

## Supporting Information

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Amine Core (#)	Name	Structure
1	3-[4-(3-([2-(4-[3-[4-(3-aminopropyl)piperazin-1-yl]propyl)piperazin-1-yl)ethyl]amino)propyl)piperazin-1-yl]propan-1-amine	
2	{2-[2-(2-aminoethoxy)ethoxy]ethyl}[2-(4-[2-[2-(2-aminoethoxy)ethoxy]ethyl)piperazin-1-yl]ethyl]amine	
3	10-(4-[2-[(10-aminodecyl)amino]ethyl]piperazin-1-yl)decan-1-amine	
4	2-[2-[4-(2-[2-(2-aminoethoxy)ethyl]amino)ethyl]piperazin-1-yl]ethoxy]ethan-1-amine	
5	1-[4-(3-[2-[2-(3-aminopropoxy)ethoxy]ethoxy]propyl)piperazin-1-yl]-7,10,13-trioxa-3-azahexadecan-16-amine	
6	3-(4-[2-[(3-amino-2-ethoxypropyl)amino]ethyl]piperazin-1-yl)-2-ethoxypropan-1-amine	
7	[1-([2-(4-[1 (aminomethyl) cyclohexyl]methyl)piperazin-1-yl) ethyl] amino)methyl]cyclohexyl]methanamine	
8	1-N-[2-[4-(4-aminocyclohexyl)piperazin-1-yl]ethyl]cyclohexane-1,4-diamine	

Table S1. Amine Cores. The named structures of the amine cores used to generate the ionizable lipid library.

<b>Formulation</b>	<b>Diameter (nm)</b>	<b>PDI</b>	<b>Luciferase mRNA (ng/uL)</b>
<b>C12-1</b>	68.85 ± 1.07	.094 ± .007	40.25 ± 0.07
<b>C12-2</b>	82.92 ± 1.21	.238 ± .006	38.65 ± 0.78
<b>C12-3</b>	51.22 ± 0.66	.158 ± .013	45.8 ± 0.28
<b>C12-4</b>	51.05 ± 2.75	.219 ± .015	36.25 ± 1.20
<b>C12-5</b>	65.6 ± 1.13	.203 ± .015	37.05 ± 0.21
<b>C12-6</b>	91.79 ± 1.17	.115 ± .018	39.8 ± 1.13
<b>C12-7</b>	64.73 ± 2.88	.219 ± .021	37.55 ± 0.07
<b>C12-8</b>	70.25 ± 0.75	.086 ± .003	47.55 ± 0.49
<b>C14-1</b>	73.15 ± 2.23	.176 ± .052	34.3 ± 0.85
<b>C14-2</b>	75.61 ± 1.21	.179 ± .136	35.25 ± 0.35
<b>C14-3</b>	74.09 ± 0.35	.238 ± .009	40.4 ± 1.13
<b>C14-4</b>	70.17 ± 0.41	.176 ± .006	35.6 ± 2.12
<b>C14-5</b>	64.57 ± 3.10	.232 ± .007	34.25 ± 0.49
<b>C14-6</b>	64.71 ± 1.73	.129 ± .020	36.8 ± 2.12
<b>C14-7</b>	67.14 ± 1.81	.175 ± .027	39.7 ± 1.56
<b>C14-8</b>	65.44 ± 0.51	.091 ± .004	40.3 ± 2.40
<b>C16-1</b>	83.82 ± 3.16	.029 ± .014	46.85 ± 0.50
<b>C16-2</b>	97.01 ± 2.57	.14 ± .023	36.1 ± 0.99
<b>C16-3</b>	82.72 ± 2.32	.113 ± .010	40.3 ± 1.84
<b>C16-4</b>	68.12 ± 1.41	.211 ± .009	38 ± 1.27
<b>C16-5</b>	75.82 ± 0.91	.284 ± .029	35.95 ± 0.64
<b>C16-6</b>	96.03 ± 1.95	.053 ± .031	37.35 ± 0.21
<b>C16-7</b>	89.46 ± 0.59	.188 ± .009	33.3 ± 4.67
<b>C16-8</b>	87.7 ± 0.33	.101 ± .004	48.3 ± 0.85

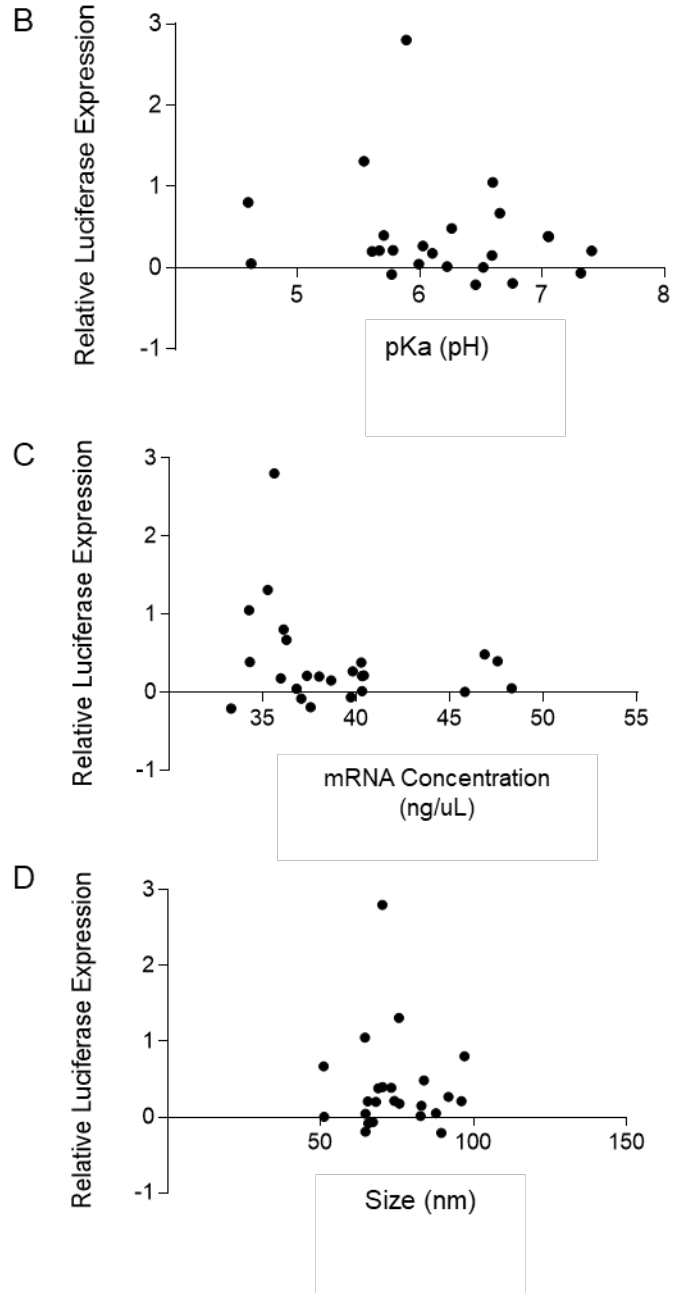
Table S2: Library Characterization. The diameter (z-average), PDI, and mRNA concentration of each LNP formulation showing a narrow range in LNP size, monodispersity, and similar mRNA loading across LNP formulations. (± standard deviation)

<b>C14-4 LNP (luciferase mRNA)</b>	<b>Crude</b>	<b>Purified</b>
mRNA (ng/uL) $\pm$ St. Dev.	35.6 $\pm$ 2.12	29.8 $\pm$ 2.31
Encapsulation Efficiency (%)	92.53	86.3
Diameter (nm) $\pm$ St. Dev.	70.17 $\pm$ 0.41	65.19 $\pm$ 0.83
Polydispersity $\pm$ St. Dev.	.176 $\pm$ .006	.189 $\pm$ .014
pKa	6.143	6.505

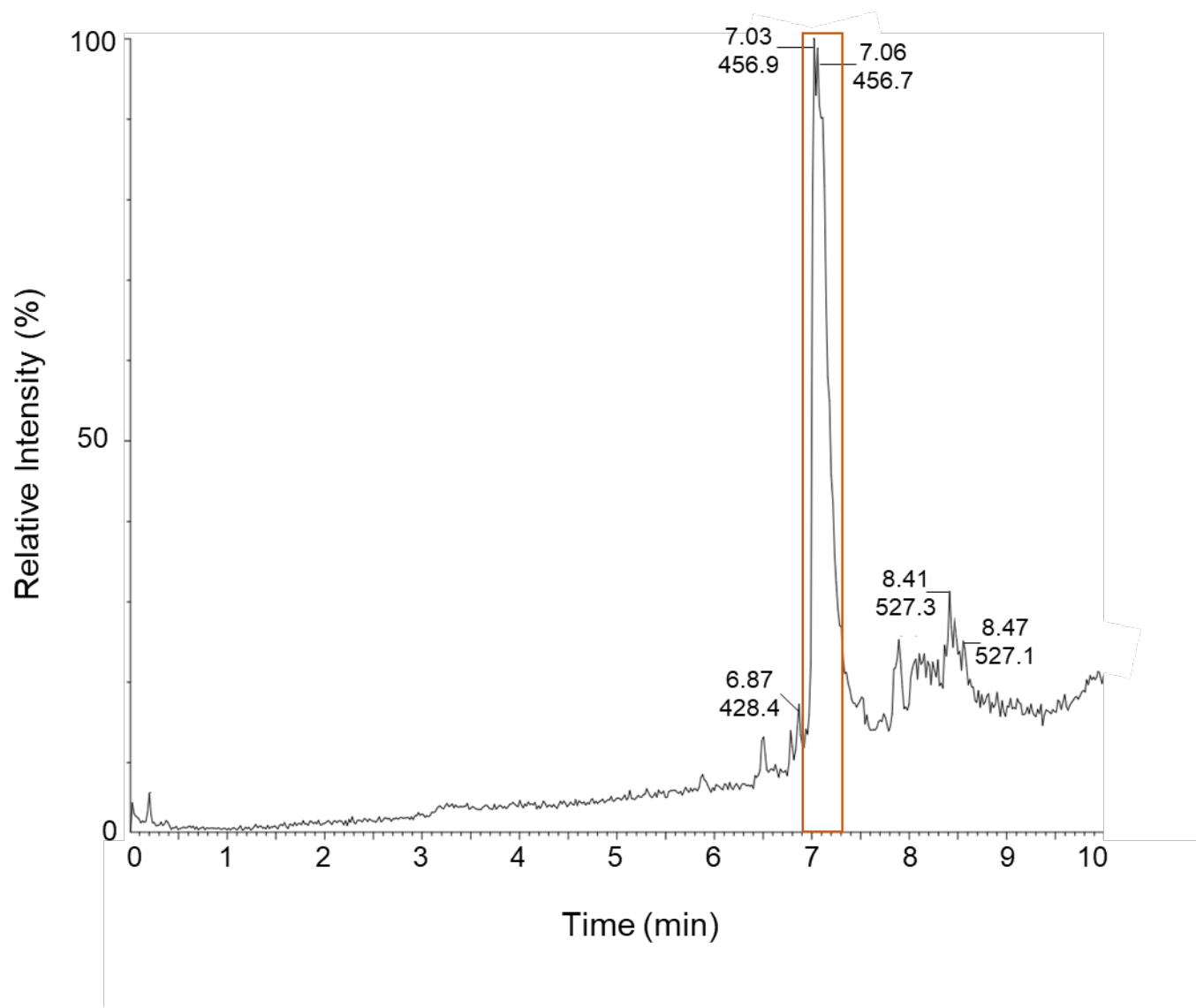
Table S3: Crude and Purified C14-4 LNPs. Comparison of characteristics of crude and purified C14-4 LNPs encapsulating luciferase mRNA.

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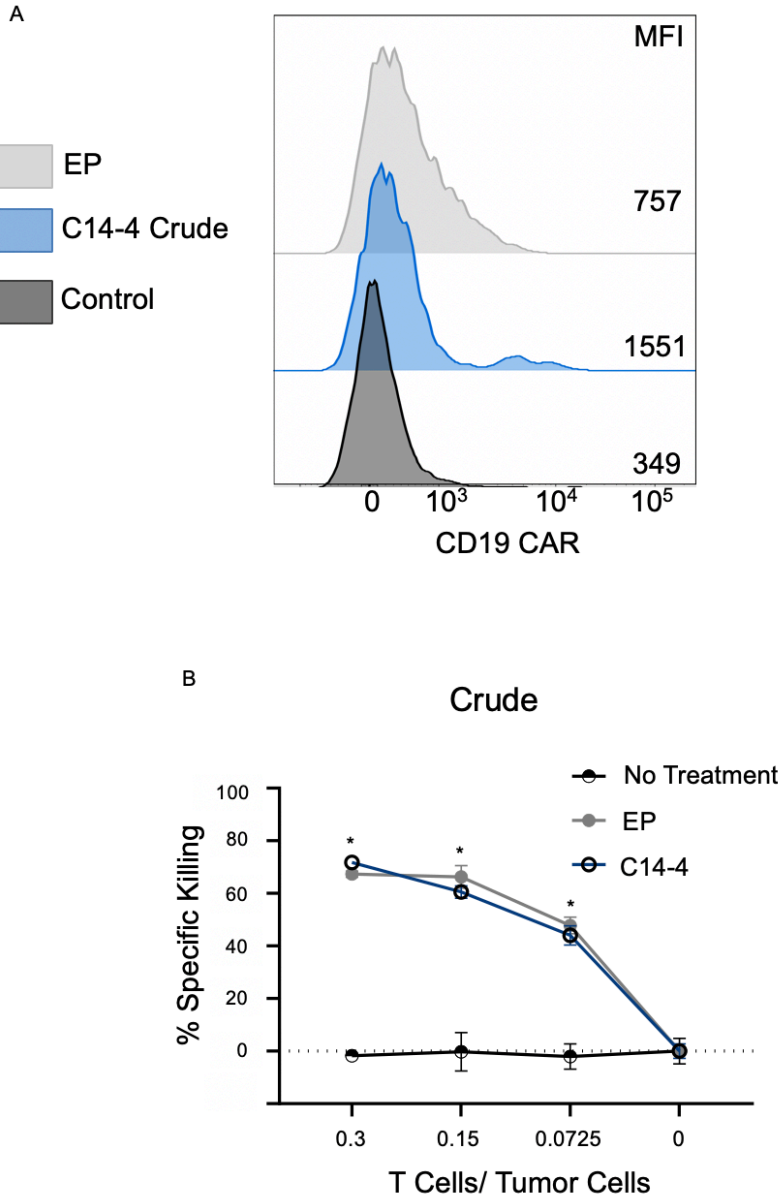
Lipid	pKa
C12-1	7.054
C12-2	6.589
C12-3	6.52
C12-4	6.655
C12-5	5.771
C12-6	6.026
C12-7	6.758
C12-8	5.706
C14-1	7.049
C14-2	5.543
C14-3	5.782
C14-4	5.889
C14-5	6.596
C14-6	5.993
C14-7	7.317
C14-8	7.406
C16-1	6.261
C16-2	4.597
C16-3	6.2228
C16-4	5.61
C16-5	6.103
C16-6	5.671
C16-7	6.457
C16-8	4.622



**Figure S1.** (A) Table of the polyamine cores and their pKa values as determined via TNS assay. (B-D) The relative luciferase expression produced by each LNP formulation (at 30 ng/ 60,000 Jurkat cells) graphed against the polyamine core pKa, LNP mRNA concentration, and LNP size, respectively. No correlations between these factors were found.



**Figure S2.** LCMS spectra of the crude C14-4 product with the fully saturated amine core highlighted with in the orange box.



**Figure S3.** (A) Surface expression of CAR on primary T cells assessed using flow cytometry (B) Results of Nalm6 and CAR T cell co-plating at different effector-to-target ratios for 48 hrs. n=3 wells. \* = p<.0001 in paired t test to No Treatment (for both EP and C14-4 LNPs)