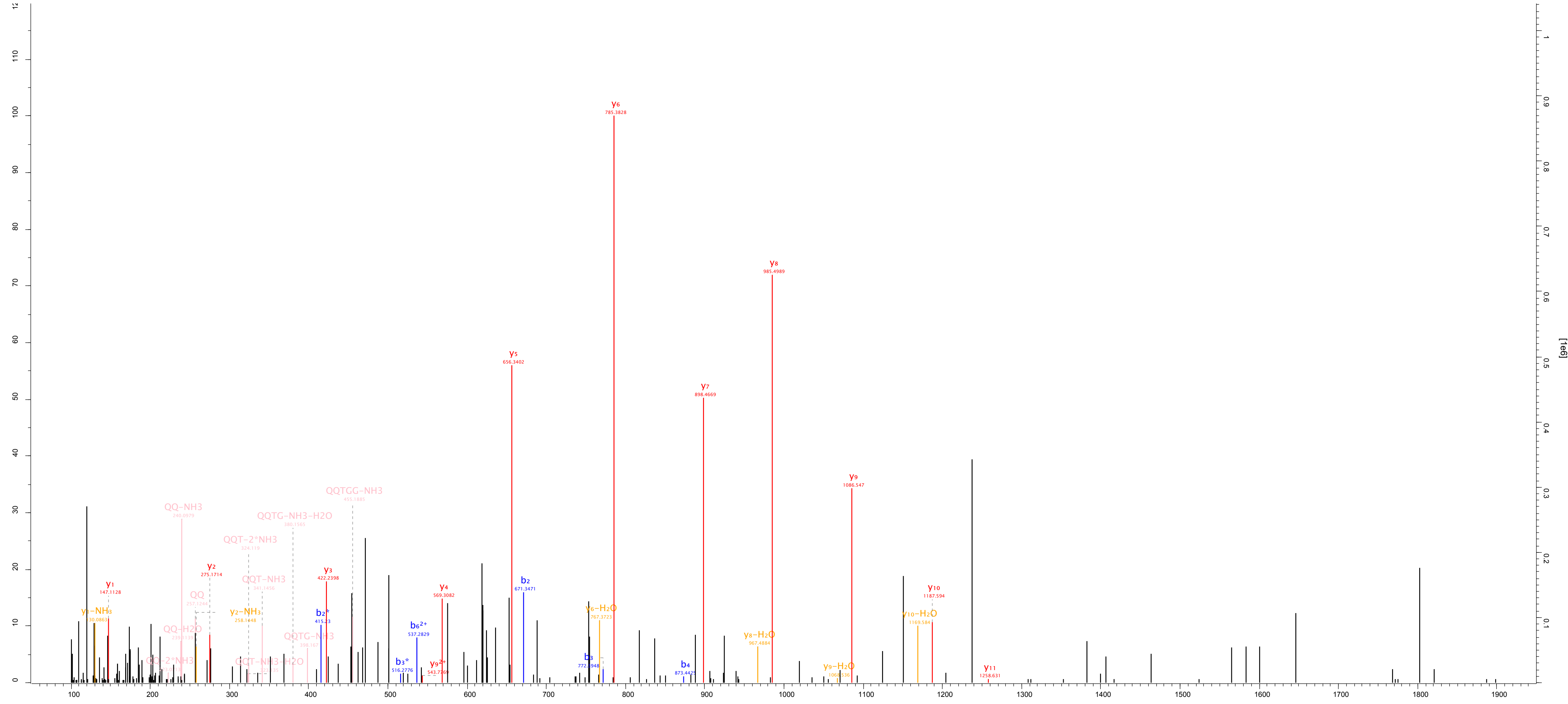




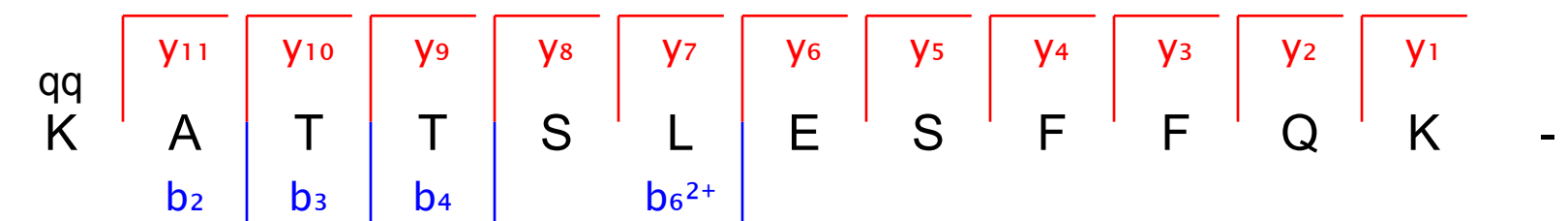
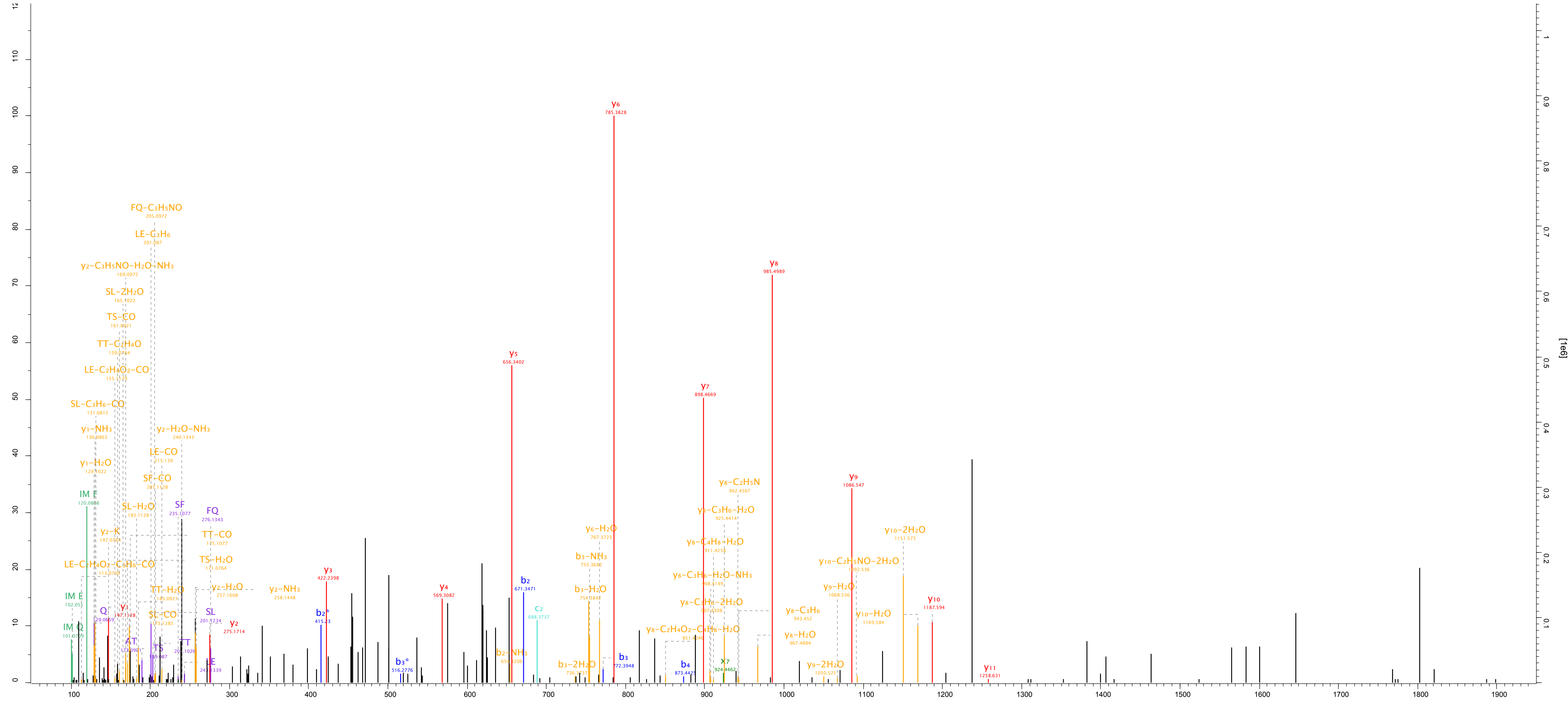
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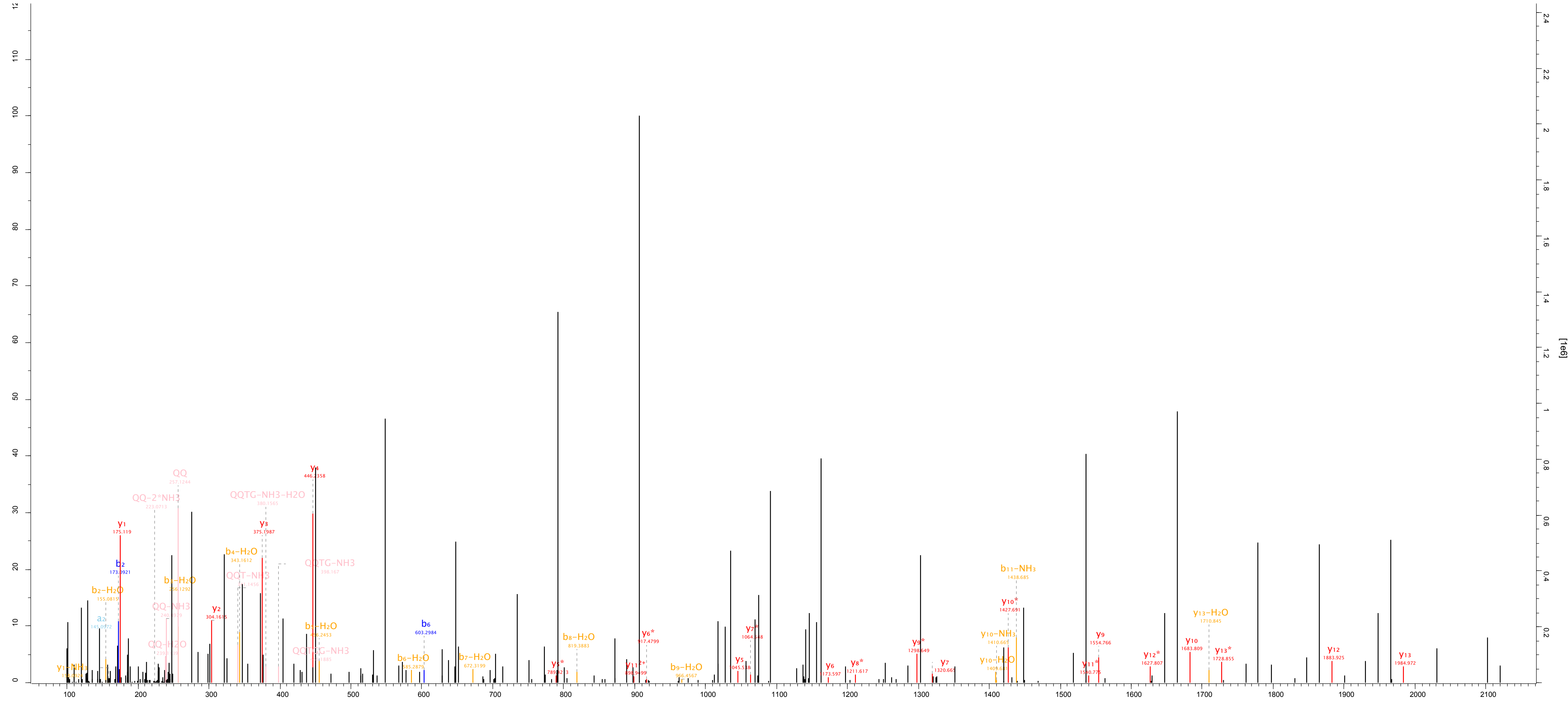
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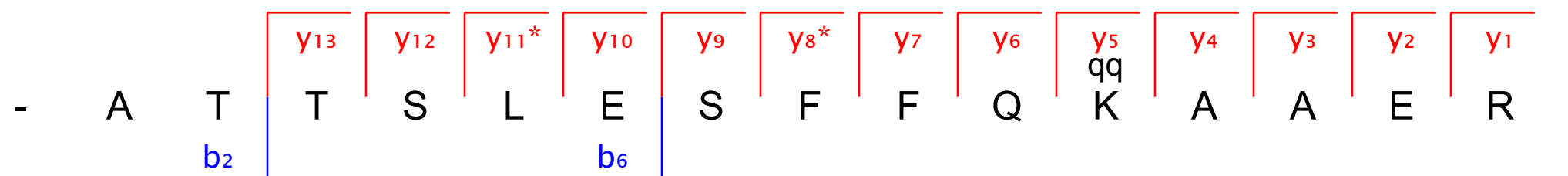
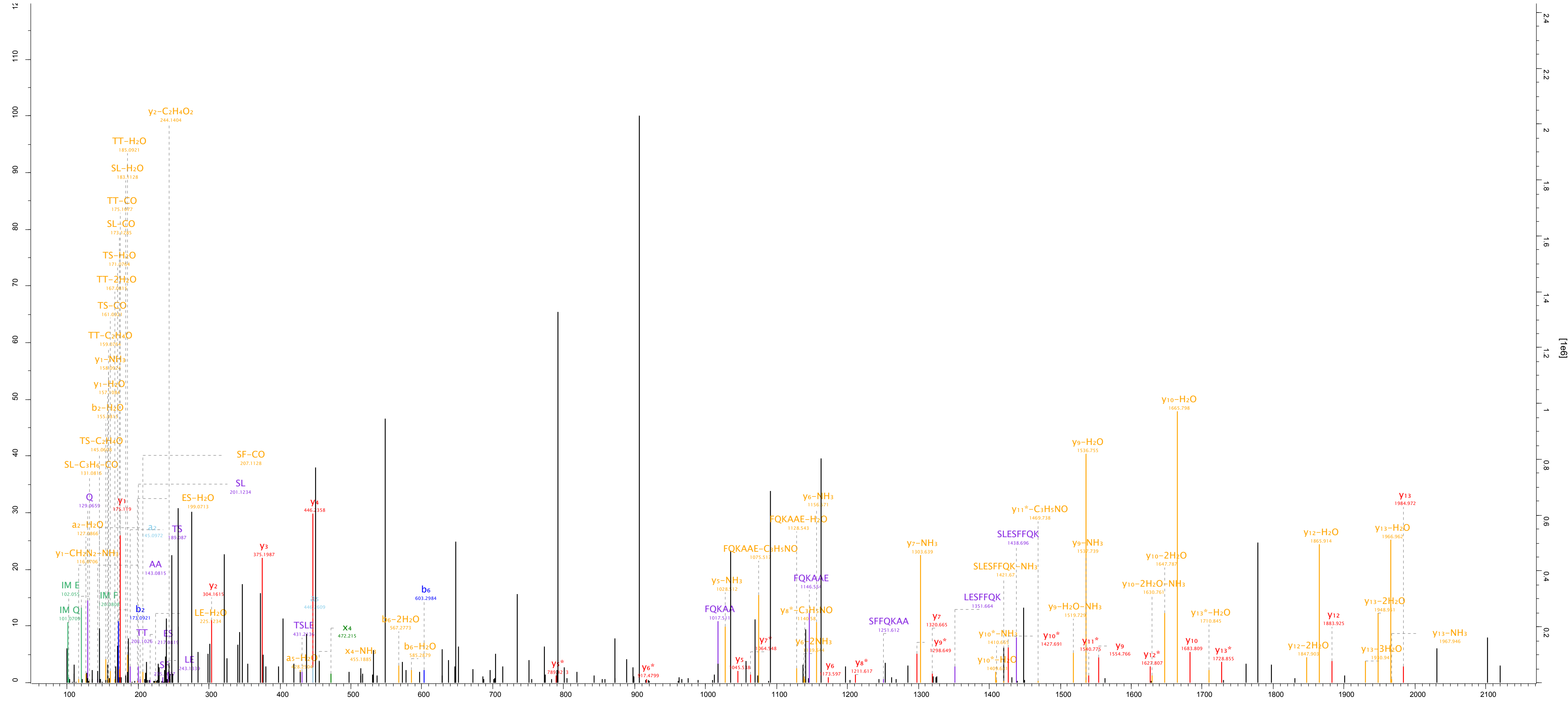
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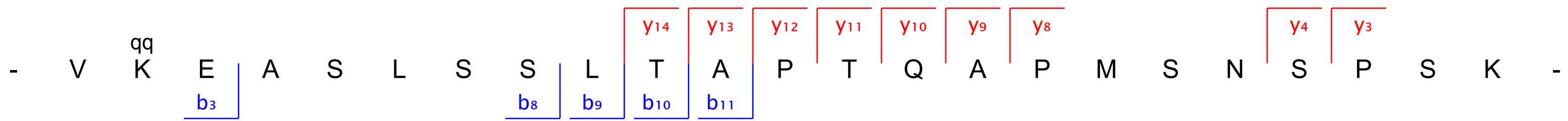
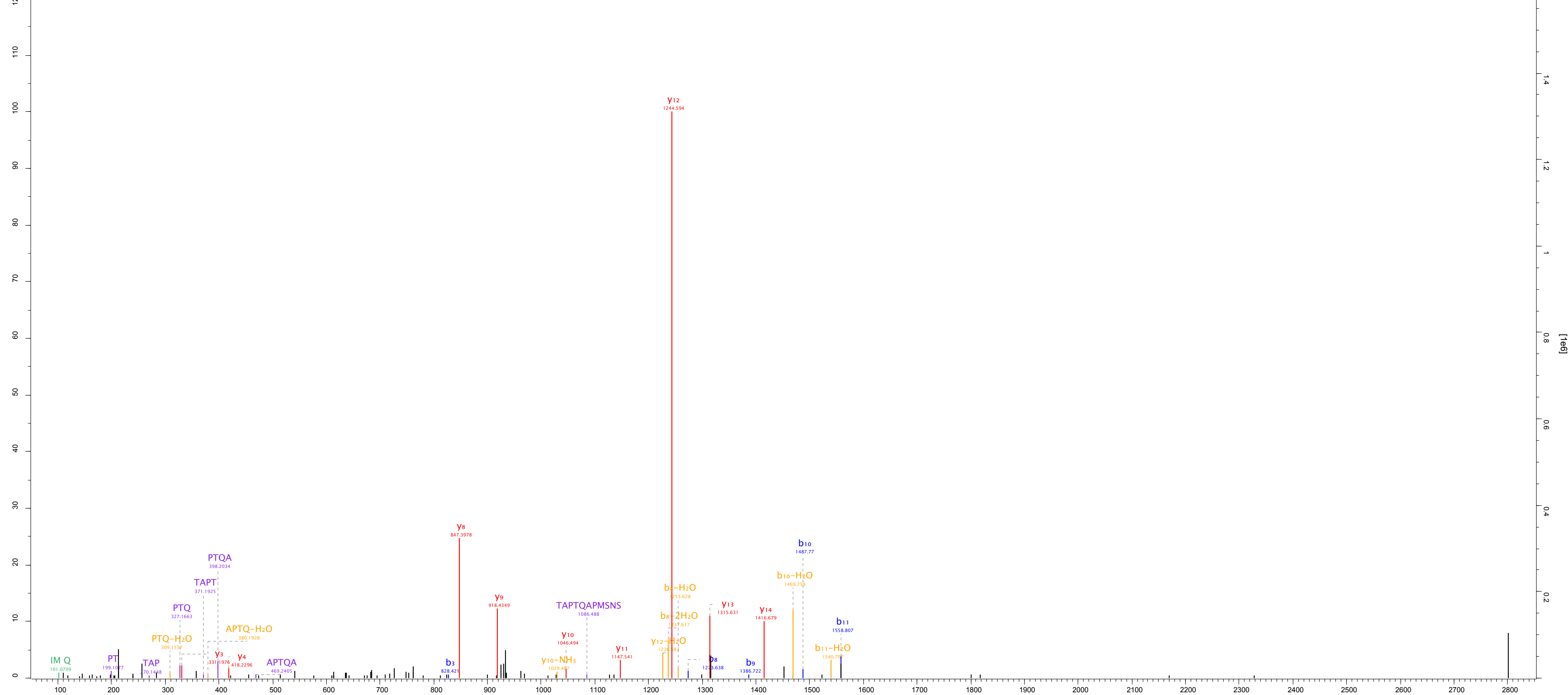
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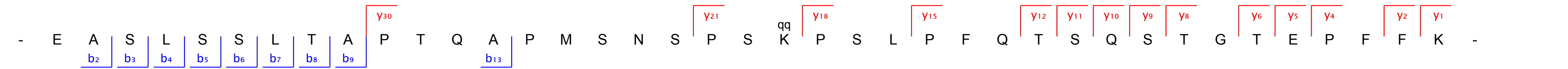
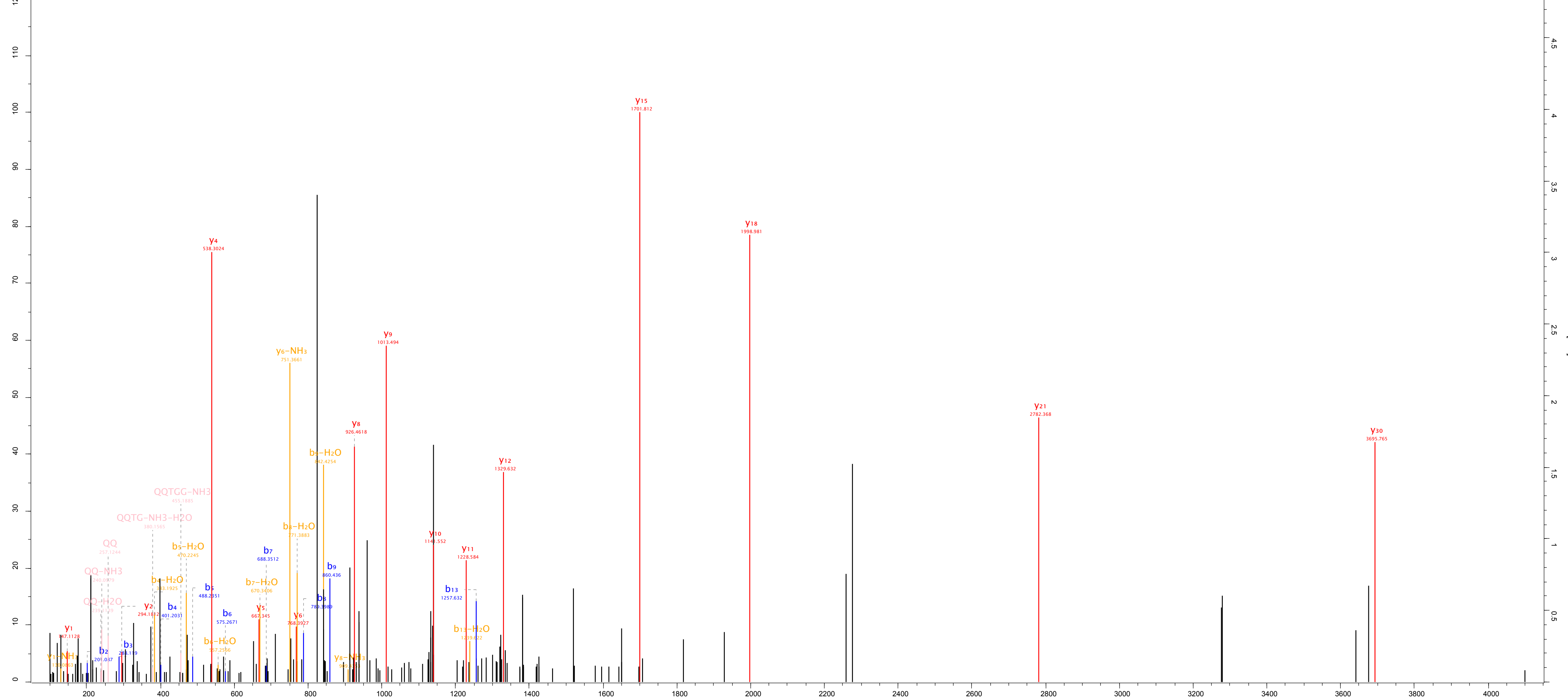
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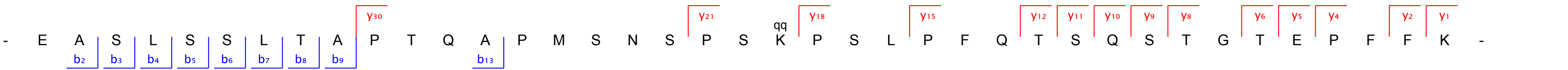
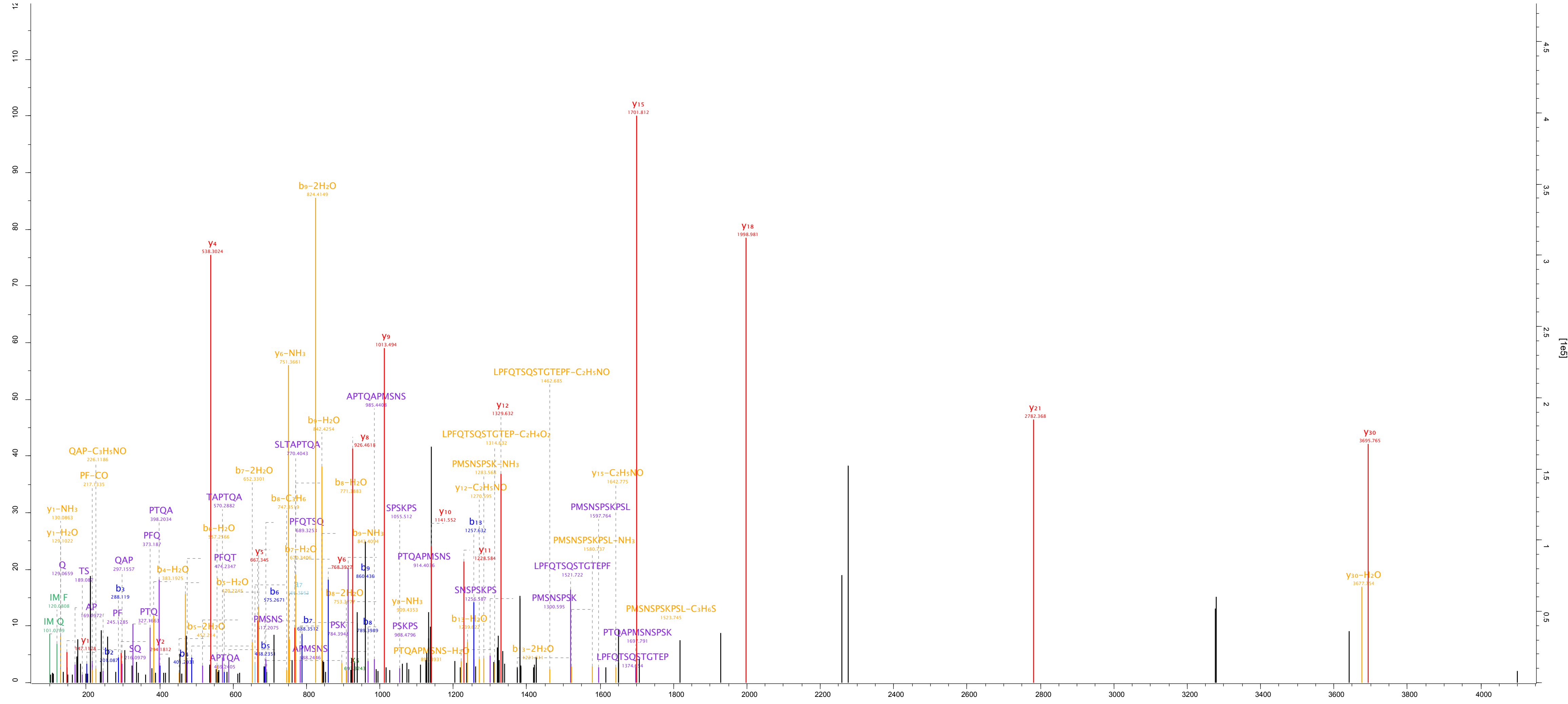
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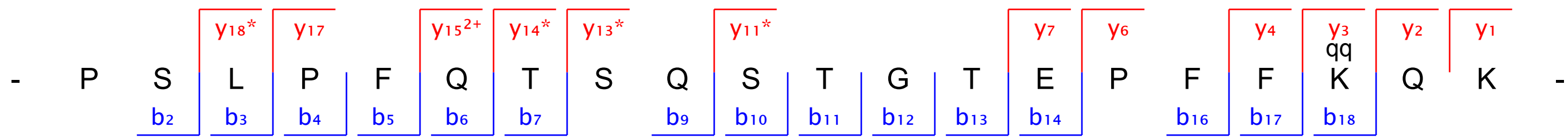
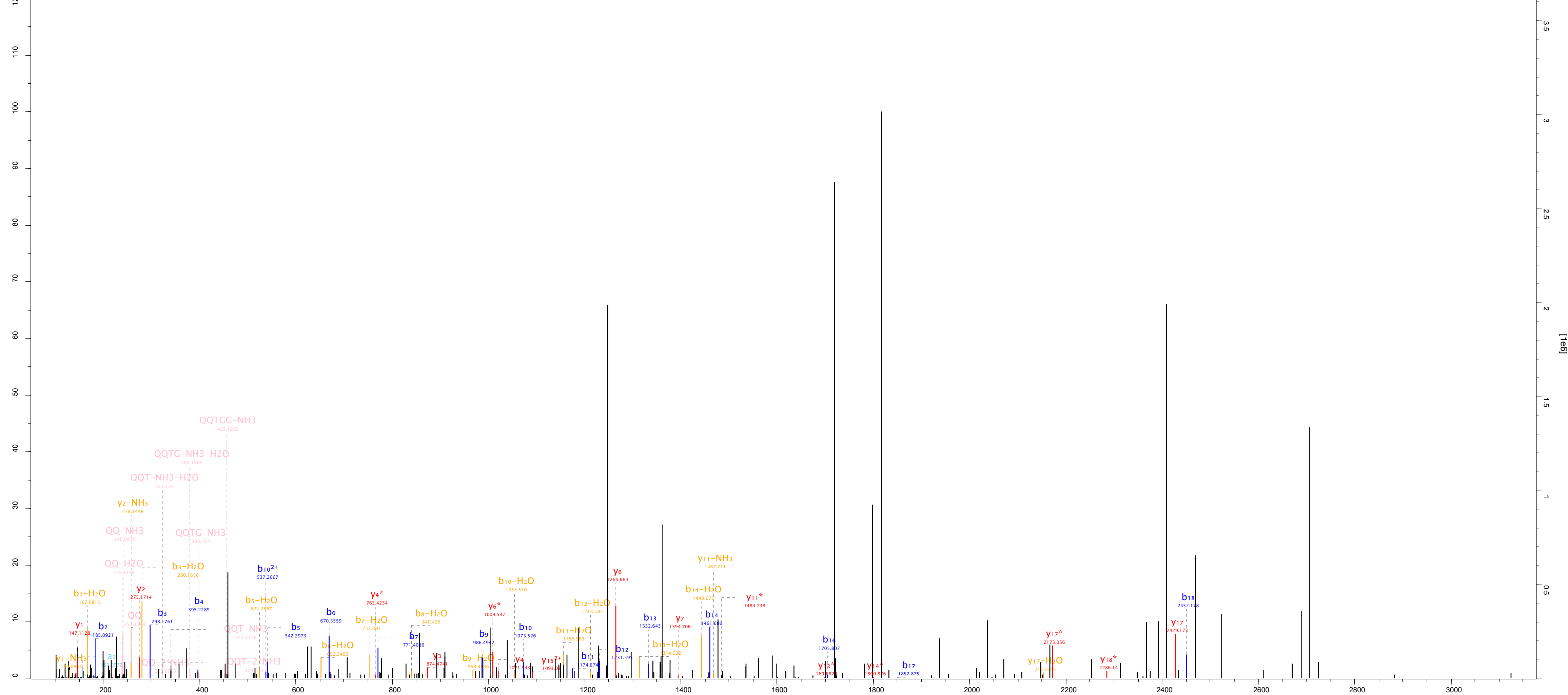




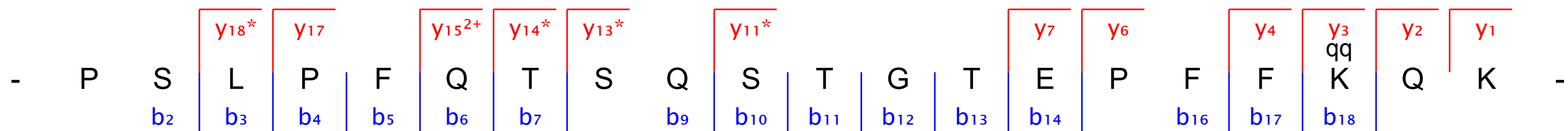
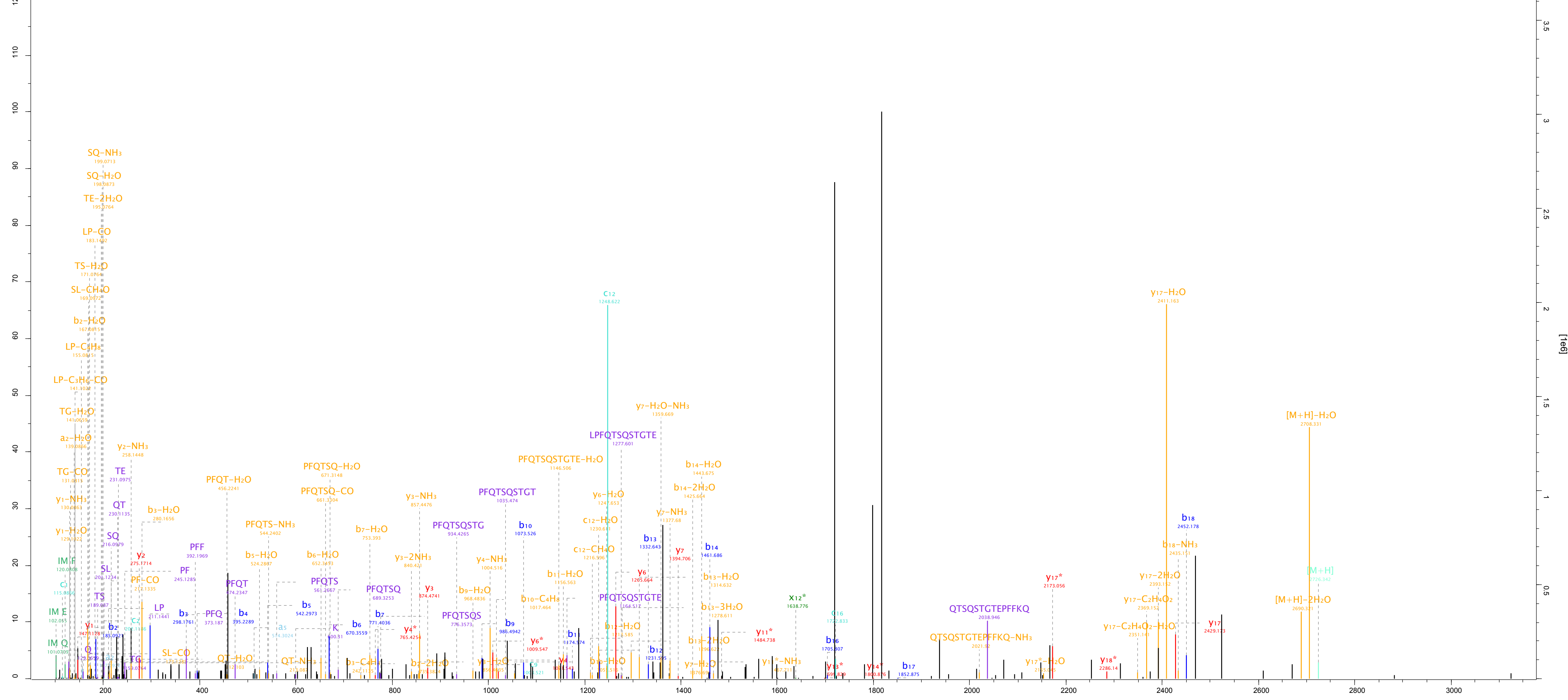
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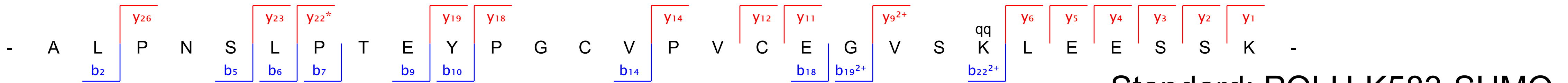
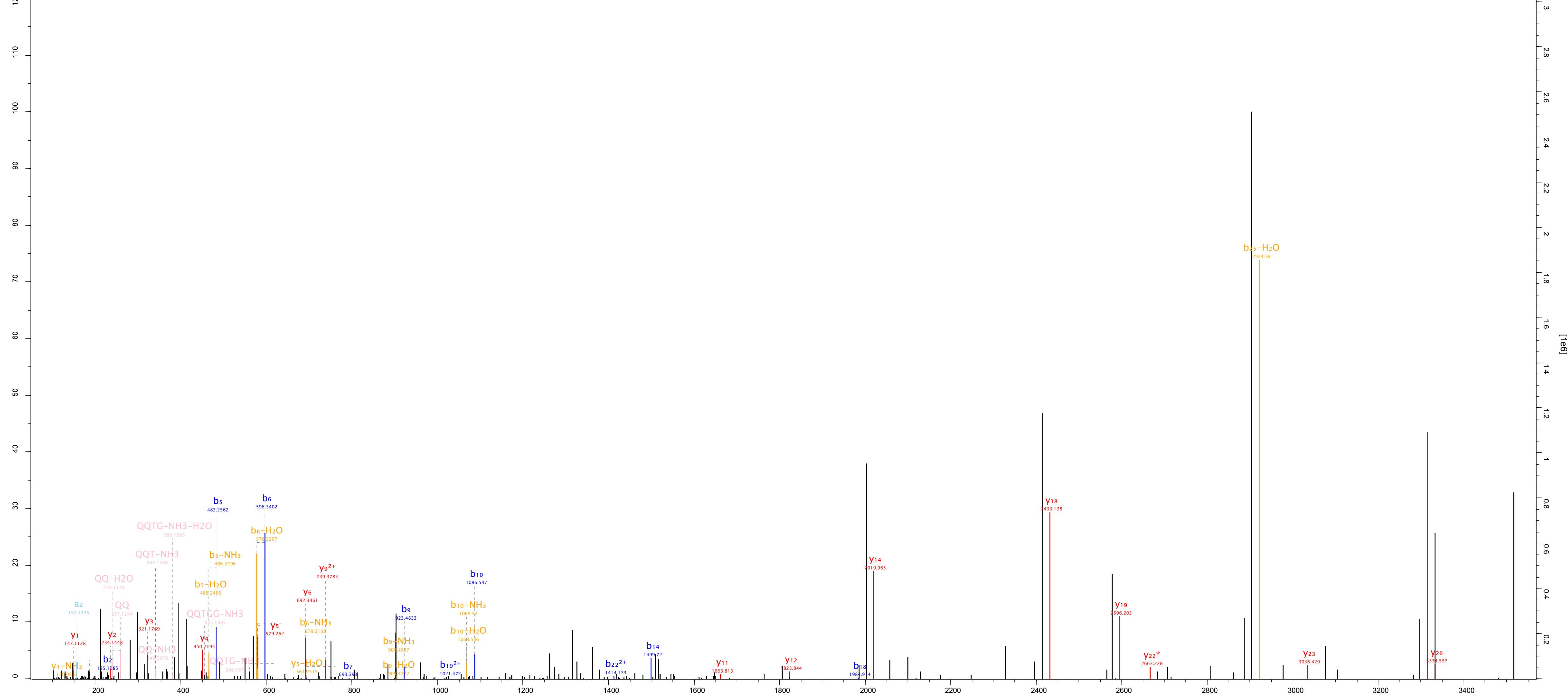
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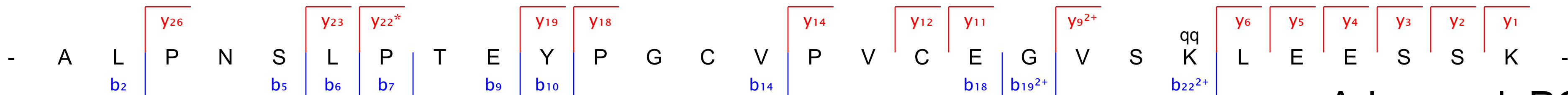
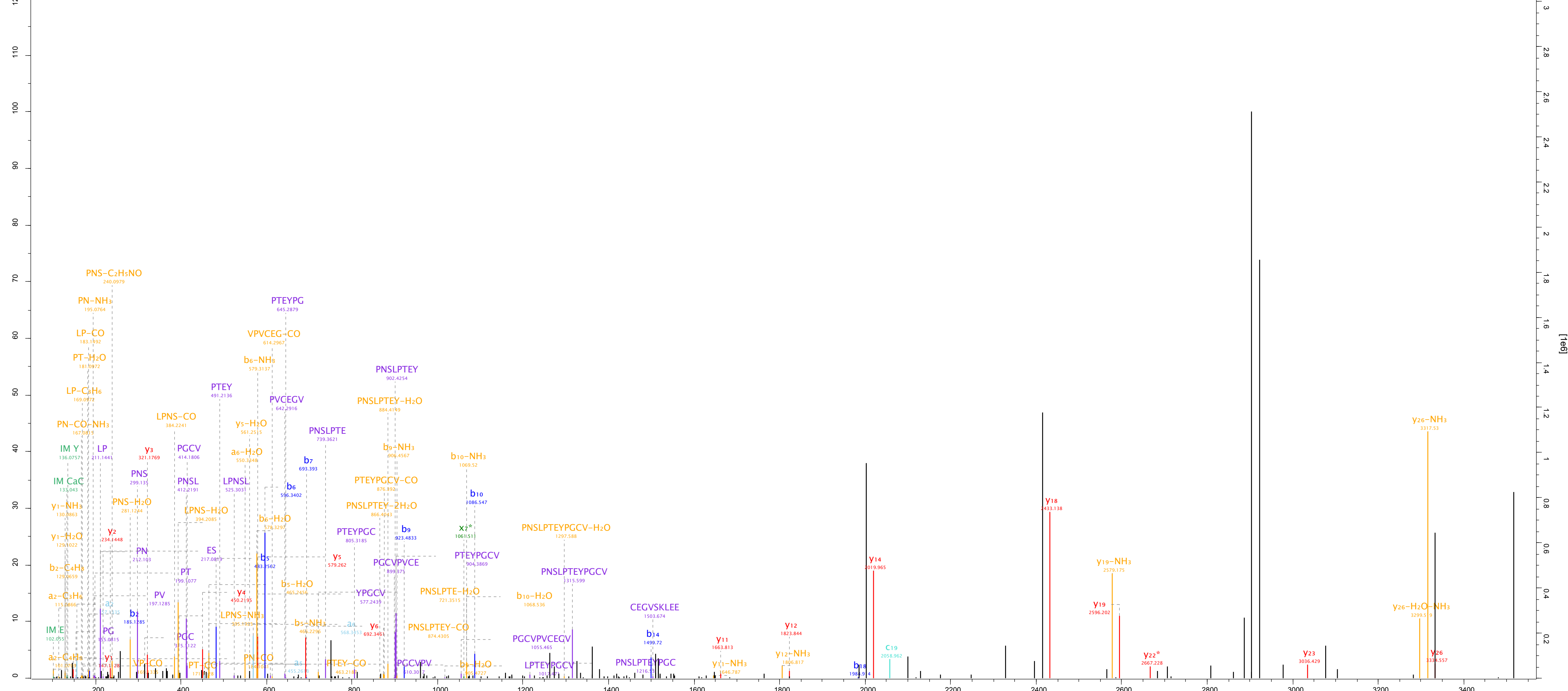
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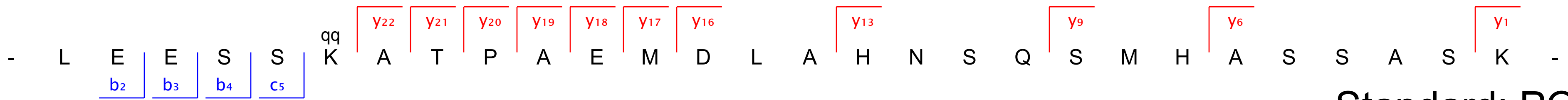
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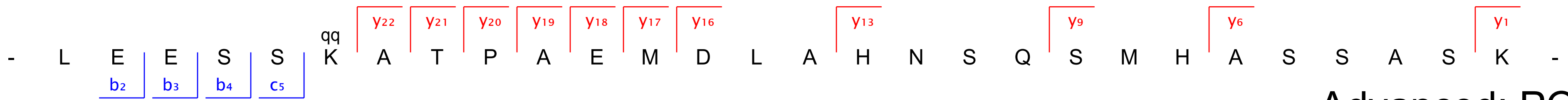
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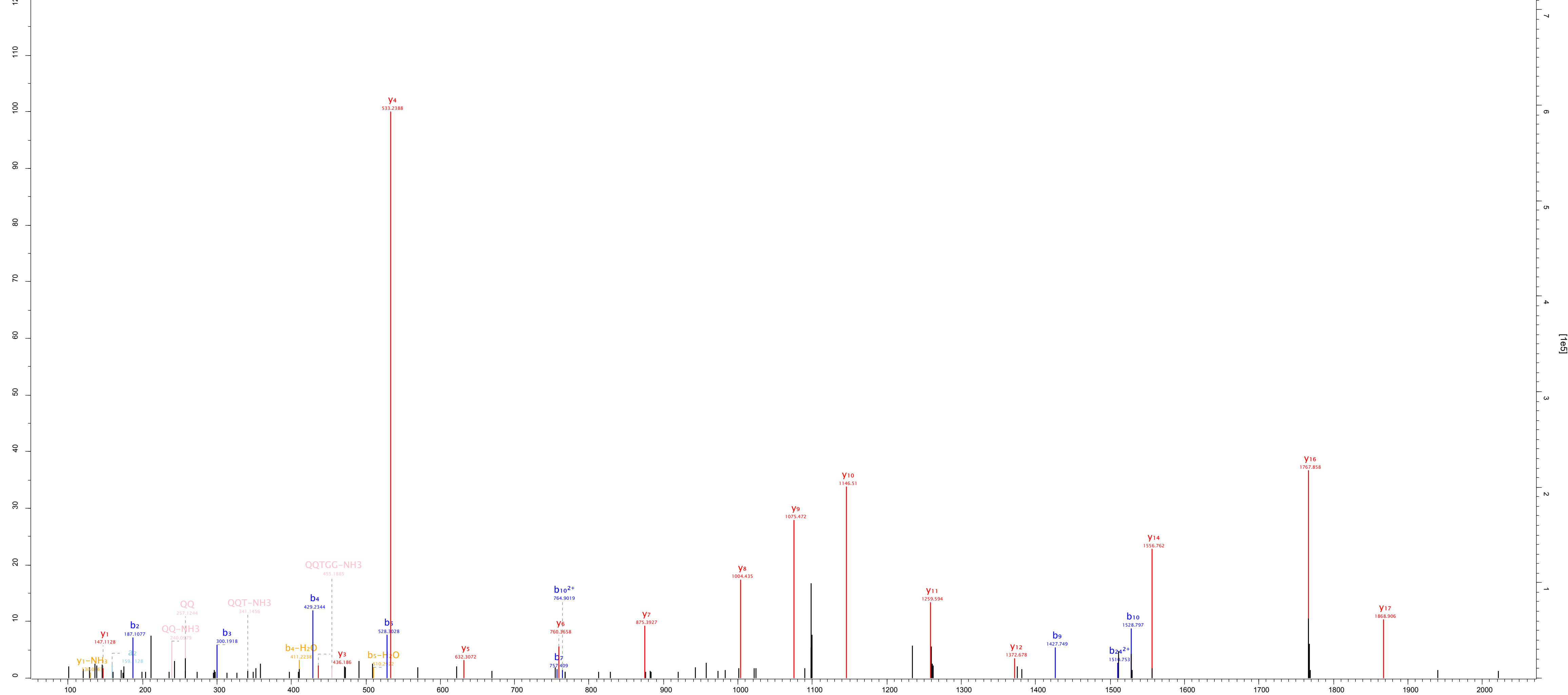


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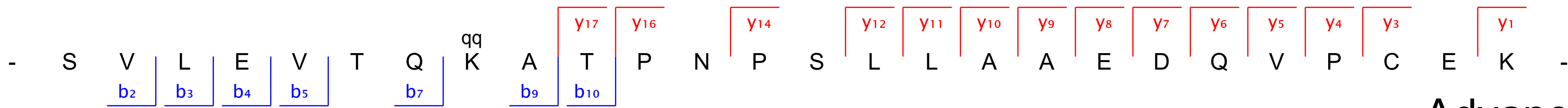
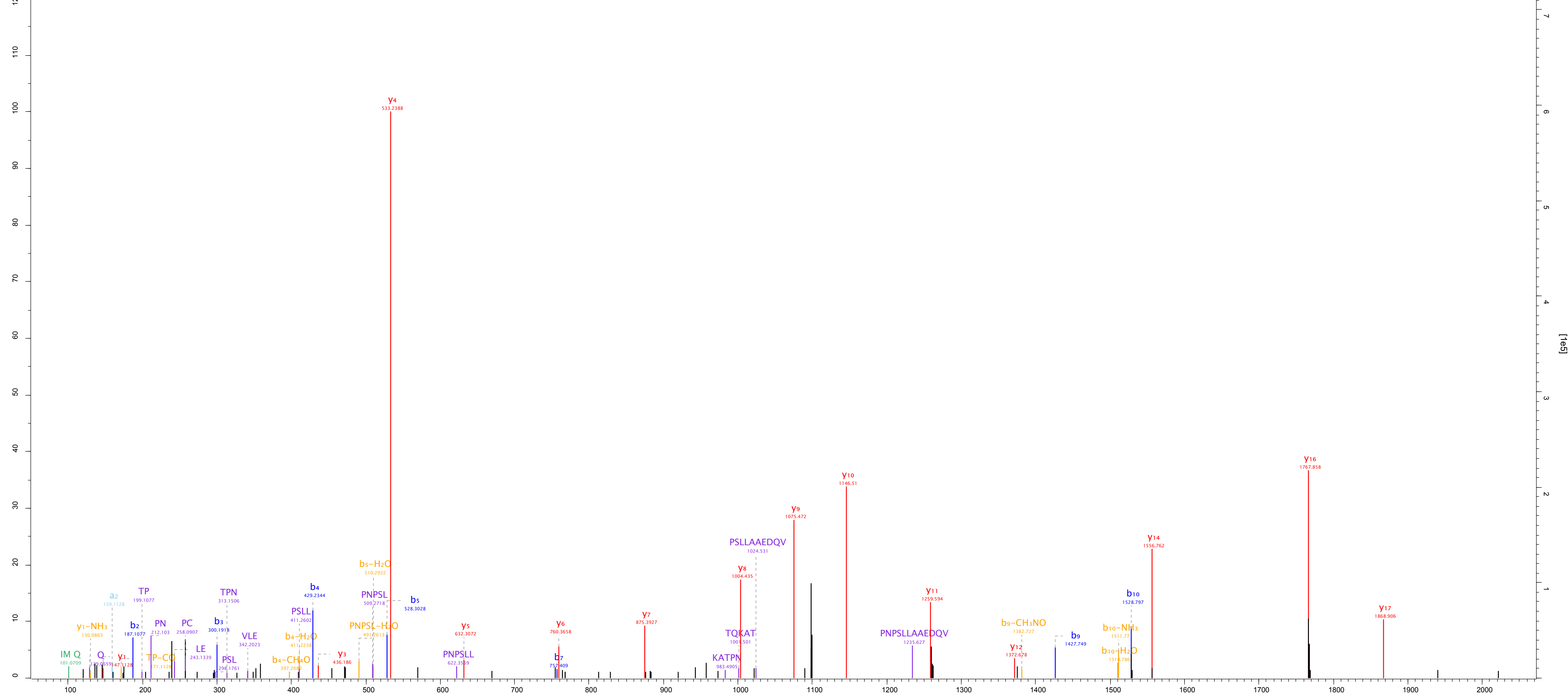


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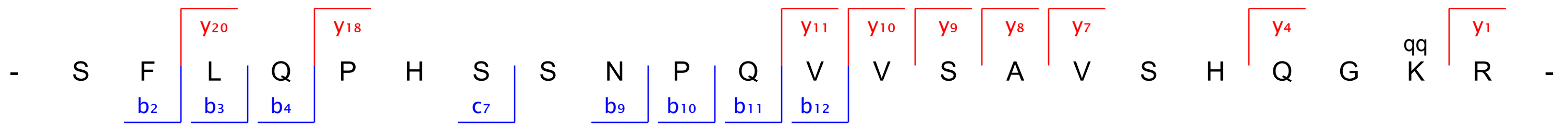
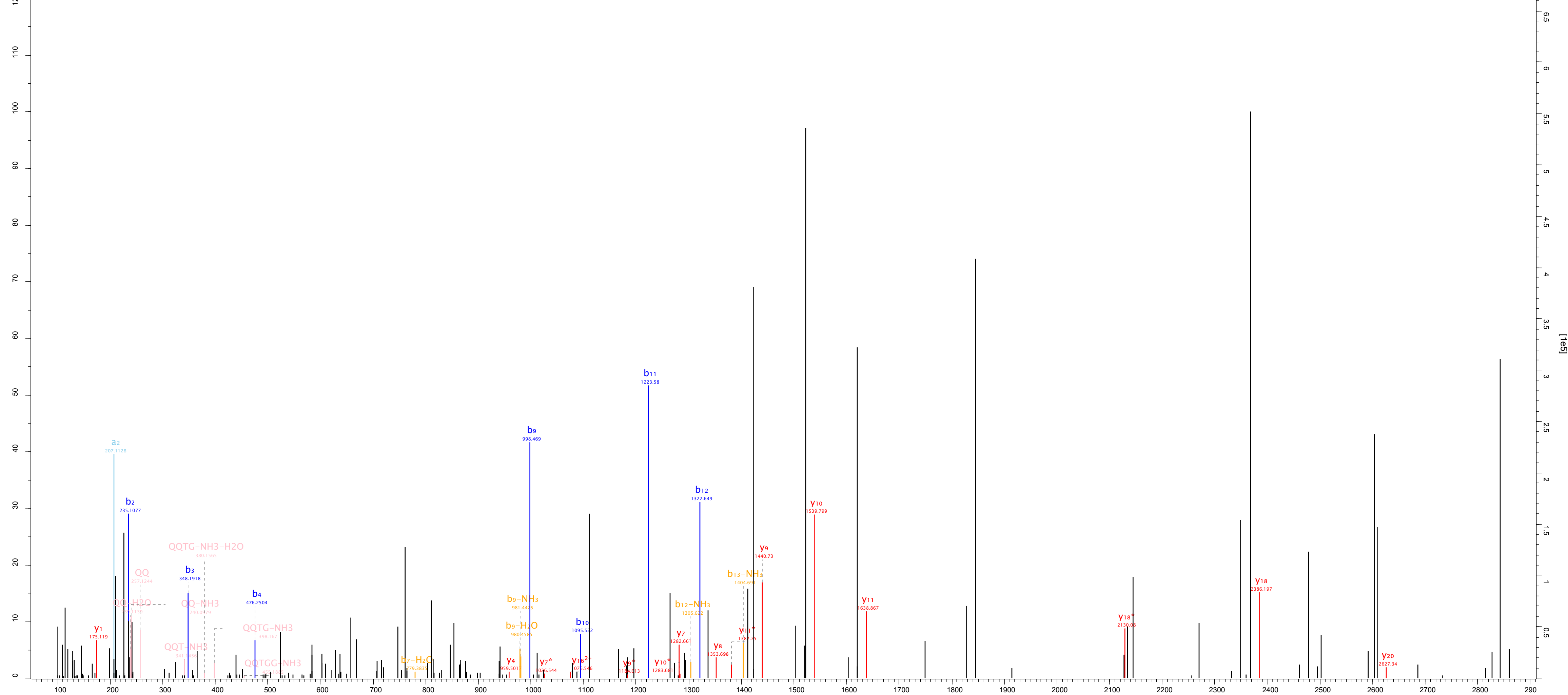
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y<sub>1</sub> y<sub>3</sub> y<sub>4</sub> y<sub>5</sub> y<sub>6</sub> y<sub>7</sub> y<sub>8</sub> y<sub>9</sub> y<sub>10</sub> y<sub>11</sub> y<sub>12</sub> y<sub>14</sub> y<sub>16</sub> y<sub>17</sub>

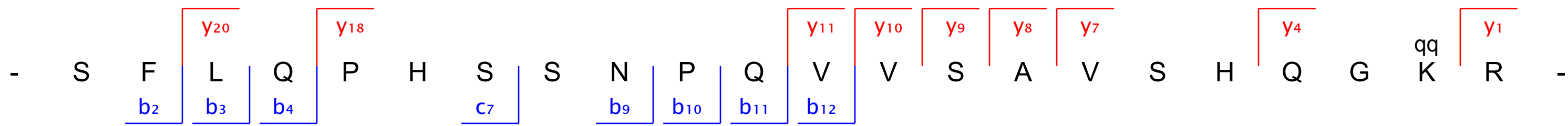
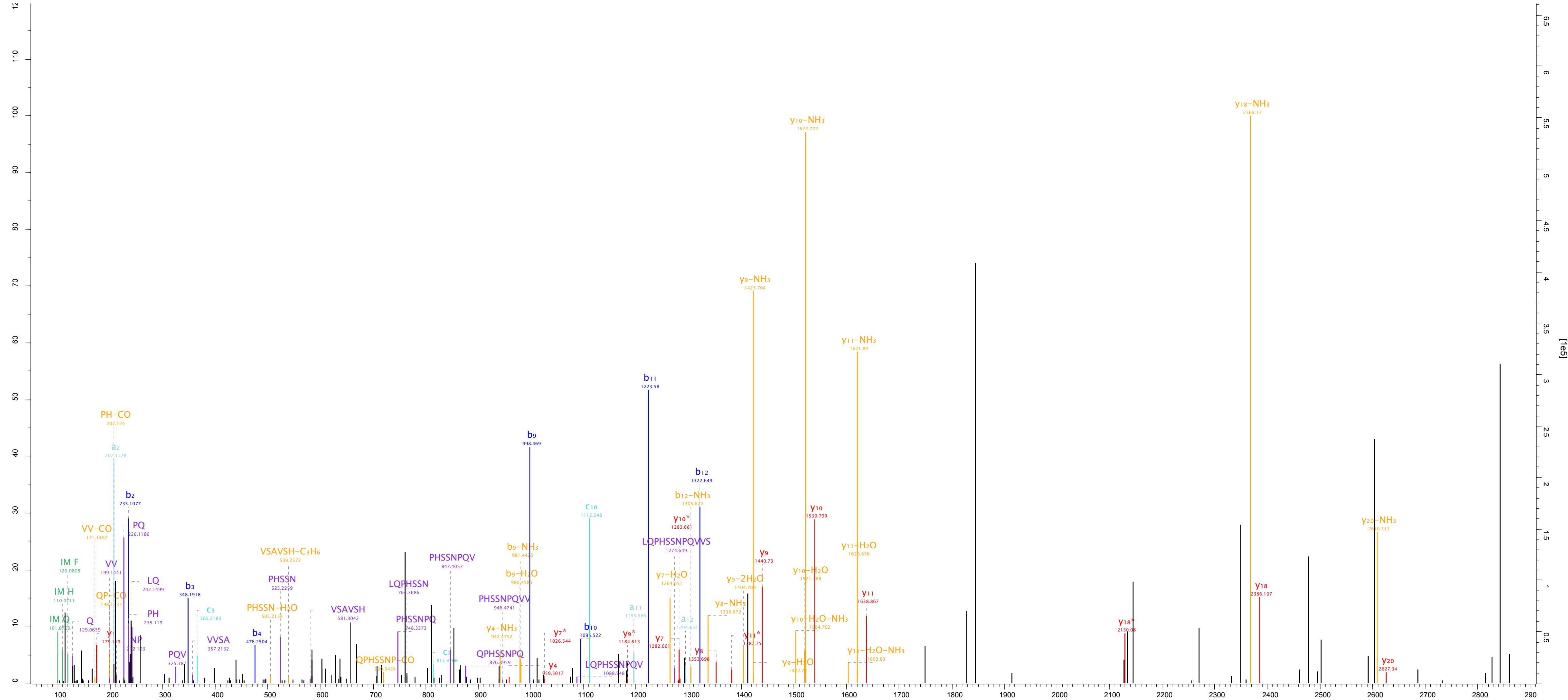
Standard: POLH-K619-SUMO



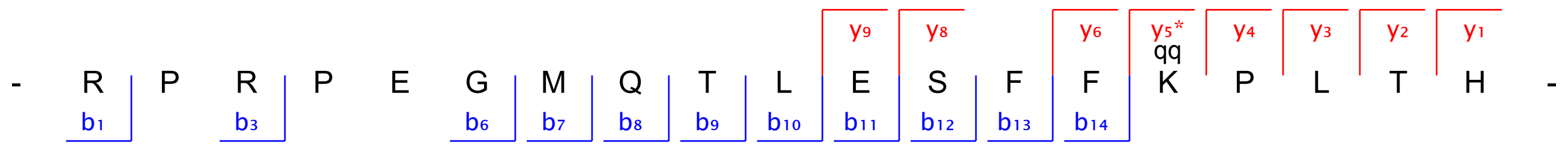
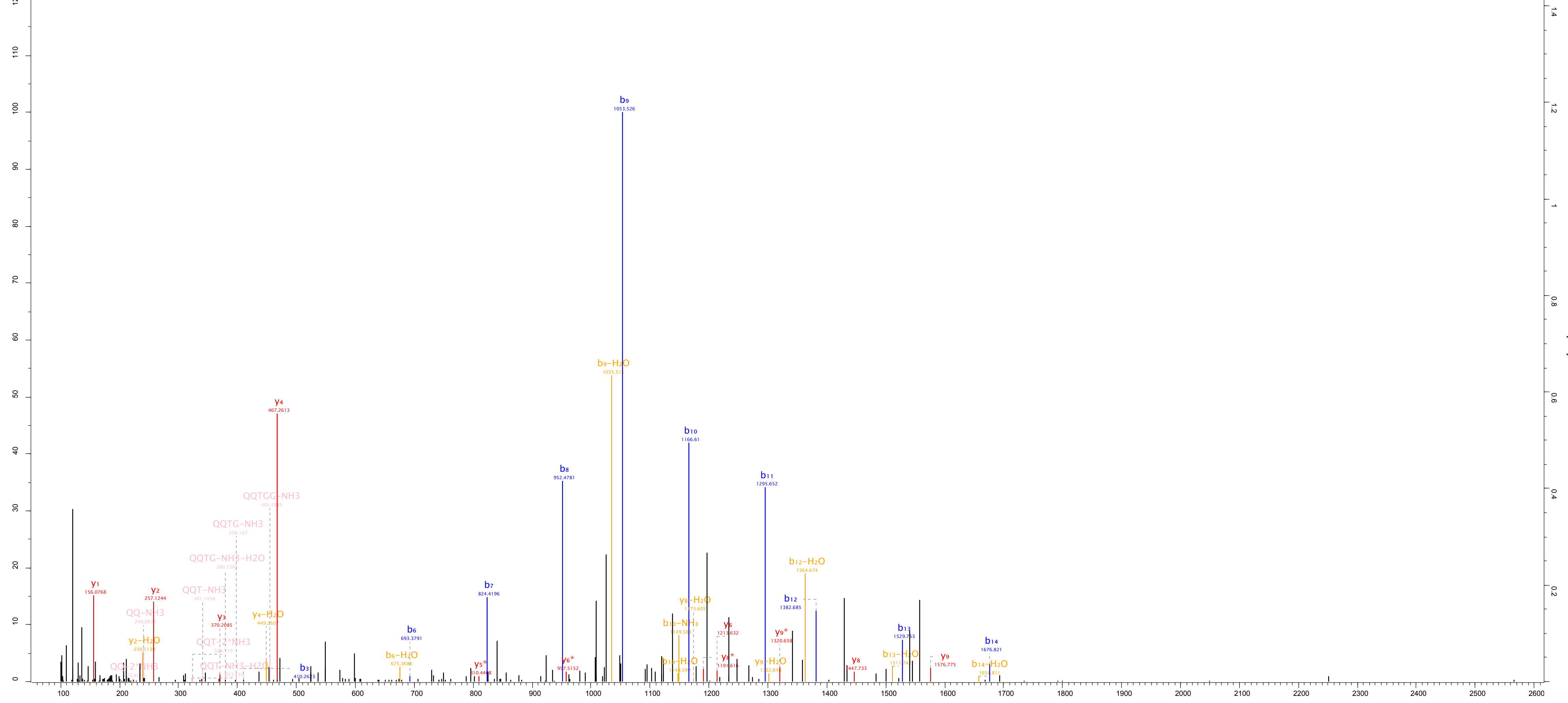
Advanced: POLH-K619-SUMO



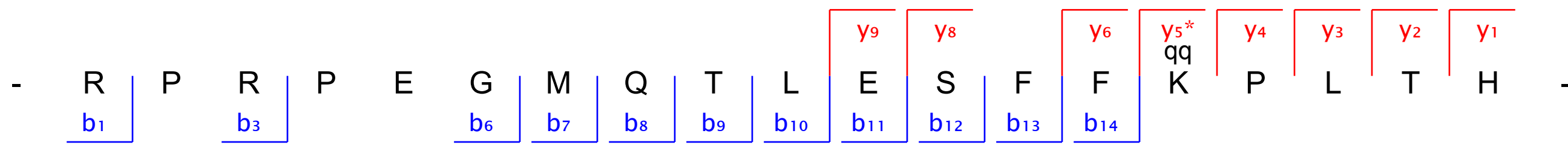
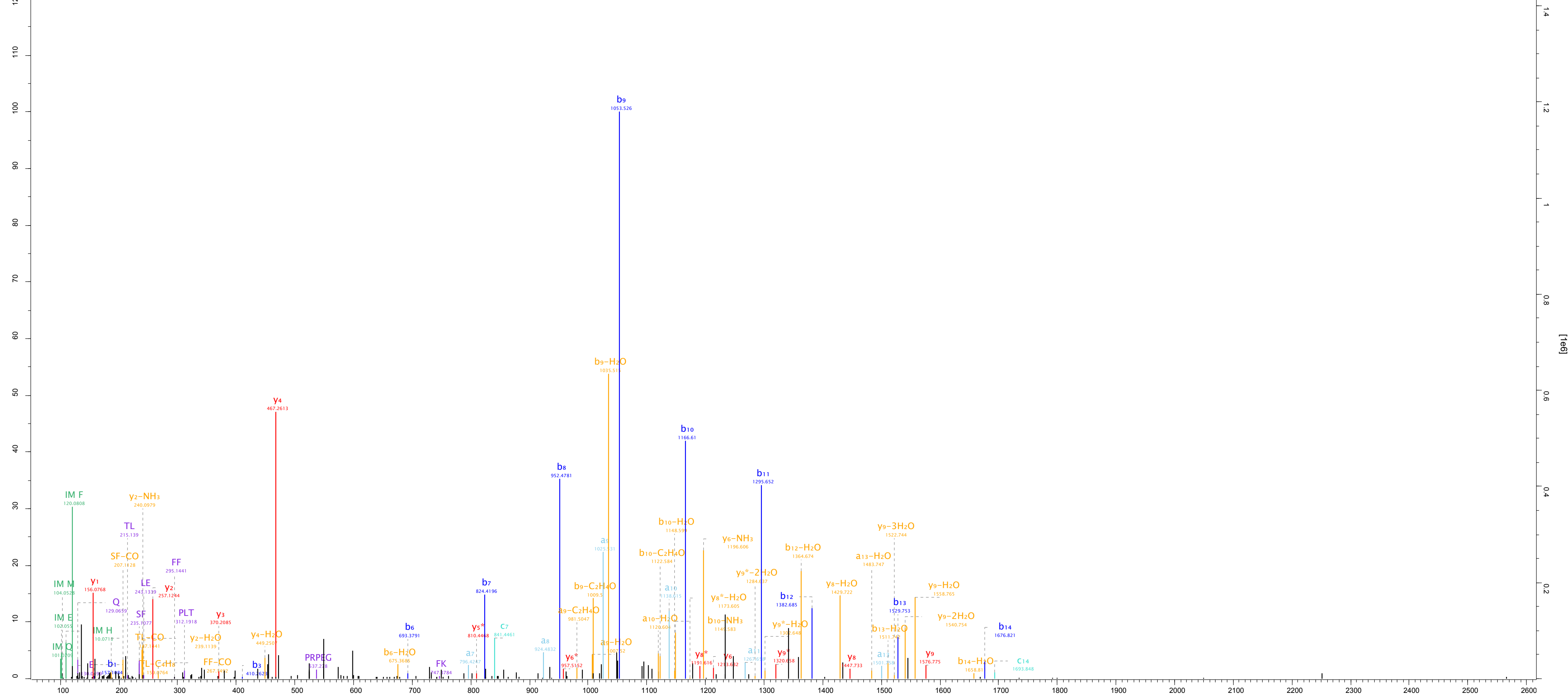
Standard: POLH-K682-SUMO



Advanced: POLH-K682-SUMO



Standard: POLH-K709-SUMO



Advanced: POLH-K709-SUMO