

# Supplementary Material

## Supplementary Tables

**Supplementary Table S1.** Recall and precision values for each broad phenotype category by submissions

		Recall	Precision
SID#1	Eye	0.33	0.50
	Neuro	0.43	0.43
	Connective	0.27	0.50
SID#2	Eye	0.17	0.17
	Neuro	0.00	0.00
	Connective	0.18	0.50
SID#3	Eye	0.17	0.14
	Neuro	0.29	0.50
	Connective	0.55	0.46
SID#4	Eye	0.50	0.50
	Neuro	0.43	0.43
	Connective	0.55	0.55
SID#5	Eye	0.50	0.38
	Neuro	0.43	0.33
	Connective	0.45	0.50
SID#6.1	Eye	0.00	0.00
	Neuro	0.14	0.14
	Connective	0.55	0.46
SID#6.2	Eye	0.00	0.00
	Neuro	0.14	0.50
	Connective	0.91	0.43
SID#7	Eye	0.33	0.33
	Neuro	0.43	0.33
	Connective	0.27	0.60
SID#8	Eye	0.50	0.43
	Neuro	0.29	0.33
	Connective	0.55	0.55

**Supplementary Table S2.** Number of gender matches for each submission

Submission ID	Gender match	Number of genomes not matched to any patient
SID#1	23	0
SID#2	<b>12</b>	12
SID#3	<b>24</b>	0
SID#4	<b>24</b>	0
SID#5	<b>12</b>	12
SID#6.1	23	0
SID#6.2	17	0
SID#7	18	3
SID#8	<b>24</b>	0

**Supplementary Table S3. Benign nominated diagnostic variants predicted with the highest probability for correct genome-patient matches**

Genome (patient)	Phenotype category	SID #	Genomic position (hg19)	Gene	Plausible gene for category?	Gene Phenotype Correlation	Transcript	Nucleotide change	Protein change	Classification	Gene disease associations
7(X)	Ophthalmologic	7	9:80647059:GC:TT	<i>GNAQ</i>	Yes	Poor	NM_002072.4	c.-908_909GC>TT	5'UTR	Benign	Somatic mutations cause Sturge Weber syndrome
		8	2:216251538:G:A	<i>FN1</i>	No	Poor	NM_212482.2	c.4486C>T	p.Arg1496Trp	Likely Benign	Glomerulopathy; Fibronectin deficiency; Spondylometaphyseal dysplasia
17(H)	Ophthalmologic	4	1:94568686:C:T	<i>ABCA4</i>	Yes	Good	NM_000350.2	c.455G>A	p.Arg152Gln	Likely Benign	Retinitis pigmentosa, red-cone dystrophy and other eye disorder (AR inheritance)
			1:94470320:C:T				NM_000350.2	c.6147+677G>A	Intronic	Likely Benign	
21(G)	Neurologic	6.1	12:7343108:GGCC TCTGAGGCAGTGA GTGTTCTTGAGGT GGAAAGCCAGGT GCA:G	<i>PEX5</i>	Yes	Poor Partial	NM_001131023 .1	c.192+32_192+76 del	None (3 repeats to 2)	Benign	Peroxisome biogenesis disorders, Rhizomelic chondrodysplasia punctata
42(O)	Ophthalmologic	3	16:1621518:GA:TT	<i>IFT140</i>	Yes	Good	NM_014714.4	c.1541_1542delTC insAA	p.Leu514Gln	Benign	Retinitis pigmentosa 80, Short-rib thoracic dysplasia 9 with or without polydactyly
56(N)	Connective	4	15:48700642:T:C	<i>FBN1</i>	Yes	Some	NM_000138.4	c.*2545A>G	3'UTR	Likely Benign	Marfan syndrome
			15:48849792:C:A				NM_000138.4	c.539-19787G>T	Deep intronic	Likely Benign	
		8	6:32012817:C:T	<i>TNXB</i>	Yes	Some	NM_019105.7	c.10887G>A	p.Lys3629=	Benign	Ehlers-Danlos like syndrome
68(J)	Neurological	8	11:108196896:C:T	<i>ATM</i>	Yes	Poor	NM_000051.3	c.6919C>T	p.Leu2307Ph e	Likely Benign	Ataxia telangiectasia
71(L)	Connective	1	*2:75811731:G:A	<i>EVA1A</i>	No	None	NM_032181.2	None	Intergenic	Likely Benign	None
78(V)	Connective	6.2	15:59443263:CGT GCACTT:C	<i>MYO1E</i>	No	None	NM_004998.3	c.3080+2518_308 0+2525delAAGTG CAC	Deep intronic	Likely Benign	Glomerulosclerosis, focal segmental 6
			1:40768483:T:TGG AG	<i>COL9A2</i>	Yes	Poor	NM_001852.4	c.1604-6_1604- 3dupCTCC	Intronic near splice	Benign	Multiple epiphyseal dysplasia 2

			6:70972993:T:C	<i>COL9A1</i>	Yes	Poor	NM_001851.4	c.1349A>G	p.Glu450Gly	Likely Benign	Stickler syndrome type IV, Epiphyseal dysplasia, multiple, 6
93(F)	Connective	8	12:2791205:A:G	<i>CACNA1C</i>	No	Poor	NM_001129830.2	c.5534A>G	p.Lys1845Arg	Benign	Brugada syndrome
95(C)	Ophthalmologic	4	1:215953583:A:G	<i>USH2A</i>	Yes	Good	NM_206933.2	c.10741-200T>C	Deep intronic	Likely Benign	Usher syndrome
			1:215964830:T:G				NM_206933.2	c.9959-1206A>C	Deep intronic	Likely Benign	
99(B)	Neurological	8	11:793588:G:A	<i>SLC25A22</i>	Yes	Mild	NM_001191061.1	c.234C>T	p.Pro78=	Benign	Epileptic encephalopathy, early infantile, 3
102(A)	Connective	8	2:189974958:G:T	<i>COL5A2</i>	Yes	Partial	NM_000393.4	c.315C>A	p.Thr105=	Benign	Ehlers-Danlos syndrome-Classic

\*Several systematic errors or differences in how variants were referenced for submission files for groups 1 and 7. All variants identified in genes on the minus strand were represented as the base of the minus strand while convention generally refers to only plus strand bases for genomic coordinates. Assumptions were made based on notes provided with submissions, gene orientation, and know variants at those loci. Assumed validated variants are listed here and do not match perfectly with original submission files.

**Supplementary Table S4. Nominated secondary variants from groups 1, 4, and 7.**

Genome	Genomic position (hg19)	Transcript	Nucleotide change	Protein change	Gene	Variant type	Classification	Disease inheritance pattern	Associated disease	Reporting suggestions
7	19:34262922:C:T	NM_001127896.1	c.229C>T	p.Arg77Trp	<i>CHST8</i>	missense	Pathogenic	AR	Peeling skin syndrome 3	do not report as secondary, only carrier status
7	1:76226846:A:G	NM_001127328.2	c.997A>G	p.Lys333Glu	<i>ACADM</i>	missense	Pathogenic	AR	Acyl-CoA dehydrogenase, medium chain, deficiency of	do not report as secondary, only carrier status
7	12:88454737:T:C	NM_025114.3	c.6392A>G	p.Glu2131Gly	<i>CEP290</i>	missense	VUS	AR	Senior-Loken syndrome 6; Joubert syndrome 5; Leber congenital amaurosis 10; Meckel syndrome 4	do not report as secondary
17	11:6411935:T:CGCTGGCGCTGGC	NM_000543.4	repeat variability	repeat variability	<i>SMPD1</i>	repeat variability	Benign	AR	Niemann-Pick disease, types A/B	do not report as secondary
17	14:88452941:A:G	NM_000153.4	c.334A>G	p.Thr112Ala	<i>GALC</i>	missense	VUS	AR	Krabbe disease	do not report as secondary
18	18:21134743:G:A	NM_000271.4	c.1532C>T	p.Thr511Met	<i>NPC1</i>	missense	VUS	AR	Niemann-Pick disease, types D/C1	do not report as secondary
18	18:21140367:G:A	NM_000271.4	c.709C>T	p.Pro237Ser	<i>NPC1</i>	missense	Benign	AR	Niemann-Pick disease, types D/C1	do not report as secondary
30	7:150644901:G:A	NM_000238.3	c.2758C>T	p.Arg920Trp	<i>KCNH2</i>	missense	VUS	AD	Long QT syndrome; Short QT syndrome	do not report as secondary
30	2:47707963:TATG:T	NM_000251.2	c.2590_25992 delATG	p.Asp864del	<i>MSH2</i>	in frame deletion	VUS	AD/AR	Mismatch repair cancer syndrome; Colorectal cancer, hereditary nonpolyposis, type 1; Muir-Torre syndrome	do not report as secondary
30	13:32911607:CCTA:C	NM_000059.3	c.3119_3121d elCTA	p.Thr1040del	<i>BRCA2</i>	in frame deletion	VUS	AD	Fanconi anemia, complementation group D1; Wilms tumor; other cancers	do not report as secondary
39	5:125887751:C:G	NM_001201377.1	c.1279G>C	p.Glu427Gln	<i>ALDH7A1</i>	missense	VLP	AR	Epilepsy, pyridoxine-dependent	do not report as secondary, only carrier status
42	2:46386838:G:C	NM_005400.2	c.2014G>C	p.Asp672His	<i>PRKCE</i>	missense	VUS	unknown	possible association with SHORT syndrome	do not report as secondary
42	*17:79767715:G:A	NM_000160.4	c.118G>A	p.Gly40Ser	<i>GCGR</i>	missense	VUS	AD	Diabetes mellitus, noninsulin-dependent	do not report as secondary
42	2:46325089:G:C	NM_005400.2	c.1592+11588 G>C	non coding	<i>PRKCE</i>	non coding	VUS	unknown	possible association with SHORT syndrome	do not report as secondary
56	16:17232391:G:A	NM_022166.3	c.1588-3C>T	splice site	<i>XYLT1</i>	splice	VLP	AR	Desbuquois dysplasia 2	do not report as secondary, only carrier status
56	4:15504459:T:G	NM_001080522.2	C.351T>G	p.Ser117Arg	<i>CC2D2A</i>	missense	VLB	AR	Joubert syndrome 9; COACH syndrome; Meckel syndrome 6	do not report as secondary

56	1:171083232:G:T	NM_006894.5	c.913G>T	p.Glu305Ter	<i>FMO3</i>	nonsense	Pathogenic	AR	Trimethylaminuria (fish odor smell)	do not report as secondary, only carrier status
68	21:35742947:T:C	NM_172201.1	c.170T>C	p.Ile57Thr	<i>KCNE2</i>	missense	VUS	AD	Long QT syndrome 6; Atrial fibrillation, familial, 4	do not report as secondary
68	7:143018525:C:G	NM_000083.2	c.501C>G	p.Phe167Leu	<i>CLCN1</i>	missense	VUS	AD/AR	Myotonia congenita; Myotonia levior	do not report as secondary
71	17:72916740:C:T	NM_173477.4	c.191G>A	p.Trp64Ter	<i>USH1G</i>	nonsense	Pathogenic	AR	Usher syndrome, type 1G	do not report as secondary, only carrier status
71	*7:117188841:TTG:	NM_000492.3	c.1360_1362delTTG	p.Leu454del	<i>CFTR</i>	in frame deletion	VLB	AR	Cystic fibrosis; Congenital bilateral absence of vas deferens; Sweat chloride elevation without CF	do not report as secondary
71	*15:45400303:C:T	NM_014080.4	c.1516G>A	p.Asp506Asn	<i>DUOX2</i>	missense	VUS	AR	Thyroid dysmorphogenesis 6	do not report as secondary
76	7:117227832:G:T	NM_000492.3	c.1624G>T	p.Gly542Ter(*)	<i>CFTR</i>	nonsense	Pathogenic	AR	Cystic fibrosis; Congenital bilateral absence of vas deferens; Sweat chloride elevation without CF	do not report as secondary, only carrier status
76	1:156108510:C:T	NM_170707.3	c.1930C>T	p.Arg644Cys	<i>LMNA</i>	missense	VUS	AD	LMNA-related diseases	do not report as secondary
78	20:52789466:CCTT:C	NM_000782.4	c.428_430delAAG	p.Glu143del	<i>CYP24A1</i>	in frame deletion	Pathogenic	AR	Hypercalcemia, infantile, 1	do not report as secondary, only carrier status
79	1:237824218:C:T	NM_001035.2	c.8407C>T	p.Arg2803Trp	<i>RYR2</i>	missense	VUS	AD	Arrhythmogenic right ventricular dysplasia 2; Ventricular tachycardia, catecholaminergic polymorphic, 1	do not report as secondary
79	11:76925023:T:C	NM_000260.3	c.6557T>C	p.Leu2186Pro	<i>MYO7A</i>	missense	VLP	AR	Usher syndrome, type 1B; Deafness, AD 11 / AR 2	do not report as secondary
81	*X:153762634:G:A	NM_000402.4	c.653C>T	p.Ser218Phe	<i>G6PD</i>	missense	Pathogenic	XLD	Hemolytic anemia, G6PD deficient (favism)	do not report as secondary, unless thought to be cause of disease, most people unaffected, common cause of favism
92	6:26093141:G:A	NM_000410.3	c.845G>A	p.Cys282Tyr	<i>HFE</i>	missense	Pathogenic	AR	Hemochromatosis	do not report as secondary, only carrier status
97	21:47542052:C:T	NM_001849.3	c.1552C>T	p.Pro518Ser	<i>COL6A2</i>	missense	VLB	AD/AR	Bethlem myopathy 1; Ullrich congenital muscular dystrophy 1	do not report as secondary
99	2:50851527:C:T	NM_004801.5	c.831-772G>A	intronic	<i>NRXN1</i>	non coding	VUS	AR	Pitt-Hopkins-like syndrome 2	do not report as secondary
99	2:51093481:T:G	NM_004801.5	c.823+56312A>C	non coding	<i>NRXN1</i>	non-coding	VUS	AR	Pitt-Hopkins-like syndrome 2	do not report as secondary
99	2:50570600:T:TA	NM_004801.5	c.3365-106493dupT	non coding	<i>NRXN1</i>	non-coding	VLB	AR	Pitt-Hopkins-like syndrome 2	do not report as secondary

21, 71	2:48027755:T:C	NM_000179.2	c.2633T>C	p.Val878Ala	<i>MSH6</i>	missense	Benign	AD	Mismatch repair cancer syndrome; Colorectal cancer, hereditary nonpolyposis, type 5	do not report as secondary
81, 91	2:47641558:GT:G	NM_000251.2	c.942+2delT	splice site	<i>MSH2</i>	splice	VLP	AD	Mismatch repair cancer syndrome; Colorectal cancer, hereditary nonpolyposis, type 1; Muir-Torre syndrome	report as secondary finding

\*Several systematic errors or differences in how variants were referenced for submission files for groups 1 and 7. All variants identified in genes on the minus strand were represented as the base of the minus strand while convention generally refers to only plus strand bases for genomic coordinates. In addition, insertions and deletions were generally off by 1 or 2 base pairs relative to convention. Assumptions were made based on notes provided with submissions, gene orientation, and known variants at those loci. Assumed validated variants are listed here and do not match perfectly with original submission files.

## Patient Phenotype descriptions

Bioinformatic groups were given the following patient phenotype information text. Some patients had pedigrees and some growth charts that are not included here-in.

### **Patient 7(X)**

Sex: Female

Indication for referral: High myopia and bilateral retinal hamartomas

### **Family history and pedigree**

Paternal ethnicity:

1. Oman
2. Middle East

Maternal ethnicity:

1. East Indian
- NO Other affected relatives

NO Consanguinity

NO Parents with at least 3 miscarriages

### **Prenatal and perinatal history**

NO Multiple gestation

Gestation at delivery (weeks) 41

Maternal age at EDD (years) 27

DELIVERY

NO Premature birth

Primary Caesarian section

NEONATAL GROWTH PARAMETERS

Abnormal birth weight

NO Small for gestational age (<-2SD)

NO Large for gestational age (>+2SD)

Notes:

Unremarkable pregnancy and emergency C-section at 41 weeks gestation. BW 2.8 kg.

Well in immediate neonatal period.

### **Medical history**

Medical and developmental history:

Nystagmus at 7 weeks, bilateral retinal hamartomas discovered at 2 months, and diagnosed with high myopia

at 6 months. Initially was assessed in London, England. There was a question of partial ocular albinism, and so

panel testing was done that was negative. After coming to Toronto, she was noted to not have blond fundi and the Ophthalmologists are not suspecting ocular albinism.

Global age of onset: Infantile onset



## Measurements

**Date:** Age: 5m

**Weight:** 2.8 kg 0th pctl (-7.51SD)

**Date:** Age: 7y 5m

**Weight:** 26.4 kg 76th pctl (+0.7SD)

**Height:** 133.1 cm 96th pctl (+1.72SD)

**BMI:** 14.9 36th pctl (-0.37SD)

**Head circumference:** 50.0 cm 9th pctl (-1.32SD)

## Clinical symptoms and physical findings

### EYE DEFECTS

Nystagmus

Onset at 7 weeks of age

Retinal hamartoma Bilateral

Identified at 2 months of age

Severe Myopia -10, diagnosed at 6 months of age

NO Hypopigmentation of the fundus

### EAR DEFECTS

Recurrent otitis media

### CUTANEOUS

NO Hypopigmentation of the skin

Multiple cafe-au-lait spots

4-5

NO Fair hair

### MUSCULOSKELETAL

Hyperextensibility of the finger joints

Hyperextensibility at elbow

### GENITOURINARY

NO Abnormality of the kidney

Normal abdominal ultrasound

### BEHAVIOR, COGNITION AND DEVELOPMENT

NO Global developmental delay

High functioning

Attention deficit hyperactivity disorder

Borderline

### NEUROLOGICAL

NO Morphological abnormality of the central nervous system

Essentially normal brain MRI. Nonspecific foci of signal abnormality are seen in the subcortical white matter of both frontal lobes.

## Diagnosis

Additional comments:

?OCULAR ALBINISM ?DEVELOPMENTAL EYE DISORDER

**Patient 9(W)**

Sex: Female

**Family history and pedigree**

Paternal ethnicity:

1. French Canadian

Maternal ethnicity:

1. British

**Medical history**

Global age of onset: Childhood onset

**Clinical symptoms and physical findings**

EYE DEFECTS

Nystagmus

CUTANEOUS

Fair hair

BLOOD AND BLOOD-FORMING TISSUES

Bruising susceptibility

**Patient 17(H)**

Sex: Male

Indication for referral: Retinitis pigmentosa

**Family history and pedigree**

Paternal ethnicity:

1. British

Maternal ethnicity:

1. Canadian

List health conditions found in family (describe the relationship with proband)

No history of eye disease

Maternal family history of sensorineural hearing loss

NO Other affected relatives

NO Consanguinity

NO Parents with at least 3 miscarriages

Global mode of inheritance: Autosomal recessive inheritance

**Medical history**

Medical and developmental history: Nyctalopia before the age of 4 years.

Global age of onset: Childhood onset

**Clinical symptoms and physical findings**

EYE DEFECTS

### NO Nystagmus

Rod-cone dystrophy

ERG

Photophobia

Mild

Nyctalopia Onset <4 years, relatively stable

Hypoplasia of the fovea

Borderline

Color vision test abnormality

Mild

EAR DEFECTS

NO Hearing impairment

### **Patient 18(U)**

Sex: Male

Indication for referral: Global developmental delay and epileptic encephalopathy.

### **Family history and pedigree**

Paternal ethnicity:

1. not known

Maternal ethnicity:

1. Portuguese

List health conditions found in family (describe the relationship with proband)

Father with bipolar disorder. Mother with chronic anemia unknown cause. Maternal side females have history of

hypothyroidism.

NO Consanguinity

NO Parents with at least 3 miscarriages

Global mode of inheritance: Autosomal recessive inheritance

### **Prenatal and perinatal history**

Gestation at delivery (weeks) 31

### **Assisted reproduction:**

NO Conception after fertility medication

NO In vitro fertilization

NO Gestational surrogacy

APGAR score (1 minute) 5

APGAR score (5 minutes) 7

PRENATAL DEVELOPMENT

NO Oligohydramnios

NO Polyhydramnios

DELIVERY

Premature birth

NEONATAL GROWTH PARAMETERS

Abnormal birth weight

NO Small for gestational age (<-2SD)

NO Large for gestational age (>+2SD)

Abnormal birth length

NO Small birth length (<-2SD)

NO Large birth length (>+2SD)

Abnormal head circumference at birth

NO Congenital microcephaly (<-3SD)

NO Congenital macrocephaly (>+2SD)

## Medical history

Medical and developmental history:

Preterm, non-reassuring fetal heart rate, premature delivery at 31 weeks. Neonatal jaundice, sepsis and GI bleeding. Global developmental delay with history of regression after 4 years of age: never walked. First word at age 2 yrs, about fifty words at age 4 yrs, regression in language after 4 years of age. Seizure onset at age 2 years, initially febrile. Severe GDD wheelchair bound, no words. MRI showed thin corpus callosum and periventricular leukomalacia.

Global age of onset: Infantile onset

## Clinical symptoms and physical findings

### GROWTH PARAMETERS

Weight for age

NO Decreased body weight (<-2SD)

Stature for age

NO Short stature (<-2SD)

NO Tall stature (>+2SD)

Head circumference for age

NO Microcephaly (<-3SD)

NO Macrocephaly (>+2SD)

NO Hemihypertrophy

NO Obesity

### CRANIOFACIAL

NO Craniosynostosis

NO Cleft upper lip

NO Cleft palate

NO Abnormal facial shape

### EYE DEFECTS

NO Visual impairment

NO Abnormality of the cornea

NO Coloboma

NO Abnormality of the anterior chamber

NO Cataract

NO Abnormality of the retina

NO Abnormality of the optic nerve

NO Microphthalmia

NO Nystagmus  
NO Strabismus  
NO Hypotelorism  
NO Hypertelorism

#### **EAR DEFECTS**

Deafness

NO Sensorineural  
NO Conductive  
NO Preauricular pit  
NO Preauricular skin tag  
NO Abnormality of the outer ear  
NO Abnormality of the inner ear

#### **CUTANEOUS**

NO Hyperpigmentation of the skin  
NO Hypopigmentation of the skin  
NO Capillary hemangiomas  
NO Vascular skin abnormality

#### **CARDIOVASCULAR**

NO Atrial septal defect  
NO Ventricular septal defect  
NO Complete atrioventricular canal defect  
NO Coarctation of aorta  
NO Tetralogy of Fallot  
NO Cardiomyopathy  
NO Arrhythmia

#### **RESPIRATORY**

NO Congenital diaphragmatic hernia  
NO Abnormality of lung morphology

#### **MUSCULOSKELETAL**

NO Skeletal dysplasia  
NO Increased susceptibility to fractures  
NO Lower limb undergrowth  
NO Upper limb undergrowth

Camptodactyly

NO Finger  
NO Toe

Syndactyly

NO Finger  
NO Toe

Polydactyly

NO Preaxial  
NO Postaxial

Oligodactyly

NO Hands  
NO Feet  
NO Scoliosis

NO Abnormality of the vertebral column  
NO Flexion contracture  
NO Talipes equinovarus

#### **GASTROINTESTINAL**

NO Esophageal atresia  
NO Tracheoesophageal fistula  
NO Gastroschisis  
NO Omphalocele  
NO Aganglionic megacolon  
NO Cholestasis  
NO Elevated hepatic transaminases  
NO Exocrine pancreatic insufficiency  
NO Diabetes mellitus

#### **GENITOURINARY**

NO Renal cyst  
NO Horseshoe kidney  
NO Abnormality of the ureter  
NO Abnormality of the urethra  
NO Ambiguous genitalia  
NO Hypospadias  
NO Cryptorchidism

#### **BEHAVIOR, COGNITION AND DEVELOPMENT**

Global developmental delay  
Intellectual disability  
Severe  
NO Attention deficit hyperactivity disorder  
NO Autism  
NO Behavioral abnormality

#### **NEUROLOGICAL**

NO Generalized hypotonia  
Seizures  
NO Ataxia  
NO Dystonia  
NO Chorea  
Spasticity  
NO Spinal dysraphism  
NO Morphological abnormality of the central nervous system

## **Patient 21(G)**

Age: 5 years

Sex: Female

Indication for referral: Infantile onset epileptic encephalopathy and global developmental delay.

### **Family history and pedigree**

Paternal ethnicity:

1. Romanian

Maternal ethnicity:

1. Romanian

List health conditions found in family (describe the relationship with proband)

None.

NO Consanguinity

NO Parents with at least 3 miscarriages

Global mode of inheritance: Autosomal recessive inheritance

### **Prenatal and perinatal history**

Gestation at delivery (weeks) 34

#### **Assisted reproduction:**

NO Conception after fertility medication

In vitro fertilization

NO Gestational surrogacy

PRENATAL DEVELOPMENT

NO Oligohydramnios

NO Polyhydramnios

DELIVERY

NO Premature birth

NEONATAL GROWTH PARAMETERS

Abnormal birth weight

NO Small for gestational age (<-2SD)

NO Large for gestational age (>+2SD)

Abnormal birth length

NO Small birth length (<-2SD)

NO Large birth length (>+2SD)

Abnormal head circumference at birth

NO Congenital microcephaly (<-3SD)

NO Congenital macrocephaly (>+2SD)

### **Medical history**

Medical and developmental history: First seizure at age 4.5 months. Hypotonic since birth. Global developmental delay(chronological age 24 months) walking at age 21 months, wide-based, no pincer grasp and no words yet. Developmental age around 9-12 months. Still ongoing seizures. MRI shows delayed myelination as well as thin corpus callosum and small hippocampi.

Global age of onset: Infantile onset

## **Clinical symptoms and physical findings**

### **GROWTH PARAMETERS**

Weight for age

NO Decreased body weight (<-2SD)

Stature for age

NO Short stature (<-2SD)

NO Tall stature (>+2SD)

Head circumference for age

NO Microcephaly (<-3SD)

NO Macrocephaly (>+2SD)

NO Hemihypertrophy

NO Obesity

### **CRANIOFACIAL**

NO Craniosynostosis

NO Cleft upper lip

NO Cleft palate

NO Abnormal facial shape

### **EYE DEFECTS**

NO Visual impairment

NO Abnormality of the cornea

NO Coloboma

NO Abnormality of the anterior chamber

NO Cataract

NO Abnormality of the retina

NO Abnormality of the optic nerve

NO Microphthalmia

NO Nystagmus

NO Strabismus

NO Hypotelorism

NO Hypertelorism

### **EAR DEFECTS**

Deafness

NO Sensorineural

NO Conductive

NO Preauricular pit

NO Preauricular skin tag

NO Abnormality of the outer ear

NO Abnormality of the inner ear

### **CARDIOVASCULAR**

NO Atrial septal defect

NO Ventricular septal defect

NO Complete atrioventricular canal defect

NO Coarctation of aorta

NO Tetralogy of Fallot



NO Cardiomyopathy

NO Arrhythmia

#### RESPIRATORY

NO Congenital diaphragmatic hernia

NO Abnormality of lung morphology

#### MUSCULOSKELETAL

NO Skeletal dysplasia

NO Increased susceptibility to fractures

NO Lower limb undergrowth

NO Upper limb undergrowth

Camptodactyly

NO Finger

NO Toe

Syndactyly

NO Finger

NO Toe

Polydactyly

NO Preaxial

NO Postaxial

Oligodactyly

NO Hands

NO Feet

NO Scoliosis

NO Abnormality of the vertebral column

NO Flexion contracture

NO Talipes equinovarus

#### GASTROINTESTINAL

NO Esophageal atresia

NO Tracheoesophageal fistula

NO Gastroschisis

NO Omphalocele

NO Aganglionic megacolon

NO Cholestasis

NO Elevated hepatic transaminases

NO Exocrine pancreatic insufficiency

NO Diabetes mellitus

#### GENITOURINARY

NO Renal cyst

NO Horseshoe kidney

NO Abnormality of the ureter

NO Abnormality of the urethra

NO Ambiguous genitalia

#### BEHAVIOR, COGNITION AND DEVELOPMENT

Global developmental delay

Delayed fine motor development

Delayed gross motor development

Delayed speech and language development  
NO Attention deficit hyperactivity disorder  
NO Autism  
NO Behavioral abnormality  
NEUROLOGICAL  
Generalized hypotonia  
Seizures  
Ataxia  
NO Dystonia  
NO Chorea  
NO Spasticity  
NO Spinal dysraphism  
NO Morphological abnormality of the central nervous system

**Patient 30(R)**

Age: 12 years

Sex: Male

Indication for referral: Global developmental delay, epileptic encephalopathy

**Family history and pedigree**

Paternal ethnicity:

1. Phillipine

Maternal ethnicity:

1. Phillipine

List health conditions found in family (describe the relationship with proband)

None

NO Consanguinity

NO Parents with at least 3 miscarriages

Global mode of inheritance: Autosomal recessive inheritance

**Prenatal and perinatal history**

**Assisted reproduction:**

NO Conception after fertility medication

NO In vitro fertilization

NO Gestational surrogacy

**PRENATAL DEVELOPMENT**

NO Oligohydramnios

NO Polyhydramnios

**DELIVERY**

NO Premature birth

**NEONATAL GROWTH PARAMETERS**

Abnormal birth weight

NO Small for gestational age (<-2SD)

NO Large for gestational age (>+2SD)

Abnormal birth length

NO Small birth length (<-2SD)

NO Large birth length (>+2SD)  
Abnormal head circumference at birth  
NO Congenital microcephaly (<-3SD)  
NO Congenital macrocephaly (>+2SD)

Notes:

Pregnancy complicated with maternal gestational diabetes mellitus.

### **Medical history**

Medical and developmental history: Infantile spasms at age 4 months. Intractable epilepsy followed. Various anti-epileptic medications, ketogenic diet non-responsive. MRI no focal lesion. Epilepsy surgery due to focal EEG features. Epilepsy surgery. Pathology reported focal cortical dysplasia type 1b in brain specimen.

Global age of onset: Infantile onset

### **Clinical symptoms and physical findings**

#### **GROWTH PARAMETERS**

Weight for age

NO Decreased body weight (<-2SD)

Stature for age

NO Short stature (<-2SD)

NO Tall stature (>+2SD)

Head circumference for age

Microcephaly (<-3SD)

NO Macrocephaly (>+2SD)

NO Obesity

#### **CRANIOFACIAL**

NO Craniosynostosis

NO Cleft upper lip

NO Cleft palate

NO Abnormal facial shape

#### **EYE DEFECTS**

NO Visual impairment

NO Abnormality of the cornea

NO Coloboma

NO Abnormality of the anterior chamber

NO Cataract

NO Abnormality of the retina

NO Abnormality of the optic nerve

NO Microphthalmia

NO Nystagmus

NO Strabismus

NO Hypotelorism

NO Hypertelorism

#### **EAR DEFECTS**

Deafness

NO Sensorineural  
NO Conductive  
NO Preauricular pit  
NO Preauricular skin tag  
NO Abnormality of the outer ear  
NO Abnormality of the inner ear

#### **CUTANEOUS**

NO Hyperpigmentation of the skin  
NO Hypopigmentation of the skin  
NO Capillary hemangiomas  
NO Vascular skin abnormality

#### **CARDIOVASCULAR**

NO Atrial septal defect  
NO Ventricular septal defect  
NO Complete atrioventricular canal defect  
NO Coarctation of aorta  
NO Tetralogy of Fallot  
NO Cardiomyopathy  
NO Arrhythmia

#### **RESPIRATORY**

NO Congenital diaphragmatic hernia  
NO Abnormality of lung morphology

#### **MUSCULOSKELETAL**

NO Skeletal dysplasia  
NO Increased susceptibility to fractures  
NO Lower limb undergrowth  
NO Upper limb undergrowth

#### **Camptodactyly**

NO Finger

NO Toe

#### **Syndactyly**

NO Finger

NO Toe

#### **Polydactyly**

NO Preaxial

NO Postaxial

#### **Oligodactyly**

NO Hands

NO Feet

NO Scoliosis

NO Abnormality of the vertebral column

NO Flexion contracture

NO Talipes equinovarus

#### **GASTROINTESTINAL**

NO Esophageal atresia

NO Tracheoesophageal fistula

NO Gastroschisis  
NO Omphalocele  
NO Aganglionic megacolon  
NO Cholestasis  
NO Elevated hepatic transaminases  
NO Exocrine pancreatic insufficiency  
NO Diabetes mellitus

#### GENITOURINARY

NO Renal cyst  
NO Horseshoe kidney  
NO Abnormality of the ureter  
NO Abnormality of the urethra  
NO Ambiguous genitalia  
NO Hypospadias  
NO Cryptorchidism

#### BEHAVIOR, COGNITION AND DEVELOPMENT

Global developmental delay  
NO Delayed fine motor development  
NO Delayed gross motor development  
NO Delayed speech and language development  
NO Specific learning disability  
Intellectual disability  
NO Severe  
NO Attention deficit hyperactivity disorder  
NO Autism  
NO Behavioral abnormality

#### NEUROLOGICAL

NO Generalized hypotonia  
Ataxia  
NO Dystonia  
NO Chorea  
NO Spasticity  
NO Spinal dysraphism  
NO Morphological abnormality of the central nervous system  
Generalized tonic-clonic seizures  
Absence seizures  
Cortical dysplasia  
Atonic seizures

### **Patient 39(P)**

Age: 12 years

Sex: Male

Indication for referral: Epileptic encephalopathy

### **Family history and pedigree**

Paternal ethnicity:

1. Caucasian

Maternal ethnicity:

1. Caucasian

List health conditions found in family (describe the relationship with proband)

Mother with ADD and celiac disease.

paternal uncle's son diagnosis of Asperger's syndrome and recent diagnosis of epilepsy

NO Consanguinity

NO Parents with at least 3 miscarriages

Global mode of inheritance: Autosomal recessive inheritance

### **Prenatal and perinatal history**

#### **Assisted reproduction:**

NO Conception after fertility medication

NO In vitro fertilization

NO Gestational surrogacy

PRENATAL DEVELOPMENT

NO Oligohydramnios

NO Polyhydramnios

DELIVERY

NO Premature birth

NEONATAL GROWTH PARAMETERS

Abnormal birth weight

NO Small for gestational age (<-2SD)

NO Large for gestational age (>+2SD)

Abnormal birth length

NO Small birth length (<-2SD)

NO Large birth length (>+2SD)

Abnormal head circumference at birth

NO Congenital microcephaly (<-3SD)

NO Congenital macrocephaly (>+2SD)

### **Medical history**

Medical and developmental history:

First seizure at age 2.5 years. Normal development until grade 1. Due to intractable seizures, in grade 1 requiring an

Individualized Educational Plan.

Ketogenic diet.

Global age of onset:

Infantile onset

### **Clinical symptoms and physical findings**

## GROWTH PARAMETERS

Weight for age

NO Decreased body weight (<-2SD)

Stature for age

NO Short stature (<-2SD)

NO Tall stature (>+2SD)

Head circumference for age

NO Microcephaly (<-3SD)

NO Macrocephaly (>+2SD)

NO Obesity

## CRANIOFACIAL

NO Craniosynostosis

NO Cleft upper lip

NO Cleft palate

NO Abnormal facial shape

## EYE DEFECTS

NO Visual impairment

NO Abnormality of the cornea

NO Coloboma

NO Abnormality of the anterior chamber

NO Cataract

NO Abnormality of the retina

NO Abnormality of the optic nerve

NO Microphthalmia

NO Nystagmus

NO Strabismus

NO Hypotelorism

NO Hypertelorism

## EAR DEFECTS

Deafness

NO Sensorineural

NO Conductive

NO Preauricular pit

NO Preauricular skin tag

NO Abnormality of the outer ear

NO Abnormality of the inner ear

## CUTANEOUS

NO Hyperpigmentation of the skin

NO Hypopigmentation of the skin

NO Capillary hemangiomas

NO Vascular skin abnormality

## CARDIOVASCULAR

NO Atrial septal defect

NO Ventricular septal defect

NO Complete atrioventricular canal defect

NO Coarctation of aorta

NO Tetralogy of Fallot

NO Cardiomyopathy

NO Arrhythmia

#### RESPIRATORY

NO Congenital diaphragmatic hernia

NO Abnormality of lung morphology

#### MUSCULOSKELETAL

NO Skeletal dysplasia

NO Increased susceptibility to fractures

NO Lower limb undergrowth

NO Upper limb undergrowth

#### Camptodactyly

NO Finger

NO Toe

#### Syndactyly

NO Finger

NO Toe

#### Polydactyly

NO Preaxial

NO Postaxial

#### Oligodactyly

NO Hands

NO Feet

NO Scoliosis

NO Abnormality of the vertebral column

NO Flexion contracture

NO Talipes equinovarus

#### GASTROINTESTINAL

NO Esophageal atresia

NO Tracheoesophageal fistula

NO Gastroschisis

NO Omphalocele

NO Aganglionic megacolon

NO Cholestasis

NO Elevated hepatic transaminases

NO Exocrine pancreatic insufficiency

NO Diabetes mellitus

#### GENITOURINARY

NO Renal cyst

NO Horseshoe kidney

NO Abnormality of the ureter

NO Abnormality of the urethra

NO Ambiguous genitalia

NO Hypospadias

NO Cryptorchidism

Nephrolithiasis



## BEHAVIOR, COGNITION AND DEVELOPMENT

NO Global developmental delay  
NO Delayed fine motor development  
NO Delayed gross motor development  
NO Delayed speech and language development  
NO Specific learning disability

Intellectual disability

NO Mild

NO Moderate

NO Severe

NO Attention deficit hyperactivity disorder

NO Autism

NO Behavioral abnormality

Dysarthria

## NEUROLOGICAL

NO Generalized hypotonia

Seizures

NO Ataxia

NO Dystonia

NO Chorea

NO Spasticity

NO Spinal dysraphism

NO Morphological abnormality of the central nervous system

Generalized tonic-clonic seizures

Absence seizures

Generalized myoclonic seizures

Focal tonic seizures

## Diagnosis

Additional comments: Epileptic encephalopathy. Normal cranial MRI.

## **Patient 42(O)**

Age: 12 years

Sex: Female

Indication for referral: Early onset retinal dystrophy

## **Family history and pedigree**

Paternal ethnicity:

1. Egyptian

Maternal ethnicity:

1. Egyptian

List health conditions found in family (describe the relationship with proband)

Mother's parents are distantly related.

Distant maternal relative with unspecified eye problem (see pedigree).

NO Other affected relatives

NO Consanguinity

NO Parents with at least 3 miscarriages  
Global mode of inheritance: Autosomal recessive inheritance

### **Medical history**

Medical and developmental history: Born with very small congenital cataracts. Later re-presented to Ophthalmology clinic with nyctalopia. Allergies - environmental, food, medication: NKDA

### **Clinical symptoms and physical findings**

#### **EYE DEFECTS**

Abnormal electroretinogram  
Severe rod and cone dysfunction  
Congenital cataract  
Bilateral  
Very small anterior pyramidal cataracts  
Retinal dystrophy  
Nyctalopia Age 5

#### **EAR DEFECTS**

**NO Hearing impairment**

#### **RESPIRATORY**

Asthma

Mild

#### **METABOLISM/HOMEOSTASIS**

Cystoid macular edema  
Persistent on serial OCT

### **Patient 56(N)**

Age: 11 years

Sex: Female

Indication for referral: Cerebral arteriovenous malformation

### **Family history and pedigree**

Paternal ethnicity:

1. French Canadian

Maternal ethnicity:

1. Chinese

List health conditions found in family (describe the relationship with proband)

Paternal family history of cerebral aneurysm

NO Consanguinity

NO Parents with at least 3 miscarriages

### **Prenatal and perinatal history**

NO Multiple gestation

Gestation at delivery (weeks) 37

Maternal age at EDD (years) 35

Paternal age at EDD (years) 36

## DELIVERY

NO Abnormal delivery (Non-NSVD)

NO Premature birth

Notes: Reportedly unremarkable pregnancy and delivery.

## Medical history

Allergies - environmental, food, medication:

NKDA

## Measurements

**Age: 17y 9m**

**Weight:** 54.3 kg 43rd pctl (-0.18SD)

**Height:** 164.5 cm 59th pctl (+0.22SD)

**BMI:** 20.07 35th pctl (-0.4SD)

**Head circumference:** 56.5 cm 91st pctl (+1.36SD)

**Age: 18y 2m**

**Weight:** 55.8 kg 49th pctl (-0.03SD)

**Height:** 164.0 cm 55th pctl (+0.14SD)

**BMI:** 20.75 43rd pctl (-0.18SD)

## Clinical symptoms and physical findings

### CARDIOVASCULAR

Aortic dilatation

Borderline

Cardiac MRI suggestive of mild dilated aortic root, ascending aorta, aortic arch, and thoracic aorta. However,

this was discordant with results from echo showing aortic root and ascending aorta z-scores within normal limits. Ultimately not felt to have significant issue. Cerebral arteriovenous malformation Discovered on brain MRI after presenting with headaches, resected from right temporal region? small associated aneurysm.

NO Arterial tortuosity

### MUSCULOSKELETAL

Joint hypermobility

Borderline

Beighton score 5

Fractures of the long bones

Left femur and right humerus fracture @ age 3 secondary to trauma.

Patellar subluxation

Bilateral knee/patellar instability for which she wears brace.

### BEHAVIOR, COGNITION AND DEVELOPMENT

NO Cognitive impairment

### NEUROLOGICAL

Headache

## **Patient 57(T)**

Age: 9 years

Sex: Female

Indication for referral: Ehlers-Danlos syndrome, hypermobility type

### **Family history and pedigree**

Paternal ethnicity:

1. Ashkenazi Jewish
2. Polish

Maternal ethnicity:

1. Ashkenazi Jewish
2. Polish

List health conditions found in family (describe the relationship with proband)

Hypermobility, joint dislocation, and IBS (see pedigree)

Other affected relatives

NO Consanguinity

NO Parents with at least 3 miscarriages

### **Prenatal and perinatal history**

NO Multiple gestation

Gestation at delivery (weeks) Term birth

Maternal age at EDD (years) 27

Paternal age at EDD (years) 30

PREGNANCY HISTORY

Hyperemesis gravidarum (excessive vomiting)

DELIVERY

NO Abnormal delivery (Non-NSVD)

NO Premature birth

NEONATAL GROWTH PARAMETERS

Abnormal birth weight

NO Small for gestational age (<-2SD)

NO Large for gestational age (>+2SD)

Notes:

Birth weight 7 lbs 6 oz.

### **Medical history**

Allergies - environmental, food, medication:

1. Adhesives / tapes (rash)

### **Measurements**

**Age: 16y 2m**

**Weight:** 47.4 kg 16th pctl (-1.0SD)

**Height:** 156.5 cm 18th pctl (-0.91SD)

**BMI:** 19.35 30th pctl (-0.52SD)

**Head circumference:** 52.0 cm 5th pctl (-1.64SD)

### **Clinical symptoms and physical findings**

EAR DEFECTS

**NO Hearing impairment**

**Audiology testing within normal limits.**

#### CARDIOVASCULAR

Patent foramen ovale

On echo at age 15 1/2 years.

Postural hypotension with compensatory tachycardia

Tilt table testing pending for query POTS.

#### RESPIRATORY

Asthma

Shoulder dislocation

Multidirectional instability with frequent subluxation and occasional dislocation. Went to OR for left shoulder.

Stridor Likely secondary to conversion disorder. ? paradoxical vocal cord movements.

Sleep apnea

CPAP at night.

#### MUSCULOSKELETAL

Joint dislocation

Knee, finger, bilateral shoulder.

Joint hypermobility

Beighton score 7.

**NO EMG abnormality**

#### GASTROINTESTINAL

Abnormality of the gastrointestinal tract

Irritable bowel syndrome. Upper and lower scopes showed only non-specific changes.

#### BEHAVIOR, COGNITION AND DEVELOPMENT

Specific learning disability

Reportedly with above average IQ but specific learning disability in processing.

#### CONSTITUTIONAL SYMPTOM

Chronic pain

#### CONNECTIVE TISSUE

Atypical scarring of skin

**NO Mastocytosis**

**Clinically suspected of having mast cell activation syndrome with history of hives/rashes  
NYD and sensitive skin.**

**No increase in mucosal mast cells on GI biopsies.**

#### VOICE

Weak voice

Likely secondary to conversion disorder.

#### IMMUNE SYSTEM

**NO Celiac disease**

### **Patient 67(M)**

Age: 13 years

Sex: Male

Indication for referral: Retinitis pigmentosa

#### **Family history and pedigree**

List health conditions found in family (describe the relationship with proband)

Mother has eye phenotype, so there was a question about X-linked RP.

NO Consanguinity

NO Parents with at least 3 miscarriages

#### **Prenatal and perinatal history**

Gestation at delivery (weeks) Term birth

DELIVERY

NO Premature birth

Notes:

Reportedly unremarkable pregnancy.

#### **Medical history**

Medical and developmental history: Glasses since age 2. Presented at ~ age 4 after noted to have chin down position and to be walking into things. Exam and investigations consistent with retinitis pigmentosa. Otherwise healthy.

#### **Clinical symptoms and physical findings**

EYE DEFECTS

Visual impairment

NO Nystagmus

Rod-cone dystrophy

Abnormality of color vision

Nyctalopia

Depigmented fundus

BEHAVIOR, COGNITION AND DEVELOPMENT

NO Cognitive impairment

### **Patient 68(J)**

Age: 17 years

Sex: Female

Indication for referral: Mitochondrial disorder (query Pearson syndrome)

#### **Family history and pedigree**

Paternal ethnicity:

1. East Indian

Maternal ethnicity:

1. East Indian

List health conditions found in family (describe the relationship with proband)

Identical twin has similar multi-system health concerns.

Strong maternal family history of diabetes.  
Other affected relatives  
NO Consanguinity  
NO Parents with at least 3 miscarriages

### **Prenatal and perinatal history**

Multiple gestation  
Gestation at delivery (weeks) 33  
Maternal age at EDD (years) 30  
Paternal age at EDD (years) 35  
PREGNANCY HISTORY  
Maternal diabetes  
**NO Maternal teratogenic exposure**  
DELIVERY  
Abnormal delivery (Non-NSVD)  
Caesarian section  
Premature birth  
Nuchal cord  
NEONATAL GROWTH PARAMETERS  
Abnormal birth weight  
**NO Small for gestational age (<-2SD)**  
**NO Large for gestational age (>+2SD)**

Notes:

BW 1.59 kg

### **Measurements**

**Age: 7y 4m**

**Weight:** 15.5 kg 0th pctl (-2.92SD)

**Height:** 107.2 cm 0th pctl (-2.85SD)

**BMI:** 13.49 8th pctl (-1.38SD)

**Age: 11y 5m**

**Weight:** 26.4 kg 2nd pctl (-1.99SD)

**Height:** 119.0 cm 0th pctl (-4.27SD)

**BMI:** 18.64 67th pctl (+0.45SD)

**Age: 14y 9m**

**Weight:** 31.0 kg 0th pctl (-3.62SD)

**Height:** 122.0 cm 0th pctl (-5.71SD)

**BMI:** 20.83 60th pctl (+0.26SD)

**Head circumference:** 54.0 cm 49th pctl (-0.03SD)

**Age: 16y 7m**

**Weight:** 34.3 kg 0th pctl (-3.7SD)

**Height:** 125.9 cm 0th pctl (-5.47SD)

**BMI:** 21.64 59th pctl (+0.24SD)

### **Clinical symptoms and physical findings**

GROWTH PARAMETERS

Weight for age  
Decreased body weight (<-2SD)  
Stature for age  
Short stature (<-2SD)  
Delayed puberty  
EYE DEFECTS  
Ptosis  
Retinal dystrophy  
Corneal dystrophy  
Corneal disease with possible endothelial dystrophy, status post right corneal transplant  
CARDIOVASCULAR  
Sinus tachycardia  
MUSCULOSKELETAL  
Scoliosis  
Mild  
Osteopenia  
Pes planus  
Bilateral  
Hallux valgus  
Bilateral  
Delayed skeletal maturation  
Vertebral compression fractures  
Myopathy  
Mild  
Proximal  
Ragged-red muscle fibers  
Abnormal mitochondria in muscle tissue  
Also in renal tissue  
GENITOURINARY  
Stage 5 chronic kidney disease  
Renal transplant  
BEHAVIOR, COGNITION AND DEVELOPMENT  
NO Global developmental delay  
Specific learning disability  
Some learning difficulties, with IEP in school. E.g., in gr. 8, at gr. 5-6 level.  
NEUROLOGICAL  
Ataxia  
Morphological abnormality of the central nervous system  
Brain MRI showed mild prominence of lateral and third ventricles, as well as mild cerebellar volume loss. No abnormal lactate peak on MRS.  
Abnormality of the peripheral nervous system  
Guillain-Barre syndrome  
BLOOD AND BLOOD-FORMING TISSUES  
Aplastic anemia  
Severe



17 pRBC transfusions  
METABOLISM/HOMEOSTASIS  
Renal tubular acidosis  
Fanconi renal syndrome in setting of sepsis  
Increased serum lactate  
Decreased activity of mitochondrial complex II  
DIGESTIVE SYSTEM  
Abnormality of pancreas morphology  
Echogenic, with fatty infiltration changes, on ultrasound  
Abnormality of exocrine pancreas physiology  
Fecal fat excretion and fecal elastase abnormal  
ENDOCRINE SYSTEM  
Type I diabetes mellitus  
IMMUNE SYSTEM  
Recurrent lower respiratory tract infections

### **Patient 71(L)**

Age: 9 years

Sex: Male

Indication for referral: Hypermobility, recurrent dislocation, query connective tissue disorder

### **Family history and pedigree**

Paternal ethnicity:

1. Argentinian

Maternal ethnicity:

1. Italian

List health conditions found in family (describe the relationship with proband)

Mother and maternal relatives with joint hypermobility, but not necessarily to same degree as the proband.

NO Other affected relatives

NO Consanguinity

NO Parents with at least 3 miscarriages

### **Prenatal and perinatal history**

NO Multiple gestation

Gestation at delivery (weeks) 38

### **Assisted reproduction:**

NO Conception after fertility medication

Maternal age at EDD (years) 31

Paternal age at EDD (years) 32

### **PRENATAL DEVELOPMENT**

Oligohydramnios

**NO Polyhydramnios**

**NO Decreased fetal movement**

### **DELIVERY**

### NO Premature birth

Primary Caesarian section

#### NEONATAL GROWTH PARAMETERS

Abnormal birth weight

Small for gestational age (<-2SD)

Notes: Unremarkable pregnancy until oligohydramnios noted in third trimester. Born via emergency C-section at 38 weeks for fetal distress. Weight 2.38 kg, labeled as SGA and kept in NICU for 10 days. No resuscitation required at birth.

### Medical history

Medical and developmental history: Noted at ~5 months to be hyperextensible. Came to Canada from Argentina in grade 1. Hypermobility has improved with age.

#### Measurements

**Age: 15y 11m**

**Weight:** 60.0 kg 44th pctl (-0.15SD)

**Height:** 164.0 cm 13th pctl (-1.11SD)

**BMI:** 22.31 74th pctl (+0.66SD)

**Head circumference:** 58.0 cm 93rd pctl (+1.47SD)

**Age: 16y 6m**

**Weight:** 62.5 kg 45th pctl (-0.11SD)

**Height:** 175.0 cm 54th pctl (+0.1SD)

**BMI:** 20.41 44th pctl (-0.16SD)

**Head circumference:** 57.5 cm 84th pctl (+1.01SD)

**Arm span:** 178.5 cm = Height + 3.5 cm

**Age: 18y 4m**

**Weight:** 66.9 kg 44th pctl (-0.16SD)

**Height:** 176.5 cm 51st pctl (+0.02SD)

**BMI:** 21.48 44th pctl (-0.15SD)

### Clinical symptoms and physical findings

#### CRANIOFACIAL

High palate

Malar flattening

Mandibular prognathia

Triangular face

Midface retrusion

#### EYE DEFECTS

Astigmatism

Left > right

Hypermetropia

NO Abnormal eye morphology

Reportedly normal dilated exam

#### CARDIOVASCULAR

NO Arrhythmia

Normal Holter

NO Orthostatic hypotension

No concern regarding POTS  
NO Abnormal heart morphology  
Reportedly normal echo

#### MUSCULOSKELETAL

Joint hypermobility

Beighton score initially 9/9, but as he aged he became less hypermobile.

Pes cavus

Right

Pes planus

Left

Hammertoe

Right

Bilateral talipes equinovarus

Congenital onset

Clinodactyly of the 5th finger

Recurrent patellar dislocation

Right

Soccer player

Ankle contracture

Mild

Bilateral

Scheuermann-like vertebral changes

On spine MRI and x-ray. Mild thoracolumbar left scoliosis.

Infantile muscular hypotonia

Few details. Apparently resolved, as exam records from Canada do not report hypotonia.

Long fingers

NO EMG abnormality

Initial EMG showed non-specific pattern consistent with increased small polyphasic low voltage motor units not

really in keeping with a muscle process. Repeat EMG was normal.

#### GENITOURINARY

Bilateral cryptorchidism

Total of 3 operations in Argentina

#### NEUROLOGICAL

NO Morphological abnormality of the central nervous system

Brain MRI showing only non-specific tiny white matter signal intensity in right frontal lobe.

NO Motor delay

#### BLOOD AND BLOOD-FORMING TISSUES

Bruising susceptibility

#### CONNECTIVE TISSUE

Inguinal hernia

Contracture of the proximal interphalangeal joint of the 5th finger

## **Patient 76(Q)**

Age: 11 years

Sex: Female

Indication for referral: Ehlers-Danlos syndrome (EDS) type III, hypermobility type

### **Family history and pedigree**

Paternal ethnicity:

1. Anglo-Celtic Australian

Maternal ethnicity:

1. Anglo-Celtic Australian

Other affected relatives

NO Consanguinity

NO Parents with at least 3 miscarriages

### **Prenatal and perinatal history**

NO Multiple gestation

Gestation at delivery (weeks) 38

PREGNANCY HISTORY

Maternal diabetes

DELIVERY

NO Abnormal delivery (Non-NSVD)

NO Premature birth

NEONATAL GROWTH PARAMETERS

Abnormal birth weight

NO Small for gestational age (<-2SD)

NO Large for gestational age (>+2SD)

Notes: Uncomplicated pregnancy and term vaginal delivery in New Zealand, BW 7 lbs 2 oz.

### **Medical history**

Allergies - environmental, food, medication:

1. Mango (anaphylaxis)

### **Measurements**

**Age: 18y 1m**

**Weight:** 61.0 kg 71st pctl (+0.56SD)

**Height:** 158.5 cm 24th pctl (-0.69SD)

**BMI:** 24.28 81st pctl (+0.87SD)

**Head circumference:** 54.5 cm 48th pctl (-0.04SD)

### **Clinical symptoms and physical findings**

CARDIOVASCULAR

Abnormal heart morphology

Chiari network in right atrium on echocardiogram.

Vasovagal syncope

Positive tilt table test response for vasovagal presyncope.

RESPIRATORY

Shoulder subluxation  
MUSCULOSKELETAL

Joint hypermobility

Beighton score  $\geq 7$ .

Pes planus

Fractured hand bones

Wrist fracture.

Multiple joint dislocation

NO Osteopenia

Normal BMD study.

NEUROLOGICAL

Migraine

Ocular migraine.

Hydromyelia

NO Motor delay

BLOOD AND BLOOD-FORMING TISSUES

NO Abnormal bleeding

DIGESTIVE SYSTEM

Constipation

Also with early satiety. Seen by GI in the past.

CONSTITUTIONAL SYMPTOM

Chronic pain

Especially in lower legs, since age 10.

CONNECTIVE TISSUE

Mastocytosis

Also with intermittent lips and tongue paresthesias and skin flushing.

NO Atypical scarring of skin

ENDOCRINE SYSTEM

Abnormality of the thyroid gland

Right posterior mid-lobe hypoechoic nodules on ultrasound, with normal thyroid function.

IMMUNE SYSTEM

Allergy

NO Celiac disease

### **Patient 78(V)**

Age: 6 years

Sex: Female

Indication for referral: Ehlers-Danlos syndrome, hypermobility type

### **Family history and pedigree**

Paternal ethnicity:

1. English

Maternal ethnicity:

1. English

2. French

List health conditions found in family (describe the relationship with proband)  
Mother, maternal aunt, and maternal grandfather with hypermobility.  
Brother and father with EDS type 3 diagnosis.  
Paternal uncle with unspecified psychiatric issues, and paternal female first-cousin with seizures and mild autism spectrum disorder.  
Other affected relatives  
NO Consanguinity  
NO Parents with at least 3 miscarriages

### **Prenatal and perinatal history**

NO Multiple gestation  
Gestation at delivery (weeks) 37  
Maternal age at EDD (years) 35  
Paternal age at EDD (years) 34  
APGAR score (1 minute) 9  
APGAR score (5 minutes) 9  
PREGNANCY HISTORY  
NO Maternal hypertension  
NO Maternal diabetes  
NO Maternal teratogenic exposure  
PRENATAL DEVELOPMENT  
NO Intrauterine growth retardation  
NO Oligohydramnios  
NO Polyhydramnios  
DELIVERY  
NO Abnormal delivery (Non-NSVD)  
NO Premature birth  
Precipitous labour  
NEONATAL GROWTH PARAMETERS  
Abnormal birth weight  
NO Small for gestational age (<-2SD)  
NO Large for gestational age (>+2SD)  
Notes: Reportedly uncomplicated pregnancy with normal ultrasounds. BW 6 lbs 11 oz.

### **Medical history**

Medical and developmental history: Followed in Hamilton in early years. Initially seen by Genetics there in because of GDD and FTT. Allergies - environmental, food, medication: NKDA

### **Measurements**

**Age:** 3y 11m  
**Weight:** 12.5 kg 3rd pctl (-1.85SD)  
**Height:** 90.0 cm 0th pctl (-2.9SD)  
**BMI:** 15.43 55th pctl (+0.12SD)  
**Head circumference:** 47.0 cm 1st pctl (-2.4SD)

**Outer canthal distance:** 7.5 cm 38th pctl (-0.29SD)  
**Inner canthal distance:** 3.0 cm 82nd pctl (+0.91SD)  
**Interpupillary distance:** 5.0 cm 60th pctl (+0.25SD)  
**Arm span:** 90.0 cm = Height

### **Clinical symptoms and physical findings**

#### GROWTH PARAMETERS

Stature for age

Short stature (<-2SD)

Failure to thrive in infancy

#### EYE DEFECTS

Hypermetropia

Borderline

Glasses. Also with "delayed visual myelination" - resolved

#### EAR DEFECTS

**NO Hearing impairment**

#### CUTANEOUS

Hyperextensible skin

Abnormality of the toenails

Short, somewhat hypoplastic, and easy breakability

**NO Eczema**

#### CARDIOVASCULAR

Urticaria

Intermittent, not always clear precipitant

**NO Abnormality of the cardiovascular system**

**Normal echocardiogram, with normal aortic measurements and no mitral valve abnormalities**

#### RESPIRATORY

Cough

Chronic

#### MUSCULOSKELETAL

Talipes equinovarus

Treated with therapy and SMO. Other reports say bilateral calcaneovalgus deformity

Joint dislocation

Joint hypermobility

Beighton score 6/9

Infantile muscular hypotonia

#### BEHAVIOR, COGNITION AND DEVELOPMENT

Global developmental delay

Early on, especially fine and gross motor. At time of last follow-up, gross motor normal.

#### NEUROLOGICAL

Seizures

Clinically suspected per parents on the basis of occasional stiffening spells beginning at 4 months of age, but

two normal EEGs, never treated with anti-epileptic medication, and resolved in childhood.

NO Morphological abnormality of the central nervous system  
Essentially normal brain MRI in Hamilton, with exception of possibly delayed myelination.

NO EEG abnormality

BLOOD AND BLOOD-FORMING TISSUES

Bruising susceptibility

DIGESTIVE SYSTEM

Constipation

CONSTITUTIONAL SYMPTOM

Chronic pain

Multisystem. Seen by Psychiatry because of disruptive behaviours that were attributed to pain

IMMUNE SYSTEM

Allergy

Multiple foods

### **Patient 79(K)**

Age: 11 years

Sex: Male

Indication for referral: query Ehlers-Danlos syndrome, hypermobility type, possible ADHD

### **Family history and pedigree**

List health conditions found in family (describe the relationship with proband)

mother - hypermobility

NO Consanguinity

### **Measurements**

**Age: 8y 6m**

**Weight:** 36.2 kg 97th pctl (+1.84SD)

**Height:** 147.7 cm 100th pctl (+2.99SD)

**BMI:** 16.59 67th pctl (+0.43SD)

### **Clinical symptoms and physical findings**

GROWTH PARAMETERS

Stature for age

Tall stature (>+2SD)

CUTANEOUS

NO Poor wound healing

CARDIOVASCULAR

Tachycardia

occasional when running

NO Abnormal echocardiogram

normal aortic root and arch, no MVP or MR, no ASD/VSD/PDA/LVOTO/RVOTO good biventricular



function, normal chamber size

#### RESPIRATORY

Pectus excavatum "slight"

Apnea

NO Cyanosis

#### MUSCULOSKELETAL

NO Increased susceptibility to fractures

NO Scoliosis

Joint hyperflexibility

particularly wrists and elbows

NO Joint dislocation

#### GENITOURINARY

NO Enuresis

#### BEHAVIOR, COGNITION AND DEVELOPMENT

NO Loss of consciousness

#### NEUROLOGICAL

Poor fine motor coordination

difficulty with buttons and zippers

#### BLOOD AND BLOOD-FORMING TISSUES

NO Abnormal bleeding

#### DIGESTIVE SYSTEM

Encopresis

NO Hepatomegaly

#### CONSTITUTIONAL SYMPTOM

Pain

daily neck, chest, knees, feet pain worse in morning, characterized as stiffness

#### IMMUNE SYSTEM

Allergy

Abnormality of the tonsils

hypertrophy

Abnormality of nasopharyngeal adenoids

Hypertrophy

### **Patient 81(I)**

Age: 11 years

Sex: Female

Indication for referral: chronic abdominal pain, dysmotility, hyperextesibility, neurogenic bladder dysmotility, superior mesenteric artery syndrome

### **Family history and pedigree**

Paternal ethnicity:

1. Scottish

Maternal ethnicity:

1. British

List health conditions found in family (describe the relationship with proband)

mother myopia and talipes

### **Clinical symptoms and physical findings**

#### GROWTH PARAMETERS

NO Disproportionate tall stature

#### CRANIOFACIAL

NO Abnormal facial shape

Malar flattening

Mild

NO Abnormality of the palate

#### EYE DEFECTS

NO Myopia

#### CUTANEOUS

NO Hyperextensible skin

NO Molluscoid pseudotumors

NO Poor wound healing

NO Abnormal elasticity of skin

#### CARDIOVASCULAR

NO Tachycardia

#### RESPIRATORY

Pectus excavatum

Mild

#### MUSCULOSKELETAL

NO Scoliosis

Arachnodactyly

Joint hypermobility

Beighton score 2/9 (elbow)

Joint laxity

NO Syndactyly

#### GASTROINTESTINAL

Gastrointestinal dysmotility

#### GENITOURINARY

Neurogenic bladder

Urinary retention

Ovarian cyst

#### BLOOD AND BLOOD-FORMING TISSUES

NO Bruising susceptibility

#### DIGESTIVE SYSTEM

Pancreatitis

Nausea and vomiting

Abdominal distention

#### CONSTITUTIONAL SYMPTOM

Abdominal pain

Chronic

Arthralgia

shoulders, hips, knees

## CONNECTIVE TISSUE

NO Scarring

### **Patient 91(E)**

Age: 11yrs

Sex: Female

Indication for referral: Global developmental delay, autism spectrum disorder, epilepsy, atrial septal defect

### **Family history and pedigree**

Paternal ethnicity:

1. Nova scotia

Maternal ethnicity:

1. Welsh

2. English

3. Irish

List health conditions found in family (describe the relationship with proband)

(1) Autism spectrum disorder: brother

(2) Depression/anxiety: mother (both), brother (anxiety)

(3) Learning difficulties: maternal female first cousins x2

(4) Query ADHD: father

(5) Query bipolar disorder: maternal female first cousin

(6) Atrial septal defect: maternal first cousin once removed

(7) Aortic aneurysm: maternal great uncle

(8) Recurrent miscarriages: maternal grandmother (up to 5, all male, including one set of twins)

NO Consanguinity

NO Parents with at least 3 miscarriages

### **Prenatal and perinatal history**

NO Multiple gestation

Gestation at delivery (weeks) 36

Maternal age at EDD (years) 28

Paternal age at EDD (years) 34

DELIVERY

Abnormal delivery (Non-NSVD)

Premature birth

Secondary Caesarian section

NEONATAL GROWTH PARAMETERS

Abnormal birth weight

NO Small for gestational age (<-2SD)

NO Large for gestational age (>+2SD)

EAR

Vertigo

Notes: BW 5 lbs 12.5 oz (2.62 kg)

## Medical history

Allergies - environmental, food, medication:

1. amoxicillin (rash)

## Measurements

**Age: 5y 5m**

**Weight:** 19.1 kg 51st pctl (+0.04SD)

**Height:** 101.8 cm 2nd pctl (-2.04SD)

**BMI:** 18.43 96th pctl (+1.76SD)

**Head circumference:** 50.4 cm 36th pctl (-0.35SD)

**Outer canthal distance:** 8.0 cm 61st pctl (+0.27SD)

**Inner canthal distance:** 3.3 cm 97th pctl (+1.95SD)

**Interpupillary distance:** 5.4 cm 86th pctl (+1.07SD)

**Left ear length:** 5.5 cm 64th pctl (+0.35SD)

**Right ear length:** 5.2 cm 29th pctl (-0.55SD)

**Age: 6y 11m**

**Weight:** 22.4 kg 52nd pctl (+0.05SD)

**Height:** 111.2 cm 4th pctl (-1.7SD)

**BMI:** 18.12 92nd pctl (+1.38SD)

**Head circumference:** 51.5 cm 48th pctl (-0.04SD)

**Age: 10y 3m**

**Weight:** 54.4 kg 98th pctl (+2.14SD)

**Height:** 139.2 cm 42nd pctl (-0.2SD)

**BMI:** 28.08 100th pctl (+2.9SD)

## Clinical symptoms and physical findings

### GROWTH PARAMETERS

Weight for age

Increased body weight (>+2SD)

Obesity

### CRANIOFACIAL

Abnormal facial shape

looks different from parents, coarse features, hooded eyelids, short upslanting palpebral fissures

High palate

One note reported bifid uvula, but other notes reported normal uvula.

Nasolacrimal duct obstruction

Left

### CUTANEOUS

Single transverse palmar crease

Left

Abnormality of the hair

Streak of blond hair in right anterior parietal area

### CARDIOVASCULAR

Secundum atrial septal defect

Noted at birth, large, surgically repaired. Also with spontaneously closed VSD.

## MUSCULOSKELETAL

Clinodactyly of the 5th finger

Bilateral

## BEHAVIOR, COGNITION AND DEVELOPMENT

Global developmental delay

MRI brain: Few tiny nonspecific periventricular white matter signals are seen, otherwise the brain parenchyma appears unremarkable.

Attention deficit hyperactivity disorder

Developed tics on Biphentin

## NEUROLOGICAL

Seizures

First diagnosed. Multiple types: GTC, absence, partial complex

Autistic behavior

Neuropsychology assessment

## **Patient 92(S)**

Age: 9 years

Sex: Female

Indication for referral: Evaluation for Ehlers-Danlos, overweight, sleep apnea, hyperlipidemia, mild limb length asymmetry, hyperextensible joints, pain, easy bruising, delayed wound healing with wide hypertrophic distended cigarette paper thin scars, sensitive to all allergens, develops hives, very sensitive to insect bites,

## **Family history and pedigree**

Paternal ethnicity:

1. English

Maternal ethnicity:

1. Italians

List health conditions found in family (describe the relationship with proband)

Mat fam hx of obesity, hypertension, hyperlipidemia, cardiac events

Other affected relatives

NO Consanguinity

NO Parents with at least 3 miscarriages

## **Medical history**

Medical and developmental history: she has hyperextensible joints, easy bruising, delayed wound healing, frequent joint subluxations in her hips, knees and elbows

Allergies - environmental, food, medication:

1. substances as normal saline and scratching herself

Global age of onset: Juvenile onset

## **Measurements**

**Age: 18y 1m**

**Weight:** 74.8 kg 96th pctl (+1.75SD)

**Height:** 161.0 cm 38th pctl (-0.32SD)

**BMI:** 28.86 97th pctl (+1.88SD)

**Clinical symptoms and physical findings**

**CUTANEOUS**

Soft skin

**MUSCULOSKELETAL**

Joint hypermobility

Beighton score 2/9

Limb pain

**CONNECTIVE TISSUE**

Cigarette-paper scars

**Patient 93(F)**

Age: 10 years

Sex: Male

Indication for referral: query Ehlers-Danlos hypermobility type,

**Family history and pedigree**

List health conditions found in family (describe the relationship with proband)

Mom dx asthma as child, feeding issues, hypotonia, dyslexia, psychiatric issues

NO Consanguinity

**Prenatal and perinatal history**

**DELIVERY**

**NO Premature birth**

Premature rupture of membranes

**NEONATAL GROWTH PARAMETERS**

Increased body weight

**CARDIOVASCULAR SYSTEM**

Syncope

**DIGESTIVE SYSTEM**

Nausea

**NERVOUS SYSTEM**

Seizures

Notes:

born at 10lbs, unusual head size

**Measurements Date:**

**Age: 18y 0m**

**Weight:** 46.9 kg 0th pctl (-2.73SD)

**Height:** 162.0 cm 3rd pctl (-1.9SD)

**BMI:** 17.87 5th pctl (-1.68SD)

**Head circumference:** 53.0 cm 1st pctl (-2.21SD)

**Outer canthal distance:** 10.0 cm 95th pctl (+1.67SD)

**Inner canthal distance:** 3.0 cm 28th pctl (-0.59SD)

**Interpupillary distance:** 6.5 cm 86th pctl (+1.08SD)

**Arm span:** 161.0 cm = Height - 1.0 cm

### **Clinical symptoms and physical findings**

#### GROWTH PARAMETERS

Weight for age

Decreased body weight (<-2SD)

Failure to thrive

was born 10lbs, lost weight in first few weeks and never gained it back.

#### CRANIOFACIAL

**NO Bifid uvula**

#### EAR DEFECTS

Hearing impairment

#### CUTANEOUS

Fragile skin

Mild

Poor wound healing

#### CARDIOVASCULAR

**NO Syncope**

**NO Palpitations**

#### RESPIRATORY

Asthma

#### MUSCULOSKELETAL

Muscular hypotonia

Joint dislocation

Infantile onset

Joint hypermobility

Knee, not elbow Beighton score 4/9

Clinodactyly of the 5th finger

Knee dislocation

Juvenile onset

multiple knee dislocations

Prominent proximal interphalangeal joints

Hypermobility of distal interphalangeal joints

#### GASTROINTESTINAL

Crohn's disease

#### NEUROLOGICAL

Motor delay

sat up at 1yr did not crawl but had bum shuffle first steps with walker at 24 months

needed stroller for long

distances at 5yrs

**NO Autonomic dysregulation**

#### BLOOD AND BLOOD-FORMING TISSUES

Bruising susceptibility

#### METABOLISM/HOMEOSTASIS

Food intolerance

## CONNECTIVE TISSUE

Scarring

"distended thin scar" right elbow

### **Patient 95(C)**

Age 13 years

Sex: Male

Indication for referral: vision problems at night, hearing issues resolved after tube insertion, delayed speech (started at age 4), mild redgreen color deficiency on testing, right iris heterochromia, normal discs, retinal pigment epithelial stippling, early pigmentary changes in mid and far periphery, photoreceptor retinal layers disrupted outside fovea, consistent with AR early onset retinal dystrophy

### **Family history and pedigree**

Paternal ethnicity:

1. Scottish

Maternal ethnicity:

1. Polish

2. Irish

List health conditions found in family (describe the relationship with proband)

Pat aunt "racing heart"

Pat grd mother d. 49 "sudden cardiac death"

NO Consanguinity

NO Parents with at least 3 miscarriages

### **Clinical symptoms and physical findings**

#### **EYE DEFECTS**

Abnormality of color vision

Mild

mild red-green color deficiency on testing

Retinal dystrophy

Abnormality of retinal pigmentation

retinal pigment epithelial stippling

Central heterochromia

**NO Optic disc hypoplasia**

### **Genotype information**

LIST OF GENES

#### **Gene Status Strategy Comments**

1 array CGH retinal dystrophy genes Negative OGT v2

2 ADH7 Negative

VUS: EYS c.3568+5T>C

VUS: GPR125 c.2107C>T (p.ARg703Trp)

VUS: MYO7A c.4159G>A (p.Asp1387Asn)

VUS: OTX2 c.707C>A (p.Thr236Asn)



## **Patient 97(D)**

Age 19 yrs

Sex: Female

Indication for referral: connective tissue disorder (EDS hypermobility), myopia, cardiovascular dysautonomia, generalized musculoskeletal pain, heaches with mydriasis and floaters, IBS, hx syncope with postural hypertension, myopia, sx excision of thyroglossal duct cyst, ?Chiari malformation, poor heat tolerance, multiple allergies, easy bruising, wide scars, chokes on food, things that aren't smooth,

## **Clinical symptoms and physical findings**

### CRANIOFACIAL

NO Bifid uvula

### EYE DEFECTS

Myopia

Mydriasis

Vitreous floaters

NO Visual loss

### CARDIOVASCULAR

Hypertension

postural

Syncope

Abnormality of the cardiovascular system

### MUSCULOSKELETAL

Joint hypermobility

Beighton 4/9

### GASTROINTESTINAL

Inflammation of the large intestine

### NEUROLOGICAL

Dysphagia

Headache

### BLOOD AND BLOOD-FORMING TISSUES

Bruising susceptibility

### METABOLISM/HOMEOSTASIS

Heat intolerance

### CONSTITUTIONAL SYMPTOM

Pain

### ENDOCRINE SYSTEM

Thyroglossal cyst

## **Patient 99(B)**

Age: 6 years

Sex: Male

Indication for referral: Seizure disorder including infantile spasms

### **Family history and pedigree**

List health conditions found in family (describe the relationship with proband)

(1) Mother with anxiety

(2) Maternal great aunt with breast cancer in her 40s

(3) Maternal great grandmother with ovarian cancer in her 50s

NO Other affected relatives

NO Consanguinity

NO Parents with at least 3 miscarriages

### **Prenatal and perinatal history**

NO Multiple gestation

Gestation at delivery (weeks) Term birth

Maternal age at EDD (years) 23

Paternal age at EDD (years) 23

PREGNANCY HISTORY

Maternal hypertension

DELIVERY

NO Abnormal delivery (Non-NSVD)

NO Premature birth

NEONATAL GROWTH PARAMETERS

Abnormal birth weight

NO Small for gestational age (<-2SD)

NO Large for gestational age (>+2SD)

METABOLISM/HOMEOSTASIS

NO Neonatal hypoglycemia

DIGESTIVE SYSTEM

NO Prolonged neonatal jaundice

NO Poor suck

RESPIRATORY SYSTEM

NO Neonatal respiratory distress

IMMUNE SYSTEM

NO Neonatal sepsis

Notes:

Pregnancy-induced hypertension did not require treatment. Maternal fever during labour, treated with antibiotics. BW ~3 kg.

### **Medical history**

Allergies - environmental, food, medication:

1. Ibuprofen (eye swelling)

2. Nuts (eye swelling)

## **Measurements**

**Age: 8m**

**Weight:** 9.3 kg 67th pctl (+0.43SD)

**Height:** 70.0 cm 21st pctl (-0.82SD)

**BMI:** 18.98 87th pctl (+1.15SD)

**Head circumference:** 41.9 cm 0th pctl (-2.81SD)

**Age: 1y 3m**

**Weight:** 12.7 kg 96th pctl (+1.72SD)

**Height:** 79.0 cm 32nd pctl (-0.46SD)

**BMI:** 20.35 99th pctl (+2.55SD)

**Head circumference:** 45.0 cm 3rd pctl (-1.92SD)

**Age: 4y 0m**

**Weight:** 17.13 kg 62nd pctl (+0.32SD)

**Height:** 104.3 cm 56th pctl (+0.15SD)

**BMI:** 15.75 63rd pctl (+0.32SD)

**Head circumference:** 49.3 cm 12th pctl (-1.16SD)

## **Clinical symptoms and physical findings**

### **MUSCULOSKELETAL**

Osteopenia

During hospitalization at 10 months of age, had multiple issues attributed to ACTH side effects: low Vitamin D, hypocalcemia, osteopenia, nephrocalcinosis, and kidney injury.

### **BEHAVIOR, COGNITION AND DEVELOPMENT**

Global developmental delay

Borderline

His development was disrupted by onset of seizures, but after appropriate treatment he caught up. At time of last assessment, there were no concerns with his development.

### **NEUROLOGICAL**

Seizures

In addition to infantile spasms, has had tonic, absence, and complex partial seizures

Infantile spasms

Onset at 5 months of age

### **ABNORMAL TEST RESULT**

Prolonged QT interval

Borderline

Seen by Cardiology, not felt to have prolonged QT syndrome

## **Patient 102(A)**

Age: 10 years

Sex: Male

Indication for referral: poor balance, hyperflexible joints Beighton score 6/9, speech and language delay, ADD, asthma, pectus carinatum, mild pes planus, high arched palate, dental crowding, triangular chin, mild micrognathia, large eyes

## **Family history and pedigree**

Paternal ethnicity:

1. Caucasian

Maternal ethnicity:

1. Caucasian

NO Consanguinity

## **Prenatal and perinatal history**

Notes: amniocentesis normal

walked ~18 months

single words by 2 years, speech delay

## **Clinical symptoms and physical findings**

### **CRANIOFACIAL**

High palate

Pointed chin

"triangular"

Micrognathia

Mild

Dental crowding

### **EYE DEFECTS**

Visual impairment

Large eyes

NO Ectopia lentis

### **EAR DEFECTS**

NO Hearing impairment

### **CARDIOVASCULAR**

NO Syncope

NO Abnormal echocardiogram

### **RESPIRATORY**

Pectus carinatum

Asthma

Childhood onset

### **MUSCULOSKELETAL**

Osteopenia

Joint hypermobility

able to put both legs around his head and spontaneously sublux his shoulder, hips and fingers Beighton score

6/9

Pes planus

Mild

**NO Pathologic fracture**

**BEHAVIOR, COGNITION AND DEVELOPMENT**

Delayed speech and language development

18-20 months behind peers expressive language more affected than receptive speech  
and language therapy

and occupational therapy

Attention deficit hyperactivity disorder

**NEUROLOGICAL**

Anxiety

**BLOOD AND BLOOD-FORMING TISSUES**

**NO Bruising susceptibility**

**NO Persistent bleeding after trauma**

**CONSTITUTIONAL SYMPTOM**

Pain

nonspecific

## Prediction method descriptions

### **CAG15**

#### **Group 1**

We first used our text-mining framework TPX [Joseph et al., 2012] for semi-automated HPO coding for each phenotype case. This resulted in a set of HPO codes for each case. We reviewed this output and came up with a final set of HPO codes by manually adding/removing/modifying as required.

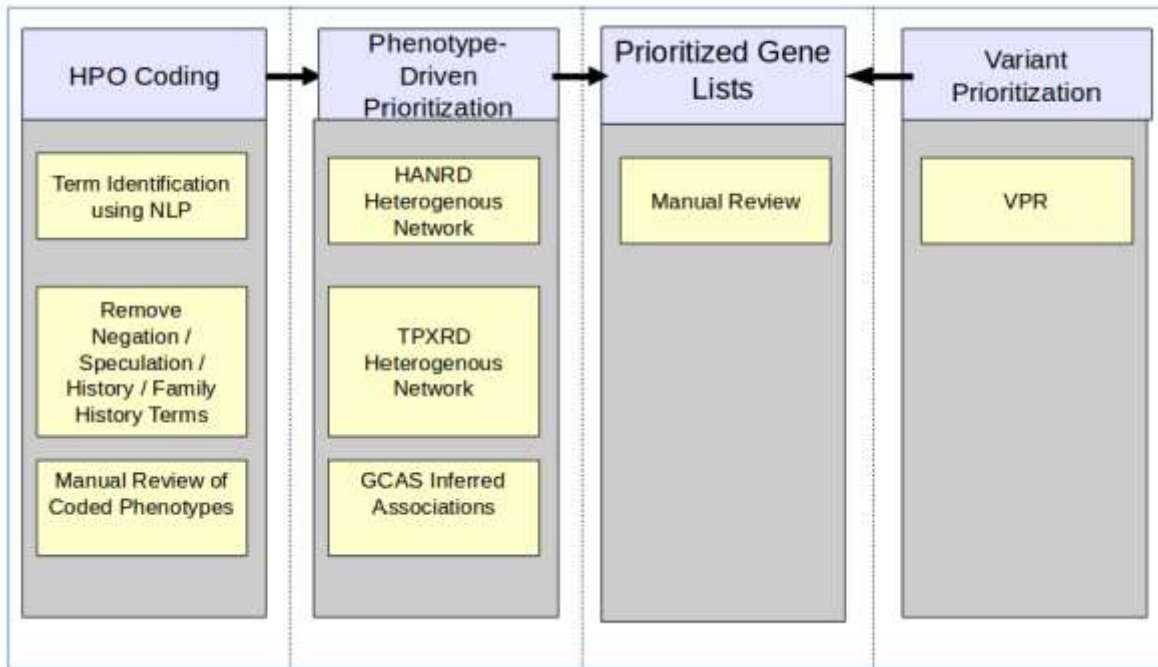
Using the set of HPO terms for each case, we queried the HANRD (*Heterogeneous Association Network for Rare Diseases*) network for a ranked list of genes. HANRD is a heterogeneous network consisting of entities such as genes, phenotypes, diseases and pathways as nodes while associations between these entities are represented as weighted edges. The weight of an edge represents the score of the association between the entity pairs. Existing association networks usually view ontological associations as distinct from the network of other heterogeneous associations [Ullah et al., 2013]. We instead combined pairwise ontological and curated associations into a single heterogeneous association network. Motivated by the recent progress in spectral graph convolutions [Hammond et al., 2011; Kipf and Welling, 2017], we developed an information propagation algorithm GCAS (Graph Convolution-based Association Scoring) that performs information propagation on the initial ontological and curated association network and infers novel binary associations between the entities of the network (Paper submitted). These inferred associations are added to the aforementioned initial network, and the resulting network of ontological, curated and inferred associations is called HANRD (paper accepted). HANRD gives us a set of ranked genes based on the input phenotype.

We also used each set of phenotypes to query TPXRD. TPXRD is a heterogeneous dataset of rare disease associations from MEDLINE abstracts generated using modules of the TPX framework. It contains association pairs such as disease-gene, phenotype-phenotype and phenotype-disease extracted from MEDLINE using TPX. We have previously described TPX, a web-based text-mining tool that

supports real-time entity assisted search and navigation of the MEDLINE repository whilst continuing to use PubMed as the underlying search engine. Although the TPX tool is primarily meant to search PubMed, specific modules of the TPX framework such as the dictionary-based named entity recognition (NER), acronym handler and association extraction (AE) were re-purposed for extracting rare disease entity association pairs from literature. Rare disease-specific dictionaries for rare diseases, phenotypes and genes from multiple sources were created, and resolution of conflicts and overlaps amongst these dictionaries was done. MEDLINE abstracts related to rare diseases were identified using terms from the disease dictionary. We then apply the information propagation algorithm GCAS (Graph Convolution-based Association Scoring) that performs information propagation on the initial ontological and curated association network and infers novel binary associations between the entities of the network. These inferred associations are added to the aforementioned initial network, and the resulting network of ontological, curated and inferred associations is called TPXRD (paper in progress). Thus, TPXRD also gives us a set of ranked genes based on the input phenotype.

Using the VCF files, we performed variant prioritization using VPR (variant prioritization), our in-house variant prioritization pipeline. Here, variants are prioritized independent of gene information using an in-house scoring scheme. The score, ranging from 0 to 1, is a weighted combination of global minor allele frequency, conservation information, and functional information. Allele frequency scores are derived from public data sources such as 1000 genomes and conservation scores from GERP, PhyloP, and PhastCons. Functional scoring is done depending on region and mutation type. Variant effect predictions are combined from sources like CADD, REVEL, LINSIGHT etc. and prior knowledge of variant from data sources like Clinvar is also considered. VPR gives us a set of ranked genes based on the input genotype.

We then manually look at the different ranked gene lists and try and match a phenotypic case to possible genotypic cases. This involved comparing the top genes from HANRD/TPXRD with those from VPR. This was done via the intermediary disease link. Figure 1 shows the overall approach of our method.



**Figure 1.** Overall Approach followed using phenotype-driven prioritization networks HANRD and TPXRD along with variant prioritization method VPR.

## References

- Hammond DK, Vanderghyest P, Gribonval R. Wavelets on graphs via spectral graph theory. *Applied and Computational Harmonic Analysis* 2011, 30(2):129{150.
- Kipf TN, Welling M: Semi-supervised classification with graph convolutional networks. *ICLR* 2017.
- Joseph, T., Saipradeep, V. G., Raghavan, G. S. V., Srinivasan, R., Rao, A., Kotte, S., and Sivadasan, N. (2012). Tpx: Biomedical literature search made easy. *Bioinformatics*, 8(12):578.
- Smedley D and Robinson PN. Phenotype-driven strategies for exome prioritization of human Mendelian disease genes. *Genome Med.* 2015 30;7:81.
- Ullah M, Aono M, Seddiqui M. Estimating a ranked list of human hereditary diseases for clinical phenotypes by using weighted bipartite network. In *Conference proceedings: Annual International Conference of the IEEE Engineering in Medicine and Biology Society. IEEE Engineering in Medicine and Biology Society. Annual Conference, Volume 2013* 2013:3475{3478.



## Group 2

To correctly associate clinical panels with genomes, we started from the symptoms and signs described in the clinical reports. We mapped them into the Human Phenotype Ontology (HPO) and the OMIM classifications. We used in-house resources, such as eDGAR (Babbi et al, 2017) and PhenPath (Babbi et al., 2019), to link phenotypes to panel-specific sets of candidate genes. We then searched in genomes for variations in these genes. To this aim, we retained from vcf files only the variations marked as “PASSED” and annotated their outcome using VEP (McLaren et al, 2016).

We scored the pathogenicity of each variant firstly by looking whether it is associated to some disease in the literature (mainly considering the UniProtKB annotation). Other missense variations in candidate genes were scored with SNPs&GO (Calabrese et al, 2009); a method based on Support Vector Machines for the prediction of deleterious single amino acid polymorphisms using protein functional annotation.

The output of SNPs&GO returns the effect (*Disease associated variant* or *Neutral variant*) associated with a Reliability Index (RI) that is a number scoring from 0 (unreliable) and 10 (reliable). We checked our predictions also considering the sex of the individual, to confirm that the genome-clinical panel association is plausible.

When no clear association emerged (e.g. when different variants in different genes had similar pathogenicity scores) we did not indicate any variant in the final submission file (since the submission of the causative variant was optional). However, due to the format of the submission file, it was not possible to leave the variant field completely empty, and thus we used the code 1:0:-:- to specify that we had no specific causative variant.

## References

- Babbi G, Martelli PL, Profiti G, Bovo S, Savojardo C, Casadio R. (2017) *eDGAR: a database of Disease-Gene Associations with annotated Relationships among genes*. BMC Genomics. 5:554
- Babbi G, Martelli PL, Casadio R. (2019) *PhenPath: a tool for characterizing biological functions underlying different phenotypes*. BMC Genomics. (accepted, in press)
- Calabrese R, Capriotti E, Fariselli P, Martelli PL, Casadio R. (2009) *Functional annotations improve the predictive score of human disease-related mutations in proteins*. Hum Mutat. 30:1237-1244

McLaren W, Gil L, Hunt SE, Riat HS, Ritchie GR, Thormann A, Flicek P, Cunningham F. (2016). *The Ensembl Variant Effect Predictor*. *Genome Biology*. 17(1):122.

### **Group 3**

VCF files were analyzed using standard parameters, including variant quality, allele frequency, functional damage prediction and gene-phenotype associations, using a variety of tools and databases. Gender was considered in phenotype-genotype matches, ethnic origin was not taken into account.

### **Group 4**

#### Collection of SickKids challenge phenotypes and the corresponding gene list

A total of 213 clinical phenotype descriptions were extracted from the CAGI phenotype data provided for 24 children. These phenotypes were used to extract a total 6239 potentially relevant genes from the Human Phenotype Ontology-based database (HPO) (Build #139) [1] and the dbNSFP database (version 3.5a) [2]. We also used the list of 319 genes from RetNet database [3] for searching for eye disorder related variants. The gene list for secondary variants was taken from the Table in the 2017 ACMG guidelines [4].

#### Annotation of VCF files and QC filters

The VCF files (including SNV and Indels) provided for this challenge from Illumina HiSeq were annotated using the Varant [5] tools, including region of occurrence (intron, exon, splice site or intergenic), observed minor allele frequencies (MAF), mutation type, predicted impact on protein function, and previously established associated phenotypes reported in ClinVar [6]. The RefGene [7] gene definition file was used for gene and transcript annotations in Varant. In addition, in-house scripts were written to further annotate the VCF files with HGMD [8] disease related variants and with dbSNV [9] variants that potentially alter splicing. We also used the Annovar annotation tool [10] to extract Genome Aggregation Database (GnomAD) frequency data [11], Eigen scores [12] and GERP++ scores [13]. Chromosome M was annotated and searched for pathogenic variants using MSeqDR mv tool [14]. We used only high quality variants for further analysis - for SNVs, and Indels, we used the 'PASS' filter.

### Diagnostic variant identification

A hierarchical scheme was used for identification of diagnostic variants, based on the strength of the evidence for disease relevance. All accepted high quality variants in the selected gene list with the highest population frequency <1% in the GnomAD exomes [11], or GnomAD genomes [11], or 1000 genome data [15] or in the ExAC database [11]) were first categorized into ordered tiers as follows:

Category 1: Variants with HGMD annotation of either DM (disease-causing mutation) or DP (disease-associated polymorphism), and/or reported in ClinVar with pathogenic or likely pathogenic clinical significance status.

Category 2: Nonsense mutation, direct splicing mutation disrupting either splice donor or acceptor site, frameshift or non-frameshift mutation, splice altering variant predicted by the dbSNV database [9], and non-synonymous mutations predicted as damaging by SNPs3D profile and stability methods [16, 17], SIFT [18], PolyPhen-2 [19], Vest [20], REVEL [21] and CADD [22]. For inclusion of a non-synonymous variant in Category 2, at least 60% of these methods were required to return a prediction of deleterious. This threshold is based on a calibration against HGMD. (Note that for any given variant not all methods may return a result, hence the non-obvious cutoff).

Category 3: Non-synonymous mutations predicted as damaging by one or more of the above non-synonymous impact prediction methods, with the deleterious prediction agreement fraction < 0.6.

Category 4: Benign non-synonymous mutations (zero reporting non-synonymous methods predicting deleterious).

Category 5: Variants annotated as close to a splice acceptor or splice donor site.

Category 6: Variants annotated as UTR and intronic. Pathogenicity of these noncoding variants are based on CADD [22], Eigen [12] and GERP++ [13] scores.

Variants from all categories were further categorized into ordered tiers according to their rarity in the population data.

Frequency bin 1: Novel mutations (not seen in any of 1000 genomes, ExAC, gnomAD exomes and genomes databases).

Frequency bin 2: Variants with population frequency > 0 and <= 0.001.

Frequency bin 3: Variants with population frequency > 0.001 and <= 0.005.

Frequency bin 4: Variants with population frequency  $> 0.005$  and  $< 0.01$ .

Binned variants were further filtered for an appropriate inheritance model using the OMIM inheritance pattern. Variants were assigned to autosomal dominant, autosomal recessive, compound heterozygous, pseudo autosomal recessive, or X-linked recessive models.

### Profile matching with genomes

For each phenotypic profile, each phenotypic term was assigned a subjective value from 0 to 1, according to its importance. For example, if a connective tissue disorder is the most serious and definitive term in the profile, it was scored the highest. If seizure is also part of that profile with borderline occurrence, then that was assigned a lower value than would be the case if the term occurred in a profile where seizure is the most serious phenotype. We then calculated a weighted matching score between the phenotypic profile and each variant-carrying gene in a genome. We selected top five scoring genomes for each clinical profile for further analysis. For each genome, we examined the evidence supporting the top five scoring variant-carrying genes, considering gender match, inheritance pattern and correspondence with the OMIM disease description.

### Searching for Predictive Secondary variants

Here we followed the rules in ACMG (2017) [4] to extract predictive secondary variants from 59 genes. We searched for clinical variants and loss of function variants in those genes according to the Table 1 in [4].

### References

1. Sebastian Köhler, Sandra C Doelken, Christopher J. Mungall, Sebastian Bauer, Helen V. Firth, et al. The Human Phenotype Ontology project: linking molecular biology and disease through phenotype data. *Nucl. Acids Res.* (1 January 2014) 42 (D1): D966-D974 doi:10.1093/nar/gkt1026
2. Liu X, Wu C, Li C and Boerwinkle E. 2015. dbNSFP v3.0: A One-Stop Database of Functional Predictions and Annotations for Human Non-synonymous and Splice Site SNVs. *Human Mutation*. Published Online First 10 NOV 2015 | DOI: 10.1002/humu.22932.

3. SP Daiger, BJB Rossiter, J Greenberg, A Christoffels, W Hide. Data services and software for identifying genes and mutations causing retinal degeneration. *Invest. Ophthalmol. Vis. Sci.* 39:S295, 1998.
4. Kalia, S. S. et al. Recommendations for reporting of secondary findings in clinical exome and genome sequencing, 2016 update (ACMG SF v2.0): a policy statement of the American College of Medical Genetics and Genomics. *Genet. Med.* 19, 249–255 (2017).
5. VARANT: <http://compbio.berkeley.edu/proj/varant/Home.html>
6. Landrum MJ, Lee JM, Riley GR, Jang W, Rubinstein WS, Church DM & Maglott DR: *ClinVar: public archive of relationships among sequence variation and human phenotype. Nucl. Acids Res.*2014; 42:D980-D985. doi:10.1093/nar/gkt1113.
7. Pruitt KD, Brown GR, Hiatt SM, Thibaud-Nissen F, Astashyn A, Ermolaeva O, et al: RefSeq: an update on mammalian reference sequences. *Nucl Acids Res.*2014; 42:756–63.
8. Stenson PD, et al: Human Gene Mutation Database (HGMD): 2003 update. *Hum Mutat.*2003; 21:577-581.
9. Jian X, Boerwinkle E and Liu X: In silico prediction of splice-altering single nucleotide variants in the human genome. *Nucleic Acids Res.*2014; 42:13534–44.
10. Wang K, Li M, Hakonarson H. ANNOVAR: Functional annotation of genetic variants from next-generation sequencing data *Nucleic Acids Research*, 38:e164, 2010.
11. Lek M. et al. Analysis of protein-coding genetic variation in 60,706 humans. *Nature*, 536, 285-291 (2016).
12. Ionita-Laza I. et al. A spectral approach integrating functional genomic annotations for coding and noncoding variants. *Nat. Genet.* 48, 214-220. (2016).
13. Davydov EV, Goode DL, Sirota M, Cooper GM, Sidow A, Batzoglou S (2010) Identifying a High Fraction of the Human Genome to be under Selective Constraint Using GERP++. *PLoS Comput Biol* 6(12): e1001025.
14. Shen L. et al. MSeqDR mvTool: A mitochondrial DNA Web and API resource for comprehensive variant annotation, universal nomenclature collation, and reference genome conversion. *Hum Mutat.* 2018 Mar 14. doi: 10.1002/humu.23422.
15. The 1000 Genome Project Consortium: An integrated map of genetic variation from 1,092 human genomes. *Nature.*2012; 491:56–65. doi:10.1038/nature11632
16. Yue P, Melamud E, Moulton J: SNPs3D: candidate gene and SNP selection for association studies. *BMC Bioinformatics.* 2006; 7:166.
17. Yue P, and Moulton J. Loss of protein structure stability as a major causative factor in monogenic disease. *J. Mol. Biol.* 353, 459-473. (2005).
18. Kumar P, Henikoff S, Ng PC: Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. *Nat Protoc.*2009; 4:1073-1081. doi:10.1038/nprot.2009.86.
19. Adzhubei I, Jordan DM, Sunyaev SR: Predicting functional effect of human missense mutations using PolyPhen-2. *Curr Protoc Hum Genet* 2013, Chapter 7:Unit7.20, doi:10.1002/0471142905.hg0720s76
20. Carter H. et al. Identifying mendelian disease genes with the variant effect scoring tool. *BMC Genomics.* 14(Suppl 3): S3. (2013).
21. Ioannidis et al. REVEL: An ensemble method for predicting the pathogenicity of rare missense variants. *Am. J. Hum Genet.* 99, 877-885. (2016).

22. Kircher M, Witten DM, Jain P, O'Roak BJ, Cooper GM, Shendure J: A general framework for estimating the relative pathogenicity of human genetic variants. Nat Genet.2014; 46:310-315. doi: 10.1038/ng.2892.

### **Group 5**

#### Evolutionary Action (EA) diffusion in a gene-disease network

In order to predict the disorder class (eye disorder, neurogenetic disease, or connective tissue disorder) for each individual's genome sequencing data set, the predictors calculated the effect of the genetic variants on the fitness of each gene (see A below). This fitness effect was used as the input of a diffusion process over a network of genes and diseases (see B below). The diffusion signal on each of the three disorder classes was used to calculate the probability of each genome to be linked to each disease.

In order to match each individual's genome sequencing data set to a clinical report file, the predictors used again the diffusion process (see B below), but the signal was measured at each specific symptom of the clinical description files. This process generated overwhelming information. To narrow down the potential matches, the predictors identified the gender and predicted the ethnic origin of each genome (see D below). Because the clinical reports varied a lot in the amount, type, and the detail level, the genomes were matched to clinical reports manually, by weighing the various accumulated links of genotype to phenotype. Only 12 of the 24 genomes were matched to clinical reports, while for the rest 12 genomes the submitted probabilities were arbitrarily set to 10% (value of 0.1) for all remaining potential matches after narrowing them down based on gender and ethnic group predictions.

No predictions of diagnostic variants nor of secondary variants were submitted.

#### Detailed Calculations

A. Gene fitness effect. The predictors computed the fitness of each gene in each individual's genome based on the predicted impact of all variants called in that gene. The variant impact was calculated according to the Evolutionary Action (EA) method for each missense variant (see C below), where a value of 0 corresponds to wild-type level gene activity and a value of 100 corresponds to complete loss of gene function. Nonsense and start loss variants were given impact of 100, while synonymous variants were given impact of 0. Insertions, deletions, and variants without a PASS filter were

given impact of 0 (although insertions and deletions may have strong impact on gene function, the ability to separate them into passenger and loss of function variants might be questionable). The fitness effect on a gene was defined to be equal to: 0 if there was no mutation,  $EA/100$  if there was one mutation, or  $1 - \prod(1 - EA_m/100)$  if there were multiple mutations in that gene ( $\prod$  indicates the product for all mutations  $m$ ). To account for the different ability of genes to tolerate mutations, the predictors weighted the fitness effect with a gene importance score (multiplied them). The ability of each gene to tolerate mutations was calculated as the average EA score of all variants in that gene found by the gnomAD project (Lek et al. 2016). This average EA value was transformed into a fraction coverage (0 means the gene can tolerate mutations with the highest impact and 1 means the gene can only tolerate mutations with the lowest impact), which coverage was used as the gene importance weighting factor of the gene fitness effect.

B. Diffusion on the gene disease network. A gene and disease network was constructed based on multiple sources (Stark et al. 2006; Gutierrez-Sacristan et al. 2015; Szklarczyk et al. 2015; Davis et al. 2017). For each individual's genome, the predictors run a diffusion process of weighted gene fitness effects (see A above) over the network and measured the diffusion signal outcome at disease labels (Lin et al. 2018). The disease labels could be either the disorder classes or specific symptoms of the clinical reports. The diffusion signal on each disease label was normalized for the 24 individual's genomes and it was reported as a fraction coverage. For example, for each disease label, the individual with the lowest diffusion signal had coverage of 0 (0/24) and the individual with the highest diffusion signal had coverage of 0.96 (23/24). This coverage was used as the probability that the individual has the particular phenotype.

C. The Evolutionary Action (EA) method predicts the fitness effect of the genetic variants (Katsonis and Lichtarge 2014). EA does not involve any training, because it relies on a formal equation of the genotype-phenotype relationship. The terms of this equation were calculated using protein homology data. Briefly, the EA equation states that the fitness effect of a mutation equals the product of the sensitivity of the mutated position with the magnitude of the change. The sensitivity of the position is calculated by quantifying the correlation of the residue variations with phylogenetic branching within

an alignment of homologous sequences (Lichtarge et al. 1996; Mihalek et al. 2004; Lichtarge and Wilkins 2010). The magnitude of the change is calculated from substitution likelihood according to numerous sequence alignments for the given context (strata of sensitivity of the position, and optionally additional stratification based on structural features). The calculated product is then normalized to represent the percentile rank of each variant within the protein in the scale of 0 (benign) to 100 (pathogenic). The EA scores are available for all human variants at:

<http://mammoth.bcm.tmc.edu/EvolutionaryAction>

D. Using gender and ethnic information. To identify the gender, we used the concordance of reads in the X chromosome. For male genome sequencing data, a large fraction of X chromosome calls appears to be homozygous, while for female sequencing data that fraction is distinctly lower. To identify the ethnic background, we calculated the proximity of each genome to the genomes of ethnic groups available in the 1000 Genomes Project (The 1000 Genomes Project Consortium et al. 2015). The predictions were at the level of major ethnic groups (EUR, AFR, AMR, EAS, and SAS).

## References

- Davis AP, Grondin CJ, Johnson RJ, Sciaky D, King BL, McMorran R, Wieggers J, Wieggers TC, Mattingly CJ. 2017. The Comparative Toxicogenomics Database: update 2017. *Nucleic Acids Res* 45: D972-D978.
- Gutierrez-Sacristan A, Grosdidier S, Valverde O, Torrens M, Bravo A, Pinero J, Sanz F, Furlong LI. 2015. PsyGeNET: a knowledge platform on psychiatric disorders and their genes. *Bioinformatics* 31: 3075-3077.
- Katsonis P, Lichtarge O. 2014. A formal perturbation equation between genotype and phenotype determines the Evolutionary Action of protein-coding variations on fitness. *Genome research* 24: 2050-2058.
- Lek M, Karczewski KJ, Minikel EV, Samocha KE, Banks E, Fennell T, O'Donnell-Luria AH, Ware JS, Hill AJ, Cummings BB et al. 2016. Analysis of protein-coding genetic variation in 60,706 humans. *Nature* 536: 285-291.



Lichtarge O, Bourne H, Cohen F. 1996. An evolutionary trace method defines binding surfaces common to protein families. *J Mol Biol* 257: 342 - 358.

Lichtarge O, Wilkins A. 2010. Evolution: a guide to perturb protein function and networks. *Curr Opin Struct Biol* 20: 351-359.

Lin CH, Konecki DM, Liu M, Wilson SJ, Nassar H, Wilkins AD, Gleich DF, Lichtarge O. 2018. Multimodal Network Diffusion Predicts Future Disease-Gene-Chemical Associations. *Bioinformatics* doi:10.1093/bioinformatics/bty858.

Mihalek I, Res I, Lichtarge O. 2004. A family of evolution-entropy hybrid methods for ranking protein residues by importance. *J Mol Biol* 336: 1265 - 1282.

Stark C, Breitkreutz BJ, Reguly T, Boucher L, Breitkreutz A, Tyers M. 2006. BioGRID: a general repository for interaction datasets. *Nucleic Acids Res* 34: D535-539.

Szklarczyk D, Franceschini A, Wyder S, Forslund K, Heller D, Huerta-Cepas J, Simonovic M, Roth A, Santos A, Tsafou KP et al. 2015. STRING v10: protein-protein interaction networks, integrated over the tree of life. *Nucleic Acids Res* 43: D447-452.

The 1000 Genomes Project Consortium, Auton A, Brooks LD, Durbin RM, Garrison EP, Kang HM, Korbel JO, Marchini JL, McCarthy S, McVean GA et al. 2015. A global reference for human genetic variation. *Nature* 526: 68-74.

## **Group 6**

Our strategy involved predicting three different pieces of information from the genomes based on what is presented in the clinical descriptions.

- A. Prediction of sex of the genomes to match to reported sex in the clinical notes
- B. Prediction of ethnicity of the genome to match to the parental ethnicities reported in clinical notes
- C. Prediction of phenotypes associated with genes carrying pathogenic mutations to match to the clinical symptoms reported in the clinical notes

For A., BCFtools +guess-ploidy function was used to guess the ploidy. Using cutoffs for the  $\log P(\text{Haploid})/n\text{Sites}$  values of male if  $>-2$  and female  $<-3$ , all but two genomes (WGS-NGS-017-03, WGS-NGS-018-03) were assigned a sex. Each genome/clinical

note pair was assigned a probability based on the predicted sex for that genome and the total occurrence of that sex in all 24 clinical notes.

For B., predictions were made using akt ancestry tools and the principal components were projected onto the components already available for 1000 genomes samples. The top 20 closest samples in the 1000 genomes were used to infer ethnicity.

For C., predictions were made using multiple criteria. First, for each genome, an in-house tool (varant) was used for annotation. All protein-altering variants with 1000G MAF < 2% were considered (PA-vars). The pdf clinical notes were converted to text using pdf2text tool and clinical terms describing patient features were extracted for each clinical note. These terms for each note was searched against a gene-phenotype database (Monarch initiative for submission 1 and eRAM for submission 2), and the genes were sorted by with highest number of matching terms. The genes matching phenotype terms from all clinical notes were then intersected with genes containing protein altering variants in all genomes. To score each pair of genome-clinical note, the following were considered.

a. Recessive genes (according to dbNSFP  $P(\text{rec}) > 0.6$ ) in the top 25 percentile of term matches with >1 PA-vars were given a score +1

b. Dominant and also essential genes (according to dbNSFP  $P(\text{rec}) < 0.3$ ) in the top 25 percentile of term matches with >0 PA-vars were given a score +1

c. Essential genes with low RVIS (according to dbNSFP) in the top 25 percentile of the term matches with >0 PA-vars were given a score +1

d. Genes with the highest number of term matches in Homo Sapiens in Monarch initiative genotype-phenotype database with >0 PA-vars were given a score +1

e. Genes with the highest number of term matches with direct mouse experimental evidence in Monarch initiative with >0 PA-vars were given a score +1

f. Genes in the top 25 percentile of term matches with StopGain, FrameShiftInsert, FrameshiftDelete were given a score of +1

For each genome/clinical note pair, the highest scoring gene was recorded in the respective comment field. The corresponding variants were recorded in the DV fields and the remaining variants from the rest of the lower scoring genes were recorded in the PSV column. For a genome, the highest score for each clinical note was gathered

and unit normalized across all clinical notes. These scores for each genome were then divided by 1/3 of the sum of scores and added to the probability scores from A. and B. to obtain the P-A, P-B, P-C..etc. reported probability values.

A second submission (group 6.2) was also made, where instead of the Monarch initiative, we used eRAM (<http://www.unimd.org/eram/>) - encyclopedia of rare disease annotation to obtain gene-phenotype relationships. Four random terms from the clinical notes were randomly selected 500 times and queried in the database together (AND of the terms) and the resulting genes were intersected with PA-vars like with the Monarch Initiative data. The second submission took a more restrictive approach, for a given genome of a predicted sex, only clinical notes with consistent matching sex were considered further. In this restricted set of clinical notes, the genome/clinical note pairs were scored by using a., b., c. and f. from above.

## **Group 7**

VCFs for each individual were uploaded into Ingenuity Variant Analysis (QIAGEN). Ingenuity Variant Analysis utilizes curated content from the literature as well as external databases for us in basic variant filtering on the following parameters: Variants were filtered based on quality (phred score of 20), population frequency (<3% in gnomAD), ACMG classification (pathogenic and likely pathogenic). Additionally, since the QIAGEN knowledgebase contains a gene-disease framework built on curated literature and clinical databases, I looked for variants in genes with a known relationship to eye disorders, connective tissue disorders, or neurological dysfunction. After reviewing all variant specific citations and data provided within Ingenuity Variant Analysis, the most compelling pathogenic and likely pathogenic variants were reported, and where possible, the variants were qualitatively matched to the medical record by visual inspection.

## **Group 8**

### Summary

To predict correspondence between phenotypes and genomes, we calculated scores for all genome-phenotype pairs and assigned the most likely connections using a bipartite matching algorithm. We obtained these scores from three independent factors: 1) pathogenic mutations (predicted by the MutPred suite) in genes related to the reported phenotype, 2) genetic similarity to the reported ancestry, and 3) the presence of Y-chromosome variants. We searched for pathogenic variants in gene sets composed of known risk genes and their putative interactors, which we predicted using a propagation algorithm on protein-protein interaction networks. Variants associated with ancestry and sex hold the strongest signal for determining the identity of a genomic sample, and they were key in guiding our matching algorithm. Beyond these guiding variants, however, matching genomes to phenotypes proved to be a challenging task. By narrowing the search for pathogenic variants to those in known disease genes, we sought to reduce false positives and simplify variant interpretation. Our approach of combining the pathogenicity scores of all variants in a single probability score per genome-phenotype implied that we lost focus on better predicting single causal variants.

## Methods

We used previously reported disease genes as seeds on the human protein-protein interaction network for running a network propagation algorithm [6]. The propagation algorithm was performed in a 5-fold cross-validation manner so as to get an initial score between 0 and 1 for all genes. We then used the AlphaMax algorithm [2, 3] to estimate the proportion of the risk genes in the human genome and calibrate those initial scores to be proper probability scores measuring the likelihood of a gene being associated with the disease [3]. We built a total of eight gene sets related to the diseases of interest: Ehlers-Danlos Syndrome (19 seed genes), autism (69), retinitis pigmentosa (52), nystagmus (9), ataxia (3), as well as general eye anomalies (DOID:102), connective tissue diseases (DOID:65), and congenital neurologic anomalies (DOID:2490). We classified each case as belonging in one of these disease categories, and considered only the corresponding gene lists when calculating their phenotype-genome scores. However, since more than one disease phenotype was reported in some cases, we

allowed those to have a secondary disease category (included in a weighed average with its primary counterpart).

We annotated protein coding variation using custom scripts to extract nucleotide sequence from the CCDS database and predict the mutant amino acid sequence. We assigned pathogenicity prediction scores to missense and stop gain variants with MutPred2 [8] and MutPred-LOF [7], respectively. For each gene in every individual genome, we included only the variant with the highest pathogenicity prediction score (which may include multiple CCDS isoforms) in further analyses. To assess significance of these pathogenic variants, we generated an empirical null distribution using the beta family based on the MutPred [5] scores of variants present in gnomAD [4], within the genes for each phenotype. The eight disease scores for each genome were computed as products of the p-values (from the beta distribution) for the individual's variants in the corresponding gene set.

We estimated genetic similarity to the reported ancestry by comparing the genomes to relevant samples from the 1000 Genomes dataset [1] with identity-by-state calculated with the SNPRelate package in R [9]. We chose 1000-G populations (or super-populations, when appropriate) that were close to the reported ethnicity of the parents and estimated relative similarity scores for each genome.

The last component of a phenotype-genotype score was the sex of the individual. We inferred sex from genomic data from SNPs mapped to the Y chromosome (a genome was assumed male if there were any Y-chromosome variants, and female otherwise).

The total score used for bipartite matching was the weighted average of the similarity and disease scores, multiplied by a sex factor (=1 only if the inferred and reported sex matched). We repeated the bipartite matching process 1000 times varying the weight assigned to the similarity score ( $p \sim U[0.2; 0.8]$ ) relative to the disease score ( $1 \sim p$ ), and the weight of the secondary disease ( $q \sim U[0; 0.3]$ ) relative to the primary category ( $1 \sim q$ ).

## References

- [1] 1000 Genomes Project Consortium, G. R. Abecasis, D. Altshuler, A. Auton, L. D. Brooks, R. M. Durbin, R. A. Gibbs, M. E. Hurles, and G. A. McVean. A map of human genome variation from population-scale sequencing. *Nature*, 467(7319):1061{1073, 2010.
- [2] S. Jain, M. White, and P. Radivojac. Estimating the class prior and posterior from noisy positives and unlabeled data. arXiv:1606.08561, 2016.
- [3] S. Jain, M. White, and P. Radivojac. Estimating the class prior and posterior from noisy positives and unlabeled data. In *Advances in Neural Information Processing Systems*, NIPS 2016, pages 2693{2701, 2016.
- [4] K. J. Karczewski et al. Variation across 141,456 human exomes and genomes reveals the spectrum of loss-of-function intolerance across human protein-coding genes. bioRxiv, page 531210, 2019.
- [5] B. Li, V. G. Krishnan, M. E. Mort, F. Xin, K. K. Kamati, D. N. Cooper, S. D. Mooney, and P. Radivojac. Automated inference of molecular mechanisms of disease from amino acid substitutions. *Bioinformatics*, 25(21):2744{2750, 2009.
- [6] E. Nabieva, K. Jim, A. Agarwal, B. Chazelle, and M. Singh. Whole-proteome prediction of protein function via graph-theoretic analysis of interaction maps. *Bioinformatics*, 21 Suppl 1:i302{310, 2005.
- [7] K. A. Pagel, V. Pejaver, G. N. Lin, H. J. Nam, M. Mort, D. N. Cooper, J. Sebat, L. M. Iakoucheva, S. D. Mooney, and P. Radivojac. When loss-of-function is loss of function: assessing mutational signatures and impact of loss-of-function genetic variants. *Bioinformatics*, 33:i389{i398, 2017.
- [8] V. Pejaver, J. Urresti, J. Lugo-Martinez, K. A. Pagel, G. N. Lin, H. J. Nam, M. Mort, D. N. Cooper, J. Sebat, L. M. Iakoucheva, S. D. Mooney, and P. Radivojac. MutPred2: inferring the molecular and phenotypic impact of amino acid variants. bioRxiv 134981, 2017.
- [9] X. Zheng, D. Levine, J. Shen, S. Gogarten, C. Laurir, and B. Weir. A high-performance computing toolset for relatedness and Principal Component Analysis of SNP data. *Bioinformatics*, 28(24):3326{3328, 2012.

## **CAG14**

### **Group 9**

Following download and unzipping of the genome data and phenotypes files the data were prepared for analysis with the Exomiser(1,2) using the following steps:

1. The ASM/vcfBeta-[ASM-ID].vcf.bz2 were uncompressed to vcf format.
2. The HPO terms in the phenotype PDF files were extracted manually into plain text format files.
3. The mode of inheritance, where provided, were also extracted manually and merged with the genome and phenotype identifiers from the clinical\_genomes\_answer\_key.txt
4. A Python script was run which used the outputs from steps 1-3 to write out a set of 25 Exomiser 7 yml format analysis scripts (sup\_1) and an exomiser batch analysis script listing the paths to 25 yml files. The exomiser requires a list of HPO ids for phenotypic analysis so this script converted the HPO terms into a list of HPO ids using the OLS beta webservice(3).

The yml script configured the Exomiser to:

- Exclude non-exonic (intergenic, intronic, upstream, downstream or intronic) or synonymous variants
- Exclude variants having a maximum minor allele frequency (MAF) of 1.0% in all the Thousand genome, ESP and ExAC datasets.
- Include variant pathogenicity predictors from the Polyphen2, MutationTaster and SIFT resources as well as scores derived from the variant type (e.g. missense or frameshift insertions)
- Score genes according to phenotypic similarity with the observed patient phenotype using the hiPHIVE prioritiser. This uses know human gene disease phenotypes, and mouse and fish knockout phenotypes along with random walk analysis of physical interaction networks.

These steps were chosen according to the guidelines in (4), which explains the steps and data sources in much full detail.

The data were analysed using the exomiser version 7.2.1 downloaded from (5) on a laptop equipped with 16GB RAM and an Intel 4th generation Core i7 processor running Windows 7 64-bit, Java version 8. This exomiser version is capable of analysing whole genomes using only a moderate amount of RAM. The analysis was run using the command:

```
java -Xms8G -Xmx12G -jar exomiser-cli-7.2.1.jar --analysis-batch {path to analysis batch file}
```

The resulting HTML files containing the top 20 candidate genes were inspected by hand and the most likely (rarest, most pathogenic) variants from the best phenotypic matching gene were included in the submission file.

## References

- (1) Improved exome prioritization of disease genes through cross-species phenotype comparison. Robinson PN, Köhler S, Oellrich A, Sanger Mouse Genetics Project, Wang K, Mungall CJ, Lewis SE, Washington N, Bauer S, Seelow D, Krawitz P, Gilissen C, Haendel M and Smedley D. Genome research 2014;24;2;340-8 PUBMED: 24162188; PMC: 3912424; DOI: 10.1101/gr.160325.113
- (2) <http://www.sanger.ac.uk/science/tools/exomiser>
- (3) <http://www.ebi.ac.uk/ols/beta/>
- (4) Next-generation diagnostics and disease-gene discovery with the Exomiser. Damian Smedley, Julius O B Jacobsen, Marten Jäger, Sebastian Köhler, Manuel Holtgrewe, Max Schubach, Enrico Siragusa, Tomasz Zemojtel, Orion J Buske, Nicole L Washington, William P Bone, Melissa A Haendel & Peter N Robinson. Nature Protocols 10, 2004–2015 (2015) DOI: 10.1038/nprot.2015.124
- (5) <ftp://ftp.sanger.ac.uk/pub/resources/software/exomiser/downloads/exomiser/>



Supplementary Information:

sup\_1 - The Exomiser YAML script template. The fields \$vcf\_file, \$inheritance\_mode, \$hpo\_list and \$out\_file\_prefix were replaced for each genome and phenotype set.

#Exomiser Analysis Template.

# These are all the possible options for running exomiser. Use this as a template for  
# your own set-up.

---

analysis:

vcf: \$vcf\_file

ped:

# AUTOSOMAL\_DOMINANT, AUTOSOMAL\_RECESSIVE, X\_RECESSIVE or  
UNDEFINED

modeOfInheritance: \$inheritance\_mode

analysisMode: PASS\_ONLY

geneScoreMode: RAW\_SCORE

hpolds: \$hpo\_list

frequencySources: [

THOUSAND\_GENOMES,

ESP\_AFRICAN\_AMERICAN, ESP\_EUROPEAN\_AMERICAN, ESP\_ALL,

EXAC\_AFRICAN\_INC\_AFRICAN\_AMERICAN, EXAC\_AMERICAN,

EXAC\_SOUTH\_ASIAN, EXAC\_EAST\_ASIAN,

EXAC\_FINNISH, EXAC\_NON\_FINNISH\_EUROPEAN,

EXAC\_OTHER

]

pathogenicitySources: [POLYPHEN, MUTATION\_TASTER, SIFT]

steps: [

variantEffectFilter: {remove: [UPSTREAM\_GENE\_VARIANT,

INTERGENIC\_VARIANT,

CODING\_TRANSCRIPT\_INTRON\_VARIANT,

```

NON_CODING_TRANSCRIPT_INTRON_VARIANT,
SYNONYMOUS_VARIANT,
DOWNSTREAM_GENE_VARIANT,
SPLICE_REGION_VARIANT]],
frequencyFilter: {maxFrequency: 1.0},
pathogenicityFilter: {keepNonPathogenic: true},
inheritanceFilter: {},
omimPrioritiser: {},
hiPhivePrioritiser: {}
]
outputOptions:
  outputPassVariantsOnly: true
  numGenes: 20
  outputPrefix: $out_file_prefix
  outputFormats: [HTML, TSV-GENE, TSV-VARIANT, VCF]

```

## Group 10

Identifying variants responsible for the phenotypic abnormalities (CAGI4 SickKids challenge) involved running our in-house outlier-phenotype predictor algorithm. The tool relies on SUPERFAMILY (1) to annotate genetic variants falling within SCOP (2) domains with FATHMM (3) pathogenicity scores as well as data on allele frequency from the 1000 Genomes project (4). Each variant is linked with the phenotype terms it affects by transferring gene ontology annotations to the SCOP domain level using dcGO (5). The prediction algorithm runs an unsupervised learning pipeline that clusters genotypes into groups exhibiting levels of high similarity (indicating that multiple individuals have the same combination of variants associated with a phenotype) and identifying outliers from these clusters (those individuals that have a rare combination of variants, which were predicted to have strong functional effects) i.e. individuals likely to have an outlier phenotype. If a “sick kid” was an outlier for a phenotype term that

matched the clinical description (subjective judgement), we prioritised the causative variants by FATHMM score and allele rarity.

## References

- (1) Matt E. Oates, Jonathan Stahlhacker, Dimitrios V. Vavoulis, Ben Smithers, Owen J.L. Rackham, Adam J. Sardar, Jan Zaucha, Natalie Thurlby, Hai Fang, Julian Gough, The SUPERFAMILY 1.75 database in 2014: a doubling of data, *Nucleic Acids Research*, Volume 43, Issue D1, 28 January 2015, Pages D227–D233.
- (2) Antonina Andreeva, Dave Howorth, John-Marc Chandonia, Steven E. Brenner, Tim J. P. Hubbard, Cyrus Chothia, Alexey G. Murzin, Data growth and its impact on the SCOP database: new developments, *Nucleic Acids Research*, Volume 36, Issue suppl\_1, 1 January 2008, Pages D419–D42.
- (3) Shihab, HA, Gough, J, Cooper, DN *et al.* Predicting the functional, molecular, and phenotypic consequences of amino acid substitutions using hidden markov models. *Hum Mutat* 2013; **34**: 57– 65.
- (4) 1000 Genomes Project Consortium, Auton A, Brooks LD, Durbin RM, Garrison EP, Kang HM, Korbel JO, Marchini JL, McCarthy S, McVean GA, Abeca
- (5) Hai Fang, Julian Gough, dcGO: database of domain-centric ontologies on functions, phenotypes, diseases and more, *Nucleic Acids Research*, Volume 41, Issue D1, 1 January 2013, Pages D536–D544.

## **Group 11**

### Phenotype to Disease/Phenotype to Genes

Since the original challenge was a matching challenge, my approach was to use the phenotypic information to find possible diseases and/or corresponding genes, if any. The phenotypic information already uses the HPO controlled-vocabulary since the referring clinical geneticist has entered this information using *Phenotips*, a Human Phenotype Ontology-based database [Girdea M et al., 2013]. However, there were certain verbatim term mentions from the phenotypic files that needed to be mapped to a HPO term.

Several tools that metrics that measure the semantic similarity between ontology annotations are present. Matching a term is not about exact match, it should also search across the HPO ontology. The Resnik measure seems the most common one when searching in the ontology [Resnik, 1995]. The HPOSim paper compares measures specific to the HPO context [Deng Y et al., 2015].

### Phenomizer

I picked Phenomizer [Köhler et al., 2009 and Köhler et al.,2013] as my choice of tool to go from a set of HPO IDs to a scored and ranked set of OMIM/Orphanet IDs that could explain these IDs. Phenomizer was run online from the website <http://compbio.charite.de/phenomizer/>.

A set of phenotypic terms could be explained by 2 (or more?) disease terms, referred to in the paper as a complex phenotype with two genetic disorders [Stavropoulous et al, 2016]. Given that there could be complex phenotypes as well as the list of HPO IDs for some cases being >8, it could be important to identify critical IDs and give them higher weight. Phenomizer allows one to mark a HPO term as mandatory or observed, with the default being observed. By default, the tool uses the Resnik measure. It automatically converts the input query to a symmetric one, with the message “It may be appropriate to use the 'symmetric' mode for queries of size larger than 5. Should 'symmetric' be checked? (You can change that in the menu later on)”. Ideally this should be asymmetric, however in order to get possible disorders, I even ran this as symmetric.

I manually looked at the mapped disease terms to see coverage. For this, I picked OMIM over Orphanet as OMIM has a section called Clinical Synopsis in which one can choose to display all HPO IDs associated with the clinical phenotypes using the Display Options->Show Clinical IDs.

### Phenolyzer

I picked Phenolyzer [Yang H et al, 2015] as my my choice of tool to go from a set of HPO IDs to a set of scored and ranked genes that could potentially explain the phenotype. Phenolyzer was run locally after installation.

I thus created the Phenomizer and Phenolyzer outputs.

## Input File(s)

I only considered the vcf format as input. This file contains the small variant, copy number variation (CNV), structural variation (SV), and mobile element insertion (MEI) calls made by the Complete Genomics Assembly Pipeline for a single genome, and conforms to the VCF 4.1 specification. This could mean ignoring crucial input information present in the Master Variations file. Also, I am relying on the tool that converted the Master Variations file to vcf format. This tool produces a vcfBeta, so not sure if there is any loss-of-information when going from the Complete Genomics format to vcf.

My first tool for gene and variant prioritization was Exomiser [Smedley D et al., 2015]. It takes a vcf file as input and a set of phenotypes encoded using the Human Phenotype Ontology (HPO) it will annotate, filter and prioritize likely causative variants using:

- The functional annotation of variants is handled by Jannovar and uses UCSC KnownGene transcript definitions and hg19 genomic coordinates.
- Predicted pathogenicity data is extracted from the dbNSFP resource. Variant frequency data is taken from the 1000 Genomes, ESP and ExAC datasets
- Variants filtered for maximum allele frequency of 1%, or in some cases 2%
- Pathogenicity Filter: Retained all non-pathogenic missense variants
- The hiPHIVE algorithm combines data on the rarity of the variant and its predicted pathogenicity along with the similarity of the human, mouse, zebrafish phenotypes as well as a guilt-by-association approach using protein–protein associations for those genes that have no data in any of the species.

A typical Exomiser command that I used was

```
java -Xmx10g -jar exomiser-cli-7.2.0.jar -F 1 -v 1099.vcf --ped 1099.ped --hpo-ids  
HP:0009889, HP:0006466, HP:0007911, HP:0005709, HP:0001773, HP:0007598,  
HP:0001263, HP:0010864, HP:0000369, HP:0002194, HP:0200055, HP:0003701,  
HP:0000219, HP:0000664, HP:0001629 --prioritiser=hiphive
```

## Genemania

Exomiser uses protein-protein interaction data to identify possible genes involved. The basis of the interaction wasn't always apparent to me. So I used Genemania [Warde-

Farley et al., 2010] to visualize the specific interactions, with emphasis on physical interactions between the entities.

### Phen-Gen

Another tool that I used for gene and variant prioritization was Phen-Gen [Javed A et al, 2014]. Phen-Gen works as follows (below summary from Smedley and Robinson, 2015): It uses a Bayesian framework to compare predicted deleterious variants in the patient's exome and known patient symptoms to prior knowledge of human disease-gene associations and gene interactions. Coding variants are analyzed using a unifying framework to predict the damaging impact of non-synonymous, splice-site and indel variants.

- Any variant that has a MAF above 1 % is removed from further analysis.
- Genes are only retained for further analysis if the predicted damaging score for the variants exceeds that seen for 99 % of the 1000 Genomes dataset.
- These remaining genes are then analyzed using the Phenomizer algorithm to match semantically the proband's phenotypes encoded using HPO to known disease-gene associations.

A typical Phen-Gen command that I used was

```
perl phen-gen.pl input_phenotype=1099_CAGIV_HPO.txt input_vcf=1099.vcf  
input_ped=1099.ped
```

### Final Variant Prioritization

Based on the inputs from Exomiser, Phen-Gen and Genemania, I picked variants based on quality, predicted pathogenicity, MAF frequencies and literature survey.

### References

1. PhenoTips: patient phenotyping software for clinical and research use. Girdea M et al. Hum Mutat. 2013;34:1057-65.
2. HPOSim: An R Package for Phenotypic Similarity Measure and Enrichment Analysis Based on the Human Phenotype Ontology. Deng Y et al. PLoS One. 2015;10:e0115692

3. Clinical diagnostics in human genetics with semantic similarity searches in ontologies. Köhler et al. *Am J Hum Genet* (2009) vol. 85 (4) pp. 457-64
4. The Human Phenotype Ontology project: linking molecular biology and disease through phenotype data. Köhler et al. *Nucleic Acids Research* (2013)
5. Using information content to evaluate semantic similarity in a taxonomy. Resnik P. *Proceedings of the 14th international joint conference on Artificial intelligence—Volume 1. IJCAI'95 San Francisco, CA, USA: Morgan Kaufmann Publishers Inc; pp. 448–453*
6. Phenolyzer: phenotype-based prioritization of candidate genes for human diseases. Yang H et al. *Nat Methods*. 2015;12:841-3
7. Next-generation diagnostics and disease-gene discovery with the Exomiser. Smedley D et al. *Nat Protoc*. 2015;10:2004-15
8. Phen-Gen: combining phenotype and genotype to analyze rare disorders. Javed A et al. *Nat Methods*. 2014;11:935-7.
9. Phenotype-driven strategies for exome prioritization of human Mendelian disease genes. Smedley D and Robinson PN. *Genome Med*. 2015 30;7:81.
10. The GeneMANIA prediction server: biological network integration for gene prioritization and predicting gene function. Warde-Farley D et al. *Nucleic Acids Res*. 2010;38 Suppl:W214-20

## **Group 12**

The genomes of 25 SickKids children were matched with their phenotypic description with the help of the published data in supplementary Table 4 of Stavropoulos, D.J., et al. (2016) [1]. Then we searched for diagnostic variants and predictive secondary variants for each child from the provided whole genome sequencing data, including SNVs, Indels, CNVs, SVs and MEI. At the end of this write-up we have included tables of the selected diagnostic variants and secondary variants, so as to provide more complete information than allowed in the submission file. In the comments fields of the submission file we have provided the gene name, mechanism, phenotype agreement, and confidence (see confidence definitions below).

### Collection of SickKids challenge phenotypes and corresponding gene list

Clinical phenotype descriptions were collected from CAGI phenotype data for 25 children with suspected genetic disorders and also from the supplementary Table 4 of Stavropoulos, D.J., et al. (2016) [1], matched by genome id of these 25 children. We extracted 3306 genes corresponding to 243 phenotypes from the Human Phenotype Ontology-based database (HPO) (Build #102) [2] and the dbNSFP database (version 3.1a) [3]. The gene list for secondary variants was taken from the Table in the 2013 ACMG guidelines [4].

### Annotation of VCF files and QC filters

The VCF files (including SNVs, Indels, CNVs, SVs and MEI) provided for this challenge from Complete Genomics assembly pipeline 2.5 were annotated using the Varant [5] tool, including region of occurrence (intron, exon, splice site or intergenic), observed minor allele frequencies (MAF), mutation type, predicted impact on protein function, and previously established associated phenotypes reported in ClinVar [6]. The RefGene [7] gene definition file was used for gene and transcript annotations in Varant. In addition, in-house scripts were written to further annotate the VCF files with HGMD [8] disease related variants and with dbSNP [9] variants that potentially alter splicing.

We used high quality data for further analysis and different QC filters were used for different types of data as mentioned in Complete Genomics assembly pipeline 2.5. For SNVs, Indels and SVs, we used the 'PASS' filter; for CNVs, we used the ploidy score and CNV type score >30 for those segments where called ploidy is not equal to 2 and regions which are not hypervariable; for MEI data, we used 'sns95' as filter status and CGA\_IS (measure of confidence that there is a mobile element insertion) > 15.

### Diagnostic variants: identification and interpretation

A hierarchical scheme was used for identification of diagnostic variants, based on the strength of the evidence for disease relevance. All accepted high quality variants in the selected gene list with population frequency <5% in either 1000 genome data [10] or in the ExAC database [11]) were first categorized into ordered tiers as follows:



Category 1: Variants with HGMD annotation of either DM (disease-causing mutation) or DP (disease-associated polymorphism), and/or reported in ClinVar with pathogenic or likely pathogenic status.

Category 2: Nonsense mutation, direct splicing mutation disrupting either splice donor or acceptor site, frameshift or non-frameshift mutation, splice altering variant predicted by the dbSNV database [9], and non-synonymous mutations predicted as damaging by one or more of SNPs3D [12], SIFT [13], PolyPhen-2[14] and CADD [15]. For inclusion of a non-synonymous variant in Category 2, at least 60% of these methods were required to return a prediction of deleterious. This threshold is based on a calibration against HGMD. (Note that for any given variant not all methods may return a result, hence the non-obvious cutoff).

Category 3: Non-synonymous mutations predicted as damaging by one or more of the above non-synonymous impact prediction methods, with the deleterious prediction agreement fraction  $< 0.6$ .

Category 4: Benign non-synonymous mutations (according to reporting of non-synonymous impact prediction methods).

Category 5: Variants annotated as close to a splice acceptor or splice donor site.

Category 6: Variants annotated as UTR and intronic.

Category 7: All CNV, SV and MEI variants overlapping with the selected gene list not included in an earlier category.

Except as noted below, only variants in Categories 1, 2 and 3 were accepted for final submission.

Variants from category 1, 2, and 3 were further categorized into ordered tiers according to their rarity in the population data.

Frequency bin 1: Novel mutations (not seen in 1000 genomes or ExAC).

Frequency bin 2: Variants with population frequency  $> 0$  and  $\leq 0.005$ .

Frequency bin 3: Variants with population frequency  $> 0.005$  and  $\leq 0.01$ .

Frequency bin 4: Variants with population frequency  $> 0.01$  and  $< 0.05$ .

Generally, variants in lower frequency bins were preferred.

Binned variants were further filtered for an appropriate inheritance model using the OMIM inheritance pattern. Variants were assigned to autosomal dominant, autosomal

recessive, compound heterozygous, pseudo autosomal recessive, or X-linked recessive models. Some of variant genotypes had phase information and for these we checked for consistency with a compound heterozygous model, where appropriate. For compound heterozygous cases where one variant belonged to category 1, 2, and 3, we also considered variants in categories 4, 5, 6 and 7 to provide the second variant.

With these criteria, we found several Category 1, 2 and 3 variants for many individuals, but with the corresponding phenotype not an exact match for the phenotypes provided. In these cases we searched for partial overlap with the reported phenotypes and made a judgment call as to whether the variant could be relevant. We tagged these variants with three different confidence levels:

Probable Match: When the OMIM disease description for the gene matches the individual's phenotype.

Possible Match: When there is a partial overlap of the OMIM disease phenotypes with the the individual's phenotype.

Speculative Match: When the variant is unlikely to be causative, for example an SV in an intergenic region near an appropriate gene or completely within an intron of an appropriate gene, such that is no obvious mechanism of action.

#### Searching for Predictive Secondary variants

Here we followed the rules in Stavropoulos, D.J., et al. (2016) [1] to extract predictive secondary variants. We followed the same protocol as for diagnostic variants, described above, using inheritance models from the Table in the of 2013 ACMG guidelines [4].