

On the parameters used in finite element modeling of compound peripheral nerves

Nicole A Pelot¹, Christina E Behrend¹, and Warren M Grill^{1,2,3,4}

¹ Duke University
Department of Biomedical Engineering
Room 1427, Fitzpatrick CIEMAS
101 Science Drive
Campus Box 90281
Durham, NC 27708

² Duke University, Department of Electrical and Computer Engineering, Durham, NC, USA

³ Duke University, Department of Neurobiology, Durham, NC, USA

⁴ Duke University School of Medicine, Department of Neurosurgery, Durham, NC, USA

Email: warren.grill@duke.edu

Journal of Neural Engineering

Short title: Parameters in neural finite element modeling

Keywords: neural computational modeling; peripheral nerve stimulation; finite element method; neuroengineering; neural engineering; vagus nerve stimulation

Supplement A – Literature Review of Perineurium Representations in Computational Models

Table 4. Compilation of implementations of perineurium in computational models, including sheet resistance, thickness, and resistivity, as applicable. The values that are underlined and italicized were calculated from information in the publication. Implementation methods (A, B, C, D) are described in the text and in Table 2; note that methods A & C produce identical results and methods B & D produce identical results if correctly implemented, but methods A & C produce different results than B & D. Publications that used constant thickness for the perineurium necessarily used both constant sheet resistance and constant resistivity (implementation methods C and D, respectively). R_s : sheet resistance.

Publication	Implementation method (see Table 2)	R_s ($\Omega\text{-m}^2$)	Thickness (μm)	Resistivity ($\Omega\text{-m}$)	Reference/justification for chosen values
(Weerasuriya et al., 1984)	N/A	0.0478	--	--	Impedance measurements of perineurium from frog sciatic nerve.
(Veltink et al., 1989)	D	--	Could estimate from Fig 2 & 3	100	Perineurial conductivity (0.01 S/m \rightarrow 100 $\Omega\text{-m}$) set to a value lower than their epineurial conductivity (0.1 S/m \rightarrow 10 $\Omega\text{-m}$), which was chosen to be higher than the conductivity of fat (0.04 S/m \rightarrow 25 $\Omega\text{-m}$).
(Meier et al., 1992)	A & B, but assumed A	0.0005	10	50	<p>“The conductance of the epineurium of frog sciatic nerve was measured by Weerasuriya et al. who found $\sigma_s = 2000$ S/m² (Weerasuriya et al. 1984)^a. They did not report the thickness of the epineurium but assuming a value of 10 μm, one calculates a specific resistance of 5000 $\Omega\text{-m}$.^b An epineurial sheath with this resistance will almost act as an insulator which is in contrast to the finding of Tasaki who found in his measurements of σ_p [the conductivity of nerve bundle in radial direction] that the resistance of the epineurium could almost be neglected (Tasaki 1964). The present authors used mostly $\sigma_s = 2000$ S/m², assuming a sheath thickness of 10 μm and a specific resistance of 50 $\Omega\text{-m}$, which is about twice the specific resistance of fat (Geddes and Baker 1967).”</p> <p>^a This is incorrect. Weerasuriya et al. found a conductance of 20 S/m².</p> <p>^b See footnote a. Using the correct Weerasuriya value of $\sigma_s = 20$ S/m² yields 5000 $\Omega\text{-m}$ (with thk = 10 μm), as stated here. But in the last sentence of the paragraph, they state that they used 50 $\Omega\text{-m}$, which results from their incorrectly stated value of $\sigma_s = 2000$ S/m².</p>
(Goodall et al., 1995)	C & D, but assumed C	<u>0.0149</u>	50	298	“T. Frieswijk, personal communication (calculated from Weerasuriya 1989 [sic]). In the model, a perineurium thickness of 50 μm was used. This was equivalent to a 35 μm perineurium with a conductivity of 0.0026 S/m [385 $\Omega\text{-m}$].”
(Koole et al., 1997)	D (& maybe C)	--	Could estimate from Fig 2	1190 (?)	“The conductivity of the perineurium was derived from Weerasuriya et al. (Weerasuriya et al. 1984), assuming a 40 μm thickness of the perineurium in their experiments. It has been suggested that the thickness of the perineurium roughly equals 5% of the diameter of a fascicle (Sunderland 1978, p. 41). The thickness of the modeled perineurium was too large in comparison with the diameters of the fascicles. Therefore, the conductivity of the perineurium, perpendicular to its surface, was increased by a correction factor. The conductivity of the perineurium, parallel to its surface, was reduced by the same factor.” Their implementation of the correction factor was unclear, especially since a single conductivity value was provided for the perineurium (i.e. modeled as isotropic tissue).
(Struijk, 1997)	C & D	<u>0.0149</u>	50	294	Goodall 1995 (Goodall et al., 1995)
(Frieswijk et al., 1998)	A?	0.011			<p>“The perineurium sheath impedance as of frog sciatic nerve is presented, in the form of an impedance locus, by Weerasuriya et al. (1984). It has a DC value of 20 S/m² and an absolute value of 45 S/m² at 10 kHz. The latter value is best suited for our model, as our stimulus is a 100 μs duration rectangular pulse, but it has to be corrected to account for the temperature difference between frog and rat, as follows: Whereas Weerasuriya et al. (1984) took measurements at a temperature of 21°C the body temperature in the rat is 37°C. By scaling with the Q_{10} factor one can account for this difference:</p> <p>$\sigma_s (37^\circ\text{C}) = \sigma_s (21^\circ\text{C}) * Q_{10}^{(37-21)/10}$</p> <p>Bostock (1983) presents $Q_{10} = 1.5$, yielding an increase to $\sigma_s = 90$ S/m².”</p>
(Deurloo et al., 1998)	C & D	<u>0.0149</u>	50	298	Frieswijk 1998 (Frieswijk et al., 1998)
(Perez-Orive and Durand, 2000)	C & D	<u>0.0398</u>	25	1592	No ref
(Rahal et al., 2000)	C & D	<u>0.0149</u>	50	294	Struijk 1997 (Struijk, 1997)

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(Choi et al., 2001)	C & D, but assumed D	<u>0.0143</u>	30	478	“The perineurium resistance was obtained from frog experiments (Weerasuriya 1984) with the assumption that the perineurium thickness in those experiments was 100 μm .” Note that the Weerasuriya perineurium thickness was assumed to be 100 μm , but then the perineurium was modeled with a thickness of 30 μm .
(Deurloo et al., 2003)	C & D	<u>0.0149</u>	50	298	Deurloo 1998 (Deurloo et al., 1998) “Although the thickness of the perineurium roughly equals 5% of the diameter of the fascicle (Sunderland 1978), the perineurium layers of all fascicles were given a thickness of 50 μm , according to the minimum grid size in the model.”
(Hennings et al., 2005)	C & D	<u>0.0149</u>	50	294	Vučković 2003 (2005?) (Vučković et al., 2005)
(Vučković et al., 2005)	C & D	<u>0.0149</u>	50	294	No ref
(Yoo and Durand, 2005)	C & D	<u>0.0238</u>	50	476	Choi 2001 (Choi et al., 2001)
(Grinberg et al., 2008)	D	--	From data: 3% * d_{fasc} Also evaluated: 0, 3, 15, 30, 50 μm 0 to 15% * d_{fasc}	476 Also evaluated 0.5 to 5000	Evaluated activation thresholds in response to different perineurium thicknesses and conductivities. Sunderland 1978 (Sunderland and Bradley, 1952) Choi 2001 (Choi et al., 2001) Other references listed for range of conductivities.
(Schiefer et al., 2008)	D	--	3% * d_{fasc}	500	Grinberg 2008 (Grinberg et al., 2008) Choi 2001 (Choi et al., 2001)
(Kent and Grill, 2013)	D	--	3% * d_{fasc}	1205	Grinberg 2008 (Grinberg et al., 2008) Weerasuriya 1984 (Weerasuriya et al., 1984) Calculation of chosen conductivity value unclear.
(Sabetian et al., 2017b)	C & D	<u>0.012</u>	25	478	Yoo 2005 (Yoo and Durand, 2005) Choi 2001 (Choi et al., 2001)
(Sabetian et al., 2017a)	C & D	<u>0.012</u>	25	478	Sabetian 2017b (Sabetian et al., 2017b)
(Raspopovic et al., 2017)	D	--	From segmentation of histology (?)	1136	Weerasuriya 1984 (Weerasuriya et al., 1984), assuming $\text{thk}_{\text{peri}} = 3\% * d_{\text{fasc}}$ and corrected from 21 to 37°C using $Q_{10} = 1.5$ See “Determination of Electrical Parameters: Correction of the Perineurium Conductivity” in the publication
(Pelot et al., 2017)	C	0.05	--	--	Weerasuriya 1984 (Weerasuriya et al., 1984)
(Elder and Yoo, 2018)	C & D	<u>0.024</u>	50	478	Yoo 2005 (Yoo and Durand, 2005)

Supplement B – Validation of Contact Impedance Boundary Condition

We used COMSOL's contact impedance boundary condition to model the perineurium in the 3D nerve models and to model the axon membranes in the 2D fascicle to estimate the bulk transverse endoneurial resistivity. For the former application, we compared thresholds when using a thin meshed perineurium to the perineurial boundary condition (see Figure 4 in the main text). For the latter application, we performed a simple validation of the contact impedance boundary condition used to model the axonal membranes. We compared $\rho_{endo-bulk-transverse}$ obtained by modeling the axon membrane as a physical annulus around the intracellular space (Figure 9(a)) and as a contact impedance (Figure 9(b)). To generate results within machine precision, i.e. to avoid numerical instabilities, the ratio of the model's largest to smallest resistivities must be less than six orders of magnitude. Since our model's smallest resistivity is $0.65 \Omega\text{-m}$ (for $\rho_{endo-micro}$) and the specific membrane resistance is $0.2 \Omega\text{-m}^2$, we set the membrane thickness to $1 \mu\text{m}$. Thus, the membrane resistivity was $2 \times 10^5 \Omega\text{-m}$, leading to $\rho_{max}/\rho_{min}=3 \times 10^5$, which is within machine precision. We placed 22 oversized axons ($10 \mu\text{m}$ diameter, plus membrane thickness) in a grid within a $105 \mu\text{m}$ fascicle with $3 \mu\text{m}$ between neighbouring axons. Since our results show that no current enters the axons, we used $12 \mu\text{m}$ diameter axons when using the contact impedance in order to encompass the same area as the $10 \mu\text{m}$ axons surrounded by $1 \mu\text{m}$ thick annuli. Both models yielded $\rho_{endo-bulk-transverse}=1.14 \Omega\text{-m}$ (see section 2.4.2), thereby validating the boundary condition.

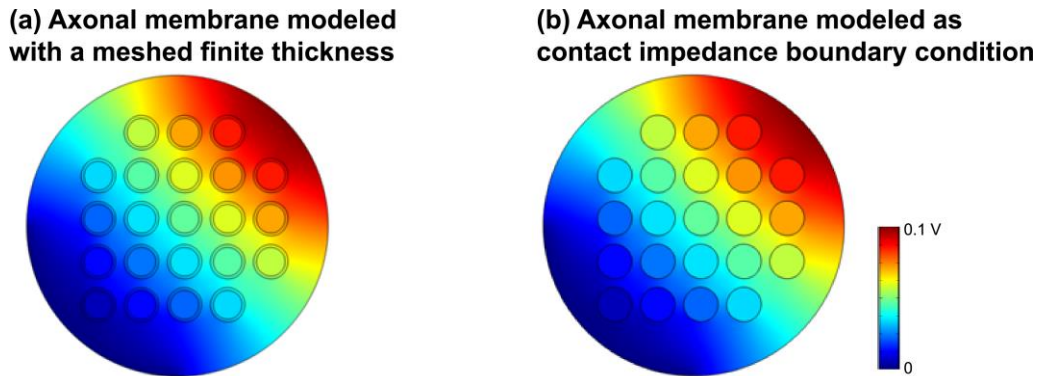


Figure 9. Potential distributions used to validate COMSOL's contact impedance boundary condition. Twenty one axons were placed in a grid within a $105 \mu\text{m}$ fascicle. We compared $\rho_{endo-bulk-transverse}$ resulting from modeling the axonal membranes as annuli with finite thickness (a) or as contact impedances (b).

Supplement C – Axonal Area Fraction Calculation

Table 5 shows calculations for estimating the axonal area fraction (*AAF*) for a single fascicle based on cat posterior abdominal vagus nerve morphology.

Table 5. Estimation of axonal area fraction in a single abdominal vagus nerve fascicle based upon cat data.

Number of fibres in cat abdominal vagus nerve (Agostini, 1957)	31,244
Total cross-sectional area of axons (Agostini, 1957; Mei et al., 1980)	$\left(\frac{31,244 \text{ fibers}}{\text{nerve}}\right) (\pi) \left(\frac{1 \mu\text{m}}{2}\right)^2 = 2.4539 \times 10^4 \mu\text{m}^2$
Fraction of epineurium in nerve diameter (Altman and Plonsey, 1989)	0.3
Diameter of cat posterior abdominal vagus nerve (Agostini, 1957)	376 μm
Cross-sectional area of nerve, neglecting epineurium (Agostini, 1957)	$(\pi) \left(\frac{(1 - 0.3) * 376 \mu\text{m}}{2}\right)^2 = 5.4437 \times 10^4 \mu\text{m}^2$
Axonal area fraction (<i>AAF</i>)	$(2.4539 \times 10^4 \mu\text{m}^2) / (5.4437 \times 10^4 \mu\text{m}^2) = \mathbf{0.45 = 45\%}$

Supplement D – Effects of Representation of the Perineurium on Thresholds

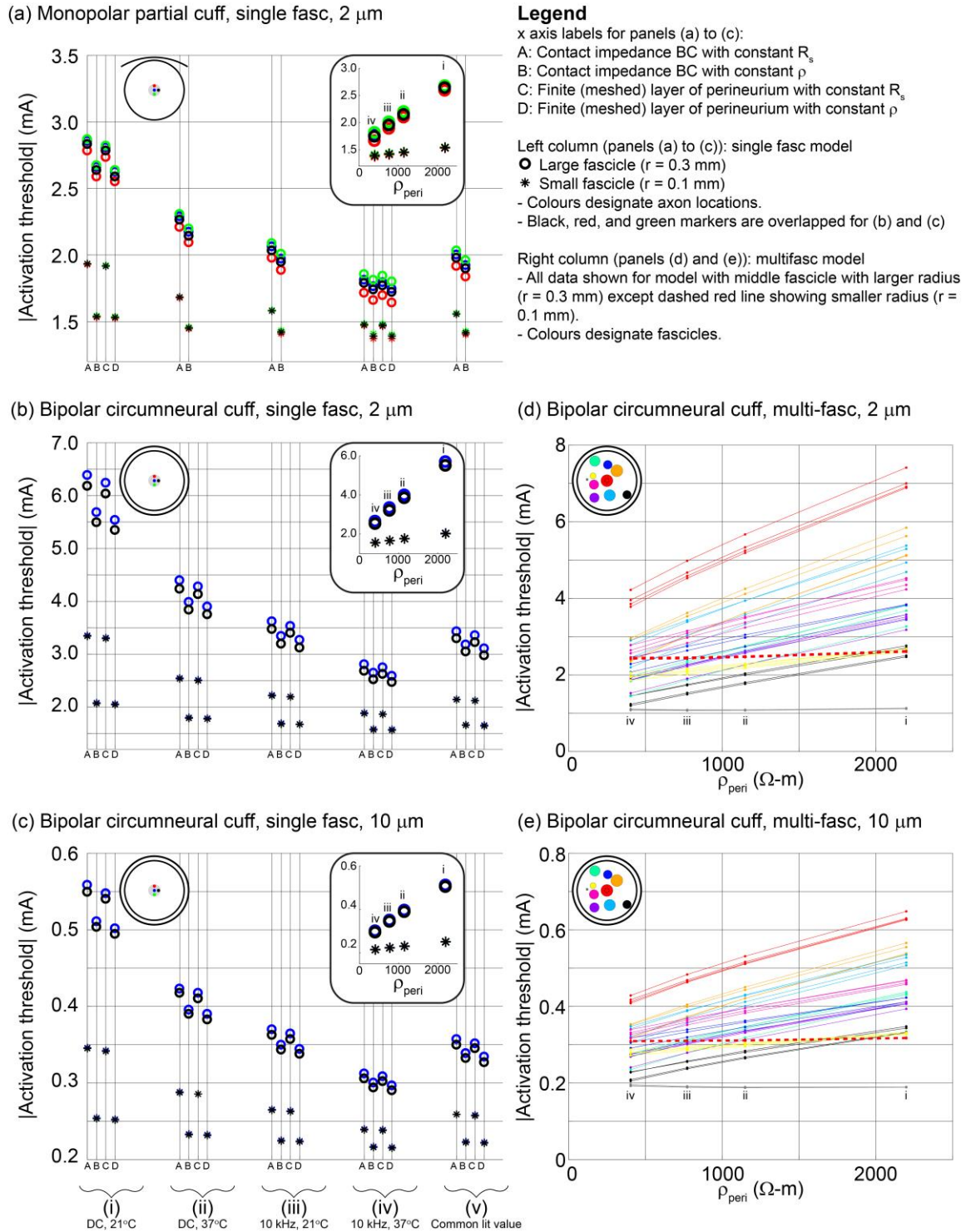


Figure 10. Activation thresholds with different representations of the perineurium (x axis labels A to D), estimates of perineurium resistivity (i to v; see Table 2), nerve models (single fascicle and multifascicular models), electrode designs (monopolar partial cuff and bipolar circumneural cuff), and fibre diameters (2 and 10 μm axons). The insets, as well as panels (d) and (e), show Method B data (constant ρ). Axon locations: centred (x_0, y_0), moved up ($x_0, y_0 + 0.75 * r_{\text{fasc}}$), moved down ($x_0, y_0 - 0.75 * r_{\text{fasc}}$), and translated laterally ($x_0 + 0.75 * r_{\text{fasc}}, y_0$). In panels (b) and (c), showing activation thresholds with the bipolar circumneural cuff, the asterisks (small fascicle thresholds) for all four axon locations (different colours) are overlapped; also, given the model's symmetry, the black, red, and green circular markers (large fascicle thresholds) are overlapped. The nerve and fascicle legends are to scale.

Supplement E – Current Density in $\rho_{\text{endo-bulk-transverse}}$ Modeling

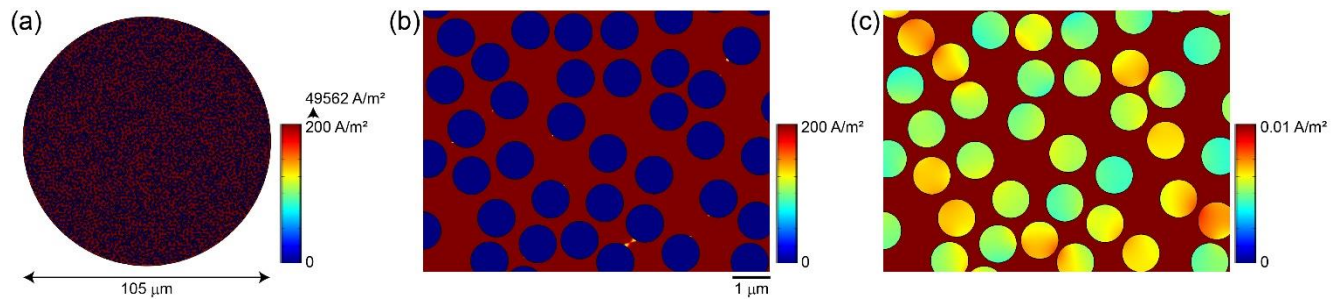


Figure 11. Magnitude of the current density showing that very little current enters the axons. Panels (b) and (c) (different colour bar bounds) are zoomed in on the centre of the entire fascicle shown in panel (a).

Supplement F – Sweeping Endoneurial Resistivity

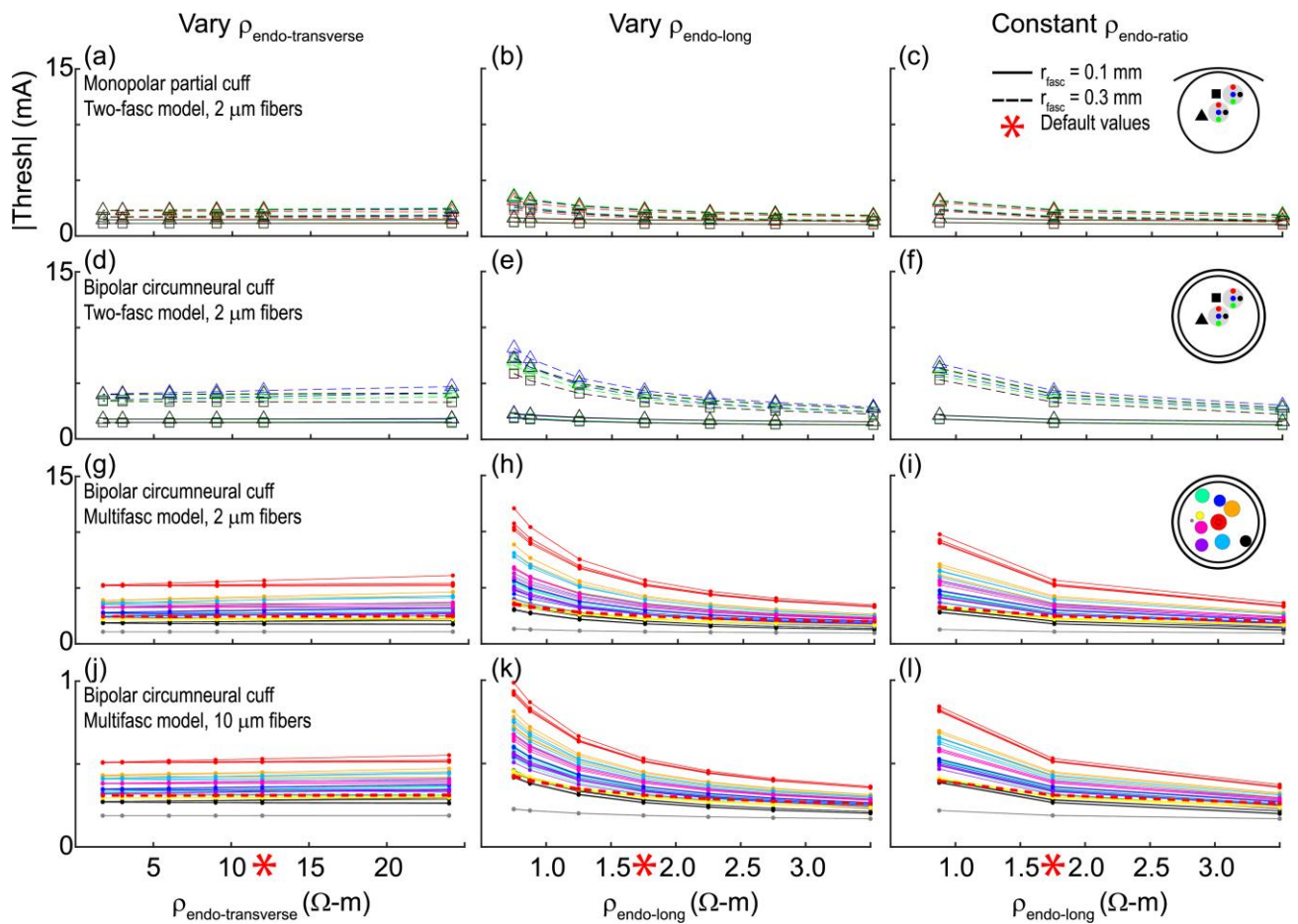


Figure 12. Activation thresholds for axons in a 3D FEM of nerve and cuff electrode across different values of endoneurial resistivity (see illustration of methods in Figure 2). The default resistivities (red asterisks) were 12 $\Omega\text{-m}$ for $\rho_{\text{endo-transverse}}$ and 1.75 $\Omega\text{-m}$ for $\rho_{\text{endo-long}}$. In the last column, the ratio of the transverse resistivity to the longitudinal resistivity was constant at 12 $\Omega\text{-m}/1.75 \Omega\text{-m} = 6.9$. All models used $\rho_{\text{peri}} = 1149 \Omega\text{-m}$ (DC, 37°C). Similar results were found for the monopolar partial cuff with $\rho_{\text{peri}} = 2198 \Omega\text{-m}$ (DC, 21°C) (data not shown). First and second rows: Thresholds for 2 μm axons for the nerve model with two fascicles for different cuff electrode geometries. The colours designate the four axon locations per fascicle, as shown in the legends in the last column (nerve cross sections not to scale). Third and fourth rows: Thresholds for 2 and 10 μm axons for the nerve model with 10 fascicles and the bipolar circumneural cuff geometry. The fascicle colours are shown in the legend in the last column (fascicles and nerve drawn to scale); the thresholds for four axons per fascicle are plotted in the same colour. The thresholds are plotted for the simulations where the centre fascicle (red) had a radius of 0.3 mm. The thresholds for axons in the centre fascicle when its radius was reduced ($r = 0.1 \text{ mm}$; dashed black circle in the legend) are plotted with the dashed red lines; the thresholds for axons in other fascicles changed less than 4% between the models with the larger and the smaller centre fascicle.

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