

H19, a Long Non-coding RNA, Mediates Transcription Factors and Target Genes through Interference of MicroRNAs in Pan-Cancer

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Long non-coding RNAs (lncRNAs) have recently been found to be important in gene regulation. lncRNA H19 has been reported to play an oncogenic role in many human cancers. Its specific regulatory role is still elusive. In this study, we developed a novel analytic approach by integrating the synergistic regulation among lncRNAs (e.g., H19), transcription factors (TFs), target genes, and microRNAs (miRNAs) and then applied it to the pan-cancer expression datasets from The Cancer Genome Atlas (TCGA). Using linear regression models, we identified 88 H19-TF-gene co-regulatory triplets, in which 93% of the TF-gene pairs were related to cancer, indicating that our approach was effective to identify disease-related lncRNA-TF-gene co-regulation mechanisms. lncRNAs can function as miRNA sponges. Our further experiments found that H19 might regulate SP1-TGFBR2 through let-7b and miR-200b, ETS1-TGFBR2 through miR-29a and miR-200b, and STAT3-KLF11 through miR-17 in breast cancer cell lines. Our work suggests that miRNA-mediated lncRNA-TF-gene co-regulation is complicated yet important in cancer.

INTRODUCTION

Long non-coding RNA (lncRNA) refers to a class of transcripts that are longer than 200 nt (bp) and are not translated to protein.¹ lncRNA has been recently found to have many biological functions such as transcriptional regulation, epigenetic modification, and cell fate determination.² It has been involved in many diseases, including cancer.³ For example, oncogenic lncRNAs may downregulate cancer cell antigen presentation and intrinsic tumor suppression,⁴ and they can serve as potential biomarkers for cancer diagnosis and therapeutic strategy development.^{5,6} Among thousands of lncRNA molecules discovered so far, H19 is one that is highly expressed.^{7–9} Accumulating data have suggested that lncRNA H19 plays a critical role in tumor initiation, progression, and recurrence in various human cancers.¹⁰ H19 has been reported to control cell cycle progression

through regulating RB-E2F signaling in colorectal cancer.¹¹ It plays an essential role in the exosome-mediated phenotype of endothelial liver cancer cells.¹² H19 competitively binds a microRNA, miR-17-5p, to regulate YES1 gene expression in thyroid cancer.¹³ Moreover, H19-derived miR-675 contributes to bladder cancer cell proliferation through regulating p53 activity.¹⁴

lncRNA can act as a competing endogenous RNA (ceRNA) to interact with other protein-coding RNA transcripts, both transcription factor (TF) and non-TF genes.^{15,16} Hereafter, we refer to genes as protein-coding genes to separate them from non-coding genes. By sharing the common miRNA-binding sites with mRNAs or directing miRNA degradation, lncRNA competes with the miRNA target genes (TFs or non-TF genes) through interacting with miRNA; consequently, the expression of miRNA-targeted genes will be upregulated. This type of lncRNA-miRNA-gene competing co-regulation (triplets) has been discovered in humans and several other species.¹⁷ lncRNA may also interfere with the classic TF-gene regulation by acting as a ceRNA.

Although lncRNA is important in cancer, how it plays its regulatory roles in the complex and dynamic cellular systems remains largely unknown, especially at the pan-cancer level. In this work, we developed

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an analytical strategy to explore the synergistic regulation among lncRNAs, miRNAs, TFs, and genes. We applied this approach to The Cancer Genome Atlas (TCGA) pan-cancer datasets. We specifically examined H19, one of the highly expressed lncRNAs in cancer (**Figure S1**). Among the 24 cancer types we examined, we found that H19 was highly expressed in 21 cancer types, although not in the other three types (lower grade glioma [LGG], prostate adenocarcinoma [PRAD], and thyroid carcinoma [THCA]). As mentioned above, lncRNA might act as miRNA sponges to play roles in TF-gene regulation. Thus, we hypothesized that H19 could mediate TF-gene regulation through controlling miRNAs. After investigating the regulatory relationship among H19, miRNAs, TFs, and genes, we pinpointed three co-regulation triplets (H19-SP1-TGFB2, H19-ETS1-TGFB2, and H19-STAT3-KLF11) to validate our hypothesis in breast cancer cell lines. Our experimental results revealed that H19 might mediate (1) SP1-TGFB2 (TF-gene) interaction through let-7b and miR-200b, (2) ETS1-TGFB2 (TF-gene) interaction through miR-29a and miR-200b, and (3) STAT3-KLF11 (TF-gene) interaction through miR-17. Such regulatory triplets can be used to predict the potential function of lncRNAs and miRNAs in cancer. Our study showed that lncRNA could interfere with miRNA-mediated TF-gene interactions. This critical regulation, if universal in various cancer types, will be important for understanding the molecular mechanisms of cancer initiation and progression, and the molecules included in the interaction can serve as biomarkers for cancer diagnosis, drug development, and therapeutic strategy development.

RESULTS

Classifying Samples by H19 Expression Level

lncRNA H19 is highly expressed in many cancer types and is actively involved in all stages of tumorigenesis.¹⁰ To better understand the role of H19 in cancer, we analyzed the expression level of H19 in TCGA pan-cancer datasets and grouped the samples into “low,” “middle,” and “high” by H19 expression level. For instance, in breast invasive carcinoma (BRCA) samples, we reordered all of the samples by H19 expression and named the top 25% as the high group and the bottom 25% as the low group of H19 expression (**Figure 1A**). Taking the same criterion, we evaluated the H19 expression level in the remaining 23 cancer types and found that 25% is an appropriate percentage to set as the threshold (**Table S1**; **Figure 2**). In most of these 24 cancer types, the H19 expression profile within the middle 50% of samples showed a steady line with a mild slope. This indicated that these samples had similar and stable levels of H19 expression. However, outside the middle section, the profile lines showed significant change at the inflection points. As shown in **Figure 2**, samples falling into the zone from zero to the end point of rapid increases (the red vertical solid line) are considered as the H19 low group, whereas samples with the H19 level in the top section, from the start of dramatic increases (the blue vertical dashed line) to the end, are designated as the H19 high group. Interestingly, in a previous study, Li et al.¹⁸ also used the 25th percentile as the threshold for dividing the top and bottom samples in their study. After having these cutoff thresholds, we further identified the H19 class labels of samples for BRCA TF expression data.

Identification of H19-TF-Gene Triplets

We first focused on explaining the results from TCGA BRCA samples; the results of other cancer types are provided in **Table S1**. Based on the FPKM (fragments per kilobase of exon per million mapped reads) score, we filtered out 694 TFs and 11,867 non-TF genes from TCGA BRCA dataset. Using two databases, TRANSFAC (release 2016.4)¹⁹ and TRRUST (version 2.0),²⁰ we obtained a total of 13,263 TF-target gene pairs (interactions), of which 8,181 TF-target gene pairs (interactions) were found to be expressed (FPKM score of at least 50% of samples was greater than 1) in these samples, corresponding to 625 unique TFs and 2,198 unique non-TF genes.

To evaluate the impact of H19 on TF-gene regulation, we used linear regression to obtain the expression profile of TFs and genes after excluding the effect of copy number variations (CNVs) on their expression (**Figure 1B**; **Equation 1**). Next, these new expression profiles were fed to the second linear regression model (**Figure 1B**; **Equation 2**) to assess the effect of H19 on TF or gene expression. In BRCA, 679 TF-gene regulation interactions were found to be affected by H19 (required both false discovery rate [$FDR]_{EXPTF,\xi;GroupH19} < 0.05$ and $p_{EXPTF,\xi} < 0.05$) (**Table 1**). Of note, to reduce the false-positive rate (FDR), we required that these significant H19-TF-gene triplets should present in at least two TCGA cancer types. By following the analysis procedure above, we obtained a total of 88 triplets (**Table S2**). **Figure 3** demonstrates the four most significant triplets ($-\log_{10}(p_{EXPTF,\xi}) > 9$) whose TF-gene regulation was affected by the change of H19 expression. The remaining triplets are presented in **Figure S2**. In **Figure 3**, all TF-gene regulations affected by the change of H19 expression were statistically significant (FDR < 0.05). For instance, in the presence of high H19 expression, the correlation between MYBL2 and COL1A1 was significantly changed (positive correlation with $p = 3.31 \times 10^{-3}$ to negative correlation with $p = 2.74 \times 10^{-10}$).

We found that 173 of 186 (93%) TF-gene pairs had direct or indirect evidence to support their relationship to cancer (**Table S3**). According to this high rate, we thought that the remaining 13 TF-gene pairs might play roles in cancer as well and warrant further studies. These 13 TF-gene pairs are as follows: LUAD, CTCF-IPO13; KIRC, SP3-EDF1; PAAD, USF1-FMRI; STAD, EZH2-DACT3; TGCT, FOXO1-HYOU1; THCA, CTCF-IPO13, NFKB1-CHUK, NFYB-EDF1, RELA-BGN, RUNX1-SYMPK, SP1-ME1, SP1-SIGIRR, and SP3-EDF1. Interestingly, most of them (8 out of 13) were found in THCA, suggesting that this cancer type might have additional regulatory roles. These results indicate that our approach is effective in identifying TFs or genes that are related to cancer.

H19 Regulates TF-Gene Function through the Related miRNAs

Given that one of the important lncRNA functions is modulating miRNAs, we speculated that H19 might mediate TF-gene interactions through the regulation of miRNAs. Through a comprehensive literature search (**Table S4**), we identified 29 miRNAs (let-7a, let-7b, let-7g, let-7i, miR-106a, miR-130b-3p, miR-138-5p, miR-139, miR-141, miR-152-3p, miR-152-5p, miR-17-5p, miR-181d-3p, miR-181d-5p,

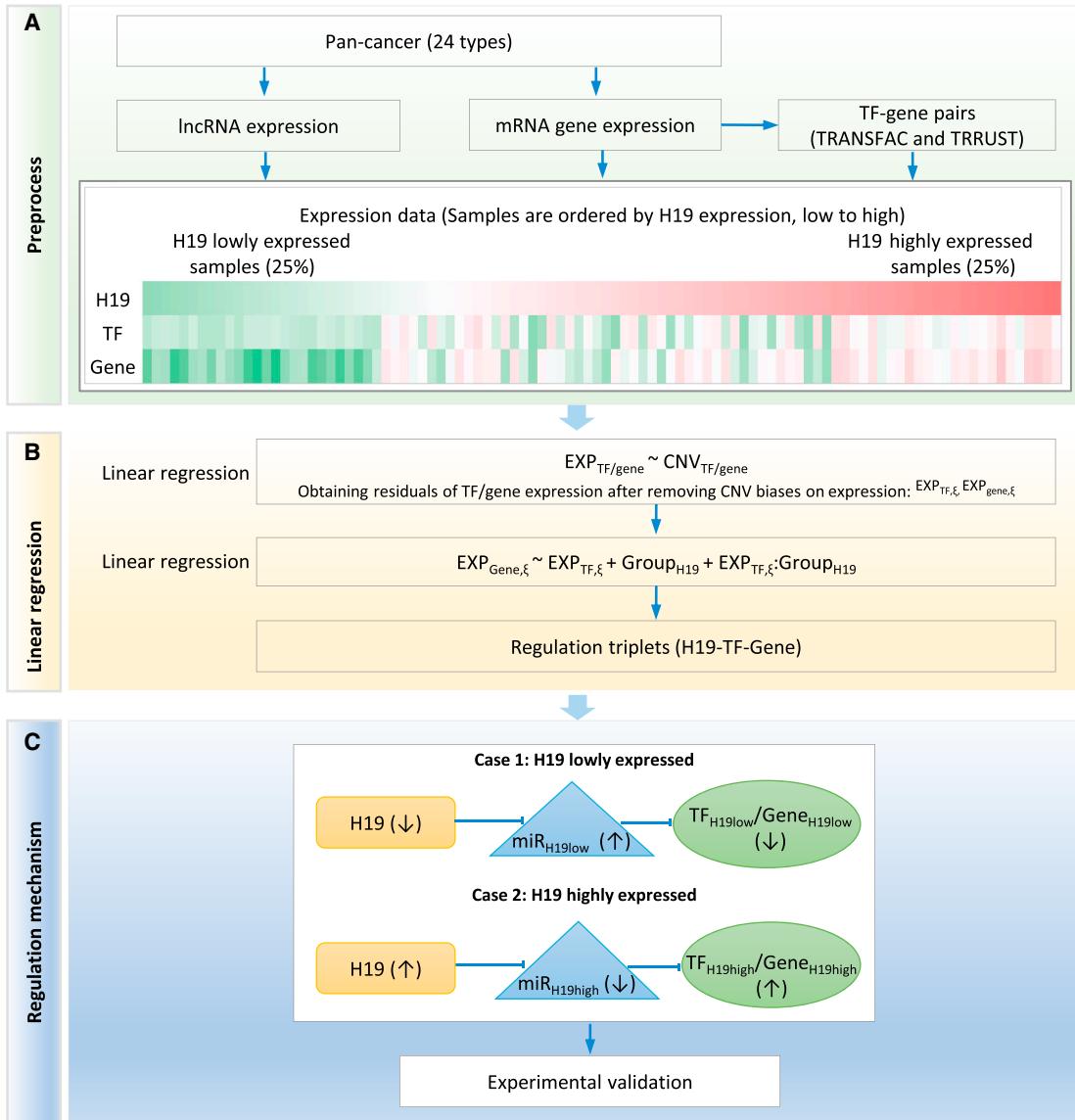
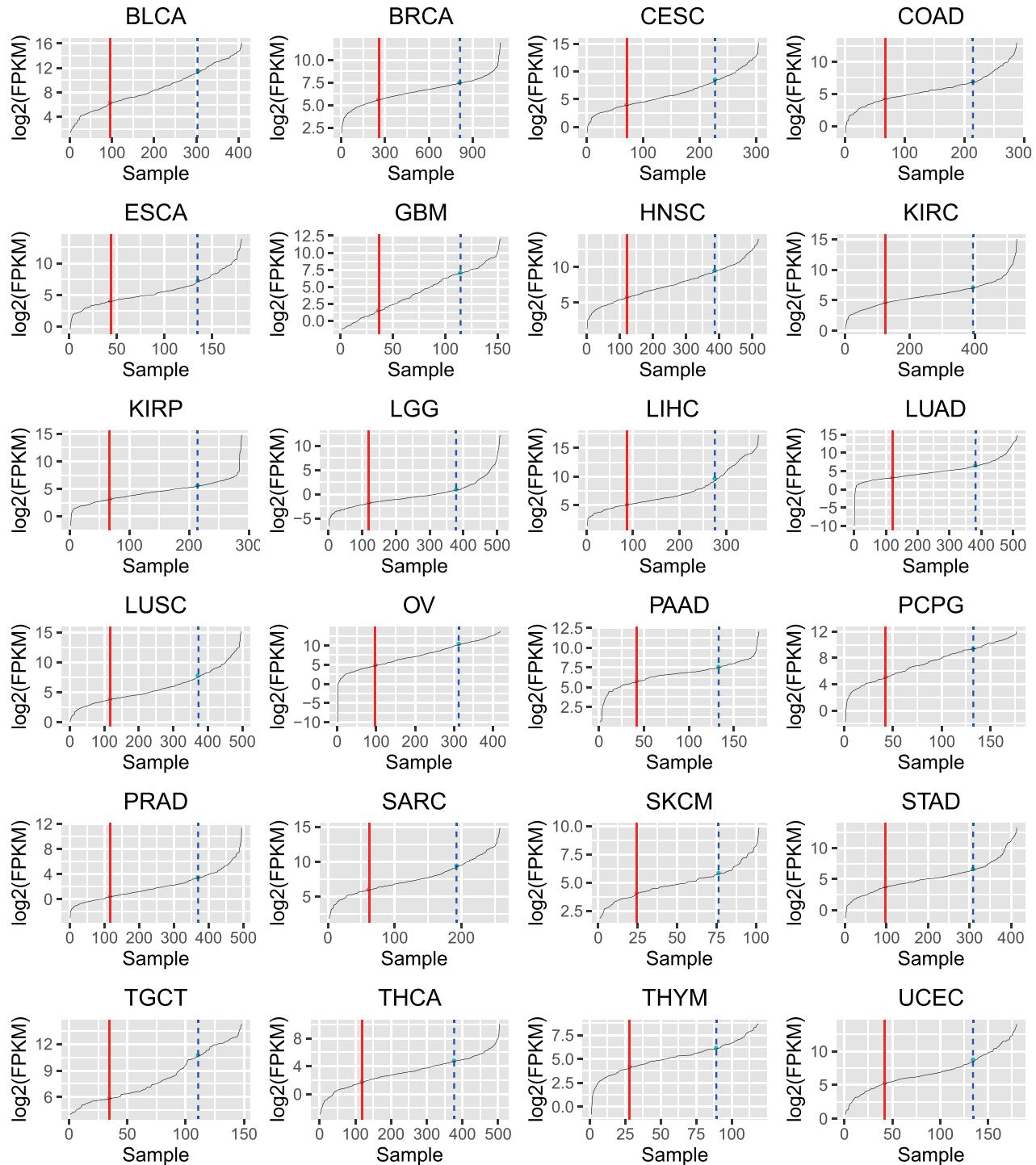


Figure 1. Analytical Pipeline for Identification of lncRNA-TF-Gene Triplets and H19 Is Used as an Example

(A) Analysis of lncRNA, TF and gene expression, followed by ordering the samples according to H19 expression. Expression data for lncRNAs, TFs, and genes in 24 cancer types were extracted from TCGA. TF-gene interaction pairs were retrieved from two databases, TRANSFAC and TRRUST. On each plot, samples were ordered by H19 expression from low to high. Samples that were within 25% of the lowly (highly) expressed samples were considered as a lowly (highly) expressed group. (B) Linear regression analysis of H19 expression with the TF-gene pairs, leading to H19-TF-gene triplets. The effect of copy number variation (CNV) on gene expression was filtered out. (C) Test of the hypothesis that H19 may act as a miRNA sponge in lncRNA-TF-gene triplet regulation. In the low expression H19 case, loss of H19 weakens the inhibition of miRNAs, leading to the downregulation of miRNA target TFs and/or genes; in the high expression H19 case, enhanced H19 inhibits miRNA expression, leading to upregulation of miRNA target TFs and/or genes. ↑, Upregulated; ↓, downregulated.

miR-18a, miR-194-5p, miR-196a, miR-19a, miR-19b-1, miR-200b, miR-200c, miR-20a, miR-22, miR-29a, miR-29b, miR-342-3p, miR-630, miR-874, and miR-92a-1) that were targeted by H19 (Table S4). Next, miRNA target genes of these 29 miRNAs were collected from the miRNA target prediction databases using the SpidermiR R tool.²¹ Among these target genes, 85 overlaid with the H19-TF-gene triplets that we identified in the earlier regression analysis. We

selected eight triplets (H19-ETS1-TGFB2, H19-FLI1-TGFB2, H19-FOXO1-TXNIP, H19-KLF6-TXNIP, H19-NFYB-SP3, H19-PPARA-KLF11, H19-SP1-TGFB2, and H19-STAT3-KLF11) and investigated the impact of these 29 miRNAs on them (Table S5). After having confirmed the target TF and non-TF genes of the H19-mediated miRNAs, we mainly focused on two cases (cases 1 and 2) in BRCA (Figure 1C).

**Figure 2. All Tested Cancer Types, with Samples Classified into Low, Moderate, and High Expression Groups by H19 Level**

Twenty-four cancer types are represented. Classifications of samples are shown as low (left of the solid line), moderate (middle), and high (right of the dotted line).

Table 1. TF-gene Regulation Interactions Significantly Affected by the Expression Alteration of H19

Cancer Type	Number			
	TFs	Genes	TF-Gene Pairs	TF-Gene Pairs (FDR < 0.05) ^a
BLCA	607	2,103	7,723	1
BRCA	625	2,198	8,181	874
CESC	634	2,161	8,057	0
COAD	631	2,187	8,123	148
ESCA	655	2,205	8,277	3
GBM	636	2,156	7,755	2
HNSC	621	2,140	7,866	78
KIRC	628	2,172	7,889	246
KIRP	591	2,030	7,130	12
LGG	589	1,996	6,837	179
LIHC	546	1,964	6,988	2
LUAD	621	2,233	8,223	610
LUSC	653	2,281	8,444	0
OV	622	2,159	8,030	0
PAAD	651	2,369	8,758	1,165
PCPG	562	1,896	6,438	1
PRAD	624	2,170	7,889	1
SARC	602	2,053	7,353	24
SKCM	585	1,982	7,055	0
STAD	648	2,252	8,460	207
TGCT	620	2,202	8,042	574
THCA	587	2,020	7,284	1,095
THYM	602	2,055	7,270	0
UCEC	625	2152	8,092	0

^aNumber of TF-gene pairs with regression FDR <0.05.^bNumber of triplets after CNV filtration.**Case 1**

First, we chose 56 matched (common) H19 low expression group samples along with the corresponding 56 matched normal samples from TCGA BRCA miRNA/TF/non-TF gene expression database. Next, we applied the limma-voom R statistical tool²² and obtained the lists of upregulated (UPR) and downregulated (DWR) miRNAs/TFs/non-TF genes from this pool. As the result (i.e., for the H19 low expression group samples), we identified five cases as UPRmiR_{H19low} (let-7b, $p = 1.51 \times 10^{-6}$; miR-29a, $p = 6.77 \times 10^{-6}$; miR-200b, $p = 2.67 \times 10^{-3}$; miR-17, $p = 1.42 \times 10^{-2}$; and miR-29b, $p = 4.49 \times 10^{-2}$), one case as DWRmiR_{H19low} (miR-130b, $p = 4.60 \times 10^{-4}$), five cases as DWRTF_{H19low} (ETS1, $p = 8.40 \times 10^{-4}$; PPARA, $p = 8.16 \times 10^{-4}$; STAT3, $p = 3.29 \times 10^{-8}$; NFYB, $p = 4.58 \times 10^{-8}$; and SP1, $p = 4.63 \times 10^{-3}$), and two cases as DWRGene_{H19low} (SP3, $p = 2.84 \times 10^{-14}$; and TGFB2, $p = 2.99 \times 10^{-3}$). Neither TFs nor genes were found as UPRTF_{H19low} or UPRGene_{H19low}, respectively (Table 2). The specific definitions of the terms such as

UPRmiR_{H19low} and DWRmiR_{H19low} are provided in Materials and Methods.

Case 2

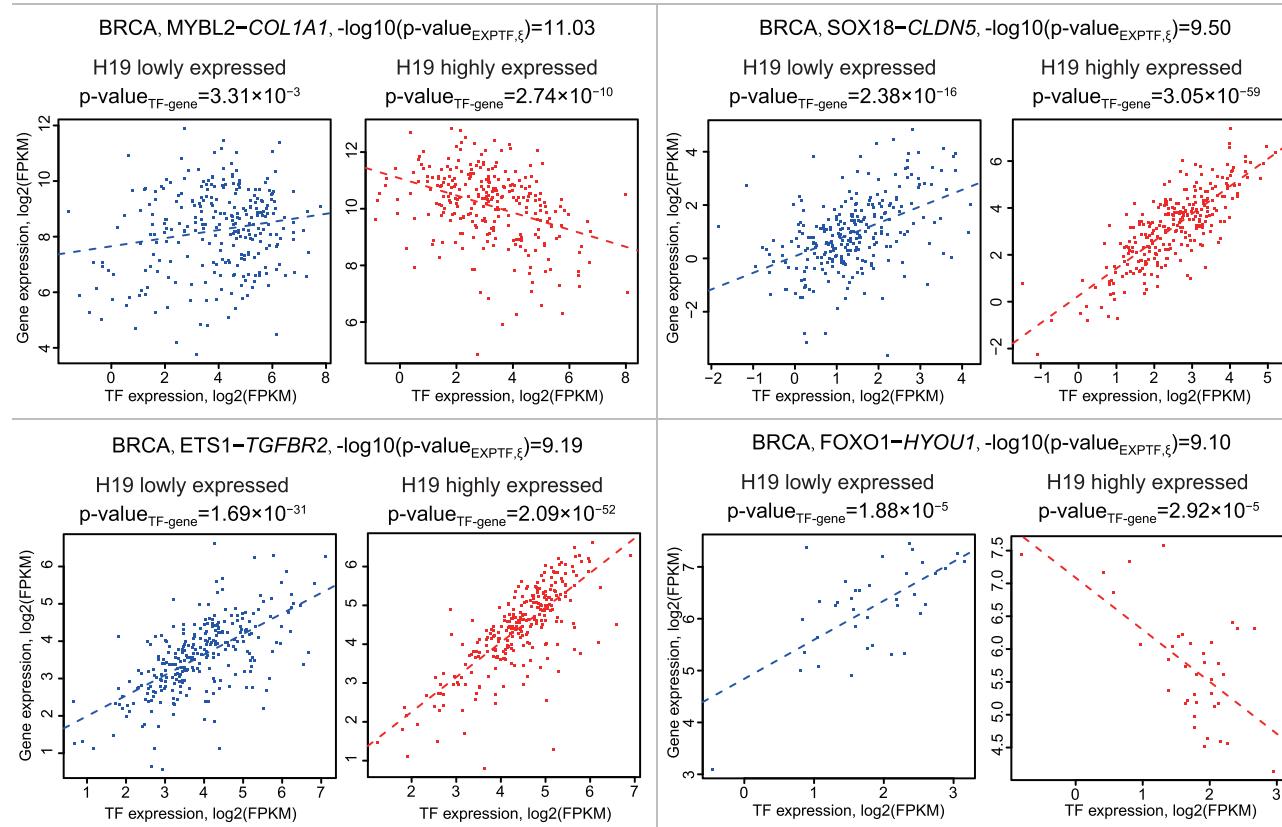
Similarly, we selected 40 matched cancer and normal samples in the H19 high expression group from TCGA BRCA miRNA/TF/non-TF gene expression database. The limma-voom tool was utilized to identify the upregulated and downregulated miRNAs/TFs/non-TF genes in this set. For this H19 high expression group samples, we identified two cases as UPRmiR_{H19high} (miR-141, $p = 5.86 \times 10^{-5}$; and miR-29b, $p = 1.42 \times 10^{-2}$), one case as DWRmiR_{H19high} (miR-200b, $p = 4.45 \times 10^{-3}$), and four cases as DWRTF_{H19high} (SP1, $p = 2.06 \times 10^{-5}$; FL11, $p = 5.65 \times 10^{-4}$; STAT3, $p = 2.86 \times 10^{-3}$; and NFYB, $p = 1.24 \times 10^{-2}$). Neither TFs nor genes were found as UPRTF_{H19high}, UPRGene_{H19high}, or DWRGene_{H19high} (Table 2).

Statistically Evaluated Interesting Gene Regulation Cascades of Triplets

We proposed two scenarios to explain the regulation between every pair of two participating biomolecules within a triplet under different conditions, as follows: (1) H19 (↓)-UPRmiR_{H19low} (↑)-DWRTF_{H19low}/DWRGene_{H19low} (↓): the downregulated expression of H19 causes the upregulation of its target miRNAs, which results in the downregulated expression of their target TFs or genes. (2) H19 (↑)-DWRmiR_{H19low} (↓)-UPRTF_{H19low}/UPRGene_{H19low} (↑): the upregulated expression of H19 causes the downregulation of its target miRNAs, which in turn causes upregulated expression of their target TFs or genes. Accordingly, we found five triplets that met the regulation scenarios above: (1) H19 (↓)-let-7b (↑)-SP1 (↓)-TGFB2 (↓); (2) H19 (↓)-miR-29a (↑)-ETS1 (↓)-TGFB2 (↓); (3) H19 (↓)-miR-200b (↑)-ETS1 (↓)-TGFB2 (↓); (4) H19 (↓)-miR-200b (↑)-SP1 (↓)-TGFB2 (↓); and (5) H19 (↓)-miR-17 (↑)-STAT3 (↓)-KLF11 (↓). If we could prove that the status changes met our prediction, it would be helpful to improve the network validation and understanding the overall effect on expression patterns of the miRNAs/TFs/genes due to different expression levels of H19.

Experimental Validation

In order to examine whether H19 serves as a mRNA sponge to regulate the expression of TFs and their downstream targets through antagonizing let-7b, miR-17, miR-200b, and miR-29a,^{13,23-28} breast cancer cell lines BT-549, HCC38, MCF7, and MDA-MB-231 were used with ectopically expressing inducible H19.²⁹ Compared to the control cells, the overexpression of H19 could upregulate the distinct levels of mRNA expression of let-7b/miR-200b-regulated SP1, miR-29a/miR-200b-regulated ETS1, and miR-17-regulated STAT3 in the breast cancer cell lines (Figures 4A–4D). Consequently, the expression of the SP1/ETS1 transcriptional target TGFB2 and STAT3 target KLF11 was also increased to different extents upon H19 induction. However, there was no obvious induction of TGFB2 detected in BT549 or MD-MB231 cells, which could be due to the low expression of SP1 and ETS in such cells or dysregulation of other transcriptional co-regulators (Figures 4A and 4C). Taken together, these lines of experimental evidence support our notion of miRNA-mediated

**Figure 3. TF-Gene Regulation as Affected by H19 Expression Level**

Linear regression was used to evaluate the association between TF expression and its target gene expression.

H19-TF-gene triplet regulation, indicating that H19 upregulates transcription factors and their downstream effects by sequestering miRNAs in cancer.

DISCUSSION

Identification of cancer-associated lncRNAs and uncovering their molecular mechanisms are currently challenging but important tasks. Traditionally, studies of gene expression deregulation and alterations in genomic sequences in tumor cells have led to the identification of cancer-associated lncRNAs.^{5,30} Subsequent *in vitro* and *in vivo* studies have directly associated some of the identified lncRNAs with specific cancer phenotypes. In this study, we identified 88 H19-TF-gene triplets based on TCGA pan-cancer data using linear regression models. Most of these TF-gene pairs (93%) had direct or indirect evidence to support their relationship to cancer (Table S3). The remaining TF-gene pairs could serve as potential candidates and warrant further investigation. Our results demonstrate that this analytical, co-regulation-based approach is promising to identify TFs or genes related to cancer. To investigate the potential regulatory mechanism, we hypothesized that H19 acts as a miRNA sponge to diminish certain miRNAs, which in turn preserves the corresponding TF-gene function. Our quantitative real-time PCR experimental results suggest that H19 mediates SP1-TGFB2 (TF-gene) regulation

through let-7b and miR-200b (miRNA), ETS1-TGFB2 regulation through miR-29a and miR-200b, and STAT3-KLF11 regulation through miR-17 in BT-549, HCC38, MCF7, and MDA-MB-231 breast cancer cell lines. Our approach can help identify lncRNAs, miRNAs, TFs, and genes that are potentially cancer associated and uncover their complex regulatory mechanisms. This approach effectively extends the previous miRNA-TF-gene co-regulation approach that has been well studied in various cancer types or other disease.^{31–33}

TRANSFAC and TRRUST are representative databases for annotations of TF-target gene pairs. The annotations are based on stringent criteria, including both experimental evidence and statistical tests. After filtration, we obtained 13,263 TF-target gene pairs from these two databases. Although the number of TF-gene pairs is smaller than that from other databases, such as ENCODE,³⁴ we decided to use them for reducing false-positive results. For the large-scale data analysis, false-positive data would have more potential problems than false-negative data, while our goal is to find those enriched signals (e.g., regulatory networks) that have reliable evidence.

The samples studied were classified into three groups by H19 expression, that is, low, middle, and high. In such a way, we could

Table 2. Genes, TFs, and miRNAs Were Upregulated or Downregulated.

Group	Molecule	Log ₂ FC	p	Regulation	
H19 lowly expressed	miRNA	let-7b miR-130b miR-17 miR-200b miR-29a miR-29b-2	0.92885 −3.6149 0.90485 2.54065 1.903 2.4692	1.51 × 10 ^{−6} 4.60 × 10 ^{−4} 1.42 × 10 ^{−2} 2.67 × 10 ^{−3} 6.78 × 10 ^{−6} 4.49 × 10 ^{−2}	upregulated downregulated upregulated upregulated upregulated upregulated
	TF	ETS1 NFYB PPARA SP1 SP3 STAT3	−1.66925 −2.8769 −2.2694 −0.89175 −1.1854 −1.9026	8.41 × 10 ^{−4} 4.58 × 10 ^{−8} 8.16 × 10 ^{−4} 4.64 × 10 ^{−3} 2.85 × 10 ^{−14} 3.29 × 10 ^{−8}	downregulated downregulated downregulated downregulated downregulated downregulated
	gene	TGFB2 miR-141 miRNA	−2.41715 1.12925 miR-200b miR-29b-2	3.00 × 10 ^{−3} 5.86 × 10 ^{−5} 4.46 × 10 ^{−3} 1.42 × 10 ^{−2}	downregulated upregulated downregulated upregulated
H19 highly expressed	TF	NFYB SP1 SP3 STAT3	−1.8413 −0.9865 0.0436 −1.3972	1.25 × 10 ^{−2} 2.07 × 10 ^{−5} 6.47 × 10 ^{−8} 2.86 × 10 ^{−3}	downregulated downregulated upregulated downregulated
	gene	KFL11	−0.992	5.65 × 10 ^{−4}	downregulated

FC, fold change.

observe the TF-gene status change along with the changes of H19 expression. For example, we used linear regression to describe the association between ETS1 (TF) expression and *TGFB2* expression. The p values of linear regression were 1.69×10^{-31} in the H19 low expression group and 2.09×10^{-52} in the H19 high expression group. Additionally, this classification method significantly improved ETS1-*TGFB2* regulation ($p < 10^{-9}$). This suggests that such an integrative grouping approach is effective in investigating the relationship between variation of lncRNA expression and TF-gene regulation.

Figure 2 shows a smooth slope of the curves for the patients with intermediate levels of H19 expression. This is because all of the samples were ordered by their H19 expression, and most of these samples had H19 expression without large fluctuation. For the H19 highly or lowly expressed groups, the curves displayed sharp slopes. These sharp slopes suggested that H19 was expressed with strong variation in these two groups, which is biologically useful to investigate the effect of H19 expression changes on TF-gene regulation.

Linear regression is a widely used approach to predict the output values of a biological process under specific conditions.^{35,36} Some studies use linear regression to predict the expression of genes.³⁷ However, to our knowledge, there are few studies that use linear regression to investigate the effect of lncRNAs on TF-gene regulation.

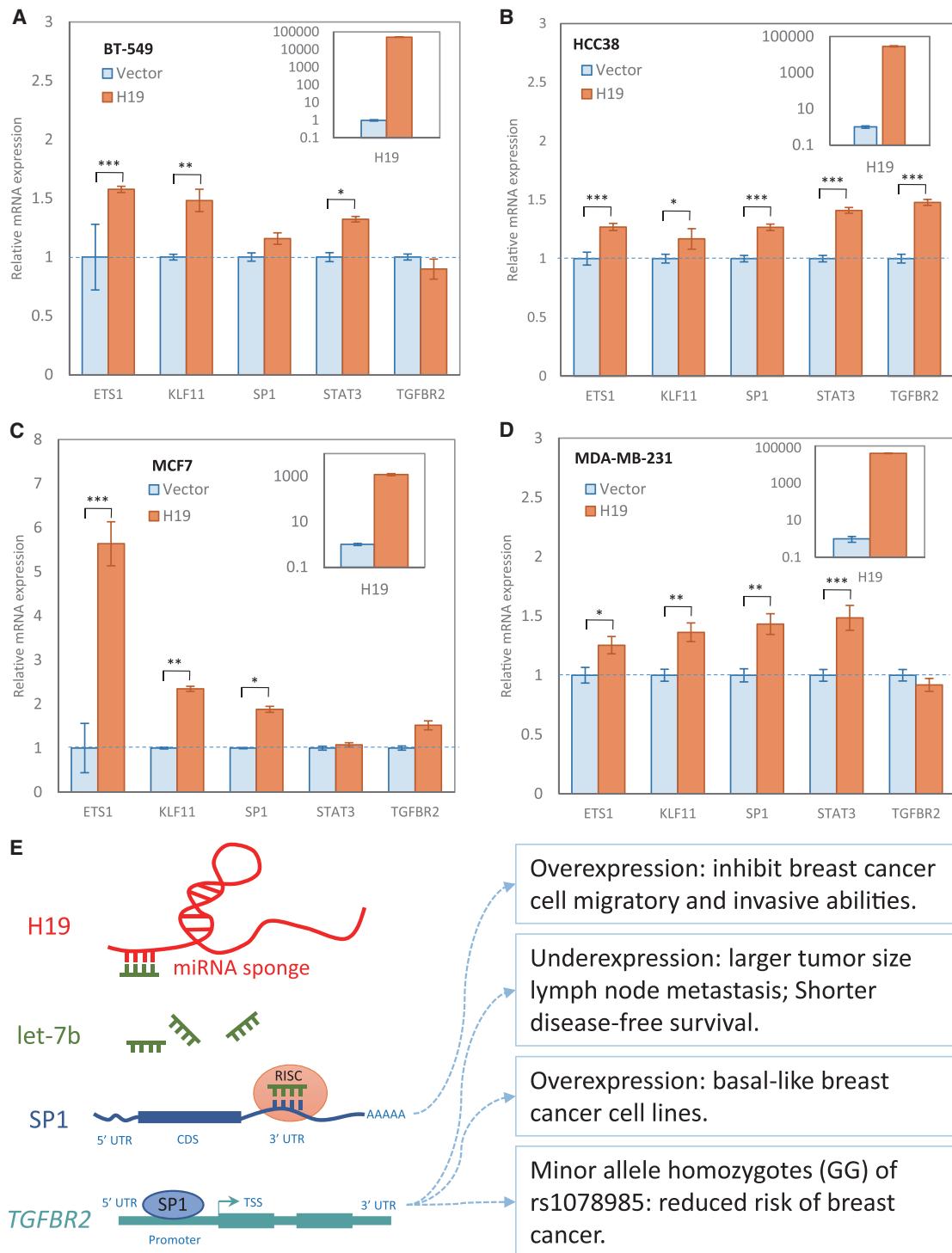
In this study, we used a linear regression model that was fit on non-TF gene expression changes in tumor samples as the response variable using a linear combination of the input variables, including TF expression, sample groups distinguished by H19 expression level, and interaction between TF and group (Equation 2). Within a H19-TF-gene triplet, this requires the TF to regulate the target gene, and H19 to affect TF expression and consequently regulate the target gene. Therefore, we required that such triplets should satisfy both $FDR_{EXPTF,\{5\};GroupH19} < 0.05$ and $p_{EXPTF,\{5\}} < 0.05$.

CNVs contribute largely to gene expression.³⁸ Structural variants, including CNVs, in cancer genomes can lead to significantly reduced or increased gene expression in cancer cells.³⁹ Therefore, we removed the effect of CNVs on gene expression and reconsidered the mediation of H19 on the TF-gene regulation relationship.

The H19-let7b-SP1-*TGFB2* interaction we identified by our bioinformatics approach was supported by actual biological experiments (Figure 4E). The lncRNA H19 can act as a sponge to bind let-7b to mediate breast cancer cell plasticity.²⁸ The let7b-SP1 interaction was verified from chimeric reads.⁴⁰ DNA precipitation, electrophoretic mobility shift assays, and promoter analysis confirmed that the *TGFB2* promoter was bound by SP1.⁴¹

We observed the expression variation of SP1 and *TGFB2* in the presence of a high level of H19 in breast cancer cell lines. The H19-let7b-SP1-*TGFB2* interaction can be used to elucidate such a correlation. The abnormal expression of H19 and let-7b might lead to abnormal expression of SP1 and *TGFB2* and, consequently, lead to the development of breast cancer. The molecules included in the interaction can provide candidate diagnosis biomarkers and targets for therapy.

The synergistic regulatory relationship between lncRNAs, miRNAs, and protein-coding RNAs implemented in our approach can better infer the potential function of non-coding RNAs. The H19-let7b-SP1-*TGFB2* interaction can be used to interpret the potential functions of lncRNAs and miRNAs (Figure 4E). In mice, an overexpression of SP1 has been reported to suppress migratory and invasive abilities of breast cancer cells.⁴² *TGFB2* is upregulated in basal-like breast cancer cell lines.⁴³ The minor allele homozygote (GG) of rs1078985, an intronic single nucleotide polymorphism (SNP) in *TGFB2*, had a 24% lower risk of having breast cancer compared with major allele carriers (AG or AA).⁴⁴ H19 and let-7b interact with SP1 and *TGFB2*. Therefore, H19 and let-7b might be involved in breast cancer in terms of cell migratory and invasive abilities, and they could serve as potential biomarkers. This regulation can also be verified by the following evidence in literature: (1) H19 enhances breast cancer cell proliferation through positive control by E2F1;⁴⁵ (2) overexpression of an ectopic H19 gene promotes the tumorigenic properties of breast cancer cells;⁴⁶ and (3) let-7b expression in breast cancer patients was inversely associated with tumor lymph node metastasis and patient overall survival.⁴⁷

**Figure 4. H19 Functions as a miRNA Sponge to Relieve miRNA-Mediated Suppression of Transcription Factors and Their Targets**

Transcription factor targets included H19-SP1-TGFBR2, H19-ETS1-TGFBR2, and H19-STAT3-KLF11. (A-D) H19 led to an upregulated expression of let-7b/miR-17/miR-29a/miR-200b-controlled SP1, ETS1 and STAT3 (three TF genes), and their transcriptional targets TGFBR2 and KLF11 in four breast cancer cell lines (A, BT-549; B, HCC38; C, MCF7; and D, MDA-MB-231). All samples were analyzed in triplicate and normalized to GAPDH expression. The top-right panel shows relative H19 expression. Quantitative real-time PCR data are presented as mean \pm SE (standard error). * p < 0.05, ** p < 0.01, *** p < 0.001. (E) The H19-let-7b-SP1-TGFBR2 interaction and its

(legend continued on next page)

The study has several limitations, and there is scope for future work to be carried out. For instance, the current work only focused on one lncRNA, H19. In fact, our approach can be applied to any other lncRNAs. We focused on H19 in this study because it is one of the most promising lncRNAs and is highly expressed in most of the cancer types we examined. Moreover, this study cannot exclude false-positive results: some H19-TF-gene triplets are possibly not the real or impactful regulatory correlations, and some TFs or genes may not directly cause or be related to the specific cancer under investigation. To address this problem, we can use a lower FDR threshold to minimize a false-positive rate or validate them using various biological experiments. Finally, cancer is highly heterogeneous, and its development is dynamic in the cellular system. Our approach, like many others, cannot consider real-time or dynamic regulation in cancer and matched normal cells. However, our approach of uncovering lncRNA molecular functions contributes to identify and functionally annotate these cancer related genes, making these genes the attractive targets. These regulatory units can also better explain cancer biology. Finally, in this study, we first examined lncRNA (H19) expression in pan-cancer and then explored how it potentially regulated genes, including TF genes. Alternatively, we may analyze H19-miRNA pairs first and examine which miRNAs might be altered by H19 in the datasets. We will explore this analytical approach in the future.

MATERIALS AND METHODS

Data Collection

TCGA pan-cancer data consisting of 24 cancer types such as BLCA, BRCA, CESC, COAD, ESCA, GBM, HNSC, KIRC, KIRP, LGG, LIHC, LUAD, LUSC, OV, PAAD, PCPG, PRAD, SARC, SKCM, STAD, TGCT, THCA, THYM, and UCEC (full names are summarized in Table S1), whose numbers of samples were at least 90, were used for this study (Figure 1A). Tissue-specific data contained RSEM⁴⁸ gene FPKM data that included the lncRNA expression profile, TF expression profile, and gene (non-TF) expression profile. The data were collected through the UCSC (University of California, Santa Cruz) Xena database (<https://xenabrowser.net/>). The BRCA RSEM gene FPKM expression data contained a total of 60,499 genes and a total of 1,212 samples. For the remaining tissue-specific data, such information was provided in the UCSC Xena database (Table S1). An expression dataset of miRNAs was also collected from the UCSC Xena database. It consisted of 744 miRNAs and 10,818 samples. The curated clinical (phenotype) data that provided the list of the primary tumor samples were collected from the UCSC Xena database. In addition, the interactions between TFs and genes were collected from TRANSFAC (release 2016.4)¹⁹ and TRRUST (version 2.0).²⁰ Furthermore, the interactions between miRNAs and (validated or predicted) target genes were obtained from the SpidermiR R tool²¹ by using six target prediction databases. Among these databases, the miRTar⁴⁹ and miRWALK⁵⁰ databases provided only validated

target genes, whereas the DIANA,⁵¹ miRanda,⁵² PicTar,⁵³ and TargetScan⁵⁴ databases supplied predicted target genes. H19-targeted miRNAs were identified from the published literature. The detailed descriptions regarding the association of H19 and its targeted miRNAs are demonstrated in Table S4.

Data Preprocessing

First, the biomolecules (lncRNAs, TFs, and genes) whose FPKM score of at least 50% of samples was greater than 1 were selected, whereas the remaining biomolecules were excluded from further analyses (Figure 1A). After this step, we partitioned the whole gene expression data into several subparts according to the category of the genes such as filtered TF expression data, filtered non-TF gene expression data, and filtered lncRNA (H19) expression data. The interactions between TFs and target genes were determined from two well-known databases, TRANSFAC (release 2016.4)¹⁹ and TRRUST (version 2.0).²⁰ We first obtained 800 TFs and 3,470 genes from the TRANSFAC and TRRUST databases and then applied these TFs and genes to filter results from these databases and obtained 13,263 TF-target gene pairs. Using the resultant interactions, we further filtered the TF expression data in a way such that the participating (interacting) TFs belonging to the TF-gene interactions (obtained by TRANSFAC and TRRUST), which were listed in the TF expression data, were only considered in the resultant filtered TF expression data. Similarly, we again filtered the non-TF gene expression data in a way such that the target genes belonging to the TF-gene interactions that were mentioned in the non-TF gene expression data were only selected in the resultant filtered non-TF gene expression data. Next, to determine the type of expression of existing samples, we ordered the expression data of H19 underlying all of these samples based on the expression values from low to high. Here, a certain percentage of the lowly expressed samples was considered as the first group (lowly expressed group), whereas the same percentage of the highly expressed samples was treated as the second group (highly expressed group). The remaining samples were used as the third group ("middle" group). According to the resultant class labels of H19 samples, the samples of the other molecules such as gene and TF had been classified.

Linear Regression and Copy Number Variation Factor

CNVs contribute largely to gene expression.³⁸ We removed the effect of CNVs on gene expression and then reconsidered the mediation of H19 on the TF-gene regulation relationship (Figure 1B). First, we obtained the residuals of expression of the TF and genes through the linear regression ("Stats" R tool)⁵⁵ using Equation 1, and then used the residuals of expression of the TF and genes to evaluate the mediation of H19 on the TF-gene regulation relationship through Equation 2 as follows:

$$\text{EXP}_{\text{TF/Non-TF Gene}} \sim \text{CNV}_{\text{TF/Non-TF Gene}}, \quad (1)$$

involvement with breast cancer. H19 acts as a miRNA sponge for let-7b that targets TF gene SP1. Let-7b inhibits the expression of SP1, whose overexpression inhibits breast cancer cell migratory and invasive abilities. SP1 regulates *TGFBR2*, whose abnormal expression or mutation is related to breast cancer. CDS, coding DNA sequence; RISC, RNA-induced silencing complex; TSS, transcription start site; UTR, untranslated region.

where $EXP_{TF/Gene}$ symbolizes the expression data of TF or gene, and $CNV_{TF/Gene}$ denotes the CNV data of the TF or gene; and

$$EXP_{Non-TF\ Gene,\xi} \sim EXP_{TF,\xi} + Group_{H19} + EXP_{TF,\xi} : Group_{H19}, \quad (2)$$

where $EXP_{Gene,\xi}$ refers to the expression data of the gene after removing the effect of CNVs, $EXP_{TF,\xi}$ denotes the expression data of TF after removing the effect of CNVs, and $Group_{H19}$ symbolizes the class labels of samples (low, middle, and high), whereas $EXP_{TF,\xi}:Group_{H19}$ refers to the interaction effect between the TF and the group. Finally, we determined the triplets using the criteria (1) the adjusted p value should be less than 0.05 ($FDR_{EXPTF,\xi:GroupH19} < 0.05$), and (2) the p value corresponding to coefficient of $EXP_{TF,\xi}$ should be less than 0.05 ($p_{EXPTF,\xi} < 0.05$).

Regulatory Mechanism

After obtaining the triplets (H19-TF-gene) through linear regression, we conducted an extensive literature search to determine the interactions between H19 and miRNAs. We identified target miRNAs that interacted with H19. Next, the target genes (including TFs as well as non-TF genes) of the employing miRNAs had been identified using the SpidermiR R tool.²¹ After finding the target genes of the H19-mediated miRNAs, we had mainly focused on two cases (Figure 1C), that is, cases 1 and 2.

Case 1

First we chose the matched (common) H19 lowly expressed group samples along with the matched normal samples from TCGA BRCA miRNA/TF/non-TF gene expression data. Then, we applied the limma-voom R statistical tool²² to determine which miRNAs/TFs/non-TF genes were upregulated (denoted as UPRmiR_{H19low}, UPRTFH_{H19low}, and UPRGene_{H19low}, respectively), downregulated (denoted as DWRmiR_{H19low}, DWRTF_{H19low}, and DWRGene_{H19low}, respectively), or not differentially expressed.

Case 2

Similarly, we selected the matched (common) H19 highly expressed group samples along with the matched normal samples from TCGA BRCA miRNA or gene expression data. Then, we used the limma-voom R statistical tool²² to determine which miRNAs or genes were upregulated (UPRMI_{H19high}, UPRTF_{H19high}, and UPRGene_{H19high}, respectively), downregulated (DWRMI_{H19high}, DWRTF_{H19high}, and DWRGene_{H19high}, respectively), or not differentially expressed.

Cell Lines

All cell lines were obtained from the American Type Culture Collection (Manassas, VA, USA), independently validated by STR DNA fingerprinting at the University of Texas MD Anderson Cancer Center (Houston, TX, USA) and determined to be negative for mycoplasma contamination. HEK293T (RRID:CVCL_0063) cells and human breast cancer cell lines BT-549 (RRID:CVCL_1092), HCC38 (RRID:CVCL_1267), MCF7 (RRID:CVCL_0031), and MDA-MB-231 (RRID:CVCL_0062) were cultured in Dulbecco's modified Ea-

gle's medium (DMEM) with 10% fetal bovine serum (FBS) supplemented with 100 IU/mL penicillin and 100 µg/mL streptomycin. Cells were maintained at 37°C in a humidified 5% CO₂ incubator.

Generation of H19 Lentivirus and Infection

2×10^6 HEK293T cells were seeded in 10-cm tissue culture plates and maintained in DMEM complete medium for 20 h. Media were discarded and replaced with 3 mL of Opti-MEM. 6 µg of pCMV-dR8.2-dvpr, 2 µg of pCMV-VSV-dvpr, 8 µg of TetO-FUW H19²⁹ or rtTA, and 32 µg of polyethyleneimine (PEI) were added into 400 µL of Opti-MEM. The transfection mixture was incubated at room temperature for 15 min and then added into HEK293T cells. After a 3-h incubation, 3 mL of DMEM complete medium was added. After overnight incubation, the medium was replaced with 6 mL of DMEM complete medium. The viruses were collected at 48–72 h after transfection and concentrated by Amicon Ultra-15 (Millipore). For viral infection, BT-549, HCC38, MCF7, and MDA-MB-231 cells were seeded, respectively, in six-well plates and infected with either control or H19 lentivirus. One µg/mL doxycycline was used to turn on the expression of H19. After 48–72 h of induction, cells were collected to examine H19, ETS1, KLF11, SP1, STAT3, and TGFB2 gene expression for quantitative real-time PCR.

Quantitative Real-Time PCR

Total RNA was extracted using TRIzol reagent (Thermo Fisher Scientific, CA, USA) following the manufacturer's instructions. The RNA samples were qualified using a NanoDrop spectrophotometer (Thermo Fisher Scientific, CA, USA). 1 µg of mRNA was used for reverse transcription using the iScript cDNA synthesis kit (Bio-Rad, CA, USA). A 20-µL quantitative real-time PCR reaction solution was composed of 1 µL of cDNA, 1 µL each of 10 µM forward and reverse qPCR primers, 10 µL of SYBR Green PCR master mix (Bio-Rad, CA, USA), and 7 µL of RT-PCR-grade water. qPCR reactions were performed on a CFX96 machine (Bio-Rad). All reactions were run in triplicate. The relative ETS1, KLF11, SP1, STAT3, and TGFB2 mRNA expression levels were normalized to their corresponding GAPDH mRNA expression. Primers used for quantitative real-time PCR detection are listed in Table S6.

Statistical Analysis

The limma-voom R statistical tool²² using an empirical Bayes statistical test was applied to identify the differentially expressed miRNAs, TFs, and non-TFs in the (common) H19 lowly or highly expressed group samples versus the matched normal samples using TCGA breast cancer gene miRNA and mRNA expression datasets. For quantitative real-time PCR, all grouped data are presented as mean ± SE (standard error). A Student's t test was used to assess statistical significance between the two groups.

SUPPLEMENTAL INFORMATION

Supplemental Information can be found online at <https://doi.org/10.1016/j.omtn.2020.05.028>.

AUTHOR CONTRIBUTIONS

Z.Z. and P.J. conceived the study. A.L. and S.M. collected the data and conducted the bioinformatics analysis. H.L. and D.-F.L. conducted laboratory experiments. A.L., S.M., H.L., D.-F.L., and Z.Z. verified the results and wrote the manuscript. All authors revised and approved the final manuscript.

CONFLICTS OF INTEREST

The authors declare no competing interests.

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REFERENCES

1. Iyer, M.K., Niknafs, Y.S., Malik, R., Singhal, U., Sahu, A., Hosono, Y., Barrette, T.R., Prensner, J.R., Evans, J.R., Zhao, S., et al. (2015). The landscape of long noncoding RNAs in the human transcriptome. *Nat. Genet.* **47**, 199–208.
2. Li, M., and Izpisua Belmonte, J.C. (2015). Roles for noncoding RNAs in cell-fate determination and regeneration. *Nat. Struct. Mol. Biol.* **22**, 2–4.
3. Mitra, R., Chen, X., Greenawalt, E.J., Maulik, U., Jiang, W., Zhao, Z., and Eischen, C.M. (2017). Decoding critical long non-coding RNA in ovarian cancer epithelial-to-mesenchymal transition. *Nat. Commun.* **8**, 1604.
4. Hu, Q., Ye, Y., Chan, L.-C., Li, Y., Liang, K., Lin, A., Egranolov, S.D., Zhang, Y., Xia, W., Gong, J., et al. (2019). Oncogenic lncRNA downregulates cancer cell antigen presentation and intrinsic tumor suppression. *Nat. Immunol.* **20**, 835–851.
5. Liu, S., Mitra, R., Zhao, M.-M., Fan, W., Eischen, C.M., Yin, F., and Zhao, Z. (2016). The potential roles of long noncoding RNAs (lncRNA) in glioblastoma development. *Mol. Cancer Ther.* **15**, 2977–2986.
6. Cui, W., Qian, Y., Zhou, X., Lin, Y., Jiang, J., Chen, J., Zhao, Z., and Shen, B. (2015). Discovery and characterization of long intergenic non-coding RNAs (lncRNA) module biomarkers in prostate cancer: an integrative analysis of RNA-seq data. *BMC Genomics* **16** (Suppl 7), S3.
7. Matouk, I.J., Raveh, E., Abu-lail, R., Mezan, S., Gilon, M., Gershstain, E., Birman, T., Gallula, J., Schneider, T., Barkali, M., et al. (2014). Oncofetal H19 RNA promotes tumor metastasis. *Biochim. Biophys. Acta* **1843**, 1414–1426.
8. Matouk, I.O.P., Ayesh, S., Sidi, A., Czerniak, A., Groot, N.D., and Hochberg, A. (2005). The oncofetal H19 RNA in human cancer, from the bench to the patient. *Cancer Ther.* **3**, 249–266.
9. Matouk, I.J., DeGroot, N., Mezan, S., Ayesh, S., Abu-lail, R., Hochberg, A., and Galun, E. (2007). The H19 non-coding RNA is essential for human tumor growth. *PLoS ONE* **2**, e845.
10. Raveh, E., Matouk, I.J., Gilon, M., and Hochberg, A. (2015). The H19 long non-coding RNA in cancer initiation, progression and metastasis—a proposed unifying theory. *Mol. Cancer* **14**, 184.
11. Ling, H., Ohtsuka, M., Ivan, C., Pichler, M., Chen, M., Slaby, O., Goel, A., Radovich, M., and Calin, G. (2017). Oncogenic function and molecular mechanism of H19 non-coding RNA in colorectal cancer. *Cancer Res.* **77** (Suppl), 2548, <https://doi.org/10.1158/1538-7445.AM2017-2548>.
12. Conigliaro, A., Costa, V., Lo Dico, A., Saieva, L., Buccheri, S., Dieli, F., Manno, M., Raccosta, S., Mancone, C., Tripodi, M., et al. (2015). CD90⁺ liver cancer cells modulate endothelial cell phenotype through the release of exosomes containing H19 lncRNA. *Mol. Cancer* **14**, 155.
13. Liu, L., Yang, J., Zhu, X., Li, D., Lv, Z., and Zhang, X. (2016). Long noncoding RNA H19 competitively binds miR-17-5p to regulate YES1 expression in thyroid cancer. *FEBS J.* **283**, 2326–2339.
14. Liu, C., Chen, Z., Fang, J., Xu, A., Zhang, W., and Wang, Z. (2016). H19-derived miR-675 contributes to bladder cancer cell proliferation by regulating p53 activation. *Tumour Biol.* **37**, 263–270.
15. Wang, P., Ning, S., Zhang, Y., Li, R., Ye, J., Zhao, Z., Zhi, H., Wang, T., Guo, Z., and Li, X. (2015). Identification of lncRNA-associated competing triplets reveals global patterns and prognostic markers for cancer. *Nucleic Acids Res.* **43**, 3478–3489.
16. Tay, Y., Rinn, J., and Pandolfi, P.P. (2014). The multilayered complexity of ceRNA crosstalk and competition. *Nature* **505**, 344–352.
17. Salmena, L., Poliseno, L., Tay, Y., Kats, L., and Pandolfi, P.P. (2011). A ceRNA hypothesis: the Rosetta Stone of a hidden RNA language? *Cell* **146**, 353–358.
18. Li, Y., Li, L., Wang, Z., Pan, T., Sahni, N., Jin, X., Wang, G., Li, J., Zheng, X., Zhang, Y., et al. (2018). LncMAP: pan-cancer atlas of long noncoding RNA-mediated transcriptional network perturbations. *Nucleic Acids Res.* **46**, 1113–1123.
19. Matys, V., Kel-Margoulis, O.V., Fricke, E., Liebich, I., Land, S., Barre-Dirrie, A., Reuter, I., Chekmenev, D., Krull, M., Hornischer, K., et al. (2006). TRANSFAC and its module TRANSCompel: transcriptional gene regulation in eukaryotes. *Nucleic Acids Res.* **34**, D108–D110.
20. Han, H., Cho, J.-W., Lee, S., Yun, A., Kim, H., Bae, D., Yang, S., Kim, C.Y., Lee, M., Kim, E., et al. (2018). TRRUST v2: an expanded reference database of human and mouse transcriptional regulatory interactions. *Nucleic Acids Res.* **46** (D1), D380–D386.
21. Cava, C., Colaprico, A., Bertoli, G., Graudenzi, A., Silva, T.C., Olsen, C., Noushmehr, H., Bontempi, G., Mauri, G., and Castiglioni, I. (2017). SpidermiR: an R/bioconductor package for integrative analysis with miRNA data. *Int. J. Mol. Sci.* **18**, E274.
22. Law, C.W., Chen, Y., Shi, W., and Smyth, G.K. (2014). voom: precision weights unlock linear model analysis tools for RNA-seq read counts. *Genome Biol.* **15**, R29.
23. Kallen, A.N., Zhou, X.-B., Xu, J., Qiao, C., Ma, J., Yan, L., Lu, L., Liu, C., Yi, J.S., Zhang, H., et al. (2013). The imprinted H19 lncRNA antagonizes let-7 microRNAs. *Mol. Cell* **52**, 101–112.
24. Huang, Z., Lei, W., Hu, H.B., Zhang, H., and Zhu, Y. (2018). H19 promotes non-small-cell lung cancer (NSCLC) development through STAT3 signaling via sponging miR-17. *J. Cell. Physiol.* **233**, 6768–6776.
25. Jia, P., Cai, H., Liu, X., Chen, J., Ma, J., Wang, P., Liu, Y., Zheng, J., and Xue, Y. (2016). Long non-coding RNA H19 regulates glioma angiogenesis and the biological behavior of glioma-associated endothelial cells by inhibiting microRNA-29a. *Cancer Lett.* **381**, 359–369.
26. He, H., Wang, N., Yi, X., Tang, C., and Wang, D. (2017). Long non-coding RNA H19 regulates E2F1 expression by competitively sponging endogenous miR-29a-3p in clear cell renal cell carcinoma. *Cell Biosci.* **7**, 65.
27. Zhou, J., Zhou, Y., and Wang, C.X. (2018). nlcRNA-MIAT regulates fibrosis in hypertrophic cardiomyopathy (HCM) by mediating the expression of miR-29a-3p. *J. Cell. Biochem.* **120**, 7265–7275.
28. Zhou, W., Ye, X.-L., Xu, J., Cao, M.-G., Fang, Z.-Y., Li, L.-Y., Guan, G.-H., Liu, Q., Qian, Y.-H., and Xie, D. (2017). The lncRNA H19 mediates breast cancer cell plasticity during EMT and MET plasticity by differentially sponging miR-200b/c and let-7b. *Sci. Signal.* **10**, eaak9557.
29. Lee, D.-F., Su, J., Kim, H.S., Chang, B., Papatsenko, D., Zhao, R., Yuan, Y., Gingold, J., Xia, W., Darr, H., et al. (2015). Modeling familial cancer with induced pluripotent stem cells. *Cell* **161**, 240–254.
30. Huarte, M. (2015). The emerging role of lncRNAs in cancer. *Nat. Med.* **21**, 1253–1261.

31. Sun, J., Gong, X., Purow, B., and Zhao, Z. (2012). Uncovering microRNA and transcription factor mediated regulatory networks in glioblastoma. *PLoS Comput. Biol.* 8, e1002488.
32. Li, A., Jia, P., Mallik, S., Fei, R., Yoshioka, H., Suzuki, A., Iwata, J., and Zhao, Z. (2019). Critical microRNAs and regulatory motifs in cleft palate identified by a conserved miRNA-TF-gene network approach in humans and mice. *Brief. Bioinform. bbz082*.
33. Mitra, R., Edmonds, M.D., Sun, J., Zhao, M., Yu, H., Eischen, C.M., and Zhao, Z. (2014). Reproducible combinatorial regulatory networks elucidate novel oncogenic microRNAs in non-small cell lung cancer. *RNA* 20, 1356–1368.
34. ENCODE Project Consortium. (2012). An integrated encyclopedia of DNA elements in the human genome. *Nature* 489, 57–74.
35. Zhu, Q.H., Stephen, S., Taylor, J., Helliwell, C.A., and Wang, M.B. (2014). Long non-coding RNAs responsive to Fusarium oxysporum infection in *Arabidopsis thaliana*. *New Phytol.* 201, 574–584.
36. GTEx Consortium (2015). Human genomics. The Genotype-Tissue Expression (GTEx) pilot analysis: multitissue gene regulation in humans. *Science* 348, 648–660.
37. Chiang, C., Scott, A.J., Davis, J.R., Tsang, E.K., Li, X., Kim, Y., Hadzic, T., Damani, F.N., Ganel, L., Montgomery, S.B., et al.; GTEx Consortium (2017). The impact of structural variation on human gene expression. *Nat. Genet.* 49, 692–699.
38. Stranger, B.E., Forrest, M.S., Dunning, M., Ingle, C.E., Beazley, C., Thorne, N., Redon, R., Bird, C.P., de Grassi, A., Lee, C., et al. (2007). Relative impact of nucleotide and copy number variation on gene expression phenotypes. *Science* 315, 848–853.
39. Jia, P., and Zhao, Z. (2017). Impacts of somatic mutations on gene expression: an association perspective. *Brief. Bioinform.* 18, 413–425.
40. Helwak, A., Kudla, G., Dudnakova, T., and Tollervey, D. (2013). Mapping the human miRNA interactome by CLASH reveals frequent noncanonical binding. *Cell* 153, 654–665.
41. Song, K., Wang, H., Krebs, T.L., Kim, S.-J., and Danielpour, D. (2008). Androgenic control of transforming growth factor- β signaling in prostate epithelial cells through transcriptional suppression of transforming growth factor- β receptor II. *Cancer Res.* 68, 8173–8182.
42. Li, L., Gao, P., Li, Y., Shen, Y., Xie, J., Sun, D., Xue, A., Zhao, Z., Xu, Z., Zhang, M., et al. (2014). JMJD2A-dependent silencing of Sp1 in advanced breast cancer promotes metastasis by downregulation of DIRAS3. *Breast Cancer Res. Treat.* 147, 487–500.
43. Breunig, C., Erdem, N., Bott, A., Greiwe, J.F., Reinz, E., Bernhardt, S., Giacomelli, C., Wachter, A., Kanthelhardt, E.J., Beißbarth, T., et al. (2018). TGF β 1 regulates HGF-induced cell migration and hepatocyte growth factor receptor MET expression via C-ets-1 and miR-128-3p in basal-like breast cancer. *Mol. Oncol.* 12, 1447–1463.
44. Ma, X., Beeghly-Fadiel, A., Lu, W., Shi, J., Xiang, Y.-B., Cai, Q., Shen, H., Shen, C.Y., Ren, Z., Matsuo, K., et al. (2012). Pathway analyses identify *TGFB2* as potential breast cancer susceptibility gene: results from a consortium study among Asians. *Cancer Epidemiol. Biomarkers Prev.* 21, 1176–1184.
45. Berteaux, N., Lottin, S., Monté, D., Pinte, S., Quatannens, B., Coll, J., Hondermarck, H., Curgy, J.J., Dugimont, T., and Adriaenssens, E. (2005). *H19* mRNA-like noncoding RNA promotes breast cancer cell proliferation through positive control by E2F1. *J. Biol. Chem.* 280, 29625–29636.
46. Lottin, S., Adriaenssens, E., Dupressoir, T., Berteaux, N., Montpellier, C., Coll, J., Dugimont, T., and Curgy, J.J. (2002). Overexpression of an ectopic *H19* gene enhances the tumorigenic properties of breast cancer cells. *Carcinogenesis* 23, 1885–1895.
47. Ma, L., Li, G.Z., Wu, Z.S., and Meng, G. (2014). Prognostic significance of let-7b expression in breast cancer and correlation to its target gene of BSG expression. *Med. Oncol.* 31, 773.
48. Li, B., and Dewey, C.N. (2011). RSEM: accurate transcript quantification from RNA-seq data with or without a reference genome. *BMC Bioinformatics* 12, 323.
49. Hsu, J.B.-K., Chiu, C.-M., Hsu, S.-D., Huang, W.-Y., Chien, C.-H., Lee, T.-Y., and Huang, H.D. (2011). miRTar: an integrated system for identifying miRNA-target interactions in human. *BMC Bioinformatics* 12, 300.
50. Dweep, H., Sticht, C., Pandey, P., and Gretz, N. (2011). miRWALK—database: prediction of possible miRNA binding sites by “walking” the genes of three genomes. *J. Biomed. Inform.* 44, 839–847.
51. Maragakis, M., Vergoulis, T., Alexiou, P., Reczko, M., Plomaritou, K., Gousis, M., Kourtis, K., Koziris, N., Dalamagas, T., and Hatzigeorgiou, A.G. (2011). DIANA-microT Web server upgrade supports Fly and Worm miRNA target prediction and bibliographic miRNA to disease association. *Nucleic Acids Res.* 39, W145–8.
52. Enright, A.J., John, B., Gaul, U., Tuschl, T., Sander, C., and Marks, D.S. (2003). MicroRNA targets in *Drosophila*. *Genome Biol.* 5, R1.
53. Krek, A., Grün, D., Poy, M.N., Wolf, R., Rosenberg, L., Epstein, E.J., MacMenamin, P., da Piedade, I., Gunsalus, K.C., Stoffel, M., and Rajewsky, N. (2005). Combinatorial microRNA target predictions. *Nat. Genet.* 37, 495–500.
54. Bartel, D.P. (2009). MicroRNAs: target recognition and regulatory functions. *Cell* 136, 215–233.
55. Pinheiro, J., Bates, D., DebRoy, S., and Sarkar, D. (2007). Linear and nonlinear mixed effects models. R package version 3, 1–89, <https://cran.r-project.org/web/packages/nlme/index.html>.

Supplemental Information

**H19, a Long Non-coding RNA, Mediates
Transcription Factors and Target Genes through
Interference of MicroRNAs in Pan-Cancer**

Aimin Li, Saurav Mallik, Haidan Luo, Peilin Jia, Dung-Fang Lee, and Zhongming Zhao

SUPPLEMENTARY FILES

Figure S1. H19 highly expressed across pan-cancer except for brain lower grade glioma (LGG), prostate adenocarcinoma (PRAD), and thyroid carcinoma (THCA).

Figure S2. TF-gene regulation was affected by H19 expression level.

Table S1. Number of samples and genes across the 24 cancer types based on TCGA data.

Table S2. Eighty-eight H19-TF-gene regulation triplets identified in at least two cancer types.

Table S3. 173 of 186 (93%) TF-gene pairs had direct or indirect evidence to support their relation to cancer (Table S3). The remaining 13 TF-gene pairs might be potential candidates for cancer research.

Table S4. The list of 29 H19 target miRNAs with evidence in literature.

Table S5. Regulation of 29 miRNAs in eight triplets. In the H19-ETS1-*TGFB2* sheet, we list all the 29 miRNAs and their targets (TFs and genes). Some of the targets were predicted and then verified. In the H19-ETS1-*TGFB2* table, TFs are marked in yellow if miRNAs target them, and genes are marked in red if miRNAs target them.

Table S6. Primers for qRT-PCR.

Figure S1. H19 highly expressed across pan-cancer except for brain lower grade glioma (LGG), prostate adenocarcinoma (PRAD), and thyroid carcinoma (THCA).

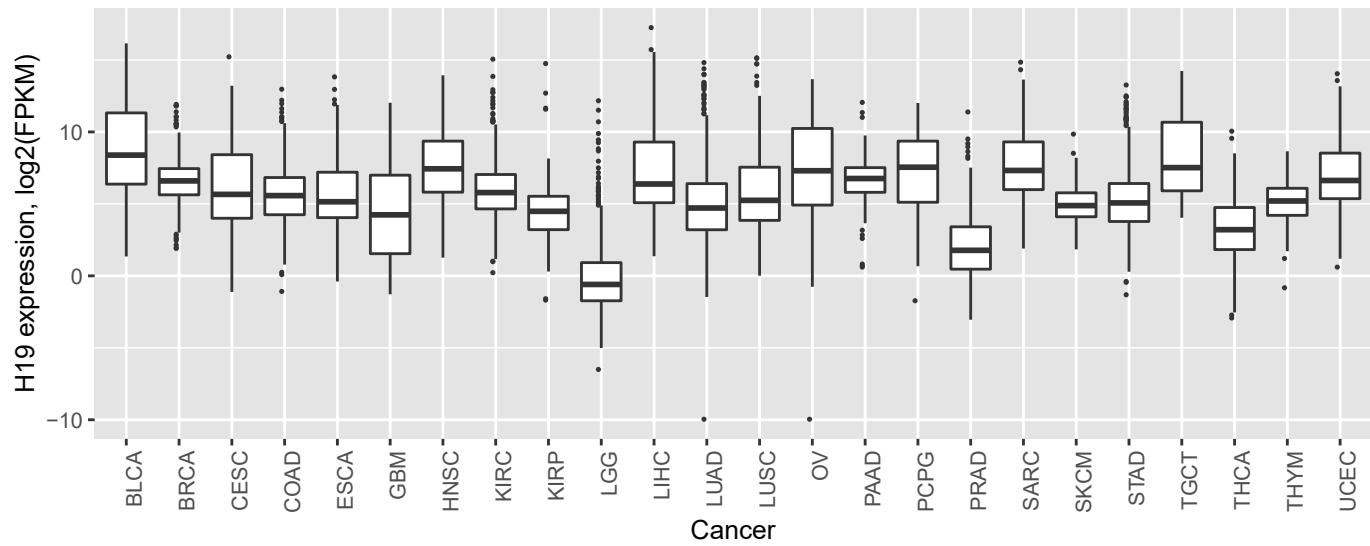
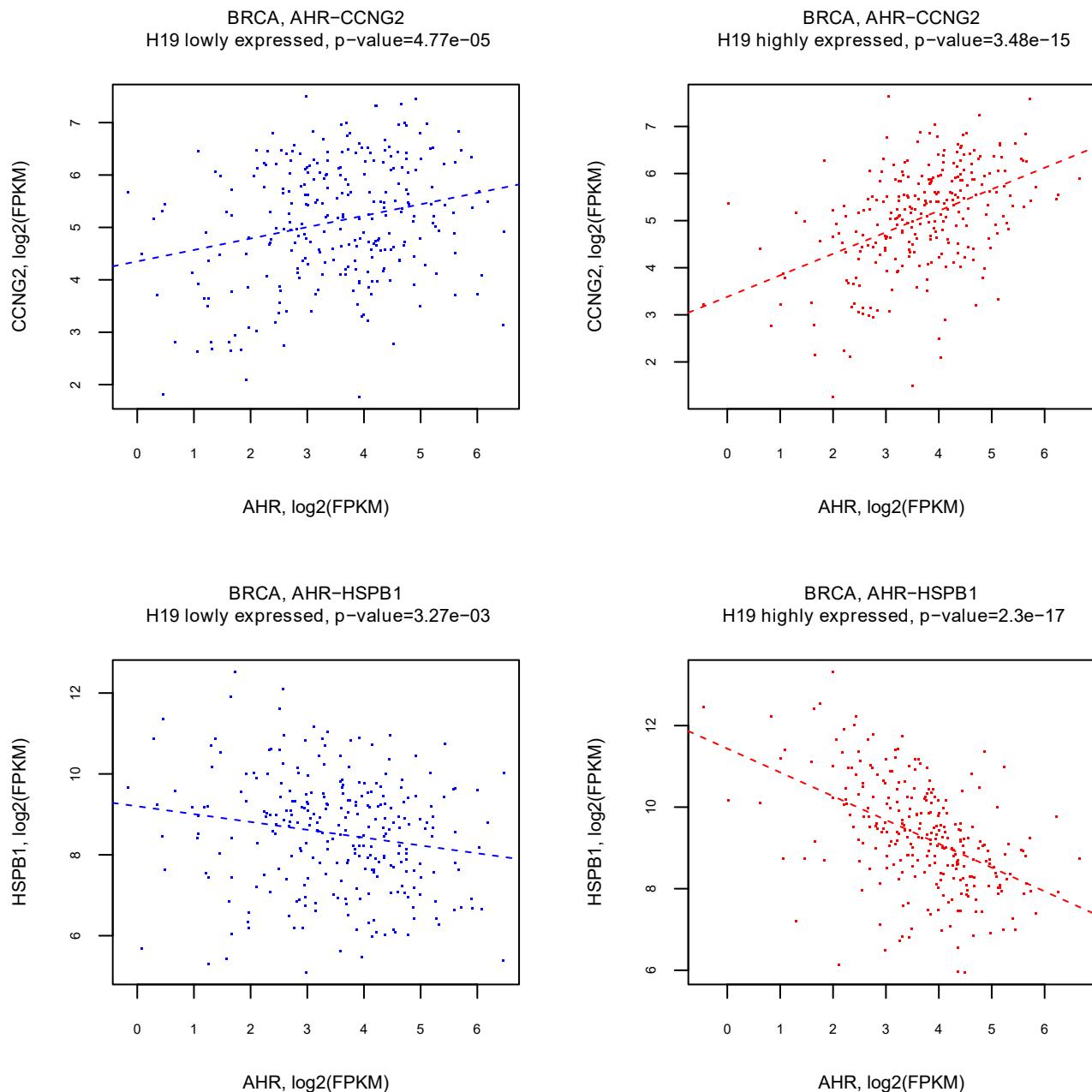
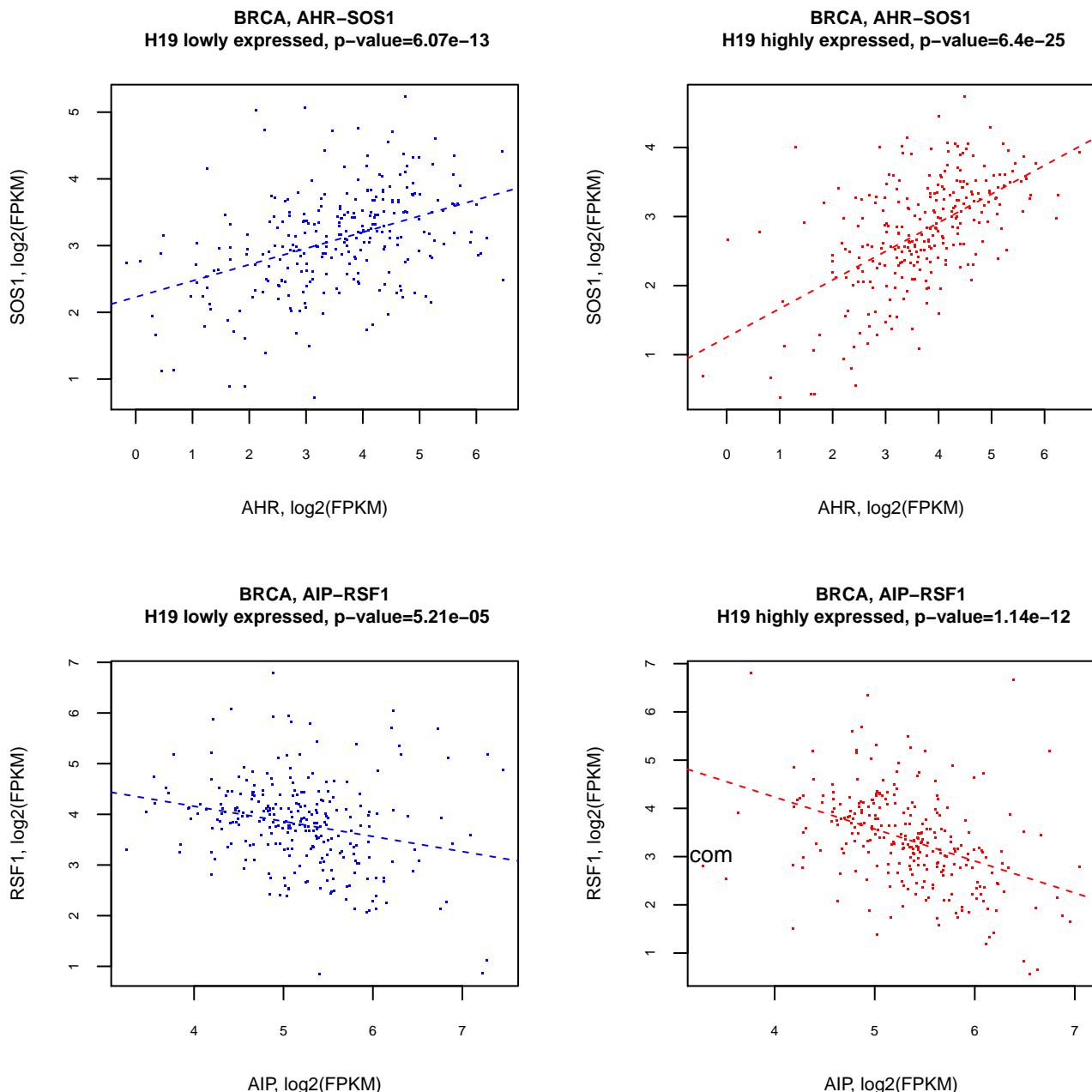
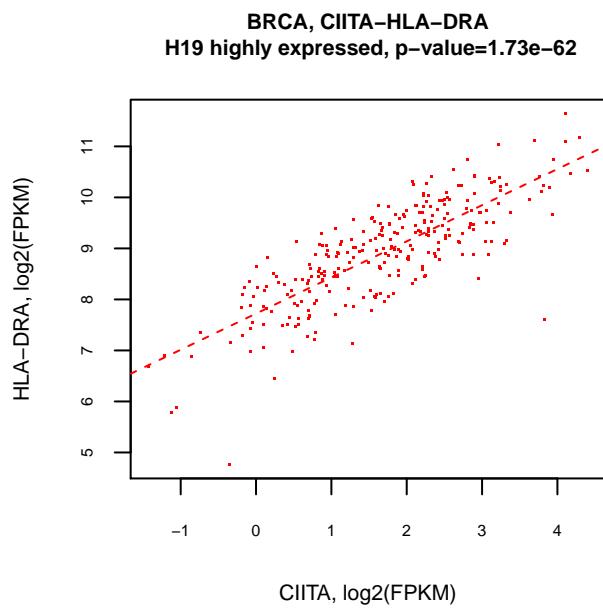
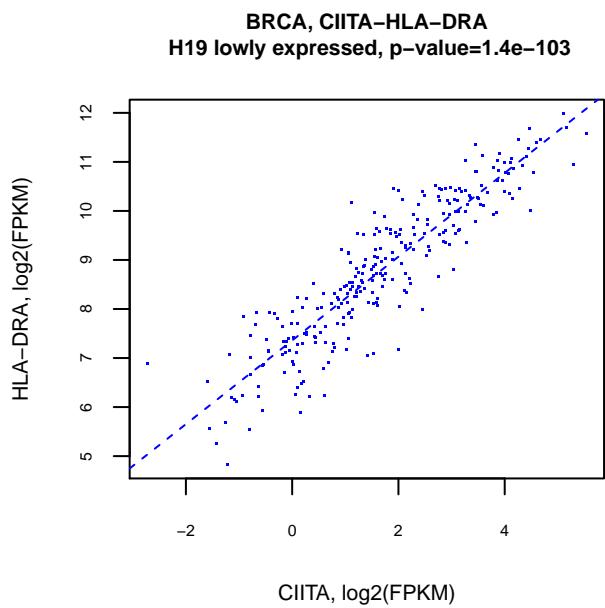
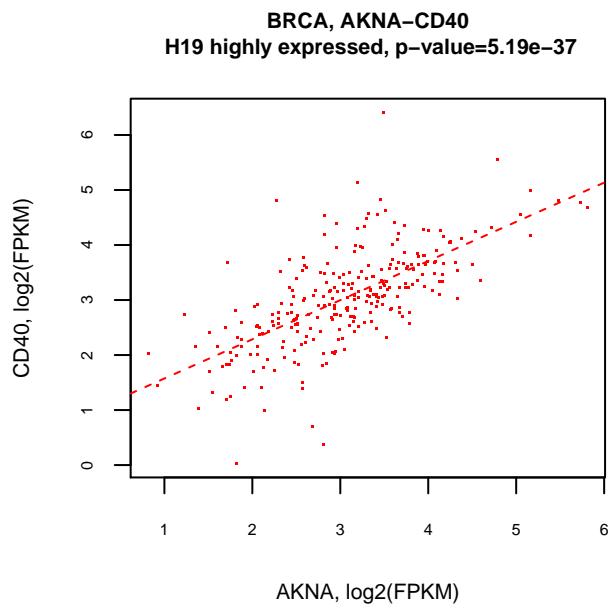
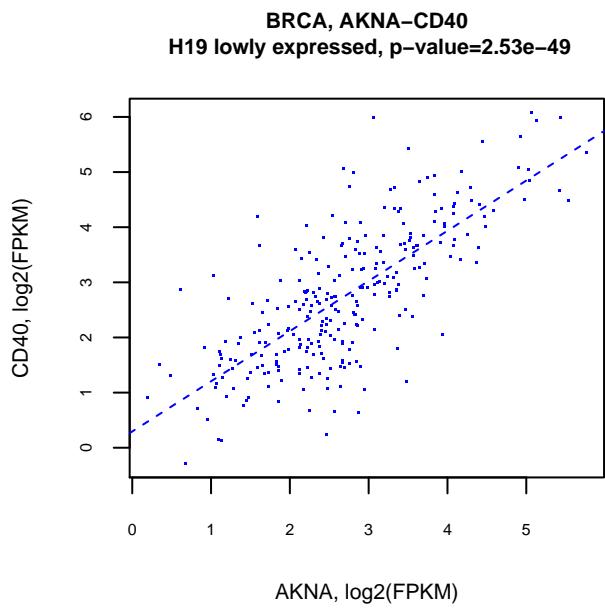
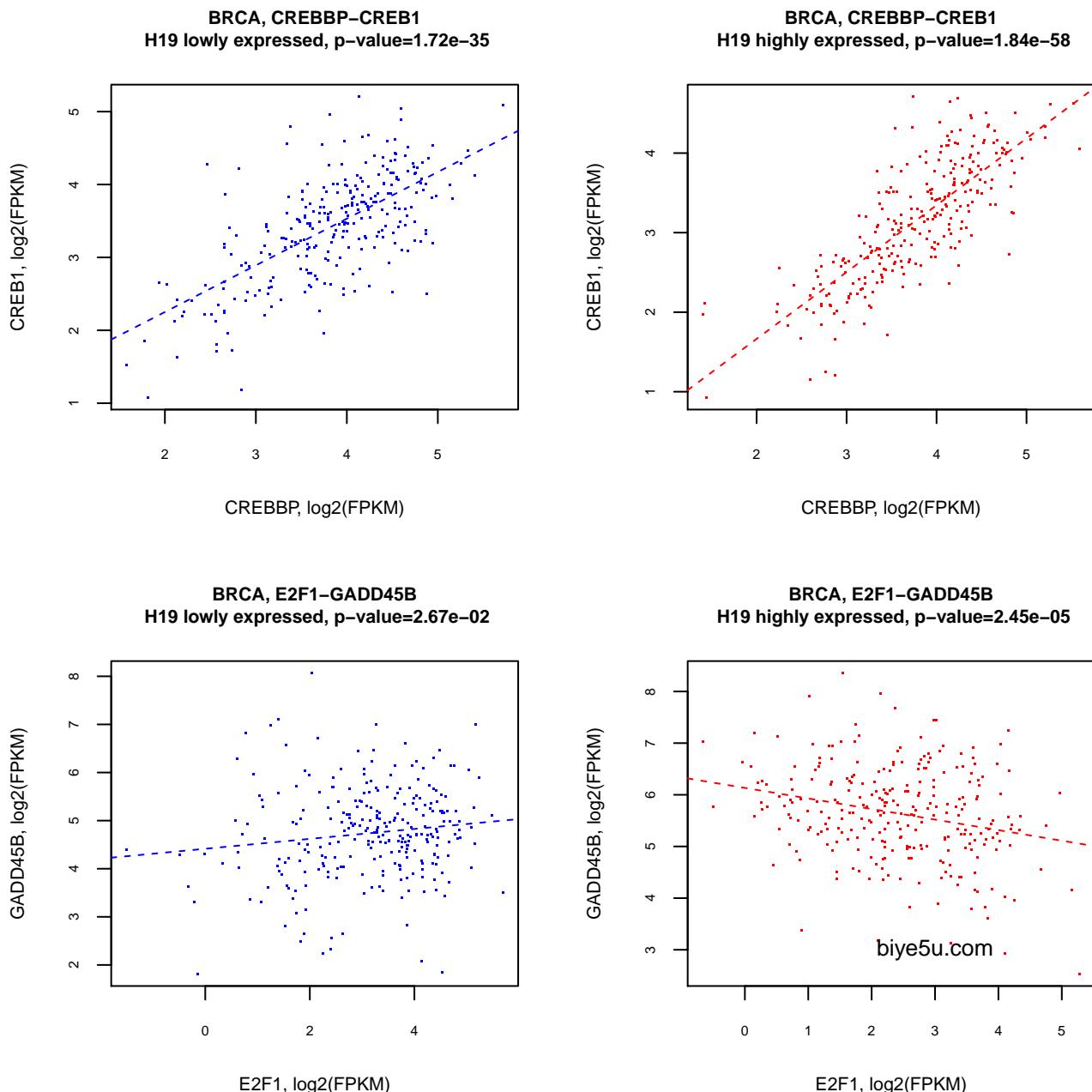


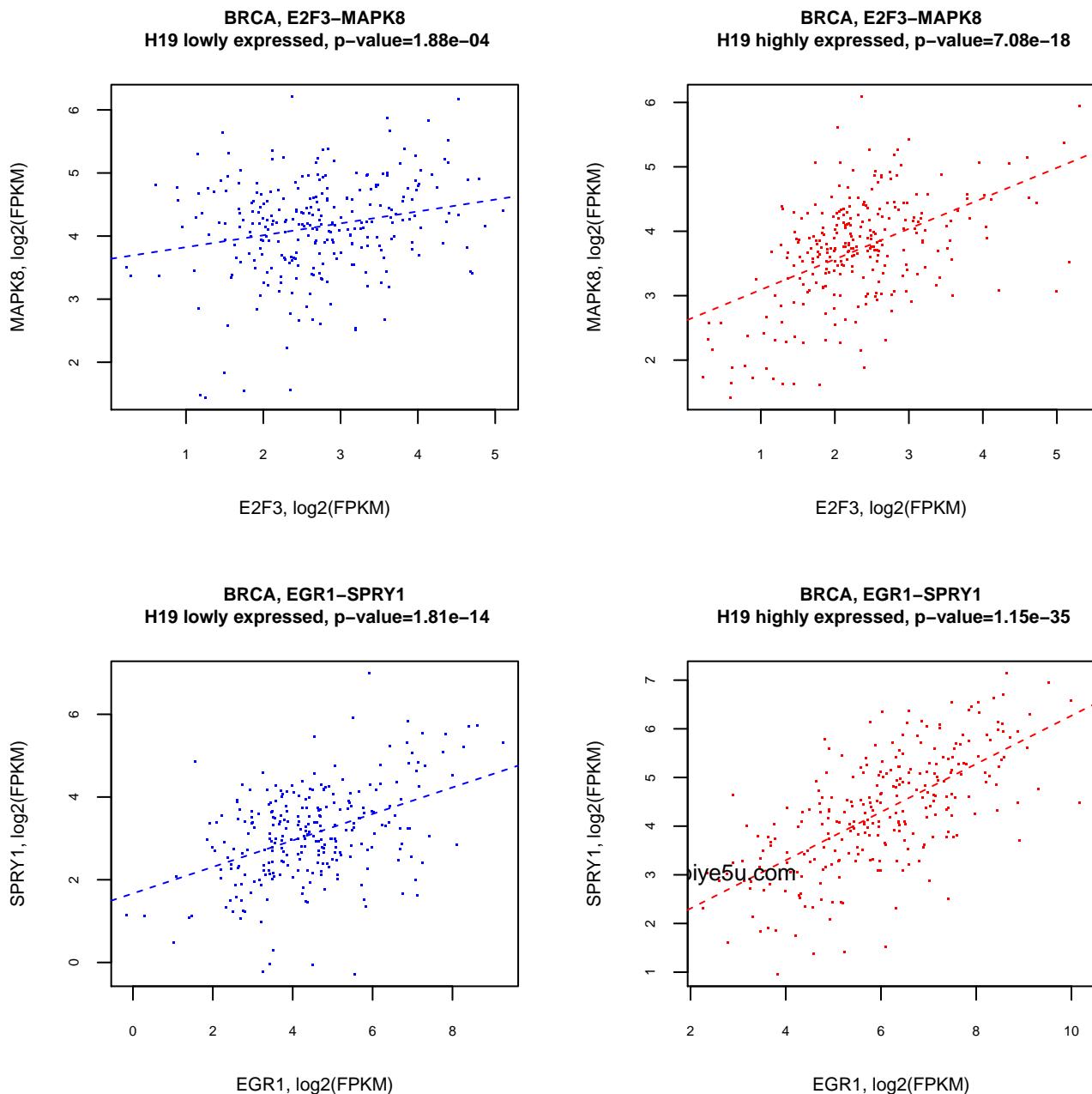
Figure S2. TF-gene regulation was affected by H19 expression level.

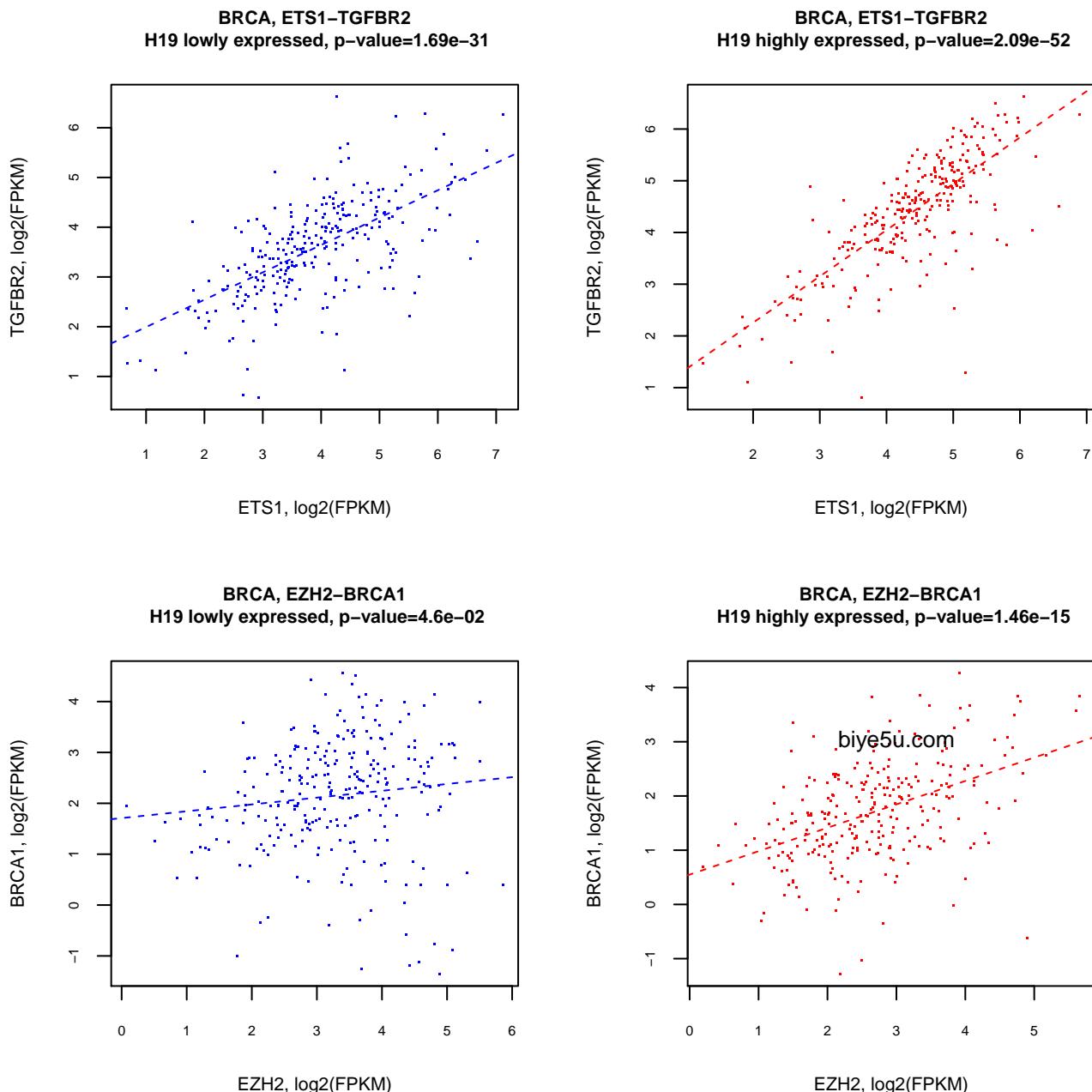


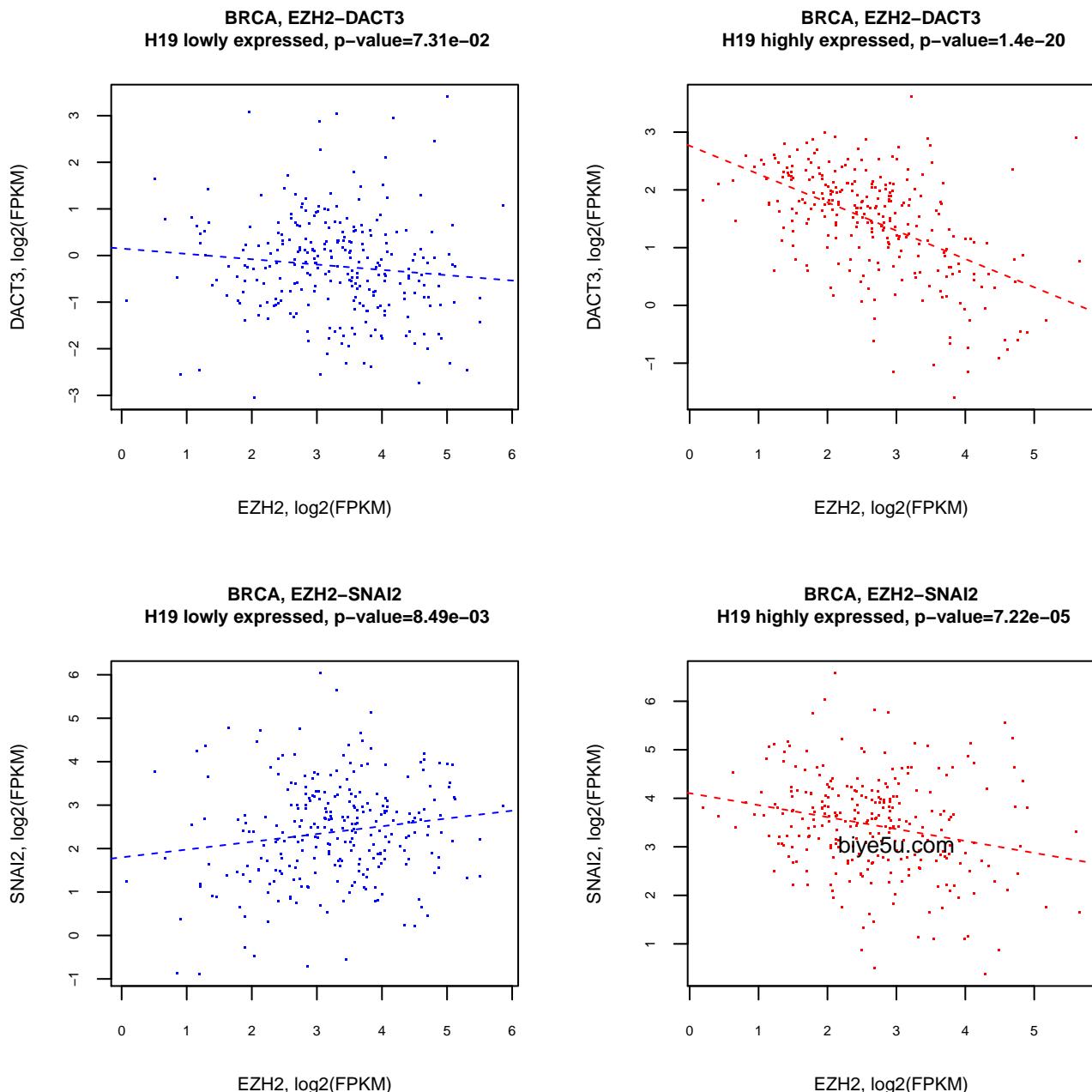


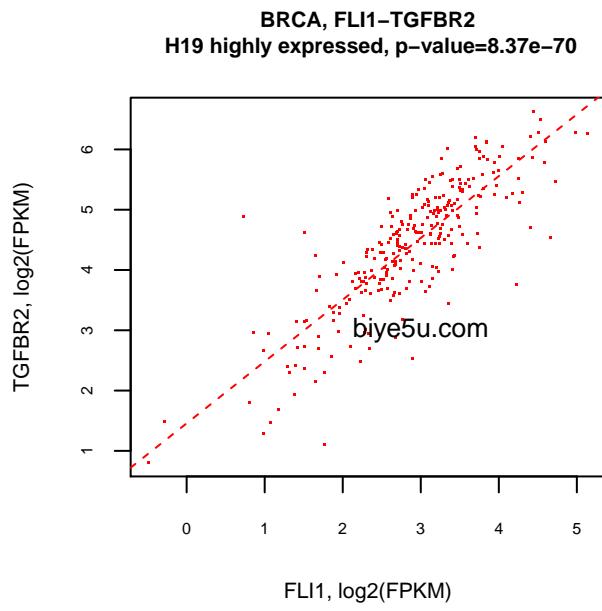
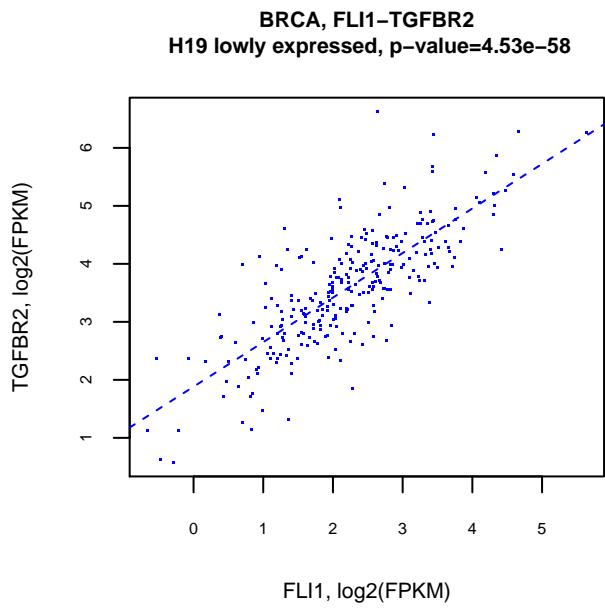
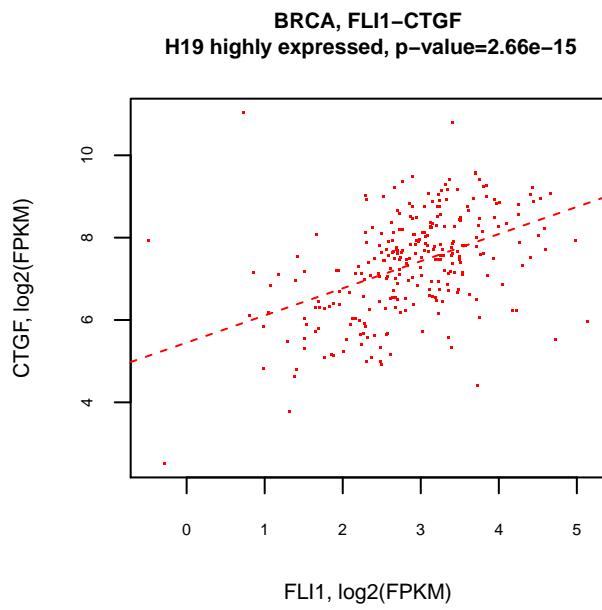
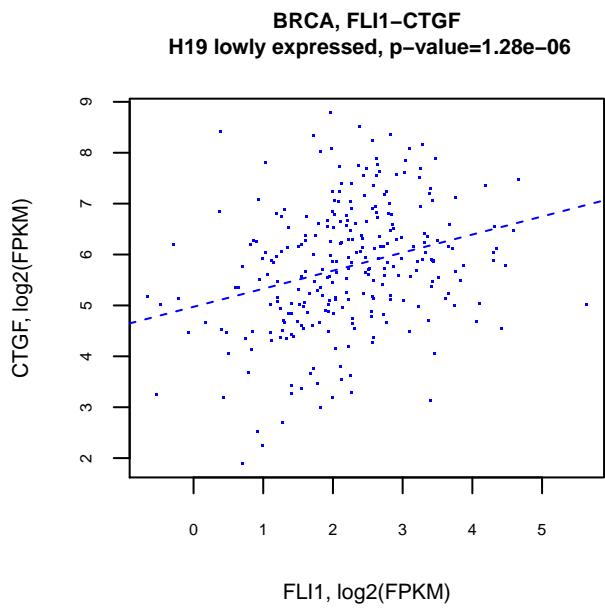


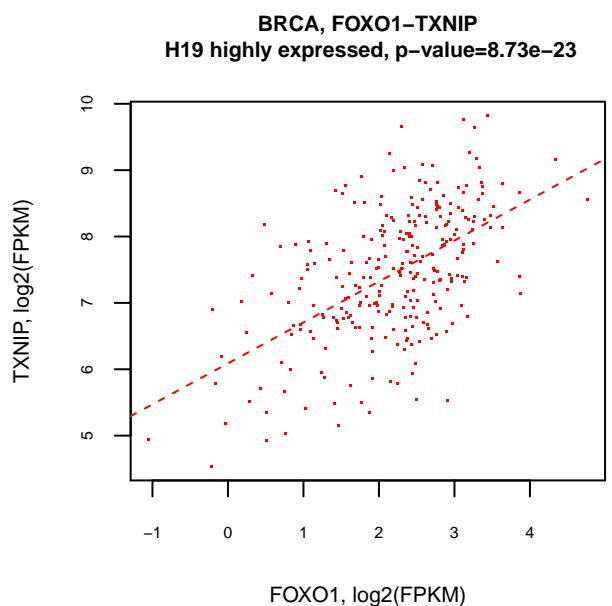
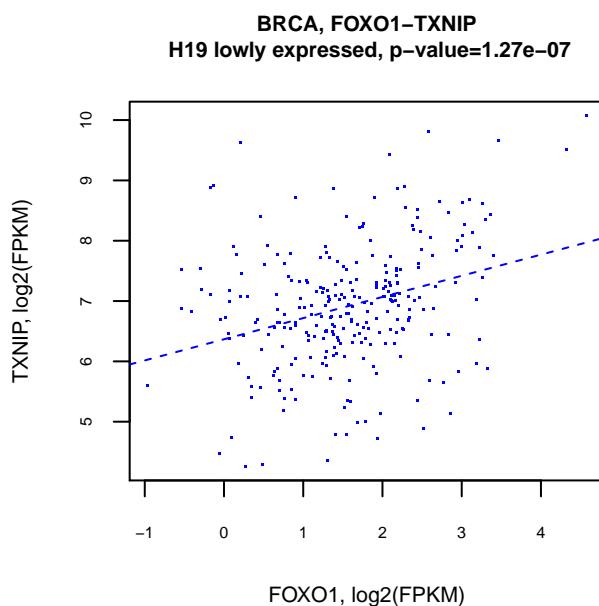
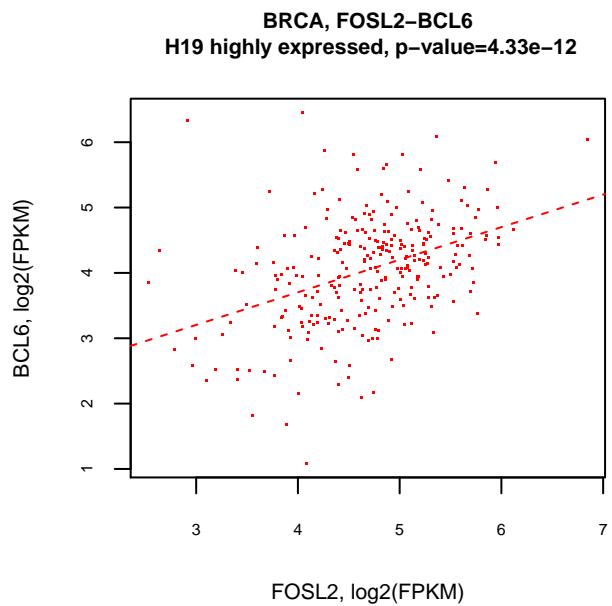
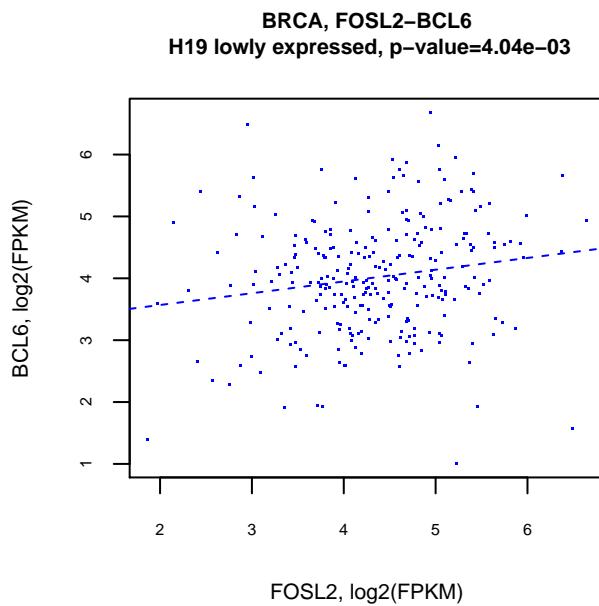


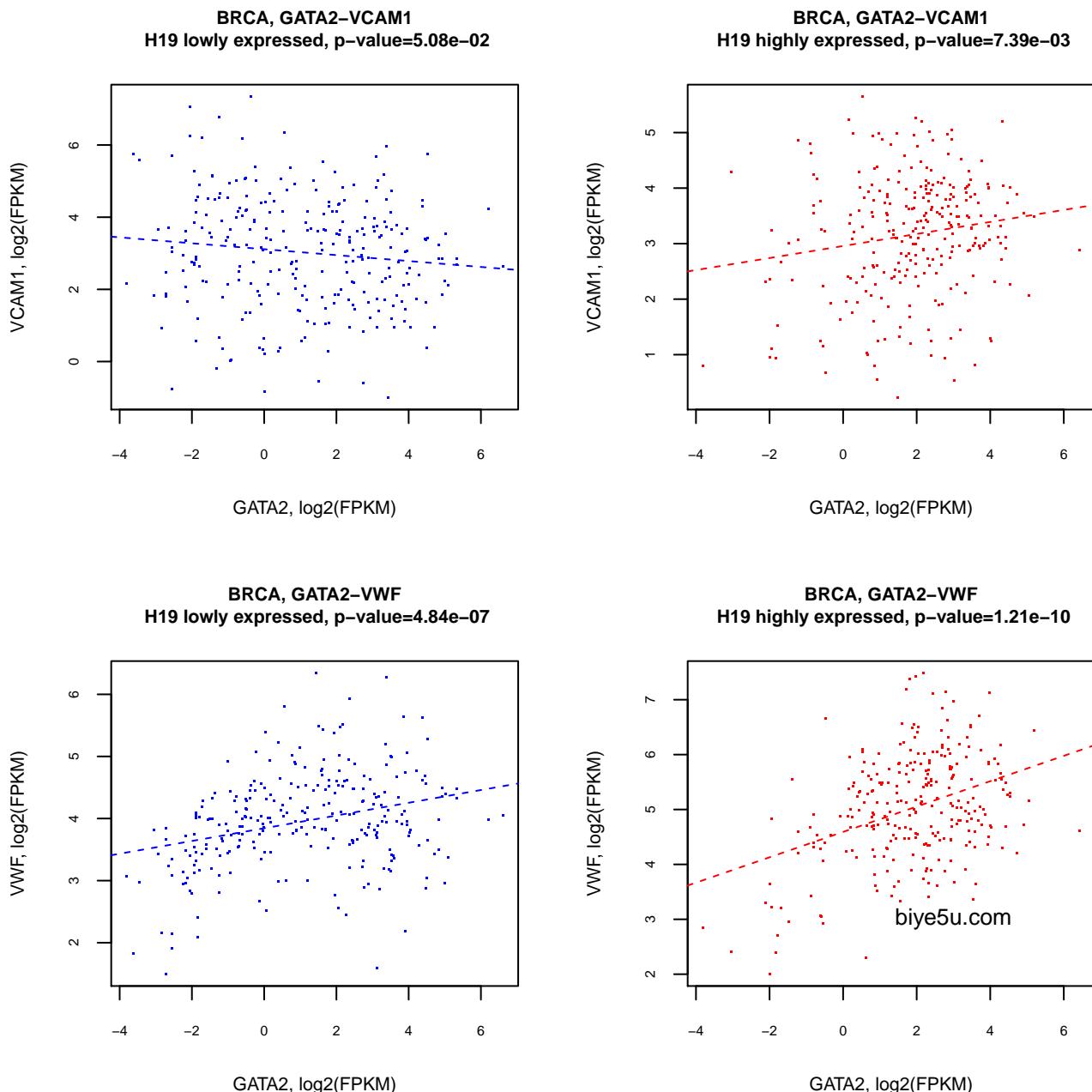


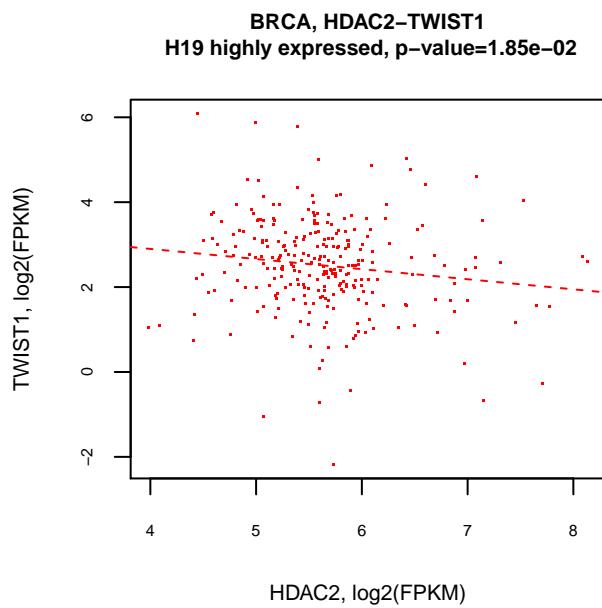
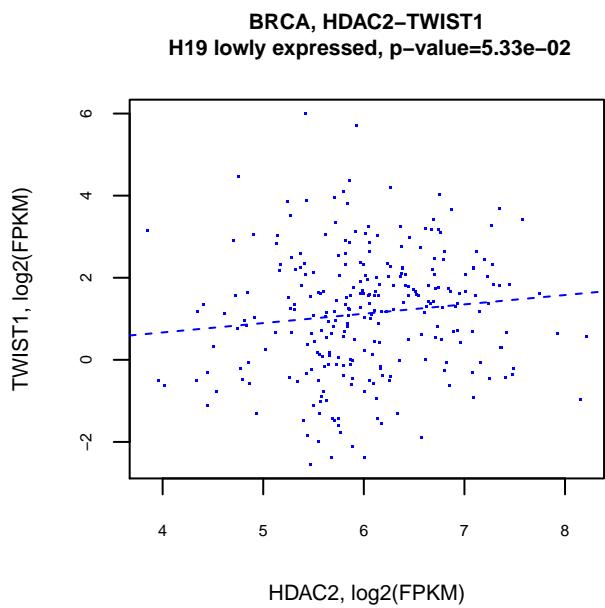
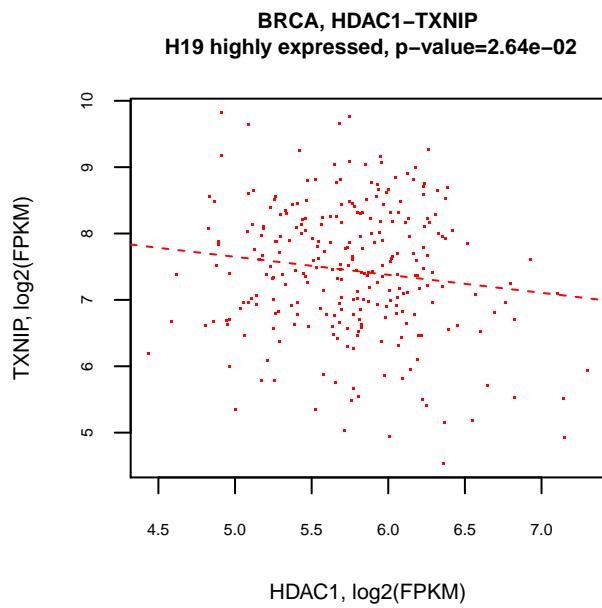
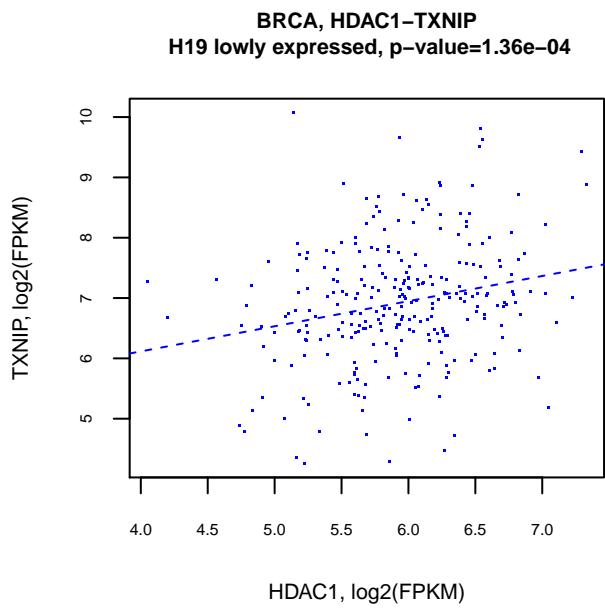


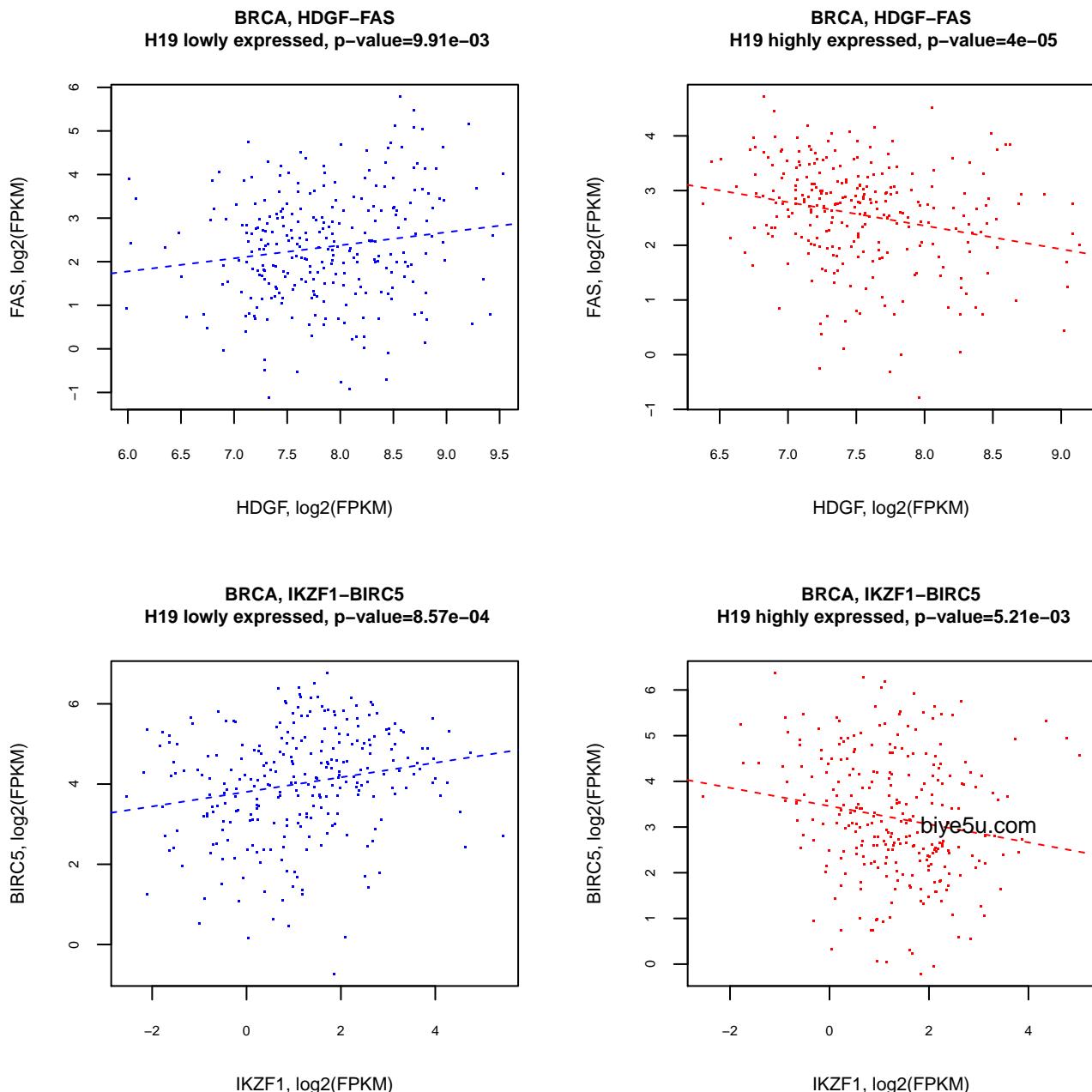


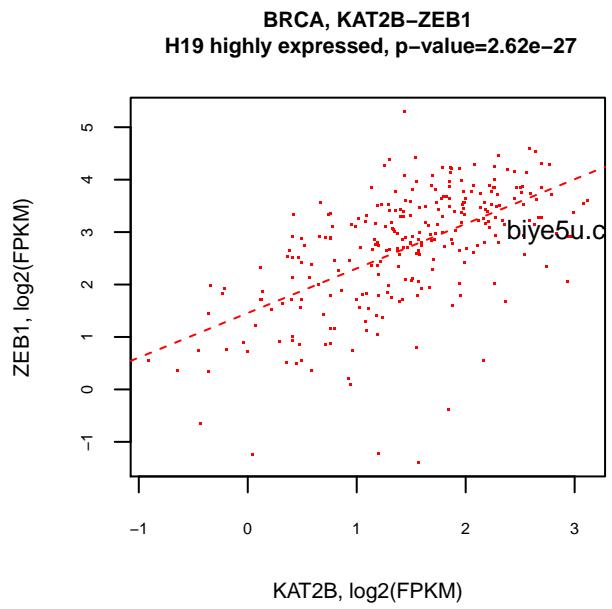
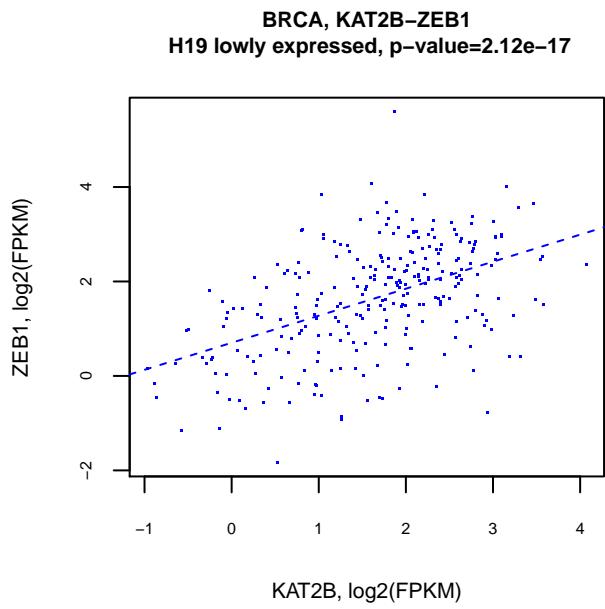
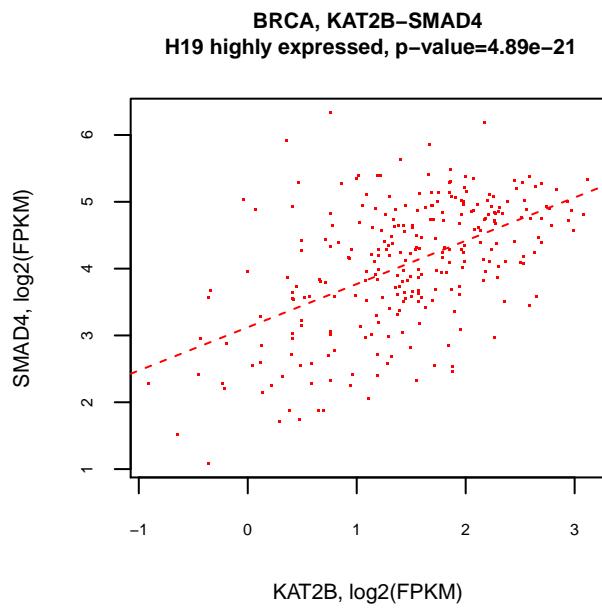
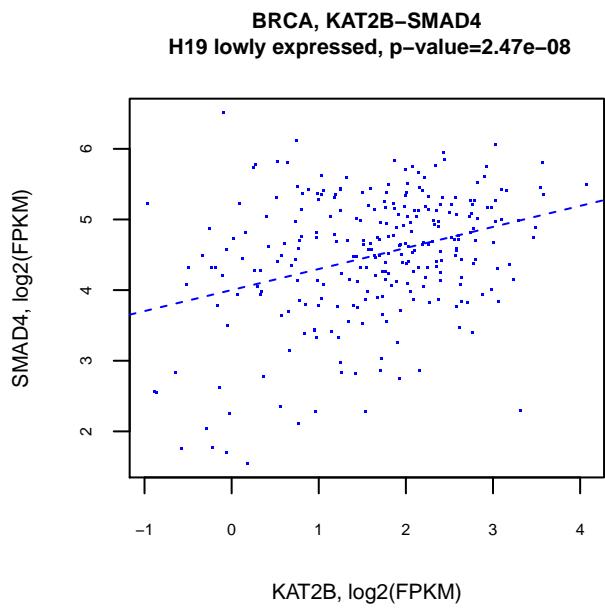


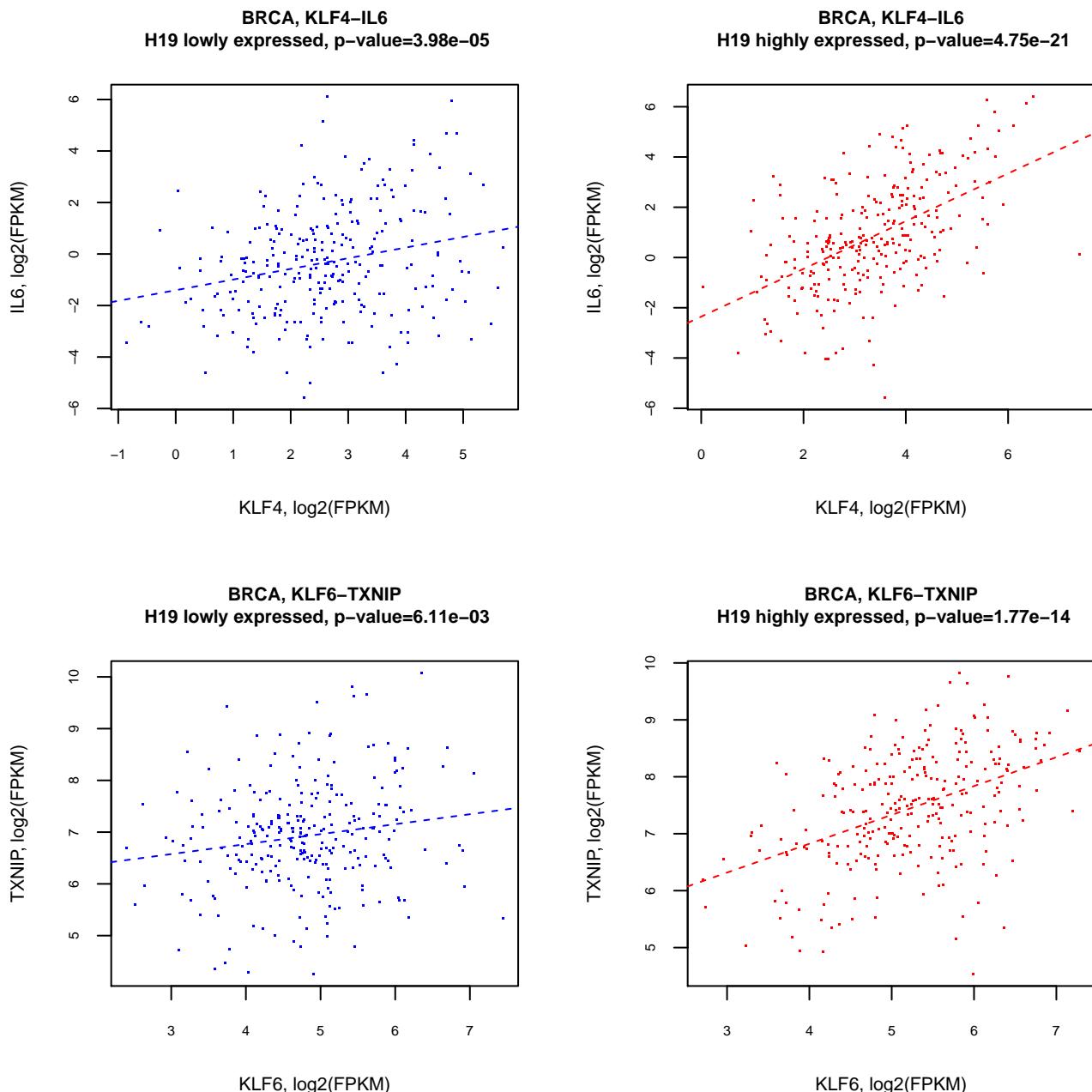


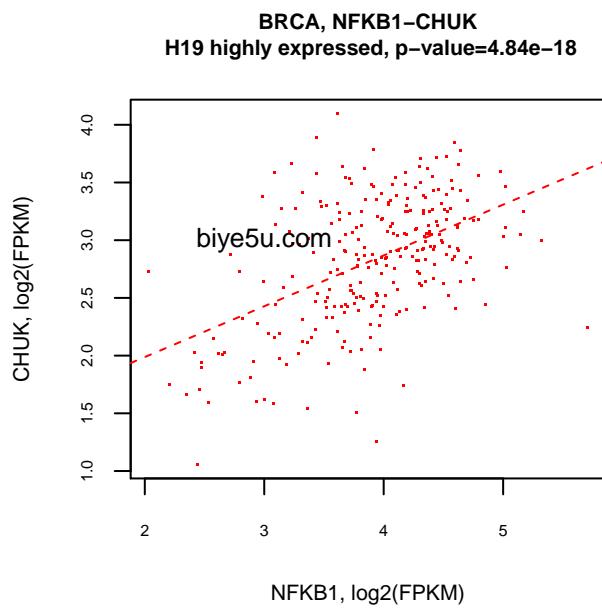
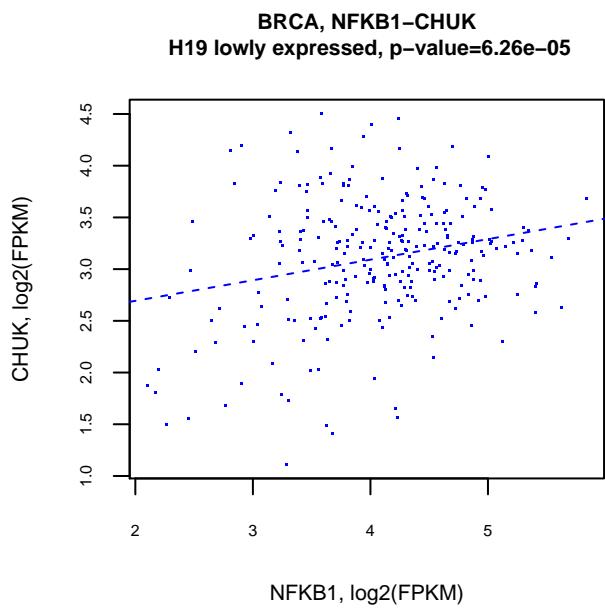
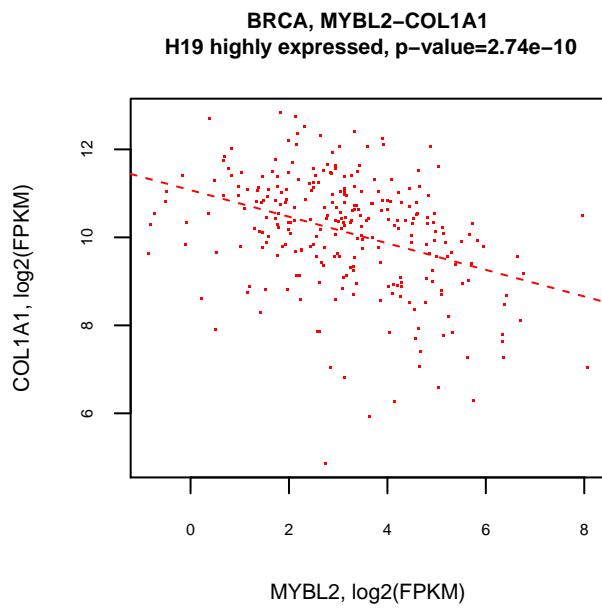
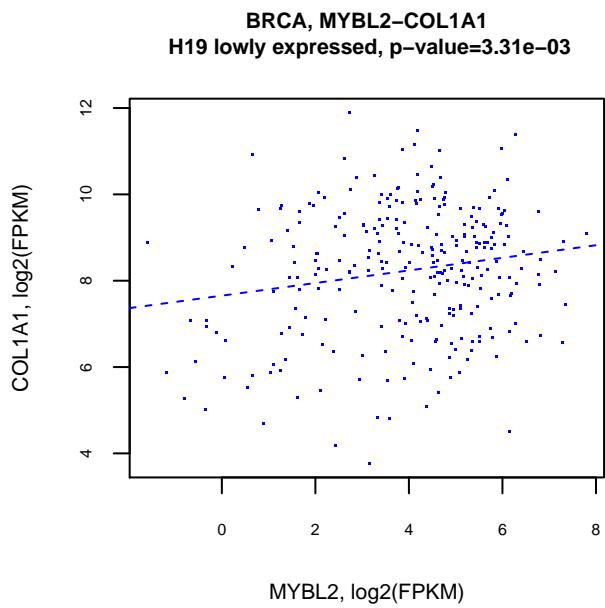


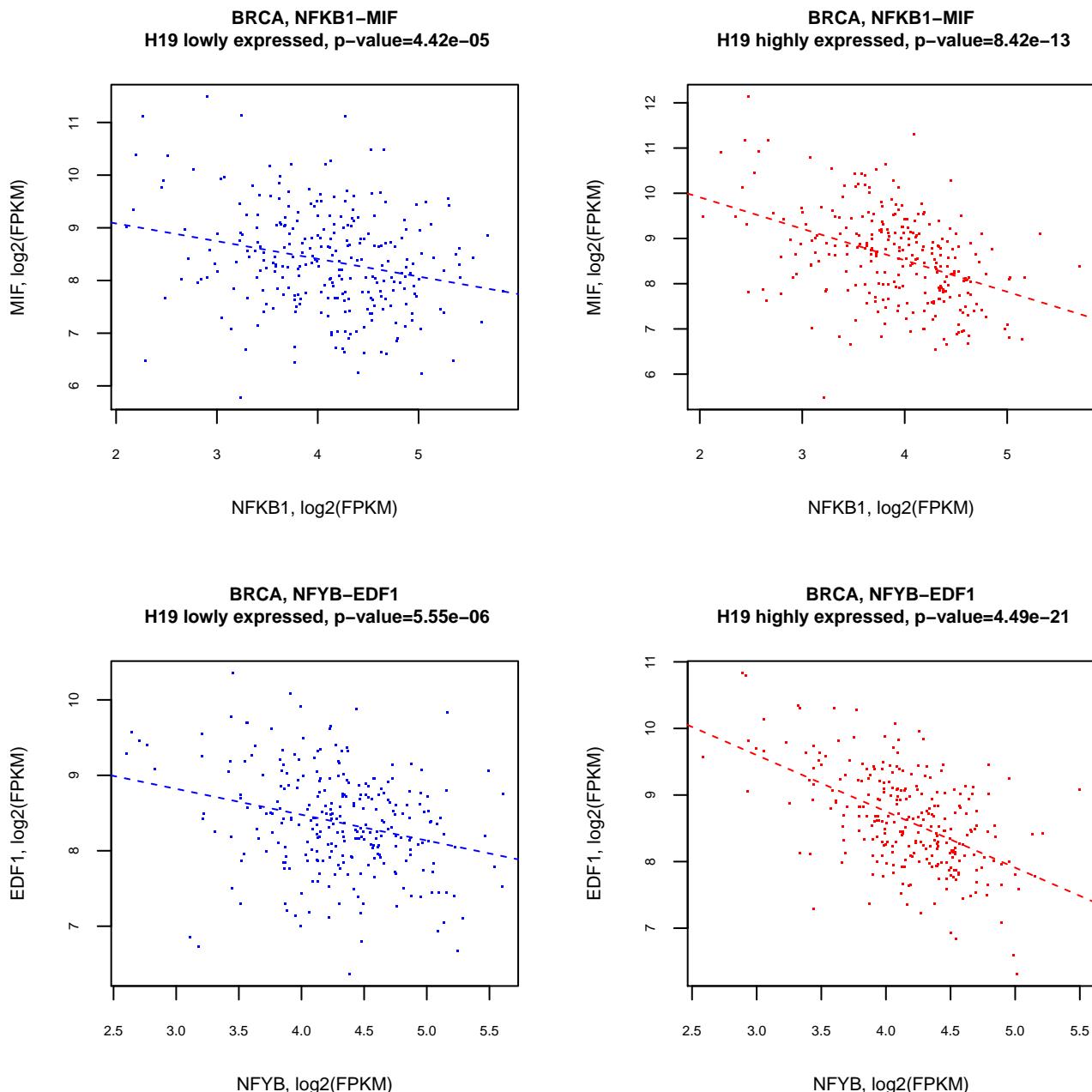


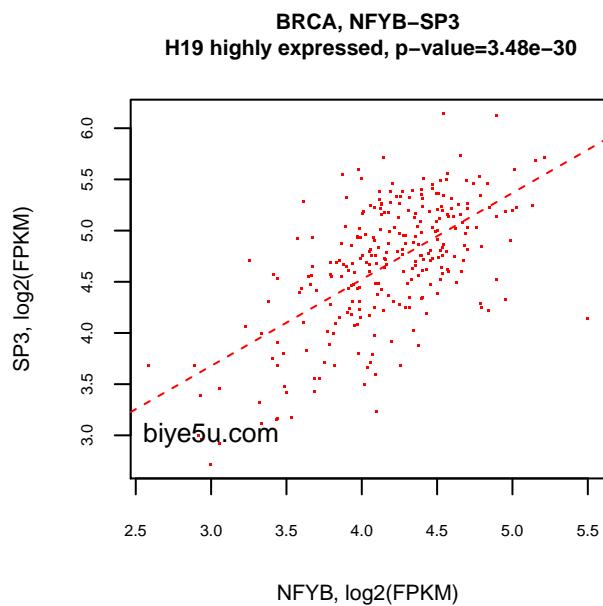
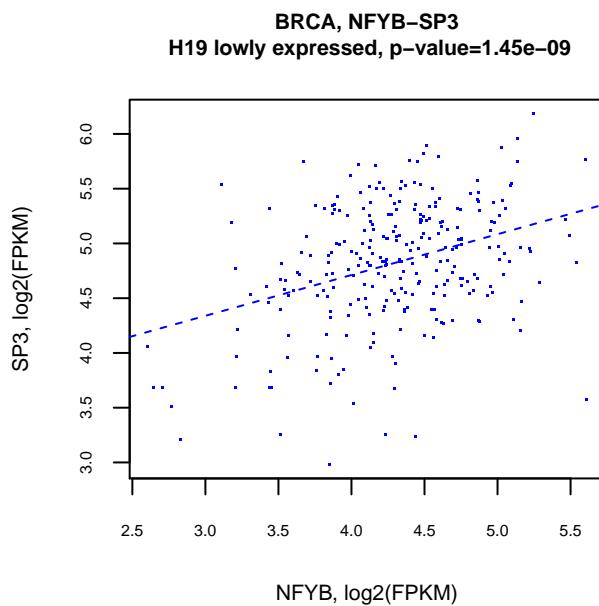
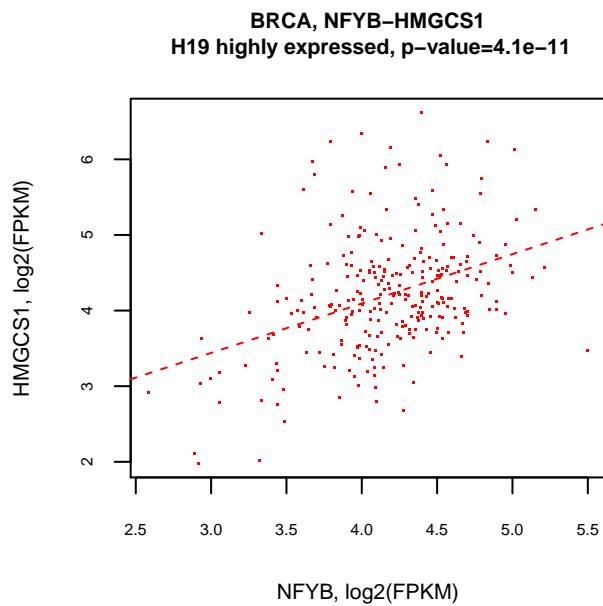
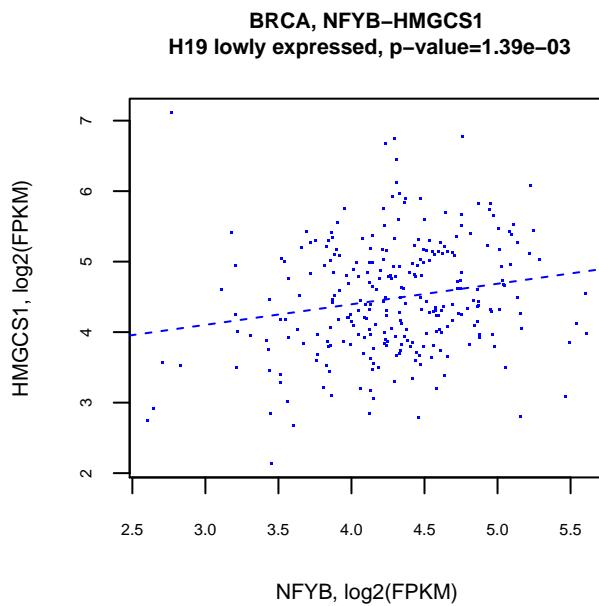


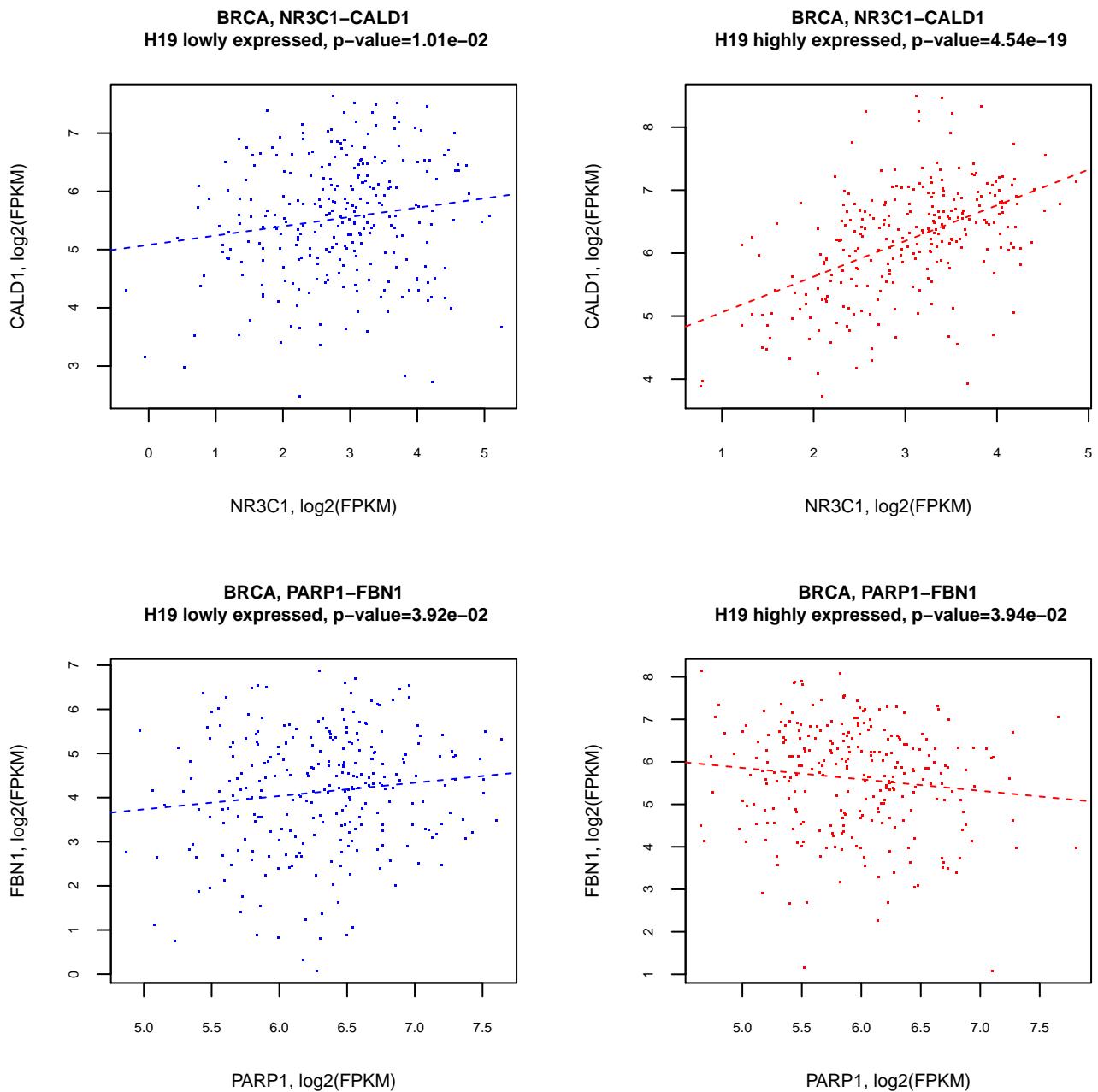


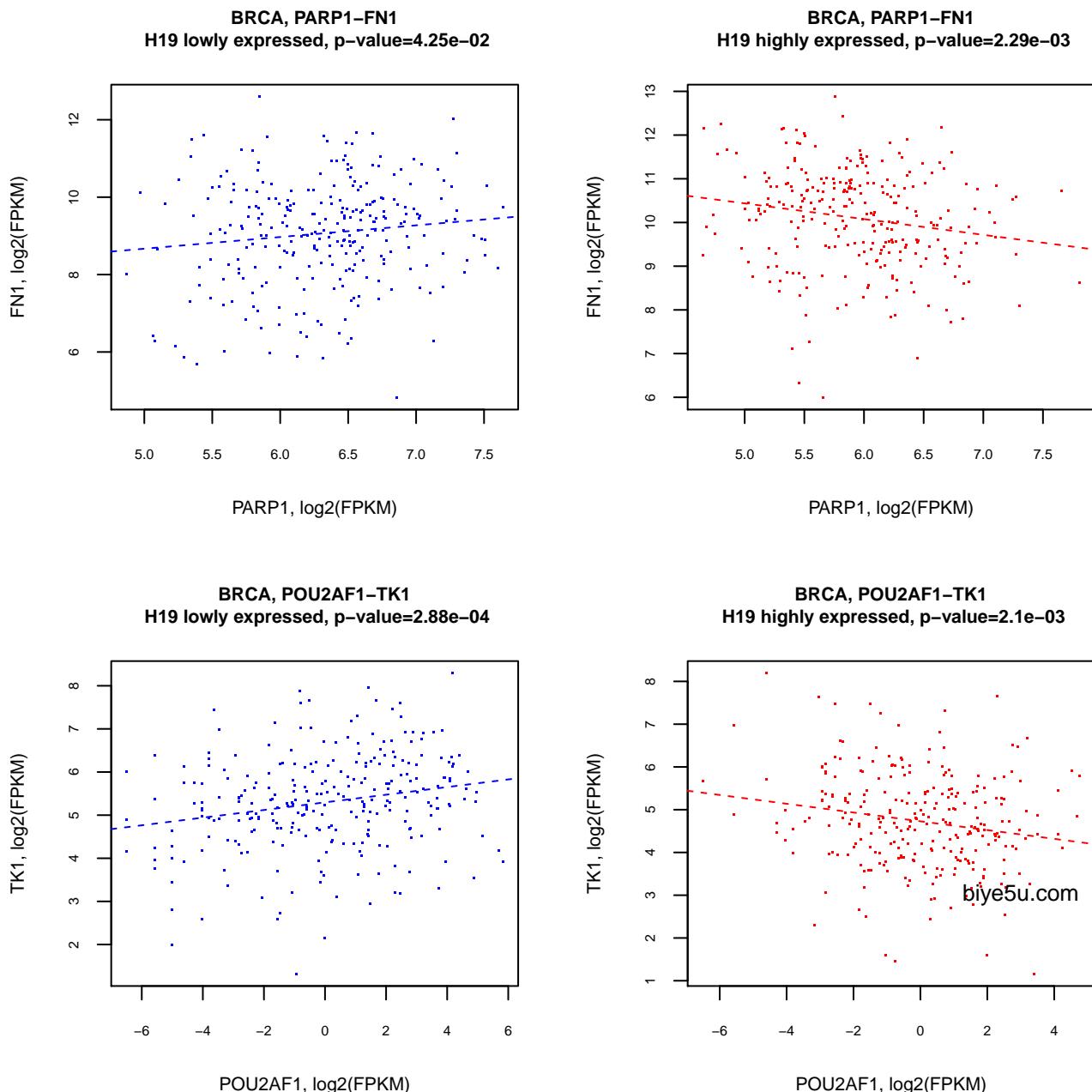


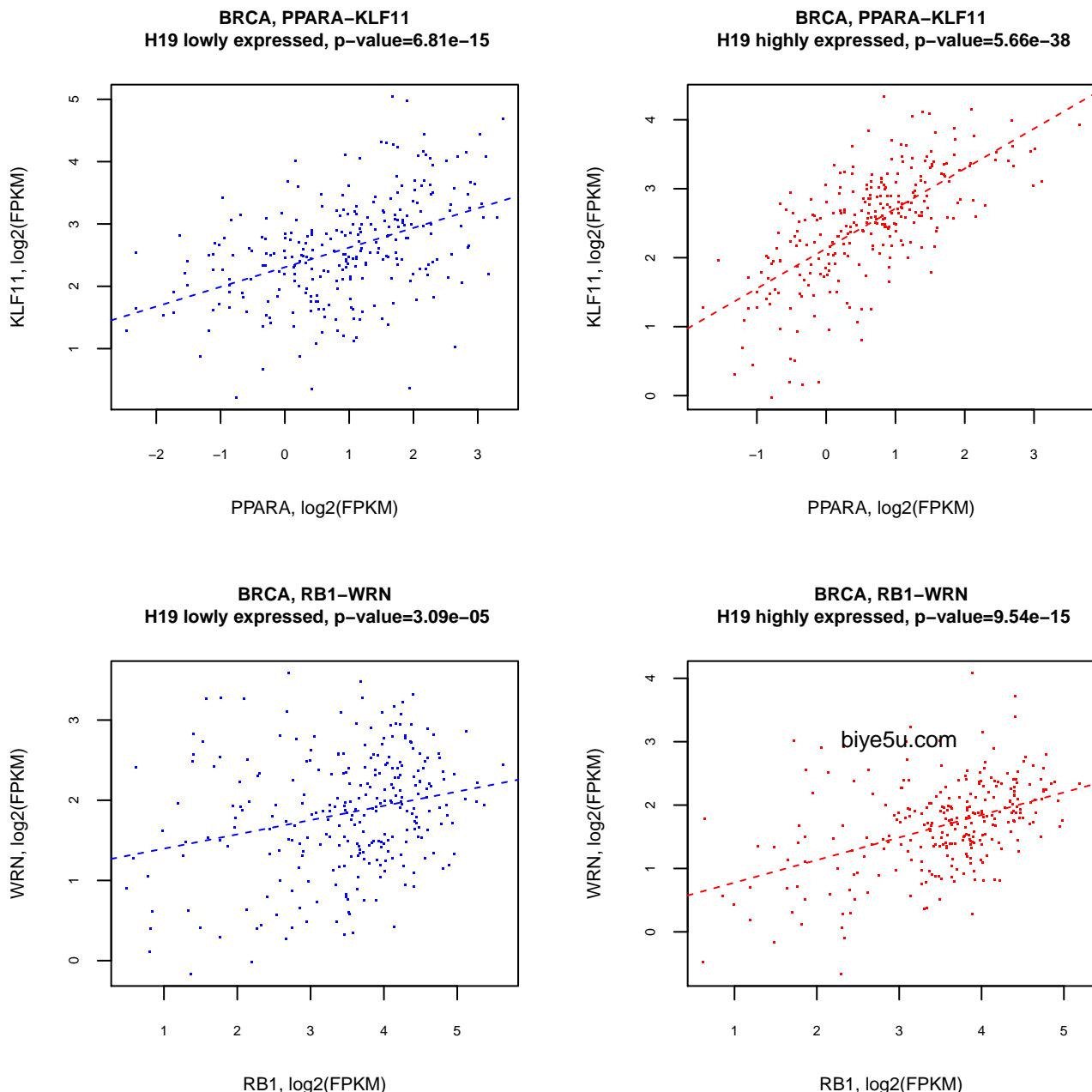


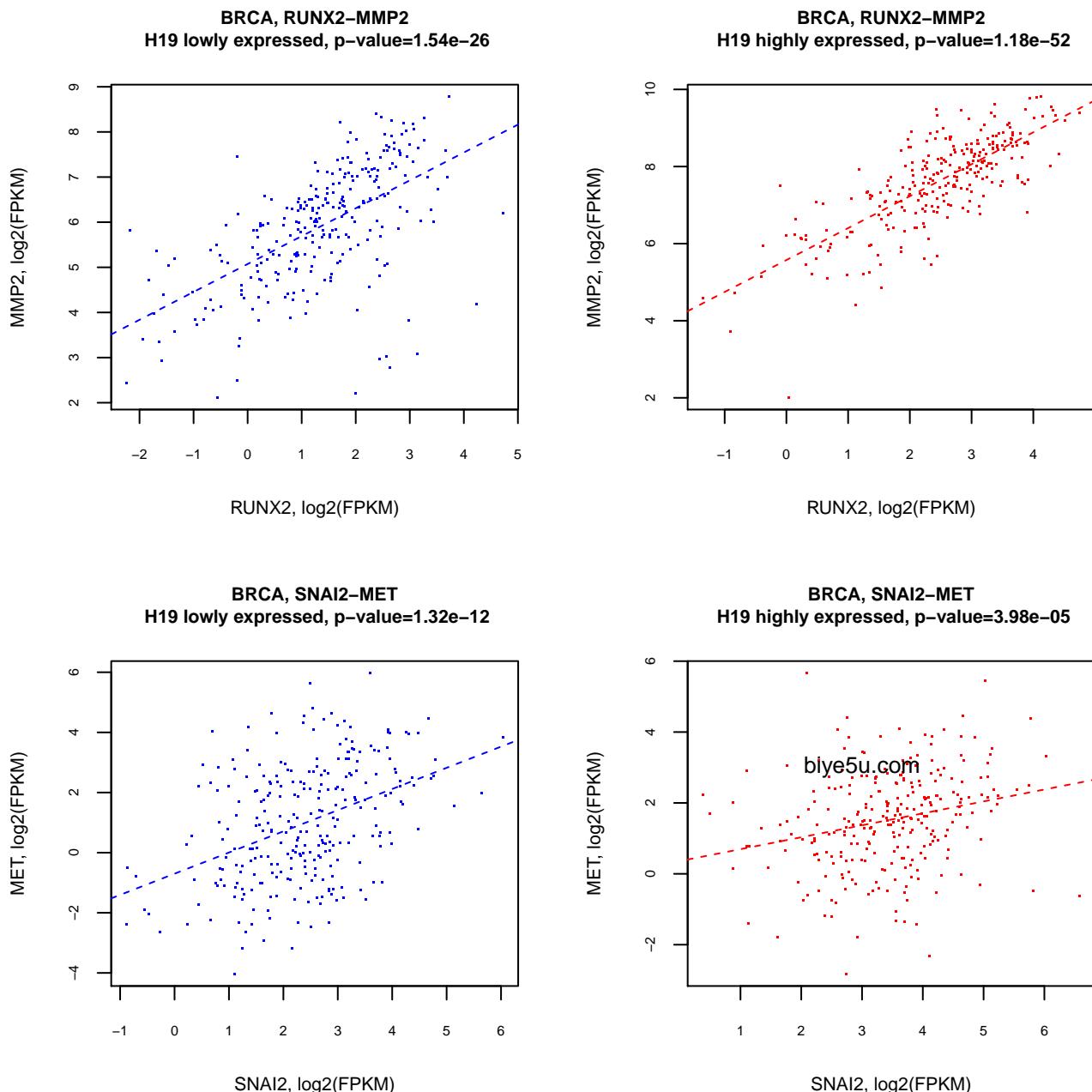


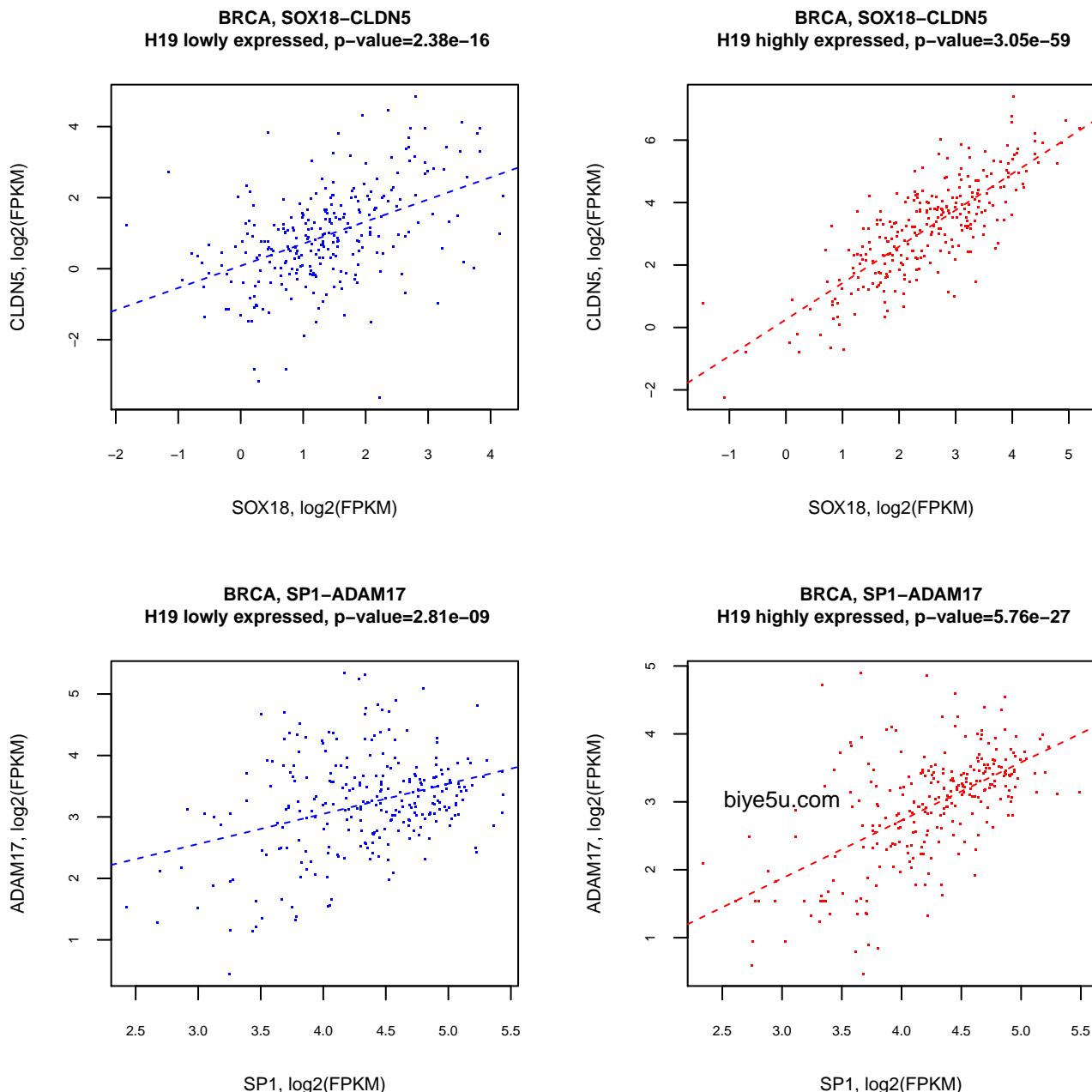


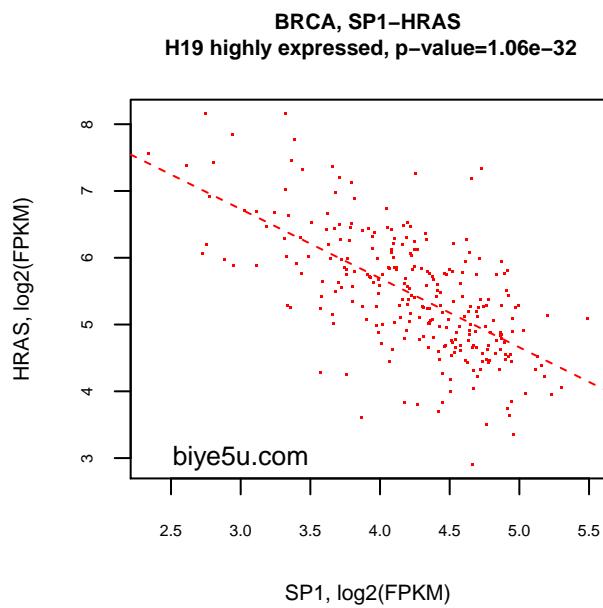
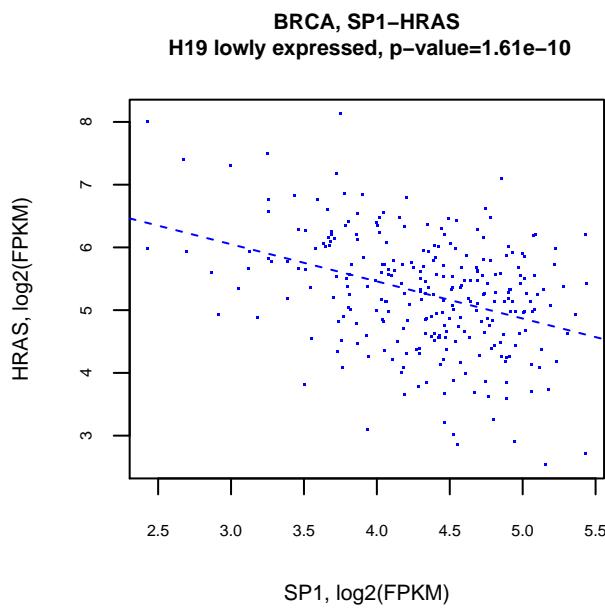
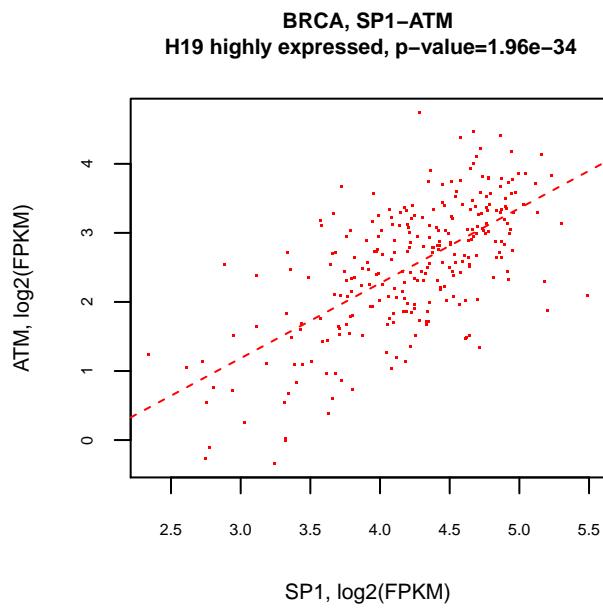
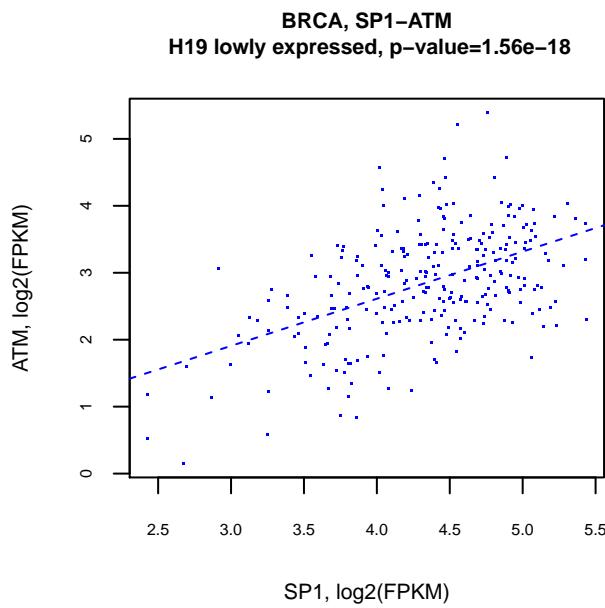


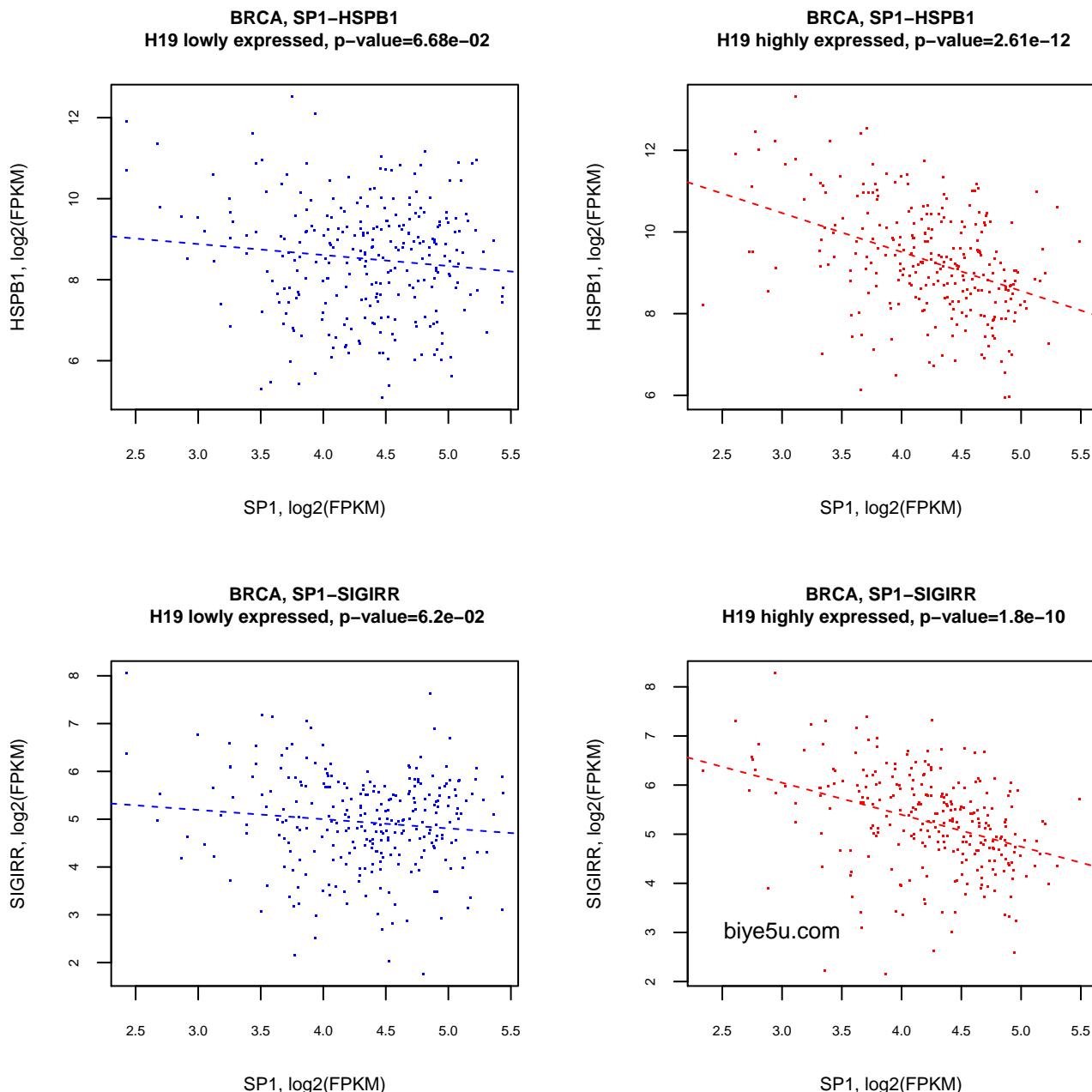


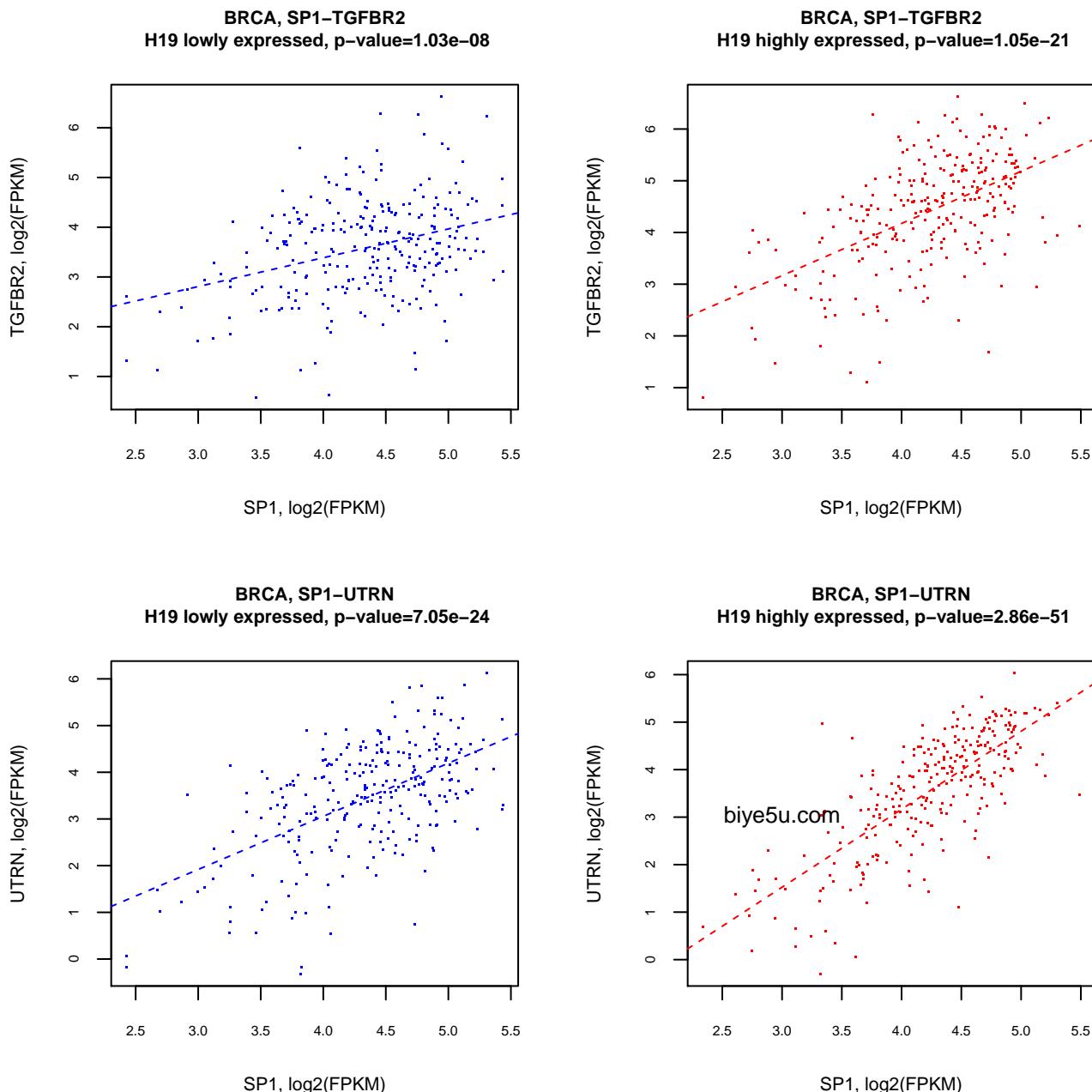


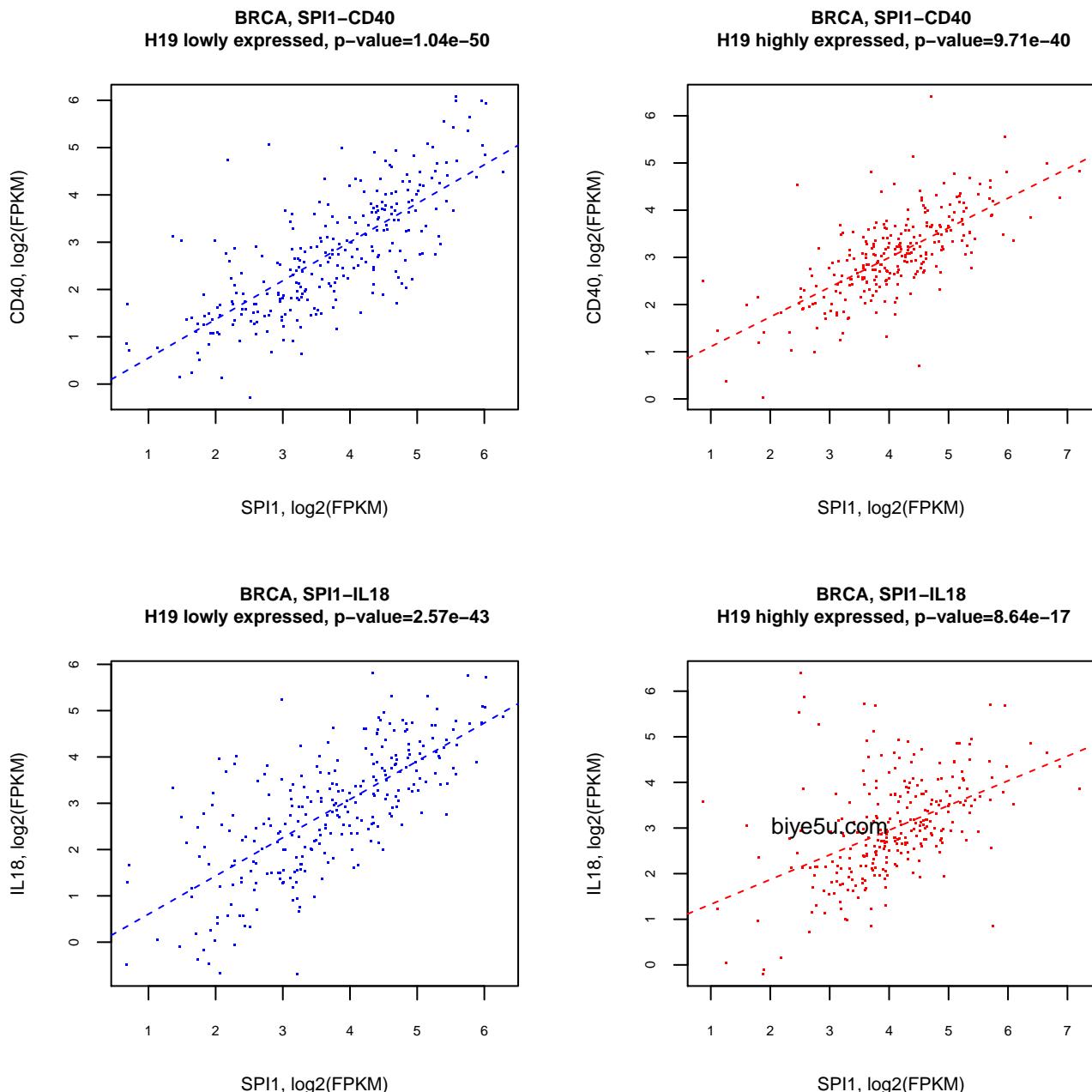


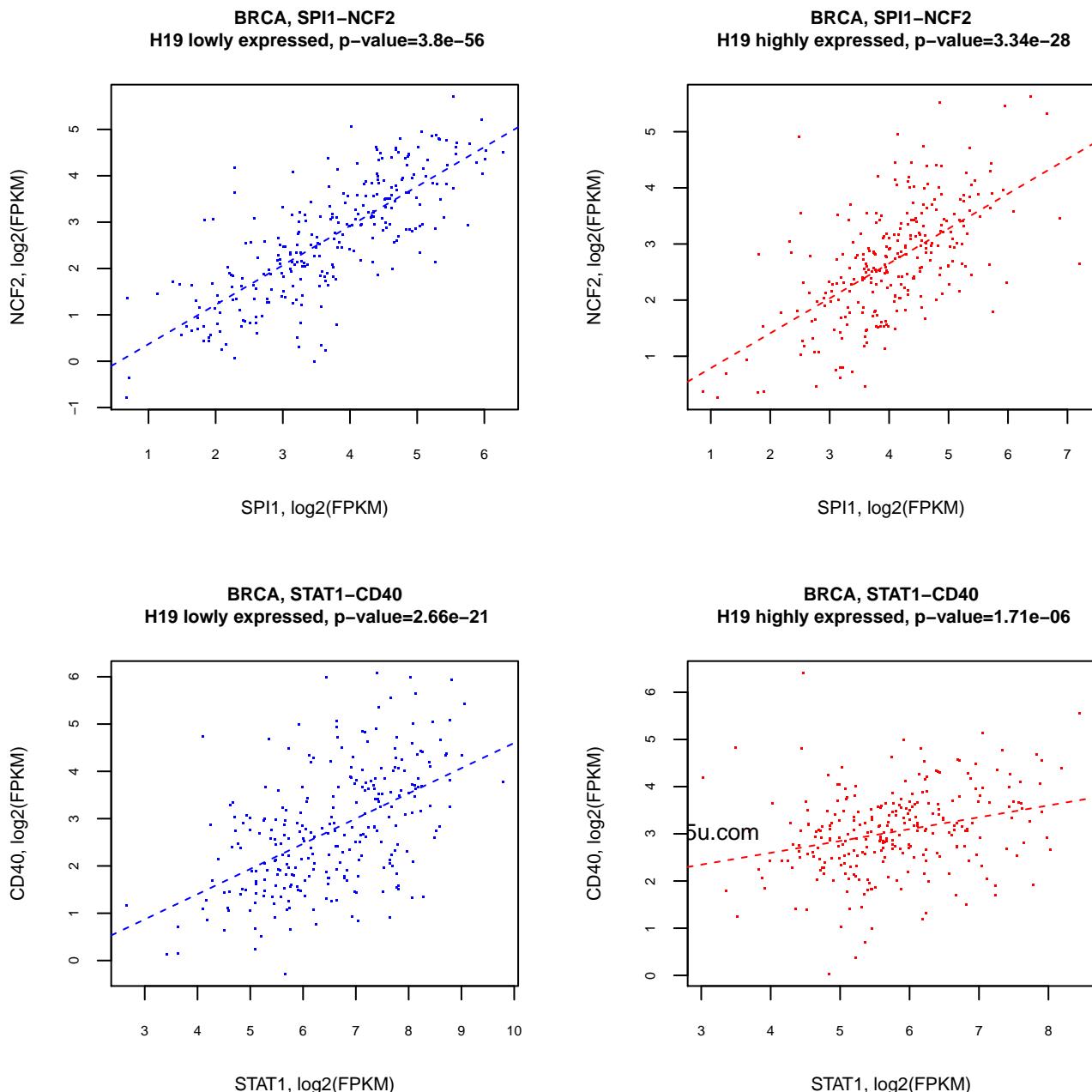


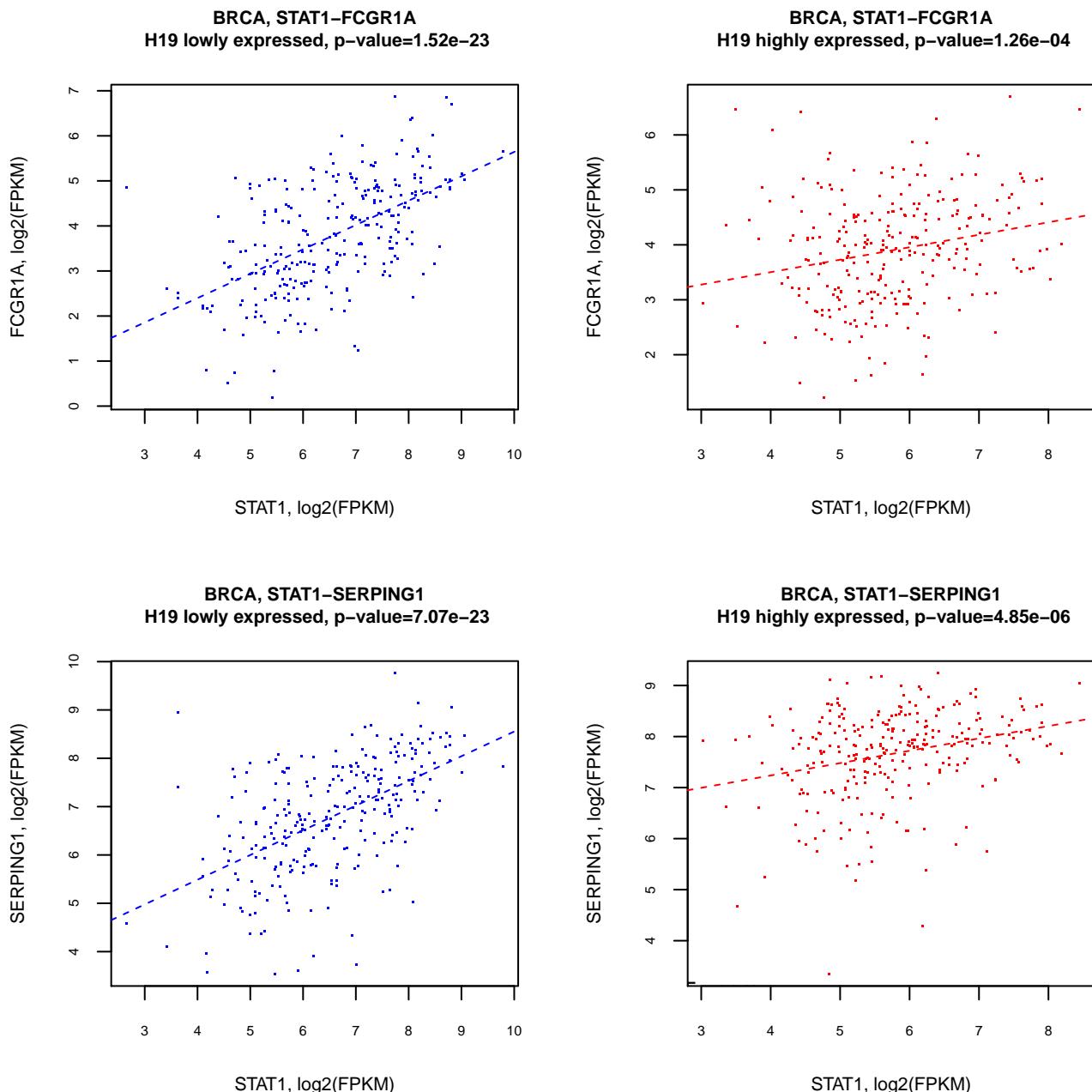


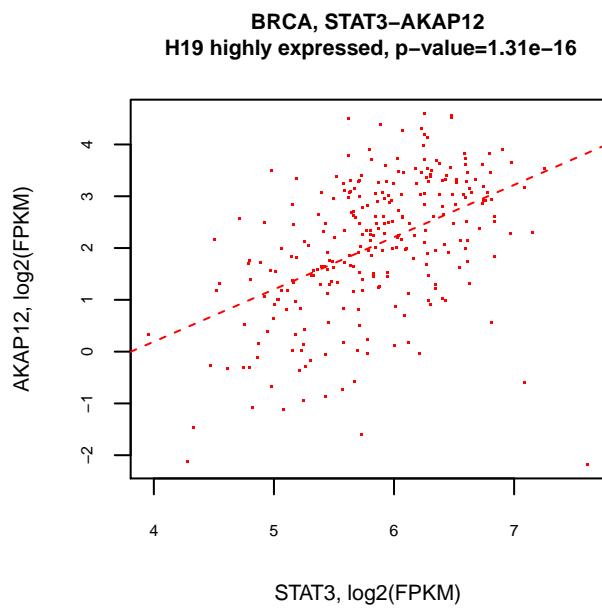
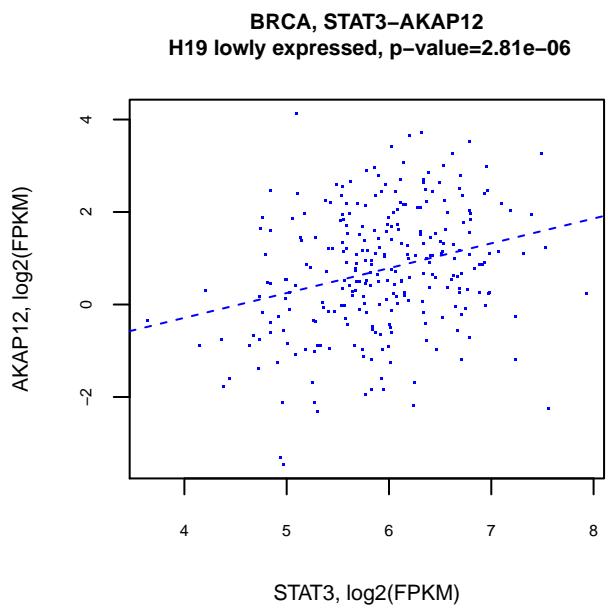
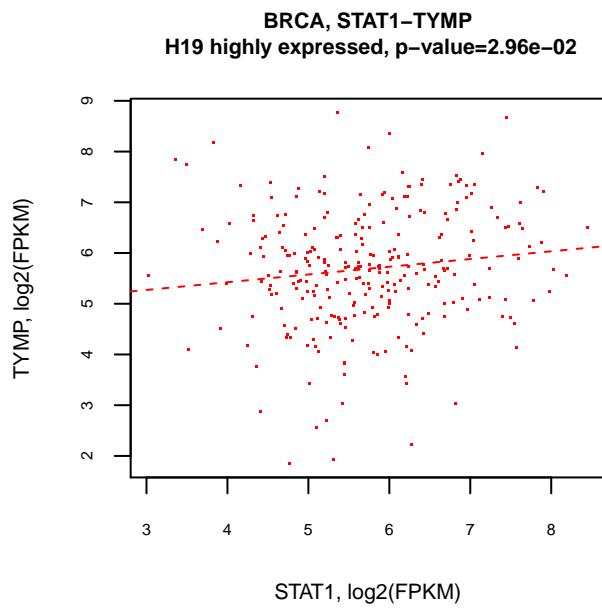
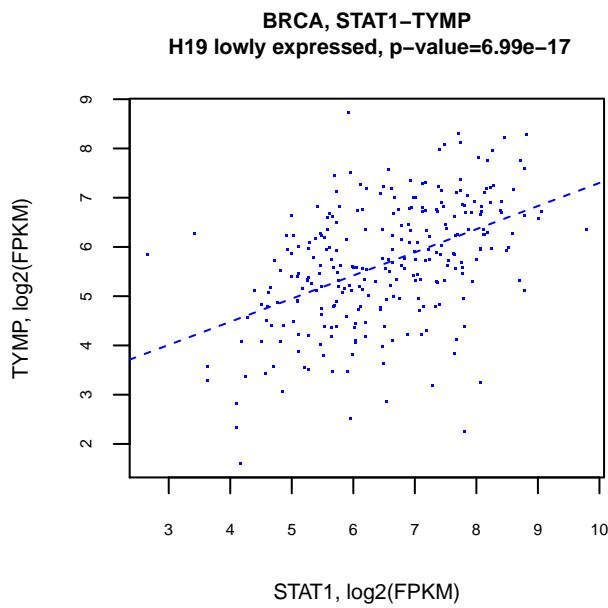


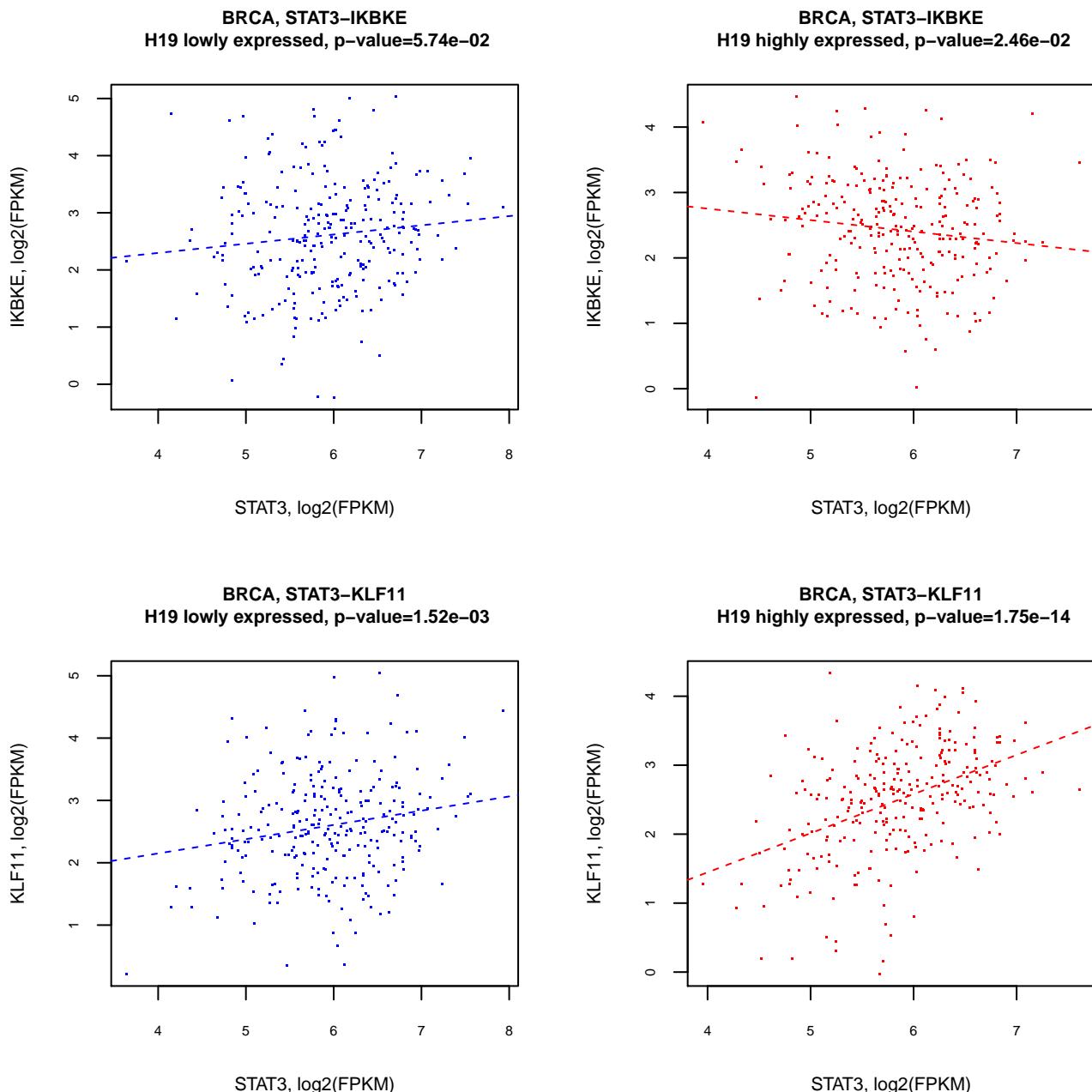












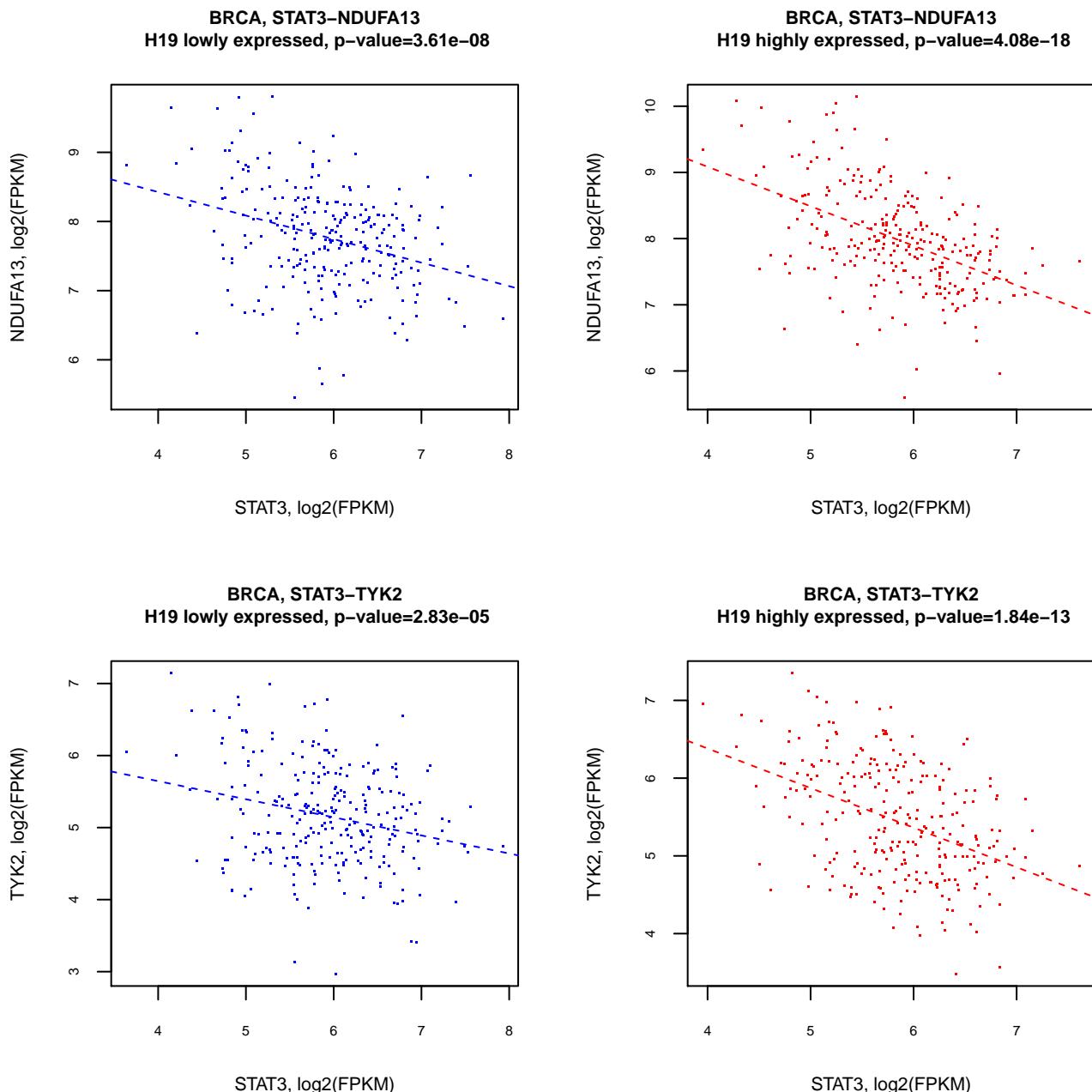


Table S1. Number of Samples and Genes

No	Cancer Type Abbreviation	Cancer Type Name	Samples				After FKPM Filtration		TF-gene Interactions		
			Total Number of Samples	# Primary Tumor Samples	# Matched Controls	# Other Samples	# Genes	Number of TFs	# Pairs (TF-gene)	# TFs	# Genes
1	BLCA	Bladder urothelial carcinoma	426	407	19	0	11481	683	7723	607	2103
2	BRCA	Breast invasive carcinoma	1212	1092	113	7	11867	694	8181	625	2198
3	CESC	Cervical squamous cell carcinoma and endocervical adenocarcinoma	309	304	3	2	11508	709	8057	634	2161
4	COAD	Colon adenocarcinoma	331	288	41	2	11537	700	8123	631	2187
5	ESCA	Esophageal carcinoma	195	181	13	1	11727	723	8277	655	2205
6	GBM	Glioblastoma multiforme	171	153	5	13	12279	717	7755	636	2156
7	HNSC	carcinoma	564	518	44	2	11406	691	7866	621	2140
8	KIRC	Kidney renal clear cell carcinoma	91	66	25	0	11881	697	7889	628	2172
9	KIRP	Kidney renal papillary cell carcinoma	321	288	32	1	11599	667	7130	591	2030
10	LGG	Brain lower grade glioma	523	509	0	14	12104	678	6837	589	1996
11	LIHC	Liver hepatocellular carcinoma	421	369	50	2	10577	622	6988	546	1964
12	LUAD	Lung adenocarcinoma	574	513	59	2	12065	695	8223	621	2233
13	LUSC	Lung squamous cell carcinoma	548	498	50	0	12002	720	8444	653	2281
14	OV	Ovarian serous cystadenocarcinoma	427	419	0	8	11838	699	8030	622	2159
15	PAAD	Pancreatic adenocarcinoma	183	178	4	1	12346	722	8758	651	2369
16	PCPG	Paraganglioma	185	177	3	5	11520	660	6438	562	1896
17	PRAD	Prostate adenocarcinoma	548	495	52	1	11785	700	7889	624	2170
18	SARC	Sarcoma	264	258	2	4	11343	690	7353	602	2053
19	SKCM	Skin cutaneous melanoma	470	102	1	367	11028	667	7055	585	1982
20	STAD	Stomach adenocarcinoma	450	414	36	0	11792	726	8460	648	2252
21	TGCT	Testicular germ cell tumors	154	148	0	6	12184	701	8042	620	2202
22	THCA	Thyroid carcinoma	571	504	59	8	11693	666	7284	587	2020
23	THYM	Thymoma	571	504	59	8	11757	680	7270	602	2055
24	UCEC	Uterine corpus endometrial carcinoma	204	180	23	1	11718	698	8092	625	2152

Table S2. Regression of H19's regulation on TF-gene pairs

Triplet	BRCA	COAD	HNSC	KIRC	LGG	LUAD	PAAD	STAD	TGCT	THCA	# of cancer types	H19 lowley expressed	H19 highly expressed
H19_AHR_HSPB1	5.05									2.98	2	→	↔
H19_AIP_RSF1	2.97									2.77	2	→	↔
H19_EZH2_DACT3	5.87							4.90			2	→	↔
H19_NFKB1_MIF	2.47					3.33					2	→	↔
H19_NFYB_EDF1	5.59									2.28	2	→	↔
H19_SP1_HRAS	4.02									4.98	2	→	↔
H19_SP1_HSPB1	3.50									2.54	2	→	↔
H19_SP1_SIGIRR	3.15									2.16	2	→	↔
H19_SP3_EDF1					5.92					4.91	2	→	↔
H19_STAT3_NDUFA13	2.54									2.93	2	→	↔
H19_STAT3_TYK2	2.57									3.18	2	→	↔
H19_AHR_CCNG2	2.99						2.38				2	→	↔↔
H19_AHR_SOS1	3.91						2.73			2.94	3	→	↔↔
H19_CREBBP_CREB1	3.13					3.94					2	→	↔↔
H19_E2F3_MAPK8	4.49									3.59	2	→	↔↔
H19_EGR1_SPRY1	3.44					3.38					2	→	↔↔
H19_ERG_EPB41L3							2.22			2.84	2	→	↔↔
H19_ETS1_TGFBR2	9.19					4.21					2	→	↔↔
H19_EZH2_BRCA1	3.70						4.87				2	→	↔↔
H19_FLI1_CTF	2.51	3.53									2	→	↔↔
H19_FLI1_TGFBR2	6.02				4.14						2	→	↔↔
H19_FOSL2_BCL6	3.02	6.40									2	→	↔↔
H19_FOXM1_CCNB1						3.83			3.39		2	→	↔↔
H19_FOXO1_TXNIP	2.93					3.12					2	→	↔↔
H19_GATA2_VWF	3.20						8.26				2	→	↔↔
H19_HNRNPK{EIF4E}							2.92			5.33	2	→	↔↔
H19_KAT2B_SMAD4	5.05									2.19	2	→	↔↔
H19_KAT2B_ZEB1	2.66							2.78			2	→	↔↔
H19_KLF4_IL6	4.16									3.96	2	→	↔↔
H19_KLF6_TXNIP	3.26						2.66				2	→	↔↔
H19_NFKB1_CHUK	3.60									5.56	2	→	↔↔

Triplet	BRCA	COAD	HNSC	KIRC	LGG	LUAD	PAAD	STAD	TGCT	THCA	# of cancer types	H19 lowley expressed	H19 highly expressed
H19_NFYB_HMGCS1	2.37									3.79	2	→	→→
H19_NFYB_SP3	7.48									2.79	2	→	→→
H19_NR3C1_CALD1	5.86									2.72	2	→	→→
H19_PPARA_KLF11	6.41									3.46	2	→	→→
H19_RB1_WRN	2.40									2.86	2	→	→→
H19_RUNX2_MMP2	3.20									2.81	2	→	→→
H19_SOX18_CLDN5	9.50					3.07					2	→	→→
H19_SP1_ADAM17	3.71									5.66	2	→	→→
H19_SP1_ATM	3.51									5.70	2	→	→→
H19_SP1_TGFBR2	2.91									4.08	2	→	→→
H19_SP1_UTRN	4.22									3.01	2	→	→→
H19_STAT3_AKAP12	2.67									3.59	2	→	→→
H19_STAT3_KLF11	3.53									3.41	2	→	→→
H19_TWIST2_SRPX						3.97	3.33				2	→	→→
H19_AHR_CYP1B1									2.45	7.59	2	→→	→
H19_AKNA_CD40	2.68					3.30					2	→→	→
H19_CIITA_HLA-DRA	3.73								3.68		2	→→	→
H19_IRF8_CD68						4.43			5.10		2	→→	→
H19_SNAI2_MET	2.58		5.20								2	→→	→
H19_SPI1_ACP5							2.42			4.55	2	→→	→
H19_SPI1_CD40	3.07							3.39			2	→→	→
H19_SPI1_IL18	3.75									3.14	2	→→	→
H19_SPI1_NCF2	3.67						2.63				2	→→	→
H19_STAT1_CD40	4.37									3.12	2	→→	→
H19_STAT1_FCGR1A	4.69									2.73	2	→→	→
H19_STAT1_SERPING1	4.06									4.74	2	→→	→
H19_STAT1_TYMP	3.88					3.72					2	→→	→
H19_GATA2_VCAM1	2.95		6.16								2	⊣	→
H19_NFKB1_NCAM1								3.51			2	⊣	→
H19_SP1_ME1							4.42			3.86	2	⊣	→
H19_TP53_IGFBP3						4.07				5.08	2	⊣	→
H19_CTCF_IPO13							3.15			3.41	2	⊣	→
H19_E2F1_GADD45B	5.75	3.86								3.26	2	→	⊣

Triplet	BRCA	COAD	HNSC	KIRC	LGG	LUAD	PAAD	STAD	TGCT	THCA	# of cancer types	H19 lowley expressed	H19 highly expressed
H19_E2F1_MYC						3.64			6.27		2	→	¬
H19_EZH2_CIITA						3.52	4.20				2	→	¬
H19_EZH2_SNAI2	6.04	4.55									2	→	¬
H19_FOXO1_HYOU1						2.64			9.10		2	→	¬
H19_HDAC1_TXNIP	4.88					4.11					2	→	¬
H19_HDAC2_TWIST1	2.77	3.76									2	→	¬
H19_HDGF_FAS	6.14						7.27				2	→	¬
H19_IKZF1_BIRC5	4.72						3.70				2	→	¬
H19_MYB_COL1A1		4.33						4.66			2	→	¬
H19_MYBL2_COL1A1	11.03	4.48						6.07			3	→	¬
H19_MYC_E2F1						4.03			5.84		2	→	¬
H19_NFKB2_HIF1A						3.54	3.57				2	→	¬
H19_PARP1_FBN1	2.70								5.57		2	→	¬
H19_PARP1_FN1	3.53	3.08									2	→	¬
H19_POU2AF1_TK1	5.63						7.38				2	→	¬
H19_POU2F1_VWF						7.74		5.91			2	→	¬
H19_RELA_BGN							3.90			3.75	2	→	¬
H19_RUNX1_SYMPK	5.54					7.74			5.65		3	→	¬
H19_SOX9_CD3E							5.49				2	→	¬
H19_SP1_ABCA2						8.08			4.60		2	→	¬
H19_SP1_FLNA						5.20			5.60		2	→	¬
H19_STAT3_DNMT1						4.92			3.38		2	→	¬
H19_STAT3_IKBKE	2.59							6.50			2	→	¬
H19_USF1_FMR1						3.09			2.91		2	→	¬

Notes: The values are transformed as $-\log_{10}(p\text{-value})$. Pattern: → activation, ¬ repression.

Table S3. TF-gene pairs modulated by H19 and their evidences that related to specific cancers.

No	TF/Gene	BRCA	COAD	HNSC	KIRC	LGG	LUAD	PAAD	STAD	TGCT	THCA
21	FLI1	[68]		[72]							
	TGFBR2	[52]		[73]							
22	FOSL2	[74]	[75]								
	BCL6	[76]	[77]								
23	FOXM1	[78]				[79]				[80]	
	CCNB1	[81]				[82]				[83]	
24	FOXO1	[84]					[85]			[86]	
	HYOU1	[87]					[88]			?	
25	FOXO1	[84]			[89]						
	TXNIP	[90]			[91]						
26	GATA2	[92]			[93]						
	VCAM1	[94]			[93]						
27	GATA2	[92]			[93]						
	VWF	[95]			[96]						
28	HDAC1	[97]				[98]					
	TXNIP	[90]				[99]					
29	HDAC2	[100]	[101]								
	TWIST1	[102]	[103]								
30	HDGF	[104]					[105]				
	FAS	[106]					[107]				
31	HNRNPK					[108]				[109]	
	EIF4E					[110]				[111]	
32	IKZF1	[112]					[113]				
	BIRC5	[114]					[115]				
33	IRF8						[116]			[117]	
	CD68						[118]			[119]	
34	KAT2B	[120]								[121]	
	SMAD4	[122]								[123]	
35	KAT2B	[120]					[124]				
	ZEB1	[125]					[126]				
36	KLF4	[127]								[128]	
	IL6	[129]								[130]	
37	KLF6	[131]				[132]					
	TXNIP	[90]				[99]					
38	MYB		[133]					[134]			
	COL1A1		[135]					[136]			
39	MYBL2	[137]	[35]					[138]			
	COL1A1	[139]	[135]					[136]			
40	MYC						[38]			[39]	
	E2F1						[36]			[37]	

No	TF/Gene	BRCA	COAD	HNSC	KIRC	LGG	LUAD	PAAD	STAD	TGCT	THCA
41	NFKB1	[140]								[141]	
	CHUK	[142]								?	
42	NFKB1	[143]					[144]				
	MIF						[145]				
43	NFKB1						[144]			[141]	
	NCAM1						[146]			[147]	
44	NFKB2					[148]	[149]				
	HIF1A					[150]	[151]				
45	NFYB	[152]								[153]	
	EDF1	[154]								?	
46	NFYB	[152]								[153]	
	HMGCS1	[155]								[156]	
47	NFYB	[152]								[153]	
	SP3	[157]								[158]	
48	NR3C1	[159]								[160]	
	CALD1	[161]								[162]	
49	PARP1	[163]							[164]		
	FBN1	[161]							[165]		
50	PARP1	[163]	[166]								
	FN1	[167]	[168]								
51	POU2AF1	[169]					[170]				
	TK1	[171]					[172]				
52	POU2F1					[173]		[174]			
	VWF					[175]		[176]			
53	PPARA	[177]								[178]	
	KLF11	[179]								[180]	
54	RB1	[181]								[182]	
	WRN	[183]								[184]	
55	RELA						[185]			[186]	
	BGN						[187]			?	
56	RUNX1		[188]			[189]				[190]	
	SYMPK		[191]			[192]				?	
57	RUNX2	[193]								[194]	
	MMP2	[195]								[196]	
58	SNAI2	[66]		[197]							
	MET	[198]		[199]							
59	SOX18	[200]				[201]					
	CLDN5	[202]				[203]					
60	SOX9				[204]		[205]				
	CD3E				[206]		[207]				

No	TF/Gene	BRCA	COAD	HNSC	KIRC	LGG	LUAD	PAAD	STAD	TGCT	THCA
81	STAT3					[254]				[251]	
	DNMT1					[255]				[256]	
82	STAT3	[250]					[257]				
	IKBKE	[258]					[259]				
83	STAT3	[250]							[251]		
	KLF11	[179]							[180]		
84	STAT3	[250]							[251]		
	NDUFA13	[260]							[261]		
85	STAT3	[250]							[251]		
	TYK2	[262]							[263]		
86	TP53					[264]				[265]	
	IGFBP3					[266]				[267]	
87	TWIST2					[268]	[269]				
	SRPX					[270]	[271]				
88	USF1						?			[272]	
	FMR1						[273]			[274]	

Notes: “?” means having not found related evidences. There are 13 TFs/genes which have not evidences to support their relation to specific cancers. TF: Transcription Factor, BRCA: Breast invasive carcinoma, COAD: Colon adenocarcinoma, HNSC: Head and Neck squamous cell carcinoma, KIRC: Kidney renal clear cell carcinoma, LGG: Brain Lower Grade Glioma, LUAD: Lung adenocarcinoma, PAAD: Pancreatic adenocarcinoma, STAD: Stomach adenocarcinoma, TGCT: Testicular Germ Cell Tumors, THCA: Thyroid carcinoma.

References

1. D'Amato NC, Rogers TJ, Gordon MA, Greene LI, Cochrane DR, Spoelstra NS, Nemkov TG, D'Alessandro A, Hansen KC, Richer JK: **A TDO2-AhR signaling axis facilitates anoikis resistance and metastasis in triple-negative breast cancer.** *Cancer research* 2015, **75**(21):4651-4664.
2. Koliopanos A, Kleeff J, Xiao Y, Safe S, Zimmermann A, Büchler MW, Friess H: **Increased arylhydrocarbon receptor expression offers a potential therapeutic target for pancreatic cancer.** *Oncogene* 2002, **21**(39):6059.
3. Li XJ, Ren ZJ, Tang JH, Yu Q: **Exosomal microRNA miR-1246 promotes cell proliferation, invasion and drug resistance by targeting CCNG2 in breast cancer.** *Cellular Physiology and Biochemistry* 2017, **44**(5):1741-1748.
4. Hasegawa S, Eguchi H, Nagano H, Konno M, Tomimaru Y, Wada H, Hama N, Kawamoto K, Kobayashi S, Nishida N: **MicroRNA-1246 expression associated with CCNG2-mediated chemoresistance and stemness in pancreatic cancer.** *British journal of cancer* 2014, **111**(8):1572.
5. Brokken LJ, Lundberg-Giwerzman Y, Meyts R-D, Eberhard J, Stahl O, Cohn-Cedermark G, Daugaard G, Arver S, Giwerzman A: **Association between polymorphisms in the aryl hydrocarbon receptor repressor gene and disseminated testicular germ cell cancer.** *Frontiers in endocrinology* 2013, **4**:4.
6. Haymart MR, Banerjee M, Yin H, Worden F, Griggs JJ: **Marginal treatment benefit in**

- anaplastic thyroid cancer.** *Cancer* 2013, **119**(17):3133-3139.
- 7. Richiardi L, Pettersson A, Akre O: **Genetic and environmental risk factors for testicular cancer.** *International journal of andrology* 2007, **30**(4):230-241.
 - 8. Poljaková J, Eckschlager T, Kizek R, Frei E, Stiborová M: **Electrochemical determination of enzymes metabolizing ellipticine in thyroid cancer cells—A tool to explain the mechanism of ellipticine toxicity to these cells.** *Int J Electrochim Sci* 2013, **8**(2):1573-1585.
 - 9. Cayado-Gutiérrez N, Moncalero VL, Rosales EM, Berón W, Salvatierra EE, Alvarez-Olmedo D, Radrizzani M, Ciocca DR: **Downregulation of Hsp27 (HSPB1) in MCF-7 human breast cancer cells induces upregulation of PTEN.** *Cell Stress and Chaperones* 2013, **18**(2):243-249.
 - 10. Li Y, Yang Q, Guan H, Shi B, Ji M, Hou P: **ZNF677 suppresses Akt phosphorylation and tumorigenesis in thyroid cancer.** *Cancer research* 2018, **78**(18):5216-5228.
 - 11. Yu Y, Nie Y, Feng Q, Qu J, Wang R, Bian L, Xia J: **Targeted covalent inhibition of Grb2–Sos1 interaction through proximity-induced conjugation in breast cancer cells.** *Molecular pharmacetics* 2017, **14**(5):1548-1557.
 - 12. Swanson KD, Winter JM, Reis M, Bentires-Alj M, Greulich H, Grewal R, Hruban RH, Yeo CJ, Yassin Y, Iartchouk O: **SOS1 mutations are rare in human malignancies: implications for Noonan Syndrome patients.** *Genes, Chromosomes and Cancer* 2008, **47**(3):253-259.
 - 13. Yoo S-K, Lee S, Kim S-j, Jee H-G, Kim B-A, Cho H, Song YS, Cho SW, Won J-K, Shin J-Y: **Comprehensive analysis of the transcriptional and mutational landscape of follicular and papillary thyroid cancers.** *PLoS genetics* 2016, **12**(8):e1006239.
 - 14. Georgitsi M, Karhu A, Winqvist R, Visakorpi T, Waltering K, Vahteristo P, Launonen V, Aaltonen L: **Mutation analysis of aryl hydrocarbon receptor interacting protein (AIP) gene in colorectal, breast, and prostate cancers.** *British journal of cancer* 2007, **96**(2):352.
 - 15. Kimmel RR, Zhao LP, Nguyen D, Lee S, Aronszajn M, Cheng C, Troshin VP, Abrosimov A, Delrow J, Tuttle RM: **Microarray comparative genomic hybridization reveals genome-wide patterns of DNA gains and losses in post-Chernobyl thyroid cancer.** *Radiation research* 2006, **166**(3):519-531.
 - 16. Mao T-L, Hsu C-Y, Yen MJ, Gilks B, Sheu JJ-C, Gabrielson E, Vang R, Cope L, Kurman RJ, Wang T-L: **Expression of Rsf-1, a chromatin-remodeling gene, in ovarian and breast carcinoma.** *Human pathology* 2006, **37**(9):1169-1175.
 - 17. Ito Y, Miyoshi E, Sasaki N, Kakudo K, Yoshida H, Tomoda C, Uruno T, Takamura Y, Miya A, Kobayashi K: **Polo-like kinase 1 overexpression is an early event in the progression of papillary carcinoma.** *British journal of cancer* 2004, **90**(2):414.
 - 18. Alvarez C, Aravena A, Tapia T, Rozenblum E, Solís L, Corvalán A, Camus M, Alvarez M, Munroe D, Maass A: **Different Array CGH profiles within hereditary breast cancer tumors associated to BRCA1 expression and overall survival.** *BMC cancer* 2016, **16**(1):219.
 - 19. Lei J, Wu Z, Jiang Z, Li J, Zong L, Chen X, Duan W, Xu Q, Zhang L, Han L: **Pancreatic carcinoma-specific immunotherapy using novel tumor specific cytotoxic T cells.** *Oncotarget* 2016, **7**(50):83601.

20. Tong AW, Papayoti MH, Netto G, Armstrong DT, Ordonez G, Lawson JM, Stone MJ: **Growth-inhibitory effects of CD40 ligand (CD154) and its endogenous expression in human breast cancer.** *Clinical cancer research* 2001, **7**(3):691-703.
21. Beatty GL, Chiorean EG, Fishman MP, Saboury B, Teitelbaum UR, Sun W, Huhn RD, Song W, Li D, Sharp LL: **CD40 agonists alter tumor stroma and show efficacy against pancreatic carcinoma in mice and humans.** *Science* 2011, **331**(6024):1612-1616.
22. Shi B, Vinyals A, Alia P, Broceño C, Chen F, Adrover M, Gelpi C, Price JE, Fabra Á: **Differential expression of MHC class II molecules in highly metastatic breast cancer cells is mediated by the regulation of the CIITA transcription: implication of CIITA in tumor and metastasis development.** *The international journal of biochemistry & cell biology* 2006, **38**(4):544-562.
23. Jo YS, Lee JC, Li S, Choi YS, Bai YS, Kim YJ, Lee IS, Rha SY, Ro Hk, Kim JM: **Significance of the expression of major histocompatibility complex class II antigen, HLA-DR and-DQ, with recurrence of papillary thyroid cancer.** *International journal of cancer* 2008, **122**(4):785-790.
24. Rangel LB, Agarwal R, Sherman-Baust CA, de Mello-Coelho V, Pizer ES, Ji H, Taub DD, Morin PJ: **Anomalous expression of the HLA-DR alpha and beta chains in ovarian and other cancers.** *Cancer biology & therapy* 2004, **3**(10):1021-1027.
25. Finn S, Smyth P, Cahill S, Streck C, O'regan E, Flavin R, Sherlock J, Howells D, Henfrey R, Cullen M: **Expression microarray analysis of papillary thyroid carcinoma and benign thyroid tissue: emphasis on the follicular variant and potential markers of malignancy.** *Virchows Archiv* 2007, **450**(3):249-260.
26. Gupta A, Patnaik MM, Naina HV: **MYST3/CREBBP rearranged acute myeloid leukemia after adjuvant chemotherapy for breast cancer. Case reports in oncological medicine** 2014, **2014**.
27. Tillinghast GW, Partee J, Albert P, Kelley JM, Burtow KH, Kelly K: **Analysis of genetic stability at the EP300 and CREBBP loci in a panel of cancer cell lines.** *Genes, Chromosomes and Cancer* 2003, **37**(2):121-131.
28. Chhabra A, Fernando H, Watkins G, Mansel RE, Jiang WG: **Expression of transcription factor CREB1 in human breast cancer and its correlation with prognosis.** *Oncology reports* 2007, **18**(4):953-958.
29. Cho JH, Hong WG, Jung Y-J, Lee J, Lee E, Hwang S-G, Um H-D, Park JK: **Γ-Ionizing radiation-induced activation of the EGFR-p38/ERK-STAT3/CREB-1-EMT pathway promotes the migration/invasion of non-small cell lung cancer cells and is inhibited by podophyllotoxin acetate.** *Tumor Biology* 2016, **37**(6):7315-7325.
30. Hong JA, Kang Y, Abdullaev Z, Flanagan PT, Pack SD, Fischette MR, Adnani MT, Loukinov DI, Vatolin S, Risinger JI: **Reciprocal binding of CTCF and BORIS to the NY-ESO-1 promoter coincides with derepression of this cancer-testis gene in lung cancer cells.** *Cancer research* 2005, **65**(17):7763-7774.
31. Wei W-J, Lu Z-W, Wang Y, Zhu Y-X, Wang Y-L, Ji Q-H: **Clinical significance of papillary thyroid cancer risk loci identified by genome-wide association studies.** *Cancer genetics* 2015, **208**(3):68-75.
32. Frietze S, Lupien M, Silver PA, Brown M: **CARM1 regulates estrogen-stimulated breast cancer growth through up-regulation of E2F1.** *Cancer research* 2008,

- 68**(1):301-306.
33. Kasahara M, Takahashi Y, Nagata T, Asai S, Eguchi T, Ishii Y, Fujii M, Ishikawa K: **Thymidylate synthase expression correlates closely with E2F1 expression in colon cancer.** *Clinical cancer research* 2000, **6**(7):2707-2711.
34. Gomis RR, Alarcón C, Nadal C, Van Poznak C, Massagué J: **C/EBP β at the core of the TGF β cytostatic response and its evasion in metastatic breast cancer cells.** *Cancer cell* 2006, **10**(3):203-214.
35. Clark-Langone KM, Sangli C, Krishnakumar J, Watson D: **Translating tumor biology into personalized treatment planning: analytical performance characteristics of the Onco type DX® Colon Cancer Assay.** *BMC cancer* 2010, **10**(1):691.
36. Rödicker F, Stiewe T, Zimmermann S, Pützer BM: **Therapeutic efficacy of E2F1 in pancreatic cancer correlates with TP73 induction.** *Cancer research* 2001, **61**(19):7052-7055.
37. Onda M, Nagai H, Yoshida A, Miyamoto S, Asaka S-i, Akaishi J, Takatsu K, Nagahama M, Ito K, Shimizu K: **Up-regulation of transcriptional factor E2F1 in papillary and anaplastic thyroid cancers.** *Journal of human genetics* 2004, **49**(6):312.
38. Sancho P, Burgos-Ramos E, Tavera A, Kheir TB, Jagust P, Schoenhals M, Barneda D, Sellers K, Campos-Olivas R, Graña O: **MYC/PGC-1 α balance determines the metabolic phenotype and plasticity of pancreatic cancer stem cells.** *Cell metabolism* 2015, **22**(4):590-605.
39. Zhu X, Zhao L, Park JW, Willingham MC, Cheng S-y: **Synergistic signaling of KRAS and thyroid hormone receptor β mutants promotes undifferentiated thyroid cancer through MYC up-regulation.** *Neoplasia* 2014, **16**(9):757-769.
40. Vimala K, Sundarraj S, Sujitha MV, Kannan S: **Curtailing overexpression of E2F3 in breast cancer using siRNA (E2F3)-based gene silencing.** *Archives of medical research* 2012, **43**(6):415-422.
41. Ziebold U, Lee EY, Bronson RT, Lees JA: **E2F3 loss has opposing effects on different pRB-deficient tumors, resulting in suppression of pituitary tumors but metastasis of medullary thyroid carcinomas.** *Molecular and cellular biology* 2003, **23**(18):6542-6552.
42. Stephens PJ, Tarpey PS, Davies H, Van Loo P, Greenman C, Wedge DC, Nik-Zainal S, Martin S, Varela I, Bignell GR: **The landscape of cancer genes and mutational processes in breast cancer.** *Nature* 2012, **486**(7403):400.
43. Bauer J, Kopp S, Schlagberger E, Grosse J, Sahana J, Riwaldt S, Wehland M, Luetzenberg R, Infanger M, Grimm D: **Proteome analysis of human follicular thyroid cancer cells exposed to the random positioning machine.** *International journal of molecular sciences* 2017, **18**(3):546.
44. Redmond K, Crawford N, Farmer H, D'costa Z, O'brien G, Buckley N, Kennedy R, Johnston P, Harkin D, Mullan P: **T-box 2 represses NDRG1 through an EGR1-dependent mechanism to drive the proliferation of breast cancer cells.** *Oncogene* 2010, **29**(22):3252.
45. Calogero A, Arcella A, De Gregorio G, Porcellini A, Mercola D, Liu C, Lombari V, Zani M, Giannini G, Gagliardi FM: **The early growth response gene EGR-1 behaves as a suppressor gene that is down-regulated independent of ARF/Mdm2 but not p53 alterations in fresh human gliomas.** *Clinical Cancer Research* 2001, **7**(9):2788-2796.

46. Fernandez S, Russo J: **Estrogen and xenoestrogens in breast cancer.** *Toxicologic pathology* 2010, **38**(1):110-122.
47. Moore LM, Zhang W: **Targeting miR-21 in glioma: a small RNA with big potential.** *Expert opinion on therapeutic targets* 2010, **14**(11):1247-1257.
48. Sarkar S, Dubaybo H, Ali S, Goncalves P, Kollepara SL, Sethi S, Philip PA, Li Y: **Down-regulation of miR-221 inhibits proliferation of pancreatic cancer cells through up-regulation of PTEN, p27kip1, p57kip2, and PUMA.** *American journal of cancer research* 2013, **3**(5):465.
49. Kim S, Park HK, Jung HY, Lee S-Y, Min K-W, Kim WY, Han HS, Kim WS, Hwang TS, Lim SD: **ERG immunohistochemistry as an endothelial marker for assessing lymphovascular invasion.** *Korean journal of pathology* 2013, **47**(4):355.
50. Ma J, Cheng L, Liu H, Zhang J, Shi Y, Zeng F, Miele L, H Sarkar F, Xia J, Wang Z: **Genistein down-regulates miR-223 expression in pancreatic cancer cells.** *Current drug targets* 2013, **14**(10):1150-1156.
51. Ciarrocchi A, Piana S, Valcavi R, Gardini G, Casali B: **Inhibitor of DNA binding-1 induces mesenchymal features and promotes invasiveness in thyroid tumour cells.** *European Journal of Cancer* 2011, **47**(6):934-945.
52. Zhang Y, Yan L-X, Wu Q-N, Du Z-M, Chen J, Liao D-Z, Huang M-Y, Hou J-H, Wu Q-L, Zeng M-S: **miR-125b is methylated and functions as a tumor suppressor by regulating the ETS1 proto-oncogene in human invasive breast cancer.** *Cancer research* 2011, **71**(10):3552-3562.
53. Kim K-R, Yoshizaki T, Miyamori H, Hasegawa K, Horikawa T, Furukawa M, Harada S, Seiki M, Sato H: **Transformation of Madin-Darby canine kidney (MDCK) epithelial cells by Epstein-Barr virus latent membrane protein 1 (LMP1) induces expression of Ets1 and invasive growth.** *Oncogene* 2000, **19**(14):1764.
54. Keklikoglou I, Koerner C, Schmidt C, Zhang J, Heckmann D, Shavinskaya A, Allgayer H, Gückel B, Fehm T, Schneeweiss A: **MicroRNA-520/373 family functions as a tumor suppressor in estrogen receptor negative breast cancer by targeting NF-κB and TGF-β signaling pathways.** *Oncogene* 2012, **31**(37):4150.
55. Lichner Z, Saleh C, Subramaniam V, Seivwright A, Prud'homme GJ, Yousef GM: **miR-17 inhibition enhances the formation of kidney cancer spheres with stem cell/tumor initiating cell properties.** *Oncotarget* 2015, **6**(8):5567.
56. Kleer CG, Cao Q, Varambally S, Shen R, Ota I, Tomlins SA, Ghosh D, Sewalt RG, Otte AP, Hayes DF: **EZH2 is a marker of aggressive breast cancer and promotes neoplastic transformation of breast epithelial cells.** *Proceedings of the National Academy of Sciences* 2003, **100**(20):11606-11611.
57. Zhang J, Chen L, Han L, Shi Z, Zhang J, Pu P, Kang C: **EZH2 is a negative prognostic factor and exhibits pro-oncogenic activity in glioblastoma.** *Cancer letters* 2015, **356**(2):929-936.
58. Miki Y, Swensen J, Shattuck-Eidens D, Futreal PA, Harshman K, Tavtigian S, Liu Q, Cochran C, Bennett LM, Ding W: **A strong candidate for the breast and ovarian cancer susceptibility gene BRCA1.** *Science* 1994, **266**(5182):66-71.
59. Rasmussen RD, Gajjar MK, Tuckova L, Jensen KE, Maya-Mendoza A, Holst CB, Møllgaard K, Rasmussen JS, Brennum J, Bartek Jr J: **BRCA1-regulated RRM2**

- expression protects glioblastoma cells from endogenous replication stress and promotes tumorigenicity.** *Nature communications* 2016, **7**:13398.
- 60. Bao B, Ali S, Banerjee S, Wang Z, Logna F, Azmi AS, Kong D, Ahmad A, Li Y, Padhye S: **Curcumin analogue CDF inhibits pancreatic tumor growth by switching on suppressor microRNAs and attenuating EZH2 expression.** *Cancer research* 2012, **72**(1):335-345.
 - 61. Carvalho J, van Grieken NC, Pereira PM, Sousa S, Tijssen M, Buffart TE, Diosdado B, Grabsch H, Santos MA, Meijer G: **Lack of microRNA-101 causes E-cadherin functional deregulation through EZH2 up-regulation in intestinal gastric cancer.** *The Journal of pathology* 2012, **228**(1):31-44.
 - 62. Xi H, Blanck G: **Interferon regulatory factor-2 point mutations in human pancreatic tumors.** *International journal of cancer* 2000, **87**(6):803-808.
 - 63. Satoh A, Toyota M, Ikeda H, Morimoto Y, Akino K, Mita H, Suzuki H, Sasaki Y, Kanaseki T, Takamura Y: **Epigenetic inactivation of class II transactivator (CIITA) is associated with the absence of interferon- γ -induced HLA-DR expression in colorectal and gastric cancer cells.** *Oncogene* 2004, **23**(55):8876.
 - 64. Ren Y, Chen Y, Liang X, Lu Y, Pan W, Yang M: **MiRNA-638 promotes autophagy and malignant phenotypes of cancer cells via directly suppressing DACT3.** *Cancer letters* 2017, **390**:126-136.
 - 65. Fussbroich B, Wagener N, Macher-Goeppinger S, Benner A, Fälth M, Sültmann H, Holzer A, Hoppe-Seyler K, Hoppe-Seyler F: **EZH2 depletion blocks the proliferation of colon cancer cells.** *PloS one* 2011, **6**(7):e21651.
 - 66. Zhang Z, Zhang B, Li W, Fu L, Fu L, Zhu Z, Dong J-T: **Epigenetic silencing of miR-203 upregulates SNAI2 and contributes to the invasiveness of malignant breast cancer cells.** *Genes & cancer* 2011, **2**(8):782-791.
 - 67. Larriba MJ, Bonilla F, Muñoz A: **The transcription factors Snail1 and Snail2 repress vitamin D receptor during colon cancer progression.** *The Journal of steroid biochemistry and molecular biology* 2010, **121**(1-2):106-109.
 - 68. Scheiber MN, Watson PM, Rumboldt T, Stanley C, Wilson RC, Findlay VJ, Anderson PE, Watson DK: **FLI1 expression is correlated with breast cancer cellular growth, migration, and invasion and altered gene expression.** *Neoplasia* 2014, **16**(10):801-813.
 - 69. Zhang J, Guo H, Zhang H, Wang H, Qian G, Fan X, Hoffman AR, Hu JF, Ge S: **Putative tumor suppressor miR-145 inhibits colon cancer cell growth by targeting oncogene friend leukemia virus integration 1 gene.** *Cancer* 2011, **117**(1):86-95.
 - 70. Pandey DP, Lappano R, Albanito L, Madeo A, Maggiolini M, Picard D: **Estrogenic GPR30 signalling induces proliferation and migration of breast cancer cells through CTGF.** *The EMBO journal* 2009, **28**(5):523-532.
 - 71. Ladwa R, Pringle H, Kumar R, West K: **Expression of CTGF and Cyr61 in colorectal cancer.** *Journal of clinical pathology* 2011, **64**(1):58-64.
 - 72. Rahman MA, Amin AR, Wang X, Zuckerman JE, Choi CHJ, Zhou B, Wang D, Nannapaneni S, Koenig L, Chen Z: **Systemic delivery of siRNA nanoparticles targeting RRM2 suppresses head and neck tumor growth.** *Journal of controlled release* 2012, **159**(3):384-392.
 - 73. Bornstein S, White R, Malkoski S, Oka M, Han G, Cleaver T, Reh D, Andersen P, Gross

- N, Olson S: **Smad4 loss in mice causes spontaneous head and neck cancer with increased genomic instability and inflammation.** *The Journal of clinical investigation* 2009, **119**(11):3408-3419.
74. He J, Mai J, Li Y, Chen L, Xu H, Zhu X, Pan Q: **miR-597 inhibits breast cancer cell proliferation, migration and invasion through FOSL2.** *Oncology reports* 2017, **37**(5):2672-2678.
75. Asting AG, Carén H, Andersson M, Lönnroth C, Lagerstedt K, Lundholm K: **COX-2 gene expression in colon cancer tissue related to regulating factors and promoter methylation status.** *BMC cancer* 2011, **11**(1):238.
76. Tran TH, Utama FE, Lin J, Yang N, Sjolund AB, Ryder A, Johnson KJ, Neilson LM, Liu C, Brill KL: **Prolactin inhibits BCL6 expression in breast cancer through a Stat5a-dependent mechanism.** *Cancer research* 2010, **70**(4):1711-1721.
77. Moos PJ, Edes K, Mullally JE, Fitzpatrick FA: **Cucumin impairs tumor suppressor p53 function in colon cancer cells.** *Carcinogenesis* 2004, **25**(9):1611-1617.
78. Kwok JM-M, Peck B, Monteiro LJ, Schwenen HD, Millour J, Coombes RC, Myatt SS, Lam EW-F: **FOXM1 confers acquired cisplatin resistance in breast cancer cells.** *Molecular cancer research* 2010, **8**(1):24-34.
79. Gong A, Huang S: **FoxM1 and Wnt/β-catenin signaling in glioma stem cells.** *Cancer research* 2012, **72**(22):5658-5662.
80. Xu X-S, Miao R-C, Wan Y, Zhang L-Q, Qu K, Liu C: **FoxM1 as a novel therapeutic target for cancer drug therapy.** *Asian Pac J Cancer Prev* 2015, **16**(1):23-29.
81. Ding K, Li W, Zou Z, Zou X, Wang C: **CCNB1 is a prognostic biomarker for ER+ breast cancer.** *Medical hypotheses* 2014, **83**(3):359-364.
82. Ajeawung NF, Faure R, Jones C, Kamnasaran D: **Preclinical evaluation of dipotassium bisperoxo (picolinato) oxovanadate V for the treatment of pediatric low-grade gliomas.** *Future Oncology* 2013, **9**(8):1215-1229.
83. Song R, Yao X, Shi L, Ren Y, Zhao H: **Effects of dietary selenium on apoptosis of germ cells in the testis during spermatogenesis in roosters.** *Theriogenology* 2015, **84**(4):583-588.
84. Li J, Yang L, Song L, Xiong H, Wang L, Yan X, Yuan J, Wu J, Li M: **Astrocyte elevated gene-1 is a proliferation promoter in breast cancer via suppressing transcriptional factor FOXO1.** *Oncogene* 2009, **28**(36):3188.
85. Li Z-C, Zhang L-M, Wang H-B, Ma J-X, Sun J-Z: **RETRACTED ARTICLE: Curcumin inhibits lung cancer progression and metastasis through induction of FOXO1.** *Tumor Biology* 2014, **35**(1):111-116.
86. Jørgensen A, Jensen MB, Nielsen JE, Juul A, Rajpert-De Meyts E: **Influence of vitamin D on cisplatin sensitivity in testicular germ cell cancer-derived cell lines and in a NTera2 xenograft model.** *The Journal of steroid biochemistry and molecular biology* 2013, **136**:238-246.
87. Stojadinovic A, Hooke JA, Shriver CD, Nissan A, Kovatich AJ, Kao T-C, Ponniah S, Peoples GE, Moroni M: **HYOU1/Orp150 expression in breast cancer.** *Medical Science Monitor* 2007, **13**(11):BR231-BR239.
88. Yoshida Y, Yamashita T, Nagano K, Imai S, Nabeshi H, Yoshikawa T, Yoshioka Y, Abe Y, Kamada H, Tsutsumi Y: **Limited expression of reticulocalbin-1 in lymphatic**

- endothelial cells in lung tumor but not in normal lung.** *Biochemical and biophysical research communications* 2011, **405**(4):610-614.
89. Zhou L, Yin B, Liu Y, Hong Y, Zhang C, Fan J: **Mechanism and function of decreased FOXO1 in renal cell carcinoma.** *Journal of surgical oncology* 2012, **105**(8):841-847.
90. Shen L, O'Shea JM, Kaadige MR, Cunha S, Wilde BR, Cohen AL, Welm AL, Ayer DE: **Metabolic reprogramming in triple-negative breast cancer through Myc suppression of TXNIP.** *Proceedings of the National Academy of Sciences* 2015, **112**(17):5425-5430.
91. Zhou J, Yu Q, Chng W-J: **TXNIP (VDUP-1, TBP-2): a major redox regulator commonly suppressed in cancer by epigenetic mechanisms.** *The international journal of biochemistry & cell biology* 2011, **43**(12):1668-1673.
92. Yan W, Cao QJ, Arenas RB, Bentley B, Shao R: **GATA3 inhibits breast cancer metastasis through the reversal of epithelial-mesenchymal transition.** *Journal of Biological Chemistry* 2010, **285**(18):14042-14051.
93. Peters I, Dubrowinskaja N, Tezval H, Kramer MW, von Klot CA, Hennenlotter J, Stenzl A, Scherer R, Kuczyk MA, Serth J: **Decreased mRNA expression of GATA1 and GATA2 is associated with tumor aggressiveness and poor outcome in clear cell renal cell carcinoma.** *Targeted oncology* 2015, **10**(2):267-275.
94. Chen Q, Zhang XH-F, Massagué J: **Macrophage binding to receptor VCAM-1 transmits survival signals in breast cancer cells that invade the lungs.** *Cancer cell* 2011, **20**(4):538-549.
95. Blann A, Gurney D, Wadley M, Bareford D, Stonelake P, Lip G: **Increased soluble P-selectin in patients with haematological and breast cancer: a comparison with fibrinogen, plasminogen activator inhibitor and von Willebrand factor.** *Blood coagulation & fibrinolysis* 2001, **12**(1):43-50.
96. Atrihi A, Mudaliar M, Zakikhani P, Lamont D, Huang JT, Bray S, Barton G, Fleming S, Nabi G: **Quantitative proteomics in resected renal cancer tissue for biomarker discovery and profiling.** *British journal of cancer* 2014, **110**(6):1622.
97. Wu M-Y, Fu J, Xiao X, Wu J, Wu R-C: **MiR-34a regulates therapy resistance by targeting HDAC1 and HDAC7 in breast cancer.** *Cancer letters* 2014, **354**(2):311-319.
98. Lin Y-C, Lin Y-C, Shih J-Y, Huang W-J, Chao S-W, Chang Y-L, Chen C-C: **DUSP1 Expression Induced by HDAC1 Inhibition Mediates Gefitinib Sensitivity in Non-Small Cell Lung Cancers.** *Clinical Cancer Research* 2015, **21**(2):428-438.
99. Li Y, Miao L-Y, Xiao Y-L, Huang M, Yu M, Meng K, Cai H-R: **Hypoxia induced high expression of thioredoxin interacting protein (TXNIP) in non-small cell lung cancer and its prognostic effect.** *Asian Pac J Cancer Prev* 2015, **16**(7):2953-2958.
100. Müller BM, Jana L, Kasajima A, Lehmann A, Prinzler J, Budczies J, Winzer K-J, Dietel M, Weichert W, Denkert C: **Differential expression of histone deacetylases HDAC1, 2 and 3 in human breast cancer-overexpression of HDAC2 and HDAC3 is associated with clinicopathological indicators of disease progression.** *BMC cancer* 2013, **13**(1):215.
101. Zhu P, Martin E, Mengwasser J, Schlag P, Janssen K-P, Göttlicher M: **Induction of HDAC2 expression upon loss of APC in colorectal tumorigenesis.** *Cancer cell* 2004, **5**(5):455-463.
102. Hong J, Zhou J, Fu J, He T, Qin J, Wang L, Liao L, Xu J: **Phosphorylation of serine 68**

- of Twist1 by MAPKs stabilizes Twist1 protein and promotes breast cancer cell invasiveness.** *Cancer research* 2011, **71**(11):3980-3990.
- 103. Gomez I, Peña C, Herrera M, Muñoz C, Larriba MJ, Garcia V, Dominguez G, Silva J, Rodriguez R, de Herreros AG: **TWIST1 is expressed in colorectal carcinomas and predicts patient survival.** *PloS one* 2011, **6**(3):e18023.
 - 104. Chen X, Yun J, Fei F, Yi J, Tian R, Li S, Gan X: **Prognostic value of nuclear hepatoma-derived growth factor (HDGF) localization in patients with breast cancer.** *Pathology-Research and Practice* 2012, **208**(8):437-443.
 - 105. Guo H, Li W, Zheng T, Liu Z: **MiR-195 targets HDGF to inhibit proliferation and invasion of NSCLC cells.** *Tumor Biology* 2014, **35**(9):8861-8866.
 - 106. Wang YY, Kuhajda FP, Li JN, Pizer ES, Han WF, Sokoll LJ, Chan DW: **Fatty acid synthase (FAS) expression in human breast cancer cell culture supernatants and in breast cancer patients.** *Cancer letters* 2001, **167**(1):99-104.
 - 107. Kornmann M, Ishiwata T, Kleeff J, Beger HG, Korc M: **Fas and Fas-ligand expression in human pancreatic cancer.** *Annals of surgery* 2000, **231**(3):368.
 - 108. Pino I, Pio R, Toledo G, Zabalegui N, Vicent S, Rey N, Lozano MD, Torre W, García-Foncillas J, Montuenga LM: **Altered patterns of expression of members of the heterogeneous nuclear ribonucleoprotein (hnRNP) family in lung cancer.** *Lung cancer* 2003, **41**(2):131-143.
 - 109. Chaker S, Kashat L, Voisin S, Kaur J, Kak I, MacMillan C, Ozcelik H, Michael Siu K, Ralhan R, Walfish PG: **Secretome proteins as candidate biomarkers for aggressive thyroid carcinomas.** *Proteomics* 2013, **13**(5):771-787.
 - 110. Yoshizawa A, Fukuoka J, Shimizu S, Shilo K, Franks TJ, Hewitt SM, Fujii T, Cordon-Cardo C, Jen J, Travis WD: **Overexpression of phospho-eIF4E is associated with survival through AKT pathway in non-small cell lung cancer.** *Clinical Cancer Research* 2010, **16**(1):240-248.
 - 111. Manfredi GI, Dicitore A, Gaudenzi G, Caraglia M, Persani L, Vitale G: **PI3K/Akt/mTOR signaling in medullary thyroid cancer: a promising molecular target for cancer therapy.** *Endocrine* 2015, **48**(2):363-370.
 - 112. Heyn H, Carmona FJ, Gomez A, Ferreira HJ, Bell JT, Sayols S, Ward K, Stefansson OA, Moran S, Sandoval J: **DNA methylation profiling in breast cancer discordant identical twins identifies DOK7 as novel epigenetic biomarker.** *Carcinogenesis* 2012, **34**(1):102-108.
 - 113. Herreros-Villanueva M, Bujanda L: **Non-invasive biomarkers in pancreatic cancer diagnosis: what we need versus what we have.** *Annals of translational medicine* 2016, **4**(7).
 - 114. Wang C, Zheng X, Shen C, Shi Y: **MicroRNA-203 suppresses cell proliferation and migration by targeting BIRC5 and LASP1 in human triple-negative breast cancer cells.** *Journal of experimental & clinical cancer research* 2012, **31**(1):58.
 - 115. Glienke W, Maute L, Wicht J, Bergmann L: **Curcumin inhibits constitutive STAT3 phosphorylation in human pancreatic cancer cell lines and downregulation of survivin/BIRC5 gene expression.** *Cancer investigation* 2009, **28**(2):166-171.
 - 116. Meyer MA, Baer JM, Knolhoff BL, Nywening TM, Panni RZ, Su X, Weilbaecher KN, Hawkins WG, Ma C, Fields RC: **Breast and pancreatic cancer interrupt IRF8-**

- dependent dendritic cell development to overcome immune surveillance.** *Nature communications* 2018, **9**(1):1250.
- 117. Melillo RM, Castellone MD, Guarino V, De Falco V, Cirafici AM, Salvatore G, Caiazzo F, Basolo F, Giannini R, Kruhoffer M: **The RET/PTC-RAS-BRAF linear signaling cascade mediates the motile and mitogenic phenotype of thyroid cancer cells.** *The Journal of clinical investigation* 2005, **115**(4):1068-1081.
 - 118. Dallal RM, Christakos P, Lee K, Egawa S, Son Y-I, Lotze MT: **Paucity of dendritic cells in pancreatic cancer.** *Surgery* 2002, **131**(2):135-138.
 - 119. Herrmann G, Schumm-Draeger P-M, Müller C, Atai E, Wenzel B, Fabian T, Usadel KH, Hübner K: **T lymphocytes, CD68-positive cells and vascularisation in thyroid carcinomas.** *Journal of cancer research and clinical oncology* 1994, **120**(11):651-656.
 - 120. Zhang G, Zhang W, Li B, Stringer-Reasor E, Chu C, Sun L, Bae S, Chen D, Wei S, Jiao K: **MicroRNA-200c and microRNA-141 are regulated by a FOXP3-KAT2B axis and associated with tumor metastasis in breast cancer.** *Breast Cancer Research* 2017, **19**(1):73.
 - 121. Zhao Y, Liu X, Zhong L, He M, Chen S, Wang T, Ma S: **The combined use of miRNAs and mRNAs as biomarkers for the diagnosis of papillary thyroid carcinoma.** *International journal of molecular medicine* 2015, **36**(4):1097-1103.
 - 122. Deckers M, van Dinther M, Buijs J, Que I, Löwik C, van der Pluijm G, ten Dijke P: **The tumor suppressor Smad4 is required for transforming growth factor β-induced epithelial to mesenchymal transition and bone metastasis of breast cancer cells.** *Cancer research* 2006, **66**(4):2202-2209.
 - 123. Geraldo M, Yamashita A, Kimura E: **MicroRNA miR-146b-5p regulates signal transduction of TGF-β by repressing SMAD4 in thyroid cancer.** *Oncogene* 2012, **31**(15):1910.
 - 124. Ansari D, Andersson R, Bauden MP, Andersson B, Connolly JB, Welinder C, Sasor A, Marko-Varga G: **Protein deep sequencing applied to biobank samples from patients with pancreatic cancer.** *Journal of cancer research and clinical oncology* 2015, **141**(2):369-380.
 - 125. Chaffer CL, Marjanovic ND, Lee T, Bell G, Kleer CG, Reinhardt F, D'Alessio AC, Young RA, Weinberg RA: **Poised chromatin at the ZEB1 promoter enables breast cancer cell plasticity and enhances tumorigenicity.** *Cell* 2013, **154**(1):61-74.
 - 126. Krebs AM, Mitschke J, Losada ML, Schmalhofer O, Boerries M, Busch H, Boettcher M, Mougiakakos D, Reichardt W, Bronsert P: **The EMT-activator Zeb1 is a key factor for cell plasticity and promotes metastasis in pancreatic cancer.** *Nature cell biology* 2017, **19**(5):518.
 - 127. Yu F, Li J, Chen H, Fu J, Ray S, Huang S, Zheng H, Ai W: **Kruppel-like factor 4 (KLF4) is required for maintenance of breast cancer stem cells and for cell migration and invasion.** *Oncogene* 2011, **30**(18):2161.
 - 128. Carina V, Zito G, Pizzolanti G, Richiusa P, Criscimanna A, Rodolico V, Tomasello L, Pitrone M, Arancio W, Giordano C: **Multiple pluripotent stem cell markers in human anaplastic thyroid cancer: the putative upstream role of SOX2.** *Thyroid* 2013, **23**(7):829-837.
 - 129. Iliopoulos D, Hirsch HA, Wang G, Struhl K: **Inducible formation of breast cancer stem**

- cells and their dynamic equilibrium with non-stem cancer cells via IL6 secretion.** *Proceedings of the National Academy of Sciences* 2011, **108**(4):1397-1402.
130. Lumachi F, Basso SM, Orlando R: **Cytokines, thyroid diseases and thyroid cancer.** *Cytokine* 2010, **50**(3):229-233.
131. Guo H, Lin Y, Zhang H, Liu J, Zhang N, Li Y, Kong D, Tang Q, Ma D: **Tissue factor pathway inhibitor-2 was repressed by CpG hypermethylation through inhibition of KLF6 binding in highly invasive breast cancer cells.** *BMC molecular biology* 2007, **8**(1):110.
132. Spinola M, Leoni VP, Galvan A, Korschning E, Conti B, Pastorino U, Ravagnani F, Columbano A, Skaug V, Haugen A: **Genome-wide single nucleotide polymorphism analysis of lung cancer risk detects the KLF6 gene.** *Cancer letters* 2007, **251**(2):311-316.
133. Hugo H, Cures A, Suraweera N, Drabsch Y, Purcell D, Mantamadiotis T, Phillips W, Dobrovic A, Zupi G, Gonda TJ: **Mutations in the MYB intron I regulatory sequence increase transcription in colon cancers.** *Genes, Chromosomes and Cancer* 2006, **45**(12):1143-1154.
134. Liang J, Liu X, Xue H, Qiu B, Wei B, Sun K: **MicroRNA-103a inhibits gastric cancer cell proliferation, migration and invasion by targeting c-Myb.** *Cell proliferation* 2015, **48**(1):78-85.
135. Suhovskikh AV, Aidagulova SV, Kashuba VI, Grigorieva EV: **Proteoglycans as potential microenvironmental biomarkers for colon cancer.** *Cell and tissue research* 2015, **361**(3):833-844.
136. Li J, Ding Y, Li A: **Identification of COL1A1 and COL1A2 as candidate prognostic factors in gastric cancer.** *World journal of surgical oncology* 2016, **14**(1):297.
137. Shi H, Bevier M, Johansson R, Enquist-Olsson K, Henriksson R, Hemminki K, Lenner P, Försti A: **Prognostic impact of polymorphisms in the MYBL2 interacting genes in breast cancer.** *Breast cancer research and treatment* 2012, **131**(3):1039-1047.
138. Buffart TE, van Grieken NC, Tijssen M, Coffa J, Ylstra B, Grabsch HI, van de Velde CJ, Carvalho B, Meijer GA: **High resolution analysis of DNA copy-number aberrations of chromosomes 8, 13, and 20 in gastric cancers.** *Virchows Archiv* 2009, **455**(3):213-223.
139. Helleman J, Jansen MP, Ruigrok-Ritstier K, van Staveren IL, Look MP, Meijer-van Gelder ME, Sieuwerts AM, Klijn JG, Sleijfer S, Foekens JA: **Association of an extracellular matrix gene cluster with breast cancer prognosis and endocrine therapy response.** *Clinical cancer research* 2008, **14**(17):5555-5564.
140. Landi S, Moreno V, Gioia-Patricola L, Guino E, Navarro M, de Oca J, Capella G, Canzian F: **Association of common polymorphisms in inflammatory genes interleukin (IL) 6, IL8, tumor necrosis factor α , NFKB1, and peroxisome proliferator-activated receptor γ with colorectal cancer.** *Cancer research* 2003, **63**(13):3560-3566.
141. Wang X, Peng H, Liang Y, Sun R, Wei T, Li Z, Gong Y, Gong R, Liu F, Zhang L: **A functional insertion/deletion polymorphism in the promoter region of the NFKB1 gene increases the risk of papillary thyroid carcinoma.** *Genetic testing and molecular biomarkers* 2015, **19**(3):167-171.
142. Lerebours F, Vacher S, Andrieu C, Espie M, Marty M, Lidereau R, Bieche I: **NF-kappa B genes have a major role in inflammatory breast cancer.** *BMC cancer* 2008, **8**(1):41.

143. Curran JE, Weinstein SR, Griffiths LR: **Polymorphic variants of NFKB1 and its inhibitory protein NFKBIA, and their involvement in sporadic breast cancer.** *Cancer letters* 2002, **188**(1-2):103-107.
144. Lu Z, Li Y, Takwi A, Li B, Zhang J, Conklin DJ, Young KH, Martin R, Li Y: **miR-301a as an NF-κB activator in pancreatic cancer cells.** *The EMBO journal* 2011, **30**(1):57-67.
145. Denz A, Pilarsky C, Muth D, Rückert F, Saeger H-D, Grützmann R: **Inhibition of MIF leads to cell cycle arrest and apoptosis in pancreatic cancer cells.** *Journal of Surgical Research* 2010, **160**(1):29-34.
146. Lunardi S, Jamieson NB, Lim SY, Griffiths KL, Carvalho-Gaspar M, Al-Assar O, Yameen S, Carter RC, McKay CJ, Spoletini G: **IP-10/CXCL10 induction in human pancreatic cancer stroma influences lymphocytes recruitment and correlates with poor survival.** *Oncotarget* 2014, **5**(22):11064.
147. Cunha LL, Morari EC, Guihen ACT, Razolli D, Gerhard R, Nonogaki S, Soares FA, Vassallo J, Ward LS: **Infiltration of a mixture of different immune cells may be related to molecular profile of differentiated thyroid cancer.** *Endocrine-related cancer* 2012, **19**(3):L31-L36.
148. Medina PP, Carretero J, Ballestar E, Angulo B, Lopez-Rios F, Esteller M, Sanchez-Cespedes M: **Transcriptional targets of the chromatin-remodelling factor SMARCA4/BRG1 in lung cancer cells.** *Human molecular genetics* 2005, **14**(7):973-982.
149. McDade TP, Perugini RA, Vittimberga Jr FJ, Carrigan RC, Callery MP: **Salicylates inhibit NF-κB activation and enhance TNF-α-induced apoptosis in human pancreatic cancer cells.** *Journal of Surgical Research* 1999, **83**(1):56-61.
150. Tantai J, Hu D, Yang Y, Geng J: **Combined identification of long non-coding RNA XIST and HIF1A-AS1 in serum as an effective screening for non-small cell lung cancer.** *International journal of clinical and experimental pathology* 2015, **8**(7):7887.
151. Hoffmann A-C, Mori R, Vallbohmer D, Brabender J, Klein E, Drebber U, Baldus SE, Cooc J, Azuma M, Metzger R: **High expression of HIF1a is a predictor of clinical outcome in patients with pancreatic ductal adenocarcinomas and correlated to PDGFA, VEGF, and bFGF.** *Neoplasia* 2008, **10**(7):674-679.
152. Verjans E, Noetzel E, Bektas N, Schütz AK, Lue H, Lennartz B, Hartmann A, Dahl E, Bernhagen J: **Dual role of macrophage migration inhibitory factor (MIF) in human breast cancer.** *BMC cancer* 2009, **9**(1):230.
153. Pan Z, Li L, Fang Q, Qian Y, Zhang Y, Zhu J, Ge M, Huang P: **Integrated Bioinformatics Analysis of Master Regulators in Anaplastic Thyroid Carcinoma.** *BioMed research international* 2019, **2019**.
154. Mihály Z, Kormos M, Lánczky A, Dank M, Budczies J, Szász MA, Győrffy B: **A meta-analysis of gene expression-based biomarkers predicting outcome after tamoxifen treatment in breast cancer.** *Breast cancer research and treatment* 2013, **140**(2):219-232.
155. Sanchez-Alvarez R, Martinez-Outschoorn UE, Lin Z, Lamb R, Hulit J, Howell A, Sotgia F, Rubin E, Lisanti MP: **Ethanol exposure induces the cancer-associated fibroblast phenotype and lethal tumor metabolism: implications for breast cancer prevention.** *Cell Cycle* 2013, **12**(2):289-301.
156. Zhu W, Li C, Ai Z: **Candidate agents for papillary thyroid cancer identified by gene**

- expression analysis.** *Pathology & Oncology Research* 2013, **19**(3):597-604.
157. Walker GE, Wilson EM, Powell D, Oh Y: **Butyrate, a histone deacetylase inhibitor, activates the human IGF binding protein-3 promoter in breast cancer cells: molecular mechanism involves an Sp1/Sp3 multiprotein complex.** *Endocrinology* 2001, **142**(9):3817-3827.
158. Chintharlapalli S, Papineni S, Lee SO, Lei P, Jin UH, Sherman SI, Santarpia L, Safe S: **Inhibition of pituitary tumor-transforming gene-1 in thyroid cancer cells by drugs that decrease specificity proteins.** *Molecular carcinogenesis* 2011, **50**(9):655-667.
159. Pan D, Kocherginsky M, Conzen SD: **Activation of the glucocorticoid receptor is associated with poor prognosis in estrogen receptor-negative breast cancer.** *Cancer research* 2011, **71**(20):6360-6370.
160. Dom G, Frank S, Floor S, Kehagias P, Libert F, Hoang C, Andry G, Spinette A, Craciun L, de Saint Aubin N: **Thyroid follicular adenomas and carcinomas: molecular profiling provides evidence for a continuous evolution.** *Oncotarget* 2018, **9**(12):10343.
161. Farmer P, Bonnefoi H, Anderle P, Cameron D, Wirapati P, Becette V, André S, Piccart M, Campone M, Brain E: **A stroma-related gene signature predicts resistance to neoadjuvant chemotherapy in breast cancer.** *Nature medicine* 2009, **15**(1):68.
162. Paricharttanakul NM, Saharat K, Chokchaichamnankit D, Punyarit P, Srisomsap C, Svasti J: **Unveiling a novel biomarker panel for diagnosis and classification of well-differentiated thyroid carcinomas.** *Oncology reports* 2016, **35**(4):2286-2296.
163. O'shaughnessy J, Osborne C, Pippen J, Yoffe M, Patt D, Monaghan G, Rocha C, Ossovskaya V, Sherman B, Bradley C: **Efficacy of BSI-201, a poly (ADP-ribose) polymerase-1 (PARP1) inhibitor, in combination with gemcitabine/carboplatin (G/C) in patients with metastatic triple-negative breast cancer (TNBC): results of a randomized phase II trial.** *Journal of Clinical Oncology* 2009, **27**(18_suppl):3-3.
164. Usanova S, Piée-Staffa A, Sied U, Thomale J, Schneider A, Kaina B, Köberle B: **Cisplatin sensitivity of testis tumour cells is due to deficiency in interstrand-crosslink repair and low ERCC1-XPF expression.** *Molecular cancer* 2010, **9**(1):248.
165. Cierna Z, Mego M, Jurisica I, Machalekova K, Chovanec M, Miskovska V, Svetlovska D, Kalavska K, Rejlekova K, Kajo K: **Fibrillin-1 (FBN-1) a new marker of germ cell neoplasia in situ.** *BMC cancer* 2016, **16**(1):597.
166. Watson JL, Hill R, Yaffe PB, Greenshields A, Walsh M, Lee PW, Giacomantonio CA, Hoskin DW: **Curcumin causes superoxide anion production and p53-independent apoptosis in human colon cancer cells.** *Cancer letters* 2010, **297**(1):1-8.
167. Vecchi M, Confalonieri S, Nuciforo P, Vigano M, Capra M, Bianchi M, Nicosia D, Bianchi F, Galimberti V, Viale G: **Breast cancer metastases are molecularly distinct from their primary tumors.** *Oncogene* 2008, **27**(15):2148.
168. Gardina PJ, Clark TA, Shimada B, Staples MK, Yang Q, Veitch J, Schweitzer A, Awad T, Sugnet C, Dee S: **Alternative splicing and differential gene expression in colon cancer detected by a whole genome exon array.** *BMC genomics* 2006, **7**(1):325.
169. Yoshimura K, Takeuchi K, Nagasaki K, Ogishima S, Tanaka H, Iwase T, Akiyama F, Kuroda Y, Miki Y: **Prognostic value of matrix Gla protein in breast cancer.** *Molecular medicine reports* 2009, **2**(4):549-553.
170. Deng L, Shang Y, Guo S, Liu C, Zhou L, Sun Y, Nie Y, Fan D, Lu Y, Guo X: **Ran GTPase**

- protein promotes metastasis and invasion in pancreatic cancer by deregulating the expression of AR and CXCR4.** *Cancer biology & therapy* 2014, **15**(8):1087-1093.
171. Zhang F, Li H, Pendleton AR, Robison JG, Monson KO, Murray BK, O'Neill KL: **Thymidine kinase 1 immunoassay: a potential marker for breast cancer.** *Cancer detection and prevention* 2001, **25**(1):8-15.
172. Paproski RJ, Young JD, Cass CE: **Predicting gemcitabine transport and toxicity in human pancreatic cancer cell lines with the positron emission tomography tracer 3'-deoxy-3'-fluorothymidine.** *Biochemical pharmacology* 2010, **79**(4):587-595.
173. Wang P, Chen D, Ma H, Li Y: **LncRNA MEG3 enhances cisplatin sensitivity in non-small cell lung cancer by regulating miR-21-5p/SOX7 axis.** *Oncotargets and therapy* 2017, **10**:5137.
174. Xu SH, Huang JZ, Xu ML, Yu G, Yin XF, Chen D, Yan GR: **ACK1 promotes gastric cancer epithelial-mesenchymal transition and metastasis through AKT-POU2F1-ECD signalling.** *The Journal of pathology* 2015, **236**(2):175-185.
175. Yano T, Tanikawa S, Fujie T, Masutani M, Horie T: **Vascular endothelial growth factor expression and neovascularisation in non-small cell lung cancer.** *European Journal of Cancer* 2000, **36**(5):601-609.
176. Ikeda M, Furukawa H, Imamura H, Shimizu J, Ishida H, Masutani S, Tatsuta M, Kawasaki T, Satomi T: **Surgery for gastric cancer increases plasma levels of vascular endothelial growth factor and von Willebrand factor.** *Gastric Cancer* 2002, **5**(3):0137-0141.
177. Golembesky AK, Gammon MD, North KE, Bensen JT, Schroeder JC, Teitelbaum SL, Neugut AI, Santella RM: **Peroxisome proliferator-activated receptor-alpha (PPARA) genetic polymorphisms and breast cancer risk: a Long Island ancillary study.** *Carcinogenesis* 2008, **29**(10):1944-1949.
178. FuÈhrer D: **A nuclear receptor in thyroid malignancy: is PAX8/PPARg the Holy Grail of follicular thyroid cancer?** *cancer* 2001, **3**:4.
179. Faryna M, Konermann C, Aulmann S, Bermejo JL, Brugger M, Diederichs S, Rom J, Weichenhan D, Claus R, Rehli M: **Genome-wide methylation screen in low-grade breast cancer identifies novel epigenetically altered genes as potential biomarkers for tumor diagnosis.** *The FASEB Journal* 2012, **26**(12):4937-4950.
180. Chang C-C, Chang Y-S, Huang H-Y, Yeh K-T, Liu T-C, Chang J-G: **Determination of the mutational landscape in Taiwanese patients with papillary thyroid cancer by whole-exome sequencing.** *Human pathology* 2018, **78**:151-158.
181. Hamann U, Herbold C, Costa S, Solomayer E-F, Kaufmann M, Bastert G, Ulmer HU, Frenzel H, Komitowski D: **Allelic imbalance on chromosome 13q: evidence for the involvement of BRCA2 and RB1 in sporadic breast cancer.** *Cancer research* 1996, **56**(9):1988-1990.
182. Takahashi C, Contreras B, Iwanaga T, Takegami Y, Bakker A, Bronson RT, Noda M, Loda M, Hunt JL, Ewen ME: **Nras loss induces metastatic conversion of Rb1-deficient neuroendocrine thyroid tumor.** *Nature genetics* 2006, **38**(1):118.
183. Ding S-l, Yu J-C, Chen S-T, Hsu G-C, Shen C-Y: **Genetic variation in the premature aging gene WRN: a case-control study on breast cancer susceptibility.** *Cancer Epidemiology and Prevention Biomarkers* 2007, **16**(2):263-269.
184. Vriens MR, Suh I, Moses W, Kebebew E: **Clinical features and genetic predisposition**

- to hereditary nonmedullary thyroid cancer.** *Thyroid* 2009, **19**(12):1343-1349.
185. Pan X, Arumugam T, Yamamoto T, Levin PA, Ramachandran V, Ji B, Lopez-Berestein G, Vivas-Mejia PE, Sood AK, McConkey DJ: **Nuclear factor-κB p65/relA silencing induces apoptosis and increases gemcitabine effectiveness in a subset of pancreatic cancer cells.** *Clinical Cancer Research* 2008, **14**(24):8143-8151.
186. Pacifico F, Leonardi A: **Role of NF-κB in thyroid cancer.** *Molecular and cellular endocrinology* 2010, **321**(1):29-35.
187. Tan AC, Jimeno A, Lin SH, Wheelhouse J, Chan F, Solomon A, Rajeshkumar N, Rubio-Viqueira B, Hidalgo M: **Characterizing DNA methylation patterns in pancreatic cancer genome.** *Molecular oncology* 2009, **3**(5-6):425-438.
188. Kourkoumpetis T, Royse KE, Chen L, Ravishankar M, Ittmann M, El-Serag HB, Jiao L: **Differential expression of tight junctions and cell polarity genes in human colon cancer.** *Exploratory Research and Hypothesis in Medicine* 2018, **3**(1):14-19.
189. Rauch TA, Wang Z, Wu X, Kernstine KH, Riggs AD, Pfeifer GP: **DNA methylation biomarkers for lung cancer.** *Tumor Biology* 2012, **33**(2):287-296.
190. Zhang H-Y, Jin L, Stilling GA, Ruebel KH, Coonse K, Tanizaki Y, Raz A, Lloyd RV: **RUNX1 and RUNX2 upregulate Galectin-3 expression in human pituitary tumors.** *Endocrine* 2009, **35**(1):101-111.
191. Wang H, Schmit SL, Haiman CA, Keku TO, Kato I, Palmer JR, van den Berg D, Wilkens LR, Burnett T, Conti DV: **Novel colon cancer susceptibility variants identified from a genome-wide association study in African Americans.** *International journal of cancer* 2017, **140**(12):2728-2733.
192. Tomoshige K, Matsumoto K, Tsuchiya T, Oikawa M, Miyazaki T, Yamasaki N, Mishima H, Kinoshita A, Kubo T, Fukushima K: **Germline mutations causing familial lung cancer.** *Journal of human genetics* 2015, **60**(10):597.
193. Javed A, Barnes GL, Pratap J, Antkowiak T, Gerstenfeld LC, Van Wijnen AJ, Stein JL, Lian JB, Stein GS: **Impaired intranuclear trafficking of Runx2 (AML3/CBFA1) transcription factors in breast cancer cells inhibits osteolysis in vivo.** *Proceedings of the National Academy of Sciences* 2005, **102**(5):1454-1459.
194. Niu D-F, Kondo T, Nakazawa T, Oishi N, Kawasaki T, Mochizuki K, Yamane T, Katoh R: **Transcription factor Runx2 is a regulator of epithelial–mesenchymal transition and invasion in thyroid carcinomas.** *Laboratory investigation* 2012, **92**(8):1181.
195. Azzam HS, Arand G, Lippman ME, Thompson EW: **Association of MMP-2 activation potential with metastatic progression in human breast cancer cell lines independent of MMP-2 production.** *JNCI: Journal of the National Cancer Institute* 1993, **85**(21):1758-1764.
196. Tian X, Cong M, Zhou W, Zhu J, Liu Q: **Relationship between protein expression of VEGF-C, MMP-2 and lymph node metastasis in papillary thyroid cancer.** *Journal of International Medical Research* 2008, **36**(4):699-703.
197. Sheu JJ, Lee C, Hua C, Li C, Lai M, Lee S, Cheng J, Chen C, Chan C, Chao SC: **LRIG1 modulates aggressiveness of head and neck cancers by regulating EGFR-MAPK-SPHK1 signaling and extracellular matrix remodeling.** *Oncogene* 2014, **33**(11):1375.
198. Gunasinghe ND, Wells A, Thompson EW, Hugo HJ: **Mesenchymal–epithelial transition (MET) as a mechanism for metastatic colonisation in breast cancer.** *Cancer and*

- Metastasis Reviews* 2012, **31**(3-4):469-478.
- 199. Seiwert TY, Jagadeeswaran R, Faoro L, Janamanchi V, Nallasura V, El Dinali M, Yala S, Kanteti R, Cohen EE, Lingen MW: **The MET receptor tyrosine kinase is a potential novel therapeutic target for head and neck squamous cell carcinoma.** *Cancer research* 2009, **69**(7):3021-3031.
 - 200. Overman J, Fontaine F, Moustaqil M, Mittal D, Sierecki E, Sacilotto N, Zuegg J, Robertson AA, Holmes K, Salim AA: **Pharmacological targeting of the transcription factor SOX18 delays breast cancer in mice.** *Elife* 2017, **6**:e21221.
 - 201. Jethon A, Pula B, Olbromski M, Werynska B, Muszczynska-Bernhard B, Witkiewicz W, Dziegiej P, Podhorska-Okolow M: **Prognostic significance of SOX18 expression in non-small cell lung cancer.** *International journal of oncology* 2015, **46**(1):123-132.
 - 202. Tókés A-M, Szász AM, Juhász É, Schaff Z, Harsányi L, Molnár IA, Baranyai Z, Besznyák I, Zaránd A, Salamon F: **Expression of tight junction molecules in breast carcinomas analysed by array PCR and immunohistochemistry.** *Pathology & Oncology Research* 2012, **18**(3):593-606.
 - 203. Paschoud S, Bongiovanni M, Pache J-C, Citi S: **Claudin-1 and claudin-5 expression patterns differentiate lung squamous cell carcinomas from adenocarcinomas.** *Modern pathology* 2007, **20**(9):947.
 - 204. Li X-L, Chen X-Q, Zhang M-N, Chen N, Nie L, Xu M, Gong J, Shen P-F, Su Z-Z, Weng X: **SOX9 was involved in TKIs resistance in renal cell carcinoma via Raf/MEK/ERK signaling pathway.** *International journal of clinical and experimental pathology* 2015, **8**(4):3871.
 - 205. Sun L, Mathews LA, Cabarcas SM, Zhang X, Yang A, Zhang Y, Young MR, Klarmann KD, Keller JR, Farrar WL: **Epigenetic regulation of SOX9 by the NF-κB signaling pathway in pancreatic cancer stem cells.** *Stem cells* 2013, **31**(8):1454-1466.
 - 206. Fergelot P, Bernhard J-C, Soulet F, Kilarski WW, Léon C, Courtois N, Deminière C, Herbert JM, Antczak P, Falciani F: **The experimental renal cell carcinoma model in the chick embryo.** *Angiogenesis* 2013, **16**(1):181-194.
 - 207. Schmielau J, Nalesnik MA, Finn OJ: **Suppressed T-cell receptor ζ chain expression and cytokine production in pancreatic cancer patients.** *Clinical cancer research* 2001, **7**(3):933s-939s.
 - 208. Lin R-K, Wu C-Y, Chang J-W, Juan L-J, Hsu H-S, Chen C-Y, Lu Y-Y, Tang Y-A, Yang Y-C, Yang P-C: **Dysregulation of p53/Sp1 control leads to DNA methyltransferase-1 overexpression in lung cancer.** *Cancer research* 2010, **70**(14):5807-5817.
 - 209. Chiefari E, Brunetti A, Arturi F, Bidart J-M, Russo D, Schlumberger M, Filetti S: **Increased expression of AP2 and Sp1 transcription factors in human thyroid tumors: a role in NIS expression regulation?** *BMC cancer* 2002, **2**(1):35.
 - 210. Boonstra R, Timmer-Bosscha H, van Echten-Arends J, van der Kolk D, van Den Berg A, De Jong B, Tew K, Poppema S, De Vries E: **Mitoxantrone resistance in a small cell lung cancer cell line is associated with ABCA2 upregulation.** *British journal of cancer* 2004, **90**(12):2411.
 - 211. Kucerova L, Feketeova L, Kozovska Z, Poturnajova M, Matuskova M, Nencka R, Babal P: **In Vivo 5FU-Exposed Human Medullary Thyroid Carcinoma Cells Contain a Chemoresistant CD133+ Tumor-Initiating Cell Subset.** *Thyroid* 2014, **24**(3):520-532.

212. Krishnan V, Wang X, Safe S: **Estrogen receptor-Sp1 complexes mediate estrogen-induced cathepsin D gene expression in MCF-7 human breast cancer cells.** *Journal of Biological Chemistry* 1994, **269**(22):15912-15917.
213. Hu B, Meng X, Zhang Y, Hossain MM, Wu L, Zhang Y, Peng X, Zhang X: **Short hairpin RNA-mediated gene silencing of ADAM17 inhibits the growth of breast cancer MCF7 cells in vitro and in vivo and its mechanism of action.** *Oncol Rep* 2018, **39**(4):1640-1648.
214. Miccichè F, Da Riva L, Fabbi M, Pilotti S, Mondellini P, Ferrini S, Canevari S, Pierotti MA, Bongarzone I: **Activated leukocyte cell adhesion molecule expression and shedding in thyroid tumors.** *PLoS One* 2011, **6**(2):e17141.
215. Ahmed M, Rahman N: **ATM and breast cancer susceptibility.** *Oncogene* 2006, **25**(43):5906-5911.
216. Gu Y, Yu Y, Ai L, Shi J, Liu X, Sun H, Liu Y: **Association of the ATM gene polymorphisms with papillary thyroid cancer.** *Endocrine* 2014, **45**(3):454-461.
217. Zhao P, Ma W, Hu Z, Zang L, Tian Z, Zhang K: **Filamin A (FLNA) modulates chemosensitivity to docetaxel in triple-negative breast cancer through the MAPK/ERK pathway.** *Tumor Biology* 2016, **37**(4):5107-5115.
218. Uramoto H, Akyuerek LM, Hanagiri T: **A positive relationship between filamin and VEGF in patients with lung cancer.** *Anticancer research* 2010, **30**(10):3939-3944.
219. Kasaian K, Wiseman SM, Walker BA, Schein JE, Zhao Y, Hirst M, Moore RA, Mungall AJ, Marra MA, Jones SJ: **The genomic and transcriptomic landscape of anaplastic thyroid cancer: implications for therapy.** *BMC cancer* 2015, **15**(1):984.
220. Garrett PA, Hulka BS, Kim YL, Farber RA: **HRAS protooncogene polymorphism and breast cancer.** *Cancer Epidemiology and Prevention Biomarkers* 1993, **2**(2):131-138.
221. Schulten H-J, Al-Maghribi J, Al-Ghamdi K, Salama S, Al-Muhayawi S, Chaudhary A, Hamour O, Abuzenadah A, Gari M, Al-Qahtani M: **Mutational screening of RET, HRAS, KRAS, NRAS, BRAF, AKT1, and CTNNB1 in medullary thyroid carcinoma.** *Anticancer research* 2011, **31**(12):4179-4183.
222. Mardente S, Mari E, Massimi I, Fico F, Faggioni A, Pulcinelli F, Antonaci A, Zicari A: **HMGB1-induced cross talk between PTEN and miRs 221/222 in thyroid cancer.** *BioMed research international* 2015, **2015**.
223. Gjerstorff MF, Benoit VM, Laenholm A-V, Nielsen O, Johansen LE, Ditzel HJ: **Identification of genes with altered expression in medullary breast cancer vs. ductal breast cancer and normal breast epithelia.** *International journal of oncology* 2006, **28**(6):1327-1335.
224. Chakrabarti G: **Mutant KRAS associated malic enzyme 1 expression is a predictive marker for radiation therapy response in non-small cell lung cancer.** *Radiation Oncology* 2015, **10**(1):145.
225. Leo JC, Wang SM, Guo CH, Aw SE, Zhao Y, Li JM, Hui KM, Lin VC: **Gene regulation profile reveals consistent anticancer properties of progesterone in hormone-independent breast cancer cells transfected with progesterone receptor.** *International journal of cancer* 2005, **117**(4):561-568.
226. Straight AM, Oakley K, Moores R, Bauer AJ, Patel A, Tuttle RM, Jimeno J, Francis GL: **Aplidin reduces growth of anaplastic thyroid cancer xenografts and the expression**

- of several angiogenic genes.** *Cancer chemotherapy and pharmacology* 2006, **57**(1):7-14.
227. Dong X-Y, Guo P, Boyd J, Sun X, Li Q, Zhou W, Dong J-T: **Implication of snoRNA U50 in human breast cancer.** *Journal of genetics and genomics* 2009, **36**(8):447-454.
228. Li Y, Huang J, Zhao Y, He J, Wang W, Davies K, Nose V, Xiao S: **UTRN on chromosome 6q24 is mutated in multiple tumors.** *Oncogene* 2007, **26**(42):6220.
229. Zhang H, Gao P, Fukuda R, Kumar G, Krishnamachary B, Zeller KI, Dang CV, Semenza GL: **HIF-1 inhibits mitochondrial biogenesis and cellular respiration in VHL-deficient renal cell carcinoma by repression of C-MYC activity.** *Cancer cell* 2007, **11**(5):407-420.
230. Lin S-F, Yu Z, Riedl C, Woo Y, Zhang Q, Yong AY, Timiryasova T, Chen N, Shah JP, Szalay AA: **Treatment of anaplastic thyroid carcinoma in vitro with a mutant vaccinia virus.** *Surgery* 2007, **142**(6):976-983.
231. Williams SA, Riel-Mehan M, Ostroff RM: **Pancreatic Cancer Biomarkers and Uses Thereof.** In.: Google Patents; 2014.
232. Zou M, Baitei EY, BinEssa HA, Al-Mohanna FA, Parhar RS, St-Arnaud R, Kimura S, Pritchard C, Alzahrani AS, Assiri AM: **Cyp24a1 attenuation limits progression of BrafV600E-induced papillary thyroid cancer cells and sensitizes them to BRAFV600E inhibitor PLX4720.** *Cancer research* 2017, **77**(8):2161-2172.
233. Scott GK, Daniel JC, Xiong X, Maki RA, Kabat D, Benz CC: **Binding of an ETS-related protein within the DNase I hypersensitive site of the HER2/neu promoter in human breast cancer cells.** *Journal of Biological Chemistry* 1994, **269**(31):19848-19858.
234. Jiang H, Yang T, Lu P, Ma Y: **Gene expression profiling of gastric cancer.** *Eur Rev Med Pharmacol Sci* 2014, **18**(15):2109-2115.
235. Futagami S, Tatsuguchi A, Hiratsuka T, Shindo T, Horie A, Hamamoto T, Ueki N, Kusunoki M, Miyake K, Gudis K: **Monocyte chemoattractant protein 1 and CD40 ligation have a synergistic effect on vascular endothelial growth factor production through cyclooxygenase 2 upregulation in gastric cancer.** *Journal of gastroenterology* 2008, **43**(3):216-224.
236. Eissa S, Zaki SA, El-Magraby SM, Kadry DY: **Importance of serum IL-18 and RANTES as markers for breast carcinoma progression.** *J Egypt Natl Canc Inst* 2005, **17**(1):51-55.
237. Abdolahi F, Dabbaghmanesh MH, Haghshenas MR, Ghaderi A, Erfani N: **A gene-disease association study of IL18 in thyroid cancer: genotype and haplotype analyses.** *Endocrine* 2015, **50**(3):698-707.
238. Blake ML, Tometsko M, Miller R, Jones JC, Dougall WC: **RANK expression on breast cancer cells promotes skeletal metastasis.** *Clinical & experimental metastasis* 2014, **31**(2):233-245.
239. Italiano D, Lena AM, Melino G, Candi E: **Identification of NCF2/p67phox as a novel p53 target gene.** *Cell cycle* 2012, **11**(24):4589-4596.
240. Khodarev N, Ahmad R, Rajabi H, Pitroda S, Kufe T, McClary C, Joshi MD, MacDermed D, Weichselbaum R, Kufe D: **Cooperativity of the MUC1 oncoprotein and STAT1 pathway in poor prognosis human breast cancer.** *Oncogene* 2010, **29**(6):920.
241. Hwang ES, Kim DW, Hwang JH, Jung HS, Suh JM, Park YJ, Chung HK, Song JH, Park KC, Park SH: **Regulation of signal transducer and activator of transcription 1**

- (STAT1) and STAT1-dependent genes by RET/PTC (rearranged in transformation/papillary thyroid carcinoma) oncogenic tyrosine kinases.** *Molecular Endocrinology* 2004, **18**(11):2672-2684.
242. Fujieda S, Sugimoto C, Seki M, Sunaga H, Saito H: **CD40 stimulation inhibits cell growth and Fas-mediated apoptosis in a thyroid cancer cell line.** *Oncology Research Featuring Preclinical and Clinical Cancer Therapeutics* 1998, **10**(9):433-439.
243. Jiang Y-Z, Liu Y-R, Xu X-E, Jin X, Hu X, Yu K-D, Shao Z-M: **Transcriptome analysis of triple-negative breast cancer reveals an integrated mRNA-lncRNA signature with predictive and prognostic value.** *Cancer research* 2016, **76**(8):2105-2114.
244. Zhao Y, Zhao L, Mao T, Zhong L: **Assessment of risk based on variant pathways and establishment of an artificial neural network model of thyroid cancer.** *BMC medical genetics* 2019, **20**(1):92.
245. Allinen M, Beroukhim R, Cai L, Brennan C, Lahti-Domenici J, Huang H, Porter D, Hu M, Chin L, Richardson A: **Molecular characterization of the tumor microenvironment in breast cancer.** *Cancer cell* 2004, **6**(1):17-32.
246. Mancikova V, Buj R, Castelblanco E, Inglada-Pérez L, Diez A, de Cubas AA, Curras-Freixes M, Maravall FX, Mauricio D, Matias-Guiu X: **DNA methylation profiling of well-differentiated thyroid cancer uncovers markers of recurrence free survival.** *International journal of cancer* 2014, **135**(3):598-610.
247. Kachroo P, Lee M-H, Zhang L, Baratelli F, Lee G, Srivastava MK, Wang G, Walser TC, Krysan K, Sharma S: **IL-27 inhibits epithelial-mesenchymal transition and angiogenic factor production in a STAT1-dominant pathway in human non-small cell lung cancer.** *Journal of Experimental & Clinical Cancer Research* 2013, **32**(1):97.
248. Marangoni E, Laurent C, Coussy F, El-Botty R, Château-Joubert S, Servely J-L, de Plater L, Assayag F, Dahmani A, Montaudon E: **Capecitabine efficacy is correlated with TYMP and RB1 expression in PDX established from triple-negative breast cancers.** *Clinical Cancer Research* 2018, **24**(11):2605-2615.
249. Airoldi I, Tupone MG, Esposito S, Russo MV, Barbarito G, Cipollone G, Di Carlo E: **Interleukin-27 re-educates intratumoral myeloid cells and down-regulates stemness genes in non-small cell lung cancer.** *Oncotarget* 2015, **6**(6):3694.
250. Zhou J, Wulffkuhle J, Zhang H, Gu P, Yang Y, Deng J, Margolick JB, Liotta LA, Petricoin E, Zhang Y: **Activation of the PTEN/mTOR/STAT3 pathway in breast cancer stem-like cells is required for viability and maintenance.** *Proceedings of the National Academy of Sciences* 2007, **104**(41):16158-16163.
251. Couto JP, Daly L, Almeida A, Knauf JA, Fagin JA, Sobrinho-Simões M, Lima J, Máximo V, Soares P, Lyden D: **STAT3 negatively regulates thyroid tumorigenesis.** *Proceedings of the National Academy of Sciences* 2012, **109**(35):E2361-E2370.
252. Zheng W, Long J, Gao Y-T, Li C, Zheng Y, Xiang Y-B, Wen W, Levy S, Deming SL, Haines JL: **Genome-wide association study identifies a new breast cancer susceptibility locus at 6q25. 1.** *Nature genetics* 2009, **41**(3):324.
253. Shiozaki A, Lodyga M, Bai X-H, Nadesalingam J, Oyaizu T, Winer D, Asa SL, Keshavjee S, Liu M: **XB130, a novel adaptor protein, promotes thyroid tumor growth.** *The American journal of pathology* 2011, **178**(1):391-401.
254. Zhang X, Yue P, Page BD, Li T, Zhao W, Namanja AT, Paladino D, Zhao J, Chen Y,

- Gunning PT: **Orally bioavailable small-molecule inhibitor of transcription factor Stat3 regresses human breast and lung cancer xenografts.** *Proceedings of the National Academy of Sciences* 2012, **109**(24):9623-9628.
255. Liu CC, Lin JH, Hsu TW, Su K, Li AFY, Hsu HS, Hung SC: **IL-6 enriched lung cancer stem-like cell population by inhibition of cell cycle regulators via DNMT1 upregulation.** *International Journal of Cancer* 2015, **136**(3):547-559.
256. Arakawa Y, Watanabe M, Inoue N, Sarumaru M, Hidaka Y, Iwatani Y: **Association of polymorphisms in DNMT1, DNMT3A, DNMT3B, MTHFR and MTRR genes with global DNA methylation levels and prognosis of autoimmune thyroid disease.** *Clinical & Experimental Immunology* 2012, **170**(2):194-201.
257. Kanda N, Seno H, Konda Y, Marusawa H, Kanai M, Nakajima T, Kawashima T, Nanakin A, Sawabu T, Uenoyama Y: **STAT3 is constitutively activated and supports cell survival in association with survivin expression in gastric cancer cells.** *Oncogene* 2004, **23**(28):4921.
258. Boehm JS, Zhao JJ, Yao J, Kim SY, Firestein R, Dunn IF, Sjostrom SK, Garraway LA, Weremowicz S, Richardson AL: **Integrative genomic approaches identify IKBKE as a breast cancer oncogene.** *Cell* 2007, **129**(6):1065-1079.
259. Ali Z, Deng Y, Tang Y, Zheng S, Ma N, He N: **Epigenetic deregulations in gastric cancer.** *Journal of nanoscience and nanotechnology* 2013, **13**(1):40-51.
260. Arce C, Pérez-Plasencia C, González-Fierro A, de la Cruz-Hernández E, Revilla-Vázquez A, Chávez-Blanco A, Trejo-Becerril C, Pérez-Cárdenas E, Taja-Chayeb L, Bargallo E: **A proof-of-principle study of epigenetic therapy added to neoadjuvant doxorubicin cyclophosphamide for locally advanced breast cancer.** *PloS one* 2006, **1**(1):e98.
261. Katoh H, Yamashita K, Enomoto T, Watanabe M: **Classification and general considerations of thyroid cancer.** *Ann Clin Pathol* 2015, **3**(1):1045.
262. Zhang Q, Sturgill JL, Kmiecik M, Szczepanek K, Derecka M, Koebel C, Graham LJ, Dai Y, Chen S, Grant S: **The role of Tyk2 in regulation of breast cancer growth.** *Journal of Interferon & Cytokine Research* 2011, **31**(9):671-677.
263. Jin S, Borkhoo O, Bao W, Yang Y-T: **Signaling pathways in thyroid cancer and their therapeutic implications.** *Journal of clinical medicine research* 2016, **8**(4):284.
264. Rasheed BA, McLendon RE, Herndon JE, Friedman HS, Friedman AH, Bigner DD, Bigner SH: **Alterations of the TP53 gene in human gliomas.** *Cancer research* 1994, **54**(5):1324-1330.
265. Rogounovitch TI, Saenko VA, Ashizawa K, Sedliarov IA, Namba H, Abrosimov AY, Lushnikov EF, Roumiantsev PO, Konova MV, Petoukhova NS: **TP53 codon 72 polymorphism in radiation-associated human papillary thyroid cancer.** *Oncology reports* 2006, **15**(4):949-956.
266. Yang CH, Yue J, Pfeffer SR, Fan M, Paulus E, Hosni-Ahmed A, Sims M, Qayyum S, Davidoff AM, Handorf CR: **MicroRNA-21 promotes glioblastoma tumorigenesis by down-regulating insulin-like growth factor-binding protein-3 (IGFBP3).** *Journal of Biological Chemistry* 2014, **289**(36):25079-25087.
267. Siegel G, Tomer Y: **Is there an association between acromegaly and thyroid carcinoma? A critical review of the literature.** *Endocrine research* 2005, **31**(1):51-58.
268. Gemmill RM, Roche J, Potiron VA, Nasarre P, Mitas M, Coldren CD, Helfrich BA,

- Garrett-Mayer E, Bunn PA, Drabkin HA: **ZEB1-responsive genes in non-small cell lung cancer.** *Cancer letters* 2011, **300**(1):66-78.
269. Yang J, Zhang X, Zhang Y, Zhu D, Zhang L, Li Y, Zhu Y, Li D, Zhou J: **HIF-2 α promotes epithelial-mesenchymal transition through regulating Twist2 binding to the promoter of E-cadherin in pancreatic cancer.** *Journal of Experimental & Clinical Cancer Research* 2016, **35**(1):26.
270. Leidinger P, Keller A, Backes C, Huwer H, Meese E: **MicroRNA expression changes after lung cancer resection: a follow-up study.** *RNA biology* 2012, **9**(6):900-910.
271. Bommer GT, Jäger C, Dürr E-M, Baehs S, Eichhorst ST, Brabletz T, Hu G, Fröhlich T, Arnold G, Kress DC: **DRO1, a gene down-regulated by oncogenes, mediates growth inhibition in colon and pancreatic cancer cells.** *Journal of Biological Chemistry* 2005, **280**(9):7962-7975.
272. Landa I, Ruiz-Llorente S, Montero-Conde C, Inglada-Pérez L, Schiavi F, Leskelä S, Pita G, Milne R, Maravall J, Ramos I: **The variant rs1867277 in FOXE1 gene confers thyroid cancer susceptibility through the recruitment of USF1/USF2 transcription factors.** *PLoS genetics* 2009, **5**(9):e1000637.
273. Wallrarr C, MÜLLER-PILLASCH F, Micha A, Wenger C, Geng M, Solinas-Toldo S, Lichter P, Frohme M, Hoheisel J, Adler G: **Strategies for the detection of disease genes in pancreatic cancer.** *Annals of the New York Academy of Sciences* 1999, **880**(1):122-146.
274. Jeong S, Lee J, Kim D, Seol M-Y, Lee WK, Jeong JJ, Nam K-H, Jung SG, Shin DY, Lee EJ: **Relationship of focally amplified long noncoding on chromosome 1 (FAL1) lncRNA with E2F transcription factors in thyroid cancer.** *Medicine* 2016, **95**(4).

Table S4. miRNAs targeted by H19

miRNA	Description	Reference
let-7a	<ul style="list-style-type: none"> ➤ The H19/let-7 double-negative feedback loop contributes to glucose metabolism in muscle cells. ➤ The imprinted H19 lncRNA antagonizes let-7 microRNAs. ➤ H19 lncRNA alters stromal cell growth via IGF signaling in the endometrium of women with endometriosis. ➤ Glycolysis gatekeeper PDK1 reprograms breast cancer stem cells under hypoxia. 	[1] [2] [3] [4]
let-7b	<ul style="list-style-type: none"> ➤ The imprinted H19 lncRNA antagonizes let-7 microRNAs. ➤ Glycolysis gatekeeper PDK1 reprograms breast cancer stem cells under hypoxia. ➤ The lncRNA H19 mediates breast cancer cell plasticity during EMT and MET plasticity by differentially sponging miR-200b/c and let-7b. 	[2] [4] [5]
let-7g	<ul style="list-style-type: none"> ➤ The imprinted H19 lncRNA antagonizes let-7 microRNAs. ➤ H19 lncRNA alters stromal cell growth via IGF signaling in the endometrium of women with endometriosis. 	[2] [3]
let-7i	<ul style="list-style-type: none"> ➤ H19/let-7/LIN28 reciprocal negative regulatory circuit promotes breast cancer stem cell maintenance. 	[6]
miR-106a	<ul style="list-style-type: none"> ➤ miR-CLIP capture of a miRNA targetome uncovers a lincRNA H19-miR-106a interaction. 	[7]
miR-130b-3p	<ul style="list-style-type: none"> ➤ H19 lncRNA regulates keratinocyte differentiation by targeting miR-130b-3p. 	[8]
miR-138-5p	<ul style="list-style-type: none"> ➤ Decreased Expression of MiR-138-5p by LncRNA H19 in Cervical Cancer Promotes Tumor Proliferation. 	[9]
miR-139	<ul style="list-style-type: none"> ➤ H19 lncRNA alters stromal cell growth via IGF signaling in the endometrium of women with endometriosis. ➤ Long Non-Coding RNA H19 Protects H9c2 Cells against Hypoxia-Induced Injury by Targeting MicroRNA-139. 	[3] [10]
miR-141	<ul style="list-style-type: none"> ➤ H19 activates Wnt signaling and promotes osteoblast differentiation by functioning as a competing endogenous RNA. 	[11]
miR-152-3p	<ul style="list-style-type: none"> ➤ Long non-coding RNA H19 promotes the proliferation and invasion of breast cancer through upregulating DNMT1 expression by sponging miR-152. 	[12]
miR-152-5p	<ul style="list-style-type: none"> ➤ Long non-coding RNA H19 promotes the proliferation and invasion of breast cancer through upregulating DNMT1 expression by sponging miR-152. 	[12]
miR-17-5p	<ul style="list-style-type: none"> ➤ Long noncoding RNA H19 competitively binds miR-17-5p to regulate YES1 expression in thyroid cancer. ➤ Long non-coding RNA H19 suppresses retinoblastoma progression 	[13] [14]

		via counteracting miR-17-92 cluster.	
miR-181d-3p	➤	Hypoxia induces H19 expression through direct and indirect Hif-1 α activity, promoting oncogenic effects in glioblastoma.	[15]
miR-181d-5p	➤	Hypoxia induces H19 expression through direct and indirect Hif-1 α activity, promoting oncogenic effects in glioblastoma.	[15]
miR-18a	➤	Long non-coding RNA H19 suppresses retinoblastoma progression via counteracting miR-17-92 cluster.	[14]
miR-194-5p	➤	Long noncoding RNA H19 contributes to gallbladder cancer cell proliferation by modulated miR-194-5p targeting AKT2.	[16]
miR-196a	➤	The lncRNA H19 Mediates Pulmonary Fibrosis by Regulating the miR-196a/COL1A1 Axis.	[17]
miR-19a	➤	Long non-coding RNA H19 suppresses retinoblastoma progression via counteracting miR-17-92 cluster.	[14]
miR-19b-1	➤	Long non-coding RNA H19 suppresses retinoblastoma progression via counteracting miR-17-92 cluster.	[14]
miR-200b	➤	The lncRNA H19 mediates breast cancer cell plasticity during EMT and MET plasticity by differentially sponging miR-200b/c and let-7b.	[5]
miR-200c	➤	The lncRNA H19 mediates breast cancer cell plasticity during EMT and MET plasticity by differentially sponging miR-200b/c and let-7b.	[5]
miR-20a	➤	Long non-coding RNA H19 suppresses retinoblastoma progression via counteracting miR-17-92 cluster.	[14]
miR-22	➤	H19 activates Wnt signaling and promotes osteoblast differentiation by functioning as a competing endogenous RNA.	[11]
miR-29a	➤	Long non-coding RNA H19 regulates glioma angiogenesis and the biological behavior of glioma-associated endothelial cells by inhibiting microRNA-29a.	[18]
miR-29b	➤	Long noncoding RNA H19 accelerates tenogenic differentiation and promotes tendon healing through targeting miR-29b-3p and activating TGF- β 1 signaling.	[19]
miR-342-3p	➤	Long non-coding RNA H19 regulates FOXM1 expression by competitively binding endogenous miR-342-3p in gallbladder cancer.	[20]
miR-630	➤	Long noncoding RNA H19 regulates EZH2 expression by interacting with miR-630 and promotes cell invasion in nasopharyngeal carcinoma.	[21]
miR-874	➤	LncRNA H19 functions as a competing endogenous RNA to regulate AQP3 expression by sponging miR-874 in the intestinal barrier.	[22]
miR-92a-1	➤	Long non-coding RNA H19 suppresses retinoblastoma progression via counteracting miR-17-92 cluster.	[14]

Reference

1. Gao Y, Wu F, Zhou J, Yan L, Jurczak MJ, Lee H-Y, Yang L, Mueller M, Zhou X-B, Dandolo L:

- The H19/let-7 double-negative feedback loop contributes to glucose metabolism in muscle cells.** *Nucleic acids research* 2014, **42**(22):13799-13811.
- 2. Kallen AN, Zhou X-B, Xu J, Qiao C, Ma J, Yan L, Lu L, Liu C, Yi J-S, Zhang H: **The imprinted H19 lncRNA antagonizes let-7 microRNAs.** *Molecular cell* 2013, **52**(1):101-112.
 - 3. Ghazal S, McKinnon B, Zhou J, Mueller M, Men Y, Yang L, Mueller M, Flannery C, Huang Y, Taylor HS: **H19 lncRNA alters stromal cell growth via IGF signaling in the endometrium of women with endometriosis.** *EMBO molecular medicine* 2015, **7**(8):996-1003.
 - 4. Peng F, Wang J-H, Fan W-J, Meng Y-T, Li M-M, Li T-T, Cui B, Wang H-F, Zhao Y, An F: **Glycolysis gatekeeper PDK1 reprograms breast cancer stem cells under hypoxia.** *Oncogene* 2018, **37**(8):1062.
 - 5. Zhou W, Ye X-l, Xu J, Cao M-G, Fang Z-Y, Li L-Y, Guan G-H, Liu Q, Qian Y-H, Xie D: **The lncRNA H19 mediates breast cancer cell plasticity during EMT and MET plasticity by differentially sponging miR-200b/c and let-7b.** *Sci Signal* 2017, **10**(483):eaak9557.
 - 6. Peng F, Li T-T, Wang K-L, Xiao G-Q, Wang J-H, Zhao H-D, Kang Z-J, Fan W-J, Zhu L-L, Li M: **H19/let-7/LIN28 reciprocal negative regulatory circuit promotes breast cancer stem cell maintenance.** *Cell death & disease* 2017, **8**(1):e2569.
 - 7. Imig J, Brunschweiger A, Brümmer A, Guennewig B, Mittal N, Kishore S, Tsikrika P, Gerber AP, Zavolan M, Hall J: **miR-CLIP capture of a miRNA targetome uncovers a lincRNA H19–miR-106a interaction.** *Nature chemical biology* 2015, **11**(2):107.
 - 8. Li C-x, Li H-g, Huang L-t, Kong Y-w, Chen F-y, Liang J-y, Yu H, Yao Z-r: **H19 lncRNA regulates keratinocyte differentiation by targeting miR-130b-3p.** *Cell death & disease* 2017, **8**(11):e3174.
 - 9. Ou L, Wang D, Zhang H, Yu Q, Hua F: **Decreased expression of MiR-138-5p by LncRNA H19 in cervical cancer promotes tumor proliferation.** *Oncology Research Featuring Preclinical and Clinical Cancer Therapeutics* 2018, **26**(3):401-410.
 - 10. Gong L-C, Xu H-M, Guo G-L, Zhang T, Shi J-W, Chang C: **Long non-coding rna h19 protects h9c2 cells against hypoxia-induced injury by targeting microrna-139.** *Cellular Physiology and Biochemistry* 2017, **44**(3):857-869.
 - 11. Liang W-C, Fu W-M, Wang Y-B, Sun Y-X, Xu L-L, Wong C-W, Chan K-M, Li G, Waye MM-Y, Zhang J-F: **H19 activates Wnt signaling and promotes osteoblast differentiation by functioning as a competing endogenous RNA.** *Scientific reports* 2016, **6**:20121.
 - 12. Li Z, Li Y, Li Y, Ren K, Li X, Han X, Wang J: **Long non-coding RNA H19 promotes the proliferation and invasion of breast cancer through upregulating DNMT1 expression by sponging miR-152.** *Journal of biochemical and molecular toxicology* 2017, **31**(9):e21933.
 - 13. Liu L, Yang J, Zhu X, Li D, Lv Z, Zhang X: **Long noncoding RNA H19 competitively binds miR-17-5p to regulate YES1 expression in thyroid cancer.** *The FEBS journal* 2016, **283**(12):2326-2339.
 - 14. Zhang A, Shang W, Nie Q, Li T, Li S: **Long non-coding RNA H19 suppresses retinoblastoma progression via counteracting miR-17-92 cluster.** *Journal of cellular biochemistry* 2018, **119**(4):3497-3509.
 - 15. Wu W, Hu Q, Nie E, Yu T, Wu Y, Zhi T, Jiang K, Shen F, Wang Y, Zhang J: **Hypoxia induces H19 expression through direct and indirect Hif-1 α activity, promoting oncogenic effects in glioblastoma.** *Scientific reports* 2017, **7**:45029.
 - 16. Wang S-H, Wu X-C, Zhang M-D, Weng M-Z, Zhou D, Quan Z-W: **Long noncoding RNA H19**

- contributes to gallbladder cancer cell proliferation by modulated miR-194-5p targeting AKT2.** *Tumor Biology* 2016, **37**(7):9721-9730.
- 17. Lu Q, Guo Z, Xie W, Jin W, Zhu D, Chen S, Ren T: **The lncRNA H19 Mediates Pulmonary Fibrosis by Regulating the miR-196a/COL1A1 Axis.** *Inflammation* 2018, **41**(3):896-903.
 - 18. Jia P, Cai H, Liu X, Chen J, Ma J, Wang P, Liu Y, Zheng J, Xue Y: **Long non-coding RNA H19 regulates glioma angiogenesis and the biological behavior of glioma-associated endothelial cells by inhibiting microRNA-29a.** *Cancer letters* 2016, **381**(2):359-369.
 - 19. Lu Y-F, Liu Y, Fu W-M, Xu J, Wang B, Sun Y-X, Wu T-Y, Xu L-L, Chan K-M, Zhang J-F: **Long noncoding RNA H19 accelerates tenogenic differentiation and promotes tendon healing through targeting miR-29b-3p and activating TGF- β 1 signaling.** *The FASEB Journal* 2016, **31**(3):954-964.
 - 20. Wang S-H, Ma F, Tang Z-h, Wu X-C, Cai Q, Zhang M-D, Weng M-Z, Zhou D, Wang J-D, Quan Z-W: **Long non-coding RNA H19 regulates FOXM1 expression by competitively binding endogenous miR-342-3p in gallbladder cancer.** *Journal of Experimental & Clinical Cancer Research* 2016, **35**(1):160.
 - 21. Li X, Lin Y, Yang X, Wu X, He X: **Long noncoding RNA H19 regulates EZH2 expression by interacting with miR-630 and promotes cell invasion in nasopharyngeal carcinoma.** *Biochemical and biophysical research communications* 2016, **473**(4):913-919.
 - 22. Su Z, Zhi X, Zhang Q, Yang L, Xu H, Xu Z: **Lnc RNA H19 functions as a competing endogenous RNA to regulate AQP 3 expression by sponging miR-874 in the intestinal barrier.** *FEBS letters* 2016, **590**(9):1354-1364.

Table S5. Regulation of 29 miRNAs in eight triplets.**H19-ETS1-TGFBR2**

miRNAs	miR-106a	miR-130b-3p	miR-138-5p	miR-139	miR-141	miR-152-3p	miR-152-5p
Predicted targets of miRNAs							
TFs	RUNX1	FOSL2	EZH2	ETS1	CREBBP	KLF6	KLF6
	E2F1	MYB	POU2F1		POU2F1	KLF4	E2F3
	PPARA	STAT3	RELA				KLF4
	RB1		SP1				
	SP1						
	STAT3						
	KAT2B						
Genes	CALD1	FMR1	HIF1A	FMR1	FMR1	CHUK	DNMT1
	CCNG2	MET	KLF11	MAPK8		DNMT1	FBN1
	E2F1	TGFBR2		ZEB1		FBN1	FMR1
	HIF1A					FMR1	SOS1
	SMAD4					SOS1	
	MMP2						
	MAPK8						
	SOS1						
	TGFBR2						
	KLF11						
	TXNIP						

Validated targets of miRNAs

TFs	E2F1	AHR	EZH2	NFKB1	E2F3	KLF4	
	MYB	E2F1	NFKB1		PPARA		
	RB1	NR3C1	SNAI2		HDGF		
	RUNX1	PPARA	SOX9				
	STAT3	RB1	TWIST2				
		STAT3					
Genes	E2F1	E2F1	HIF1A	HRAS	ZEB1	DNMT1	
	FAS	KLF11	SNAI2	MET	EIF4E	ADAM17	
	TGFBR2	SMAD4			KLF11		
	ATM	TGFBR2			ZEB1		
	HIF1A	ZEB1					
		FMR1					
		MMP2					

Notes: In the H19-ETS1-TGFBR2 sheet, we list all the 29 miRNAs and their targets (TFs and genes). Some of the targets were predicted and then verified. In the H19-ETS1-TGFBR2 table, TFs are marked in yellow if miRNAs target them, and genes are marked in red if miRNAs target them.

miRNAs	miR-17	miR-181d-3p	miR-181d-5p	miR-18a	miR-194-5p	miR-196a	miR-19a
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**Predicted targets
of miRNAs**

TFs	RUNX1		RUNX1	RUNX1	E2F3	ERG	
	E2F1		KLF6	KLF6	FLI1	FLI1	
	RB1		ETS1	NR3C1	SP3		
	STAT3		NR3C1	POU2F1			
	KAT2B		KAT2B				
Genes	CALD1		ATM	ATM	ACP5	FAS	FMR1
	CCNG2		SOS1	CTGF	FMR1	COL1A1	IL18
	E2F1		VCAM1	HIF1A	SP3	MAPK8	
	HIF1A		EPB41L3	HMGCS1	RSF1	ZEB1	
	MMP2			MAPK8			
	SOS1						
	TGFBR2						
	KLF11						
	TXNIP						

**Validated targets
of miRNAs**

TFs	E2F1			NR3C1	FOXM1	FOXO1	KAT2B
	E2F3			RUNX1			
	KAT2B						
	MYC						
	RB1						
	RUNX1						
	STAT3						
Genes	E2F1		HRAS	CTGF		FLNA	CTGF
	MMP2			EDF1		TGFBR2	SMAD4
	MYC			ATM			TGFBR2
	SMAD4			HIF1A			DNMT1
	TGFBR2			HMGCS1			
	HIF1A			SMAD4			
	DNMT1			TGFBR2			
	IGFBP3			DNMT1			

miRNAs	miR-19b-1	miR-200b	miR-200c	miR-20a	miR-22	miR-29a	miR-29b
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**Predicted targets
of miRNAs**

TFs	KLF6	E2F1	FOXO1	ETS1	MYBL2
	SP3	E2F3	HNRNPK	KAT2B	
		EZH2			
		PPARA			
		RB1			
		STAT3			
		KAT2B			
Genes	NCF2	CALD1	CD68		DNMT1
	MAPK8	CCNG2	SOS1		FBN1
	SP3	E2F1	UTRN		EDF1
		HIF1A	VCAM1		
		SMAD4	WRN		
		MMP2			
		MAPK8			
		SOS1			
		TGFBR2			
		KLF11			
		TXNIP			

**Validated targets
of miRNAs**

TFs	KAT2B	E2F3	MYB	E2F1	PPARA	KLF4	SP1
NR3C1	ETS1	ETS1	E2F3		AHR	STAT3	
	MYB	SP1	MYC		MYC	SP1	
	SP1	E2F3	RB1				STAT3
	EZH2	FOXO1	RUNX1				MYC
		E2F3	STAT3				
Genes	CTGF	CREB1	FLNA	CCNB1	HIF1A	MMP2	COL1A1
FMR1	FN1	FN1	E2F1		DNMT1	DNMT1	
ATM	KLF11	KLF11	FLNA		FBN1	MMP2	
HMGCS1	ZEB1	ZEB1	HIF1A		MYC	FBN1	
SMA4	DNMT1	NCAM1	MYC			MYC	
TGFBR2		CYP1B1	SMAD4				
DNMT1			TGFBR2				
			DNMT1				

miRNAs	miR-342-3p	miR-630	miR-874	miR-92a-1	let-7a	let-7b	let-7g
Predicted targets of miRNAs							
TFs	E2F3	KLF6	POU2F1		EZH2	EZH2	EZH2
		EZH2	STAT3		POU2F1	POU2F1	POU2F1
					RB1	RB1	RB1
					TP53	TP53	TP53
Genes	DNMT1		FBN1	COL1A1	FAS	FAS	FAS
	ZEB1		FMR1	DACT3	COL1A1	COL1A1	COL1A1
			SIGIRR		IL6	IL6	IL6
					MAPK8	MAPK8	MAPK8
					UTRN	UTRN	UTRN

Validated targets of miRNAs

TFs	FOSL2	SNAI2	PARP1	HDAC1	E2F1	CTCF	MYC
	E2F1	FOXM1	HDAC1	HDGF	EZH2	E2F3	
			STAT3	KAT2B	MYC	EZH2	
			E2F3	MYBL2	NFKB1	MYC	
				NFKB1	PARP1	SOX9	
				STAT3	SP1	SP1	
				HDAC2	STAT3		
				KLF4			
				IKZF1			
Genes	DNMT1	SNAI2		ATM	MYC	BIRC5	MYC
	E2F1			CCNB1	HRAS	CCNB1	FN1
				EPB41L3	E2F1	EPB41L3	
				FLNA	IL6	FLNA	
				HMGCS1		HIF1A	
				SMAD4		HMGCS1	
				TGFBR2		MYC	
				TYMP		UTRN	
				MAPK8		HRAS	
				DNMT1			

miRNAs	let-7i
Predicted targets of miRNAs	
TFs	EZH2
	POU2F1
	RB1
	TP53
Genes	FAS
	CHUK
	COL1A1
	IL6
	MAPK8
	UTRN

Validated targets of miRNAs

H19-FLI1-TGFB2

miRNAs	miR-106a	miR-130b-3p	miR-138-5p	miR-139	miR-141	miR-152-3p	miR-152-5p	miR-17
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Predicted targets of miRNAs

TFs	RUNX1	FOSL2	EZH2	ETS1	CREBBP	KLF6	KLF6	RUNX1
	E2F1	MYB	POU2F1		POU2F1	KLF4	E2F3	E2F1
	PPARA	STAT3	RELA				KLF4	RB1
	RB1		SP1					STAT3
	SP1							KAT2B
	STAT3							
	KAT2B							
Genes	CALD1	FMR1	HIF1A	FMR1	FMR1	CHUK	DNMT1	CALD1
	CCNG2	MET	KLF11	MAPK8		DNMT1	FBN1	CCNG2
	E2F1	TGFB2		ZEB1		FBN1	FMR1	E2F1
	HIF1A					FMR1	SOS1	HIF1A
	SMAD4					SOS1		MMP2
	MMP2							SOS1
	MAPK8							TGFB2
	SOS1							KLF11
	TGFB2							TXNIP
	KLF11							
	TXNIP							

Validated targets of miRNAs

TFs	E2F1	AHR	EZH2	NFKB1	E2F3	KLF4		E2F1
	MYB	E2F1	NFKB1		PPARA			E2F3
	RB1	NR3C1	SNAI2		HDGF			KAT2B
	RUNX1	PPARA	SOX9					MYC
	STAT3	RB1	TWIST2					RB1
		STAT3						RUNX1
								STAT3
Genes	E2F1	E2F1	HIF1A	HRAS	ZEB1	DNMT1		E2F1
	FAS	KLF11	SNAI2	MET	EIF4E	ADAM17		MMP2
	TGFB2	SMAD4			KLF11			MYC
	ATM	TGFB2			ZEB1			SMAD4
	HIF1A	ZEB1						TGFB2
		FMR1						HIF1A
		MMP2						DNMT1
								IGFBP3

miRNAs	miR-181d-3p	miR-181d-5p	miR-18a	miR-194-5p	miR-196a	miR-19a	miR-19b-1	miR-200b
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**Predicted targets
of miRNAs**

TFs		RUNX1	RUNX1	E2F3	ERG			KLF6
		KLF6	KLF6	FLI1	FLI1			SP3
		ETS1	NR3C1	SP3				
		NR3C1	POU2F1					
		KAT2B						
Genes		ATM	ATM	ACP5	FAS	FMR1		NCF2
		SOS1	CTGF	FMR1	COL1A1	IL18		MAPK8
		VCAM1	HIF1A	SP3	MAPK8			SP3
		EPB41L3	HMGCS1	RSF1	ZEB1			
			MAPK8					

**Validated targets
of miRNAs**

TFs			NR3C1	FOXM1	FOXO1	KAT2B	KAT2B	E2F3
			RUNX1				NR3C1	ETS1
								MYB
								SP1
								EZH2
Genes		HRAS	CTGF		FLNA	CTGF	CTGF	CREB1
			EDF1		TGFBR2	SMAD4	FMR1	FN1
			ATM			TGFBR2	ATM	KLF11
			HIF1A			DNMT1	HMGCS1	ZEB1
			HMGCS1				SMAD4	DNMT1
			SMAD4				TGFBR2	
			TGFBR2				DNMT1	
			DNMT1					

miRNAs	miR-200c	miR-20a	miR-22	miR-29a	miR-29b	miR-342-3p	miR-630	miR-874
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**Predicted targets
of miRNAs**

TFs		E2F1	FOXO1	ETS1	MYBL2	E2F3	KLF6	POU2F1
		E2F3	HNRNPK	KAT2B			EZH2	STAT3
		EZH2						
		PPARA						
		RB1						
		STAT3						
		KAT2B						
Genes		CALD1	CD68		DNMT1	DNMT1		FBN1
		CCNG2	SOS1		FBN1	ZEB1		FMR1
		E2F1	UTRN		EDF1			SIGIRR
		HIF1A	VCAM1					
		SMAD4	WRN					
		MMP2						
		MAPK8						
		SOS1						
		TGFBR2						
		KLF11						
		TXNIP						

**Validated targets
of miRNAs**

TFs	MYB	E2F1	PPARA	KLF4	SP1	FOSL2	SNAI2	PARP1
	ETS1	E2F3		AHR	STAT3	E2F1	FOXM1	HDAC1
	SP1	MYC		MYC	SP1			STAT3
	E2F3	RB1			STAT3			E2F3
	FOXO1	RUNX1			MYC			
	E2F3	STAT3						
Genes	FLNA	CCNB1	HIF1A	MMP2	COL1A1	DNMT1	SNAI2	
	FN1	E2F1		DNMT1	DNMT1	E2F1		
	KLF11	FLNA		FBN1	MMP2			
	ZEB1	HIF1A		MYC	FBN1			
	NCAM1	MYC			MYC			
	CYP1B1	SMAD4						
		TGFBR2						
		DNMT1						

miRNAs	miR-92a-1	let-7a	let-7b	let-7g	let-7i
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**Predicted targets
of miRNAs**

TFs		EZH2	EZH2	EZH2	EZH2
		POU2F1	POU2F1	POU2F1	POU2F1
		RB1	RB1	RB1	RB1
		TP53	TP53	TP53	TP53
Genes	COL1A1	FAS	FAS	FAS	FAS
	DACT3	COL1A1	COL1A1	COL1A1	CHUK
		IL6	IL6	IL6	COL1A1
		MAPK8	MAPK8	MAPK8	IL6
		UTRN	UTRN	UTRN	MAPK8
					UTRN

**Validated targets
of miRNAs**

TFs	HDAC1	E2F1	CTCF	MYC	
	HDGF	EZH2	E2F3		
	KAT2B	MYC	EZH2		
	MYBL2	NFKB1	MYC		
	NFKB1	PARP1	SOX9		
	STAT3	SP1	SP1		
	HDAC2	STAT3			
	KLF4				
	IKZF1				
Genes	ATM	MYC	BIRC5	MYC	
	CCNB1	HRAS	CCNB1	FN1	
	EPB41L3	E2F1	EPB41L3		
	FLNA	IL6	FLNA		
	HMGCS1		HIF1A		
	SMAD4		HMGCS1		
	TGFBR2		MYC		
	TYMP		UTRN		
	MAPK8		HRAS		
	DNMT1				

H19-FOXO1-TXNIP

miRNAs	miR-106a	miR-130b-3p	miR-138-5p	miR-139	miR-141	miR-152-3p	miR-152-5p
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Predicted targets of miRNAs

TFs	RUNX1	FOSL2	EZH2	ETS1	CREBBP	KLF6	KLF6
	E2F1	MYB	POU2F1		POU2F1	KLF4	E2F3
	PPARA	STAT3	RELA				KLF4
	RB1		SP1				
	SP1						
	STAT3						
	KAT2B						
Genes	CALD1	FMR1	HIF1A	FMR1	FMR1	CHUK	DNMT1
	CCNG2	MET	KLF11	MAPK8		DNMT1	FBN1
	E2F1	TGFBR2		ZEB1		FBN1	FMR1
	HIF1A					FMR1	SOS1
	SMAD4					SOS1	
	MMP2						
	MAPK8						
	SOS1						
	TGFBR2						
	KLF11						
	TXNIP						

Validated targets of miRNAs

TFs	E2F1	AHR	EZH2	NFKB1	E2F3	KLF4	
	MYB	E2F1	NFKB1		PPARA		
	RB1	NR3C1	SNAI2		HDGF		
	RUNX1	PPARA	SOX9				
	STAT3	RB1	TWIST2				
		STAT3					
Genes	E2F1	E2F1	HIF1A	HRAS	ZEB1	DNMT1	
	FAS	KLF11	SNAI2	MET	EIF4E	ADAM17	
	TGFBR2	SMAD4			KLF11		
	ATM	TGFBR2			ZEB1		
	HIF1A	ZEB1					
		FMR1					
		MMP2					

miRNAs	miR-17	miR-181d-3p	miR-181d-5p	miR-18a	miR-194-5p	miR-196a	miR-19a
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**Predicted targets
of miRNAs**

TFs	RUNX1		RUNX1	RUNX1	E2F3	ERG	
	E2F1		KLF6	KLF6	FLI1	FLI1	
	RB1		ETS1	NR3C1	SP3		
	STAT3		NR3C1	POU2F1			
	KAT2B		KAT2B				
Genes	CALD1		ATM	ATM	ACP5	FAS	FMR1
	CCNG2		SOS1	CTGF	FMR1	COL1A1	IL18
	E2F1		VCAM1	HIF1A	SP3	MAPK8	
	HIF1A		EPB41L3	HMGCS1	RSF1	ZEB1	
	MMP2			MAPK8			
	SOS1						
	TGFBR2						
	KLF11						
	TXNIP						

**Validated targets
of miRNAs**

TFs	E2F1			NR3C1	FOXM1	FOXO1	KAT2B
	E2F3			RUNX1			
	KAT2B						
	MYC						
	RB1						
	RUNX1						
	STAT3						
Genes	E2F1		HRAS	CTGF		FLNA	CTGF
	MMP2			EDF1		TGFBR2	SMAD4
	MYC			ATM			TGFBR2
	SMAD4			HIF1A			DNMT1
	TGFBR2			HMGCS1			
	HIF1A			SMAD4			
	DNMT1			TGFBR2			
	IGFBP3			DNMT1			

miRNAs	miR-19b-1	miR-200b	miR-200c	miR-20a	miR-22	miR-29a	miR-29b
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**Predicted targets
of miRNAs**

TFs		KLF6		E2F1	FOXO1	ETS1	MYBL2
		SP3		E2F3	HNRNPK	KAT2B	
				EZH2			
				PPARA			
				RB1			
				STAT3			
				KAT2B			
Genes		NCF2		CALD1	CD68		DNMT1
		MAPK8		CCNG2	SOS1		FBN1
		SP3		E2F1	UTRN		EDF1
				HIF1A	VCAM1		
				SMAD4	WRN		
				MMP2			
				MAPK8			
				SOS1			
				TGFBR2			
				KLF11			
				TXNIP			

**Validated targets
of miRNAs**

TFs	KAT2B	E2F3	MYB	E2F1	PPARA	KLF4	SP1
	NR3C1	ETS1	ETS1	E2F3		AHR	STAT3
		MYB	SP1	MYC		MYC	SP1
		SP1	E2F3	RB1			STAT3
		EZH2	FOXO1	RUNX1			MYC
			E2F3	STAT3			
Genes	CTGF	CREB1	FLNA	CCNB1	HIF1A	MMP2	COL1A1
	FMR1	FN1	FN1	E2F1		DNMT1	DNMT1
	ATM	KLF11	KLF11	FLNA		FBN1	MMP2
	HMGCS1	ZEB1	ZEB1	HIF1A		MYC	FBN1
	SMAD4	DNMT1	NCAM1	MYC			MYC
	TGFBR2		CYP1B1	SMAD4			
	DNMT1			TGFBR2			
				DNMT1			

miRNAs	miR-342-3p	miR-630	miR-874	miR-92a-1	let-7a	let-7b	let-7g
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**Predicted targets
of miRNAs**

TFs	E2F3	KLF6	POU2F1		EZH2	EZH2	EZH2
		EZH2	STAT3		POU2F1	POU2F1	POU2F1
					RB1	RB1	RB1
					TP53	TP53	TP53
Genes	DNMT1		FBN1	COL1A1	FAS	FAS	FAS
	ZEB1		FMR1	DACT3	COL1A1	COL1A1	COL1A1
			SIGIRR		IL6	IL6	IL6
					MAPK8	MAPK8	MAPK8
					UTRN	UTRN	UTRN

**Validated targets
of miRNAs**

TFs	FOSL2	SNAI2	PARP1	HDAC1	E2F1	CTCF	MYC
	E2F1	FOXM1	HDAC1	HDGF	EZH2	E2F3	
			STAT3	KAT2B	MYC	EZH2	
			E2F3	MYBL2	NFKB1	MYC	
				NFKB1	PARP1	SOX9	
				STAT3	SP1	SP1	
				HDAC2	STAT3		
				KLF4			
				IKZF1			
Genes	DNMT1	SNAI2		ATM	MYC	BIRC5	MYC
	E2F1			CCNB1	HRAS	CCNB1	FN1
				EPB41L3	E2F1	EPB41L3	
				FLNA	IL6	FLNA	
				HMGCS1		HIF1A	
				SMAD4		HMGCS1	
				TGFBR2		MYC	
				TYMP		UTRN	
				MAPK8		HRAS	
				DNMT1			

miRNAs	let-7i
Predicted targets of miRNAs	
TFs	EZH2
	POU2F1
	RB1
	TP53
Genes	FAS
	CHUK
	COL1A1
	IL6
	MAPK8
	UTRN

Validated targets of miRNAs

H19-KLF6-TXNIP

miRNAs	miR-106a	miR-130b-3p	miR-138-5p	miR-139	miR-141	miR-152-3p	miR-152-5p
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**Predicted targets
of miRNAs**

TFs	RUNX1	FOSL2	EZH2	ETS1	CREBBP	KLF6	KLF6
	E2F1	MYB	POU2F1		POU2F1	KLF4	E2F3
	PPARA	STAT3	RELA				KLF4
	RB1		SP1				
	SP1						
	STAT3						
	KAT2B						
Genes	CALD1	FMR1	HIF1A	FMR1	FMR1	CHUK	DNMT1
	CCNG2	MET	KLF11	MAPK8		DNMT1	FBN1
	E2F1	TGFBR2		ZEB1		FBN1	FMR1
	HIF1A					FMR1	SOS1
	SMAD4					SOS1	
	MMP2						
	MAPK8						
	SOS1						
	TGFBR2						
	KLF11						
	TXNIP						

**Validated targets
of miRNAs**

TFs	E2F1	AHR	EZH2	NFKB1	E2F3	KLF4	
	MYB	E2F1	NFKB1		PPARA		
	RB1	NR3C1	SNAI2		HDGF		
	RUNX1	PPARA	SOX9		KLF6????		
	STAT3	RB1	TWIST2				
		STAT3					
Genes	E2F1	E2F1	HIF1A	HRAS	ZEB1	DNMT1	
	FAS	KLF11	SNAI2	MET	EIF4E	ADAM17	
	TGFBR2	SMAD4			KLF11		
	ATM	TGFBR2			ZEB1		
	HIF1A	ZEB1					
		FMR1					
		MMP2					

miRNAs	miR-17	miR-181d-3p	miR-181d-5p	miR-18a	miR-194-5p	miR-196a	miR-19a
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**Predicted targets
of miRNAs**

TFs	RUNX1		RUNX1	RUNX1	E2F3	ERG	
	E2F1		KLF6	KLF6	FLI1	FLI1	
	RB1		ETS1	NR3C1	SP3		
	STAT3		NR3C1	POU2F1			
	KAT2B		KAT2B				
Genes	CALD1		ATM	ATM	ACP5	FAS	FMR1
	CCNG2		SOS1	CTGF	FMR1	COL1A1	IL18
	E2F1		VCAM1	HIF1A	SP3	MAPK8	
	HIF1A		EPB41L3	HMGCS1	RSF1	ZEB1	
	MMP2			MAPK8			
	SOS1						
	TGFBR2						
	KLF11						
	TXNIP						

**Validated targets
of miRNAs**

TFs	E2F1			NR3C1	FOXM1	FOXO1	KAT2B
	E2F3			RUNX1			
	KAT2B						
	MYC						
	RB1						
	RUNX1						
	STAT3						
Genes	E2F1		HRAS	CTGF		FLNA	CTGF
	MMP2			EDF1		TGFBR2	SMAD4
	MYC			ATM			TGFBR2
	SMAD4			HIF1A			DNMT1
	TGFBR2			HMGCS1			
	HIF1A			SMAD4			
	DNMT1			TGFBR2			
	IGFBP3			DNMT1			

miRNAs	miR-19b-1	miR-200b	miR-200c	miR-20a	miR-22	miR-29a	miR-29b
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**Predicted targets
of miRNAs**

TFs		KLF6		E2F1	FOXO1	ETS1	MYBL2
		SP3		E2F3	HNRNPK	KAT2B	
				EZH2			
				PPARA			
				RB1			
				STAT3			
				KAT2B			
Genes		NCF2		CALD1	CD68		DNMT1
		MAPK8		CCNG2	SOS1		FBN1
		SP3		E2F1	UTRN		EDF1
				HIF1A	VCAM1		
				SMAD4	WRN		
				MMP2			
				MAPK8			
				SOS1			
				TGFBR2			
				KLF11			
				TXNIP			

**Validated targets
of miRNAs**

TFs	KAT2B	E2F3	MYB	E2F1	PPARA	KLF4	SP1
	NR3C1	ETS1	ETS1	E2F3		AHR	STAT3
		MYB	SP1	MYC		MYC	SP1
		SP1	E2F3	RB1			STAT3
		EZH2	FOXO1	RUNX1			MYC
			E2F3	STAT3			
Genes	CTGF	CREB1	FLNA	CCNB1	HIF1A	MMP2	COL1A1
	FMR1	FN1	FN1	E2F1		DNMT1	DNMT1
	ATM	KLF11	KLF11	FLNA		FBN1	MMP2
	HMGCS1	ZEB1	ZEB1	HIF1A		MYC	FBN1
	SMAD4	DNMT1	NCAM1	MYC			MYC
	TGFBR2		CYP1B1	SMAD4			
	DNMT1			TGFBR2			
				DNMT1			

miRNAs	miR-342-3p	miR-630	miR-874	miR-92a-1	let-7a	let-7b	let-7g
Predicted targets of miRNAs							
TFs	E2F3	KLF6	POU2F1		EZH2	EZH2	EZH2
		EZH2	STAT3		POU2F1	POU2F1	POU2F1
					RB1	RB1	RB1
					TP53	TP53	TP53
Genes	DNMT1		FBN1	COL1A1	FAS	FAS	FAS
	ZEB1		FMR1	DACT3	COL1A1	COL1A1	COL1A1
			SIGIRR		IL6	IL6	IL6
					MAPK8	MAPK8	MAPK8
					UTRN	UTRN	UTRN

Validated targets of miRNAs

TFs	FOSL2	SNAI2	PARP1	HDAC1	E2F1	CTCF	MYC
	E2F1	FOXM1	HDAC1	HDGF	EZH2	E2F3	
			STAT3	KAT2B	MYC	EZH2	
			E2F3	MYBL2	NFKB1	MYC	
				NFKB1	PARP1	SOX9	
				STAT3	SP1	SP1	
				HDAC2	STAT3		
				KLF4			
				IKZF1			
Genes	DNMT1	SNAI2		ATM	MYC	BIRC5	MYC
	E2F1			CCNB1	HRAS	CCNB1	FN1
				EPB41L3	E2F1	EPB41L3	
				FLNA	IL6	FLNA	
				HMGCS1		HIF1A	
				SMAD4		HMGCS1	
				TGFBR2		MYC	
				TYMP		UTRN	
				MAPK8		HRAS	
				DNMT1			

miRNAs	let-7i
Predicted targets of miRNAs	
TFs	EZH2
	POU2F1
	RB1
	TP53
Genes	FAS
	CHUK
	COL1A1
	IL6
	MAPK8
	UTRN

Validated targets of miRNAs

H19-PPARA-KLF11

miRNAs	miR-106a	miR-130b-3p	miR-138-5p	miR-139	miR-141	miR-152-3p	miR-152-5p
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Predicted targets of miRNAs

TFs	RUNX1	FOSL2	EZH2	ETS1	CREBBP	KLF6	KLF6
	E2F1	MYB	POU2F1		POU2F1	KLF4	E2F3
	PPARA	STAT3	RELA				KLF4
	RB1		SP1				
	SP1						
	STAT3						
	KAT2B						
Genes	CALD1	FMR1	HIF1A	FMR1	FMR1	CHUK	DNMT1
	CCNG2	MET	KLF11	MAPK8		DNMT1	FBN1
	E2F1	TGFBR2		ZEB1		FBN1	FMR1
	HIF1A					FMR1	SOS1
	SMAD4					SOS1	
	MMP2						
	MAPK8						
	SOS1						
	TGFBR2						
	KLF11						
	TXNIP						

Validated targets of miRNAs

TFs	E2F1	AHR	EZH2	NFKB1	E2F3	KLF4	
	MYB	E2F1	NFKB1		PPARA		
	RB1	NR3C1	SNAI2		HDGF		
	RUNX1	PPARA	SOX9				
	STAT3	RB1	TWIST2				
		STAT3					
Genes	E2F1	E2F1	HIF1A	HRAS	ZEB1	DNMT1	
	FAS	KLF11	SNAI2	MET	EIF4E	ADAM17	
	TGFBR2	SMAD4			KLF11		
	ATM	TGFBR2			ZEB1		
	HIF1A	ZEB1					
		FMR1					
		MMP2					

miRNAs	miR-17	miR-181d-3p	miR-181d-5p	miR-18a	miR-194-5p	miR-196a	miR-19a
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**Predicted targets
of miRNAs**

TFs	RUNX1		RUNX1	RUNX1	E2F3	ERG	
	E2F1		KLF6	KLF6	FLI1	FLI1	
	RB1		ETS1	NR3C1	SP3		
	STAT3		NR3C1	POU2F1			
	KAT2B		KAT2B				
Genes	CALD1		ATM	ATM	ACP5	FAS	FMR1
	CCNG2		SOS1	CTGF	FMR1	COL1A1	IL18
	E2F1		VCAM1	HIF1A	SP3	MAPK8	
	HIF1A		EPB41L3	HMGCS1	RSF1	ZEB1	
	MMP2			MAPK8			
	SOS1						
	TGFBR2						
	KLF11						
	TXNIP						

**Validated targets
of miRNAs**

TFs	E2F1			NR3C1	FOXM1	FOXO1	KAT2B
	E2F3			RUNX1			
	KAT2B						
	MYC						
	RB1						
	RUNX1						
	STAT3						
Genes	E2F1		HRAS	CTGF		FLNA	CTGF
	MMP2			EDF1		TGFBR2	SMAD4
	MYC			ATM			TGFBR2
	SMAD4			HIF1A			DNMT1
	TGFBR2			HMGCS1			
	HIF1A			SMAD4			
	DNMT1			TGFBR2			
	IGFBP3			DNMT1			

miRNAs	miR-19b-1	miR-200b	miR-200c	miR-20a	miR-22	miR-29a	miR-29b
Predicted targets of miRNAs							
TFs		KLF6		E2F1	FOXO1	ETS1	MYBL2
		SP3		E2F3	HNRNPK	KAT2B	
				EZH2			
				PPARA			
				RB1			
				STAT3			
				KAT2B			
Genes		NCF2		CALD1	CD68		DNMT1
		MAPK8		CCNG2	SOS1		FBN1
		SP3		E2F1	UTRN		EDF1
				HIF1A	VCAM1		
				SMAD4	WRN		
				MMP2			
				MAPK8			
				SOS1			
				TGFBR2			
				KLF11			
				TXNIP			

Validated targets of miRNAs							
TFs	KAT2B	E2F3	MYB	E2F1	PPARA	KLF4	SP1
	NR3C1	ETS1	ETS1	E2F3		AHR	STAT3
		MYB	SP1	MYC		MYC	SP1
		SP1	E2F3	RB1			STAT3
		EZH2	FOXO1	RUNX1			MYC
			E2F3	STAT3			
Genes	CTGF	CREB1	FLNA	CCNB1	HIF1A	MMP2	COL1A1
	FMR1	FN1	FN1	E2F1		DNMT1	DNMT1
	ATM	KLF11	KLF11	FLNA		FBN1	MMP2
	HMGCS1	ZEB1	ZEB1	HIF1A		MYC	FBN1
	SMAD4	DNMT1	NCAM1	MYC			MYC
	TGFBR2		CYP1B1	SMAD4			
	DNMT1			TGFBR2			
				DNMT1			

miRNAs	miR-342-3p	miR-630	miR-874	miR-92a-1	let-7a	let-7b	let-7g
Predicted targets of miRNAs							
TFs	E2F3	KLF6	POU2F1		EZH2	EZH2	EZH2
	EZH2	STAT3		POU2F1	POU2F1	POU2F1	POU2F1
				RB1	RB1	RB1	
					TP53	TP53	TP53
Genes	DNMT1		FBN1	COL1A1	FAS	FAS	FAS
	ZEB1		FMR1	DACT3	COL1A1	COL1A1	COL1A1
			SIGIRR		IL6	IL6	IL6
					MAPK8	MAPK8	MAPK8
					UTRN	UTRN	UTRN

Validated targets of miRNAs

TFs	FOSL2	SNAI2	PARP1	HDAC1	E2F1	CTCF	MYC
	E2F1	FOXM1	HDAC1	HDGF	EZH2	E2F3	
			STAT3	KAT2B	MYC	EZH2	
			E2F3	MYBL2	NFKB1	MYC	
				NFKB1	PARP1	SOX9	
				STAT3	SP1	SP1	
				HDAC2	STAT3		
				KLF4			
				IKZF1			
Genes	DNMT1	SNAI2		ATM	MYC	BIRC5	MYC
	E2F1			CCNB1	HRAS	CCNB1	FN1
				EPB41L3	E2F1	EPB41L3	
				FLNA	IL6	FLNA	
				HMGCS1		HIF1A	
				SMAD4		HMGCS1	
				TGFBR2		MYC	
				TYMP		UTRN	
				MAPK8		HRAS	
				DNMT1			

miRNAs	let-7i
Predicted targets of miRNAs	
TFs	EZH2
	POU2F1
	RB1
	TP53
Genes	FAS
	CHUK
	COL1A1
	IL6
	MAPK8
	UTRN

Validated targets of miRNAs

H19-SP1-TGFBR2

miRNAs	miR-106a	miR-130b-3p	miR-138-5p	miR-139	miR-141	miR-152-3p	miR-152-5p
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Predicted targets of miRNAs

TFs	RUNX1	FOSL2	EZH2	ETS1	CREBBP	KLF6	KLF6
	E2F1	MYB	POU2F1		POU2F1	KLF4	E2F3
	PPARA	STAT3	RELA				KLF4
	RB1		SP1				
	SP1						
	STAT3						
	KAT2B						
Genes	CALD1	FMR1	HIF1A	FMR1	FMR1	CHUK	DNMT1
	CCNG2	MET	KLF11	MAPK8		DNMT1	FBN1
	E2F1	TGFBR2		ZEB1		FBN1	FMR1
	HIF1A					FMR1	SOS1
	SMAD4					SOS1	
	MMP2						
	MAPK8						
	SOS1						
	TGFBR2						
	KLF11						
	TXNIP						

Validated targets of miRNAs

TFs	E2F1	AHR	EZH2	NFKB1	E2F3	KLF4	
	MYB	E2F1	NFKB1		PPARA		
	RB1	NR3C1	SNAI2		HDGF		
	RUNX1	PPARA	SOX9				
	STAT3	RB1	TWIST2				
		STAT3					
Genes	E2F1	E2F1	HIF1A	HRAS	ZEB1	DNMT1	
	FAS	KLF11	SNAI2	MET	EIF4E	ADAM17	
	TGFBR2	SMAD4			KLF11		
	ATM	TGFBR2			ZEB1		
	HIF1A	ZEB1					
		FMR1					
		MMP2					

miRNAs	miR-17	miR-181d-3p	miR-181d-5p	miR-18a	miR-194-5p	miR-196a	miR-19a
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**Predicted targets
of miRNAs**

TFs	RUNX1		RUNX1	RUNX1	E2F3	ERG	
	E2F1		KLF6	KLF6	FLI1	FLI1	
	RB1		ETS1	NR3C1	SP3		
	STAT3		NR3C1	POU2F1			
	KAT2B		KAT2B				
Genes	CALD1		ATM	ATM	ACP5	FAS	FMR1
	CCNG2		SOS1	CTGF	FMR1	COL1A1	IL18
	E2F1		VCAM1	HIF1A	SP3	MAPK8	
	HIF1A		EPB41L3	HMGCS1	RSF1	ZEB1	
	MMP2			MAPK8			
	SOS1						
	TGFBR2						
	KLF11						
	TXNIP						

**Validated targets
of miRNAs**

TFs	E2F1			NR3C1	FOXM1	FOXO1	KAT2B
	E2F3			RUNX1			
	KAT2B						
	MYC						
	RB1						
	RUNX1						
	STAT3						
Genes	E2F1		HRAS	CTGF		FLNA	CTGF
	MMP2			EDF1		TGFBR2	SMAD4
	MYC			ATM			TGFBR2
	SMAD4			HIF1A			DNMT1
	TGFBR2			HMGCS1			
	HIF1A			SMAD4			
	DNMT1			TGFBR2			
	IGFBP3			DNMT1			

miRNAs	miR-19b-1	miR-200b	miR-200c	miR-20a	miR-22	miR-29a	miR-29b
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**Predicted targets
of miRNAs**

TFs	KLF6	E2F1	FOXO1	ETS1	MYBL2
	SP3	E2F3	HNRNPK	KAT2B	
		EZH2			
		PPARA			
		RB1			
		STAT3			
		KAT2B			
Genes	NCF2	CALD1	CD68		DNMT1
	MAPK8	CCNG2	SOS1		FBN1
	SP3	E2F1	UTRN		EDF1
		HIF1A	VCAM1		
		SMAD4	WRN		
		MMP2			
		MAPK8			
		SOS1			
		TGFBR2			
		KLF11			
		TXNIP			

**Validated targets
of miRNAs**

TFs	KAT2B	E2F3	MYB	E2F1	PPARA	KLF4	SP1
	NR3C1	ETS1	ETS1	E2F3		AHR	STAT3
		MYB	SP1	MYC		MYC	SP1
		SP1	E2F3	RB1			STAT3
	EZH2	FOXO1	RUNX1				MYC
		E2F3	STAT3				
Genes	CTGF	CREB1	FLNA	CCNB1	HIF1A	MMP2	COL1A1
	FMR1	FN1	FN1	E2F1		DNMT1	DNMT1
	ATM	KLF11	KLF11	FLNA		FBN1	MMP2
	HMGCS1	ZEB1	ZEB1	HIF1A		MYC	FBN1
	SMAD4	DNMT1	NCAM1	MYC			MYC
	TGFBR2		CYP1B1	SMAD4			
	DNMT1			TGFBR2			
				DNMT1			

miRNAs	miR-342-3p	miR-630	miR-874	miR-92a-1	let-7a	let-7b	let-7g
Predicted targets of miRNAs							
TFs	E2F3	KLF6	POU2F1		EZH2	EZH2	EZH2
	EZH2	STAT3		POU2F1	POU2F1	POU2F1	POU2F1
				RB1	RB1	RB1	
					TP53	TP53	TP53
Genes	DNMT1		FBN1	COL1A1	FAS	FAS	FAS
ZEB1		FMR1	DACT3	COL1A1	COL1A1	COL1A1	COL1A1
		SIGIRR		IL6	IL6	IL6	
				MAPK8	MAPK8	MAPK8	MAPK8
				UTRN	UTRN	UTRN	UTRN

Validated targets of miRNAs

TFs	FOSL2	SNAI2	PARP1	HDAC1	E2F1	CTCF	MYC
	E2F1	FOXM1	HDAC1	HDGF	EZH2	E2F3	
			STAT3	KAT2B	MYC	EZH2	
			E2F3	MYBL2	NFKB1	MYC	
				NFKB1	PARP1	SOX9	
				STAT3	SP1	SP1	
				HDAC2	STAT3		
				KLF4			
				IKZF1			
Genes	DNMT1	SNAI2		ATM	MYC	BIRC5	MYC
	E2F1			CCNB1	HRAS	CCNB1	FN1
				EPB41L3	E2F1	EPB41L3	
				FLNA	IL6	FLNA	
				HMGCS1		HIF1A	
				SMAD4		HMGCS1	
				TGFBR2		MYC	
				TYMP		UTRN	
				MAPK8		HRAS	
				DNMT1			

miRNAs	let-7i
Predicted targets of miRNAs	
TFs	EZH2
	POU2F1
	RB1
	TP53
Genes	FAS
	CHUK
	COL1A1
	IL6
	MAPK8
	UTRN

Validated targets of miRNAs

H19-STAT3-KLF11

miRNAs	miR-106a	miR-130b-3p	miR-138-5p	miR-139	miR-141	miR-152-3p	miR-152-5p	miR-17
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Predicted targets of miRNAs

TFs	RUNX1	FOSL2	EZH2	ETS1	CREBBP	KLF6	KLF6	RUNX1
	E2F1	MYB	POU2F1		POU2F1	KLF4	E2F3	E2F1
	PPARA	STAT3	RELA				KLF4	RB1
	RB1		SP1					STAT3
	SP1							KAT2B
	STAT3							
	KAT2B							
Genes	CALD1	FMR1	HIF1A	FMR1	FMR1	CHUK	DNMT1	CALD1
	CCNG2	MET	KLF11	MAPK8		DNMT1	FBN1	CCNG2
	E2F1	TGFBR2		ZEB1		FBNI	FMR1	E2F1
	HIF1A					FMR1	SOS1	HIF1A
	SMAD4					SOS1		MMP2
	MMP2							SOS1
	MAPK8							TGFBR2
	SOS1							KLF11
	TGFBR2							TXNIP
	KLF11							
	TXNIP							

Validated targets of miRNAs

TFs	E2F1	AHR	EZH2	NFKB1	E2F3	KLF4		E2F1
	MYB	E2F1	NFKB1		PPARA			E2F3
	RB1	NR3C1	SNAI2		HDGF			KAT2B
	RUNX1	PPARA	SOX9					MYC
	STAT3	RB1	TWIST2					RB1
		STAT3						RUNX1
								STAT3
Genes	E2F1	E2F1	HIF1A	HRAS	ZEB1	DNMT1		E2F1
	FAS	KLF11	SNAI2	MET	EIF4E	ADAM17		MMP2
	TGFBR2	SMAD4			KLF11			MYC
	ATM	TGFBR2			ZEB1			SMAD4
	HIF1A	ZEB1						TGFBR2
		FMR1						HIF1A
		MMP2						DNMT1
								IGFBP3

Validated targets of miRNAs

TFs			NR3C1	FOXM1	FOXO1	KAT2B	KAT2B
		RUNX1				NR3C1	
Genes	HRAS	CTGF		FLNA	CTGF	CTGF	
		EDF1		TGFBR2	SMAD4	FMR1	
		ATM			TGFBR2	ATM	
		HIF1A			DNMT1	HMGCS1	
		HMGCS1				SMAD4	
		SMAD4				TGFBR2	
		TGFBR2				DNMT1	
		DNMT1					

miRNAs	miR-200b	miR-200c	miR-20a	miR-22	miR-29a	miR-29b	miR-342-3p
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**Predicted targets
of miRNAs**

TFs	KLF6		E2F1	FOXO1	ETS1	MYBL2	E2F3
	SP3		E2F3	HNRNPK	KAT2B		
			EZH2				
			PPARA				
			RB1				
			STAT3				
			KAT2B				
Genes	NCF2		CALD1	CD68		DNMT1	DNMT1
	MAPK8		CCNG2	SOS1		FBN1	ZEB1
	SP3		E2F1	UTRN		EDF1	
			HIF1A	VCAM1			
			SMAD4	WRN			
			MMP2				
			MAPK8				
			SOS1				
			TGFBR2				
			KLF11				
			TXNIP				

**Validated targets
of miRNAs**

TFs	E2F3	MYB	E2F1	PPARA	KLF4	SP1	FOSL2
	ETS1	ETS1	E2F3		AHR	STAT3	E2F1
	MYB	SP1	MYC		MYC	SP1	
	SP1	E2F3	RB1			STAT3	
	EZH2	FOXO1	RUNX1			MYC	
		E2F3	STAT3				
Genes	CREB1	FLNA	CCNB1	HIF1A	MMP2	COL1A1	DNMT1
	FN1	FN1	E2F1		DNMT1	DNMT1	E2F1
	KLF11	KLF11	FLNA		FBN1	MMP2	
	ZEB1	ZEB1	HIF1A		MYC	FBN1	
	DNMT1	NCAM1	MYC			MYC	
		CYP1B1	SMAD4				
			TGFBR2				
			DNMT1				

miRNAs	miR-630	miR-874	miR-92a-1	let-7a	let-7b	let-7g	let-7i
Predicted targets of miRNAs							
TFs	KLF6	POU2F1		EZH2	EZH2	EZH2	EZH2
	EZH2	STAT3		POU2F1	POU2F1	POU2F1	POU2F1
				RB1	RB1	RB1	RB1
				TP53	TP53	TP53	TP53
Genes	FBN1	COL1A1	FAS	FAS	FAS	FAS	FAS
	FMR1	DACT3	COL1A1	COL1A1	COL1A1	COL1A1	CHUK
	SIGIRR		IL6	IL6	IL6	IL6	COL1A1
			MAPK8	MAPK8	MAPK8	MAPK8	IL6
			UTRN	UTRN	UTRN	UTRN	MAPK8
							UTRN

Validated targets of miRNAs

TFs	SNAI2	PARP1	HDAC1	E2F1	CTCF	MYC	
	FOXM1	HDAC1	HDGF	EZH2	E2F3		
		STAT3	KAT2B	MYC	EZH2		
		E2F3	MYBL2	NFKB1	MYC		
			NFKB1	PARP1	SOX9		
			STAT3	SP1	SP1		
			HDAC2	STAT3			
			KLF4				
			IKZF1				
Genes	SNAI2		ATM	MYC	BIRC5	MYC	
			CCNB1	HRAS	CCNB1	FN1	
			EPB41L3	E2F1	EPB41L3		
			FLNA	IL6	FLNA		
			HMGCS1		HIF1A		
			SMAD4		HMGCS1		
			TGFBR2		MYC		
			TYMP		UTRN		
			MAPK8		HRAS		
			DNMT1				

H19-NFYB-SP3

miRNAs	miR-106a	miR-130b-3p	miR-138-5p	miR-139	miR-141	miR-152-3p	miR-152-5p
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**Predicted targets
of miRNAs**

TFs	RUNX1	FOSL2	EZH2	ETS1	CREBBP	KLF6	KLF6
E2F1	MYB	POU2F1		POU2F1	KLF4	E2F3	
PPARA	STAT3	RELA				KLF4	
RB1		SP1					
SP1							
STAT3							
KAT2B							
Genes	CALD1	FMR1	HIF1A	FMR1	FMR1	CHUK	DNMT1
CCNG2	MET	KLF11	MAPK8			DNMT1	FBN1
E2F1	TGFBR2		ZEB1			FBN1	FMR1
HIF1A						FMR1	SOS1
SMAD4						SOS1	
MMP2							
MAPK8							
SOS1							
TGFBR2							
KLF11							
TXNIP							

**targets of
miRNAs**

TFs	E2F1	AHR	EZH2	NFKB1	E2F3	KLF4	
MYB	E2F1	NFKB1		PPARA			
RB1	NR3C1	SNAI2		HDGF			
RUNX1	PPARA	SOX9					
STAT3	RB1	TWIST2					
	STAT3						
Genes	E2F1	E2F1	HIF1A	HRAS	ZEB1	DNMT1	
FAS	KLF11	SNAI2	MET	EIF4E	ADAM17		
TGFBR2	SMAD4			KLF11			
ATM	TGFBR2			ZEB1			
HIF1A	ZEB1						
	FMR1						
	MMP2						

miRNAs	miR-17	miR-181d-3p	miR-181d-5p	miR-18a	miR-194-5p	miR-196a	miR-19a
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**Predicted targets
of miRNAs**

TFs	RUNX1		RUNX1	RUNX1	E2F3	ERG	
	E2F1		KLF6	KLF6	FLI1	FLI1	
	RB1		ETS1	NR3C1	SP3		
	STAT3		NR3C1	POU2F1			
	KAT2B		KAT2B				
Genes	CALD1		ATM	ATM	ACP5	FAS	FMR1
	CCNG2		SOS1	CTGF	FMR1	COL1A1	IL18
	E2F1		VCAM1	HIF1A	SP3	MAPK8	
	HIF1A		EPB41L3	HMGCS1	RSF1	ZEB1	
	MMP2			MAPK8			
	SOS1						
	TGFBR2						
	KLF11						
	TXNIP						

**targets of
miRNAs**

TFs	E2F1			NR3C1	FOXM1	FOXO1	KAT2B
	E2F3			RUNX1			
	KAT2B						
	MYC						
	RB1						
	RUNX1						
	STAT3						
Genes	E2F1		HRAS	CTGF		FLNA	CTGF
	MMP2			EDF1		TGFBR2	SMAD4
	MYC			ATM			TGFBR2
	SMAD4			HIF1A			DNMT1
	TGFBR2			HMGCS1			
	HIF1A			SMAD4			
	DNMT1			TGFBR2			
	IGFBP3			DNMT1			

miRNAs	miR-19b-1	miR-200b	miR-200c	miR-20a	miR-22	miR-29a	miR-29b
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**Predicted targets
of miRNAs**

TFs		KLF6		E2F1	FOXO1	ETS1	MYBL2
		SP3		E2F3	HNRNPK	KAT2B	
				EZH2			
				PPARA			
				RB1			
				STAT3			
				KAT2B			
Genes		NCF2		CALD1	CD68		DNMT1
		MAPK8		CCNG2	SOS1		FBN1
		SP3		E2F1	UTRN		EDF1
				HIF1A	VCAM1		
				SMAD4	WRN		
				MMP2			
				MAPK8			
				SOS1			
				TGFBR2			
				KLF11			
				TXNIP			

**targets of
miRNAs**

TFs	KAT2B	E2F3	MYB	E2F1	PPARA	KLF4	SP1
NR3C1	ETS1	ETS1	E2F3		AHR	STAT3	
	MYB	SP1	MYC		MYC	SP1	
	SP1	E2F3	RB1				STAT3
	EZH2	FOXO1	RUNX1				MYC
		E2F3	STAT3				
Genes	CTGF	CREB1	FLNA	CCNB1	HIF1A	MMP2	COL1A1
FMR1	FN1	FN1	E2F1		DNMT1	DNMT1	
ATM	KLF11	KLF11	FLNA		FBN1	MMP2	
HMGCS1	ZEB1	ZEB1	HIF1A		MYC	FBN1	
SMAD4	DNMT1	NCAM1	MYC			MYC	
TGFBR2		CYP1B1	SMAD4				
DNMT1			TGFBR2				
			DNMT1				

miRNAs	miR-342-3p	miR-630	miR-874	miR-92a-1	let-7a	let-7b	let-7g
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Predicted targets

of miRNAs

TFs	E2F3	KLF6	POU2F1		EZH2	EZH2	EZH2
		EZH2	STAT3		POU2F1	POU2F1	POU2F1
					RB1	RB1	RB1
					TP53	TP53	TP53
Genes	DNMT1		FBN1	COL1A1	FAS	FAS	FAS
	ZEB1		FMR1	DACT3	COL1A1	COL1A1	COL1A1
			SIGIRR		IL6	IL6	IL6
					MAPK8	MAPK8	MAPK8
					UTRN	UTRN	UTRN

**targets of
miRNAs**

TFs	FOSL2	SNAI2	PARP1	HDAC1	E2F1	CTCF	MYC
	E2F1	FOXM1	HDAC1	HDGF	EZH2	E2F3	
			STAT3	KAT2B	MYC	EZH2	
			E2F3	MYBL2	NFKB1	MYC	
				NFKB1	PARP1	SOX9	
				STAT3	SP1	SP1	
				HDAC2	STAT3		
				KLF4			
				IKZF1			
Genes	DNMT1	SNAI2		ATM	MYC	BIRC5	MYC
	E2F1			CCNB1	HRAS	CCNB1	FN1
				EPB41L3	E2F1	EPB41L3	
				FLNA	IL6	FLNA	
				HMGCS1		HIF1A	
				SMAD4		HMGCS1	
				TGFBR2		MYC	
				TYMP		UTRN	
				MAPK8		HRAS	
				DNMT1			

miRNAs	let-7i
Predicted targets of miRNAs	
TFs	EZH2
	POU2F1
	RB1
	TP53
Genes	FAS
	CHUK
	COL1A1
	IL6
	MAPK8
	UTRN

targets of miRNAs

Table S6. Primers for qRT-PCR experiment

Gene	Forward sequence	Reverse sequence
GAPDH	5'-CCACTCCTCCACCTTGAC-3'	5'-ACCCTGTTGCTGTAGCCA-3'
H19	5'-GTGGACTTGGTGACGCTGTA-3'	5'-CACCATCCTCCCTCCTGAGA-3'
SP1	5'-TGGCAGCAGTACCAATGGC-3'	5'-CCAGGTAGTCCTGTCAGAACTT-3'
ETS1	5'-GATAGTTGTGATCGCCTCACC-3'	5'-GTCCTCTGAGTCGAAGCTGTC-3'
STAT3	5'-ACCAGCAGTATAAGCCGCTTC-3'	5'-GCCACAATCCGGGCAATCT-3'
TGFBR2	5'-GTAGCTCTGATGAGTGCAATGAC-3'	5'-CAGATATGGCAACTCCCAGTG-3'
KLF11	5'-GTTGCGGATAAGACCCCTCAC-3'	5'-TGGAATCTGTTACTTGGGGAGA-3'