

Corresponding author(s):	Yael David
Last updated by author(s):	May 26, 2020

Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, seeAuthors & Referees and theEditorial Policy Checklist.

_				
Si	ta	ŤΙ	l¢†	ICS

For	all st	atistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.			
n/a	Confirmed				
	×	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement			
	×	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly			
x	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.				
x	A description of all covariates tested				
x	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons				
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)				
x	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give P values as exact values whenever suitable.				
x	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings				
x	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes				
x	Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated				
Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.					
Software and code					
Poli	y in	formation about <u>availability of computer code</u>			
Da	ita c	ollection No special software was used for data collection.			
Data analysis Microsoft Excel, Gra		nalysis Microsoft Excel, GraphPad Prism 7			
For m	anus	cripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers.			

Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data $% \left(1\right) =\left(1\right) \left(1\right) \left($
- A description of any restrictions on data availability

All the data are available within the article and its Supplementary Information files or from the corresponding author upon reasonable request. A reporting summary for this article is available as a Supplementary Information file. The source data underlying Figures 2, 3 and Supplementary Figures 2, 3, 4, 7, 8, 10, 12, 13, 14, 15 are provided as a Source Data file.

Field-specific reporting

Life scier	ices study design		
All studies must dis	close on these points even when the disclosure is negative.		
Sample size	Sample size was chosen based on our previous work using the same samples and models (Nat. Commun., 2019, 10: 1289).		
Data exclusions	No data was excluded from the analysis.		
Replication	All the experiments in this research were repeated independently three times with similar results.		
Randomization	All the samples were allocated into experimental groups randomly. Animals were randomized according to weight at the start of studies.		
Blinding	Blinding was not necessary as we were observing large differences in staining or morphology.		
We require information system or method list Materials & exponsion Involved in th X Antibodies X Eukaryotic X Palaeontolo X Animals an	Cell lines Ex ChIP-seq Cell lines Ex Flow cytometry Day MRI-based neuroimaging d other organisms earch participants		
Antibodies used Validation	Mouse Anti-Methylglyoxal Monoclonal Antibody (Cell Biolabs, STA-011), Chicken Anti-H3 Polyclonal Antibody (Abcam, ab134198), Mouse Anti-H3 Monoclonal Antibody (Abcam, ab10799), Mouse Anti-Actin Monoclonal Antibody (CST, 3700S), Rabbit Anti-H3R8Me2 Polyclonal Antibody (Abcam, ab194692), Rabbit Anti-H3Cti (R2, R8, R17) Polyclonal Antibody (Abcam, ab5103), Mouse Anti-Citrulline Monoclonal Antibody (Sigma, SAB5202274), Rabbit Anti-PAD4 Polyclonal Antibody (Abcam, ab50332), Chicken Anti-H4 Polyclonal Antibody (Abcam, ab134212), Rabbit Anti-H4Cit3 Polyclonal Antibody (MilliporeSigma, 07-596), Rabbit Anti-H4R3me2 Polyclonal Antibody (Abcam, ab194683), Rabbit Anti-HA Monoclonal Antibody (CST, 3724S), Rabbit Anti-MEK ½ Polyclonal Antibody (CST, 9122S). All the secondary antibodies (listed in Supplementary Table 2) used in this manuscript are obtained from Li-Cor. Each primary antibody used in this study was validated for species (human or mouse) and application (immunblotting and IP) by		
vanuatiOH	the above-listed manufacturer using recombinant protein and/or expressing cell types as a positive control; these validation		

Eukaryotic cell lines

Cell line source(s)

Policy information about **cell lines**

MCF7 is obtained from DSMZ. HEK293T, T47D, BT474, ZR75-1, and Cama-1 cell lines used in this study are all sourced from

species and application (immunblotting and IP) in our experiments using recombinant proteins and/or expressing cell types as a positive control and are reported in main text and supplemental figures (see our previous work Nat. Commun., 2019, 10: 1289). For more validation data of the antibodies used in this study, see the product information of the corresponding suppliers'

All cell lines used in this manuscript were authenticated by the vendors where we acquired these cell lines. ATCC used Authentication morphology, kryotyping, and PCR-based approaches.

Mycoplasma contamination All cell lines were tested for mycoplasma monthly (MycoFluor Mycoplasma detection kit, Invitrogen) and tested negative for contamination.

Commonly misidentified lines (See <u>ICLAC</u> register)

No commonly misidentified cell lines were used in this study.

Animals and other organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research

Laboratory animals Six-to-8-week-old nu/nu athymic BALB/c female mice (Harlan Laboratories, Inc.) were used for the breast cancer xenografts

experiments and maintained in pressurized ventilated caging. All these studies were performed in compliance with institutional

guidelines under an Institutional Animal Care and Use Committee-approved protocol (MSKCC#12-10-016).

Wild animals The study did not involve any wild animals.

The study did not involve any samples collected from the field. Field-collected samples

All the animal studies were performed in compliance with institutional guidelines under an Institutional Animal Care and Use Ethics oversight

Committee-approved protocol (MSKCC#12-10-016).

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Human research participants

Policy information about <u>studies involving human research participants</u>

Population characteristics N/A

Recruitment N/A

Ethics oversight

The clinical samples (MSKCC set) used in this study were obtained from the Biobank of MSKCC. The patients with breast cancer and either recurrence of disease after receiving adjuvant therapy or WHO-defined progression of metastatic disease on therapy were prospectively enrolled on an IRB approved tissue collection protocol (IRB#06-163).

Note that full information on the approval of the study protocol must also be provided in the manuscript.