

FIGURE S1 | Promoter of *IbBT4* showing different *cis*-acting regulatory elements associated with abiotic stress responses.

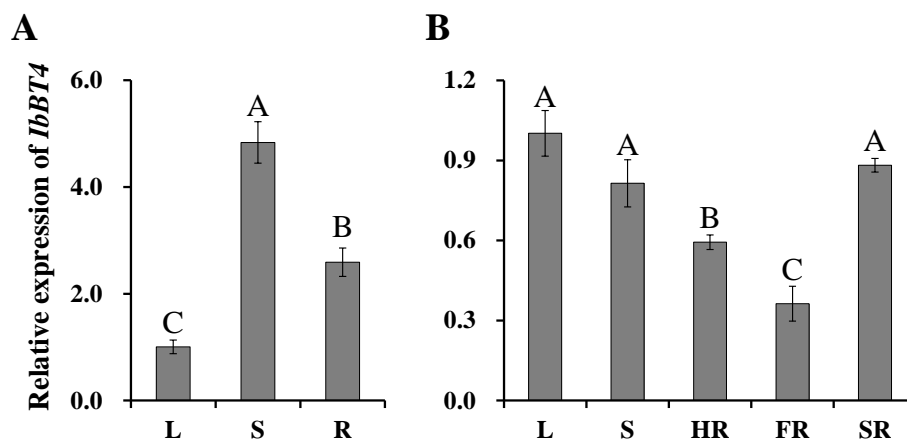


FIGURE S2 | Expression analysis of *IbBT4* in different tissues of Xushu55-2. The data are presented as the means \pm SEs ($n = 3$). The different capital letters indicate a significant difference at $P < 0.01$ according to Student's *t*-test. L: Leaf; S: Stem; HR: Hairy root; FR: Fibrous root; SR: Storage root.

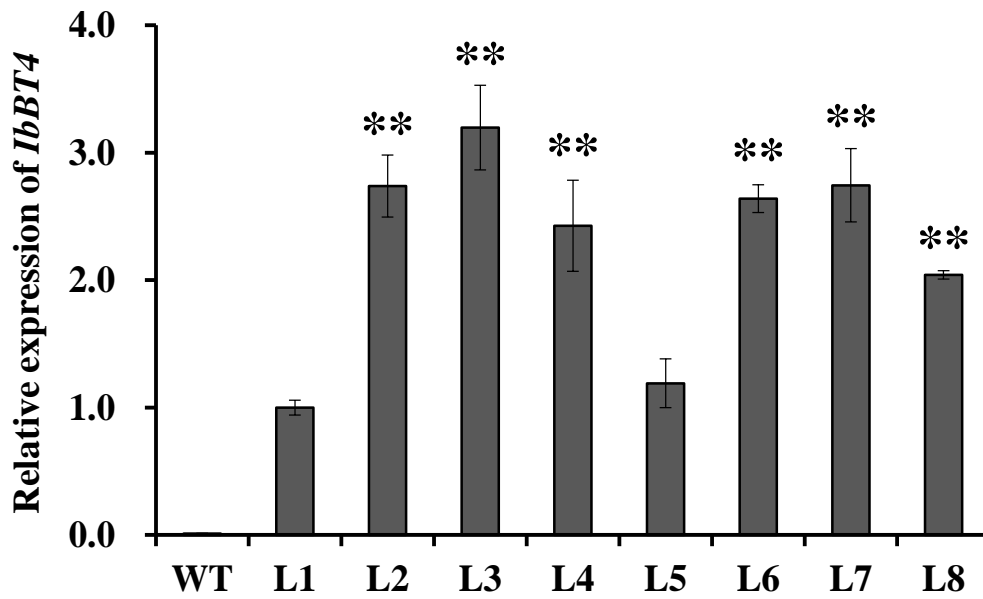


FIGURE S3 | Expression analysis of *IbBT4* in transgenic *Arabidopsis* plants. *Atactin* was used as an internal control. The data are presented as the means \pm SEs (n=3). ** indicates a significant difference from L1 at $P < 0.01$ according to Student's *t*-test.

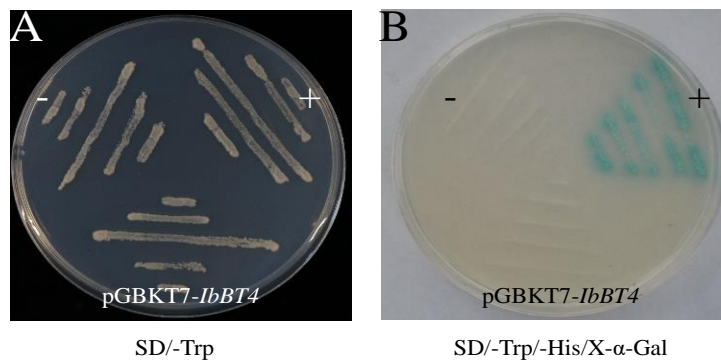


FIGURE S4 | Transactivation activity assay of *IbBT4* in yeast. **(A)** Transformed yeast cells harbouring different expression vectors were drawn onto SD/-Trp media. pBD (-) and pGAL4 (+) were used as negative and positive controls, respectively. **(B)** Transformed yeast cells harbouring different expression vectors were drawn onto SD/-Trp/-His media supplemented with X- α -Gal. pBD (-) and pGAL4 (+) were used as negative and positive controls, respectively.

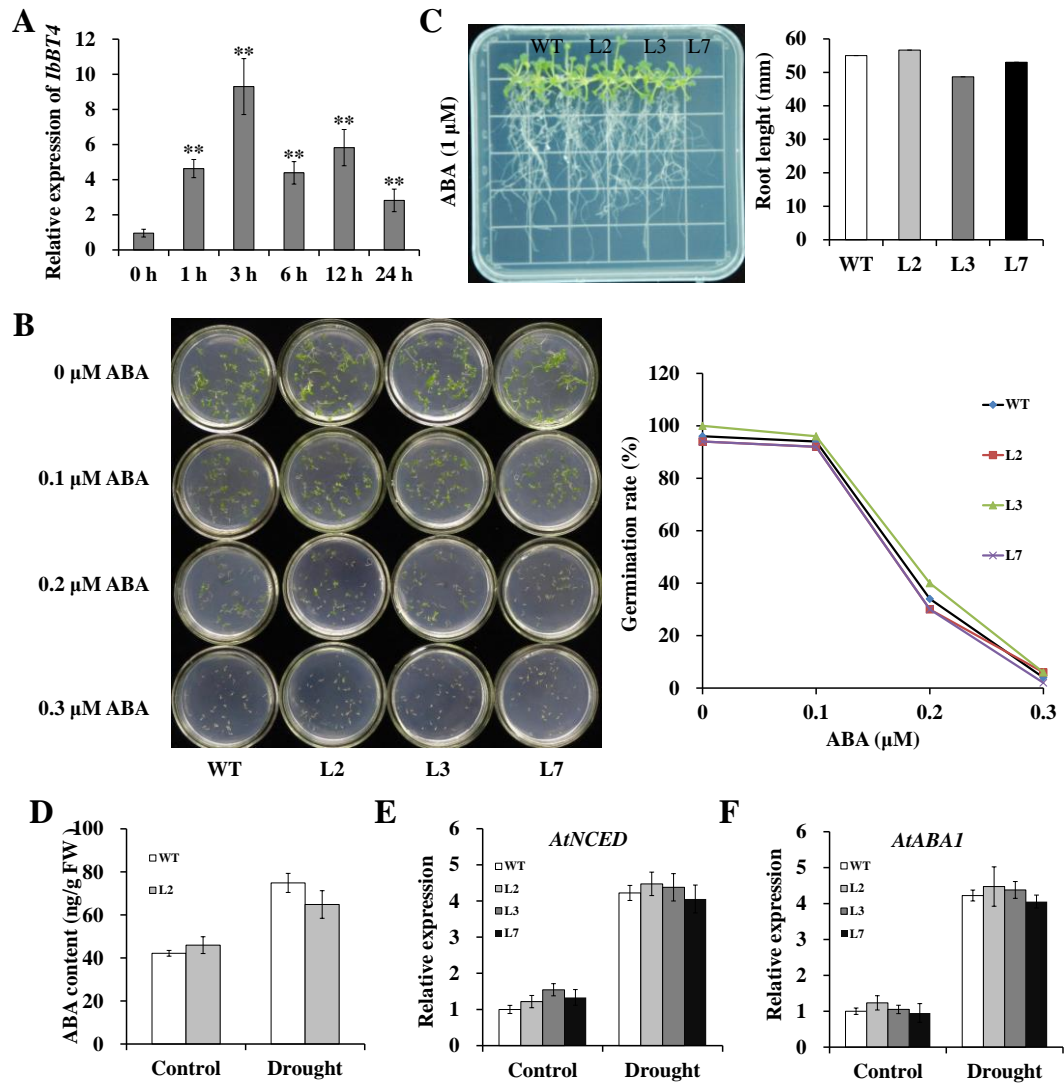


FIGURE S5 | Analysis of the function of *IbBT4* in the ABA signalling pathway. **(A)** Expression analysis of *IbBT4* in *in vitro*-grown Xushu55-2 plants after different time points (h) in response to 100 μ M ABA. **(B)** Responses of transgenic *Arabidopsis* and WT seeds sown on 1/2 MS media with 0, 0.1, 0.2 and 0.3 μ M ABA for 1 week. **(C)** Responses of transgenic *Arabidopsis* and WT seedlings cultured for 2 weeks on 1/2 MS media supplemented with 1 μ M ABA. **(D)** ABA content in the leaves of transgenic *Arabidopsis* and WT plants grown for 2 weeks under normal conditions followed by 2 weeks of drought stress and for 4 weeks under normal conditions (control), respectively. **(E-F)** Transcript levels of ABA-related genes in the leaves of transgenic *Arabidopsis* and WT plants grown for 2 weeks under normal conditions followed by 2 weeks of drought stress and for 4 weeks under normal conditions (control), respectively. The data are presented as the means \pm SEs (n = 3). ** indicates a significant difference at $P < 0.01$ according to Student's *t*-test.

Table S1 Primers used in this study

Primer name	Primer sequence (5'-3')
Primers for 5'/3' RACE	
5GSP1	CGAAGACACACCGAGAACAC
5GSP2	GCGAACAAAAATGGAGACAG
3GSP1	CGAAACTTTAGGCAGCAGG
3GSP2	AGTCCGTTTCCTTCACCGAA
Primers for 5' - promoter region	
GW1	TTACTCTGCTCCGACGATGA
GW2	CATCATAGCGGGAAGAATACA
Pro-F	CTCAACTCCCAAGTCCCATC
Pro-R	CTTCCGATCCTTAAATTTCTGC
Primers for vector construction	
IbBT4-F	ATGGGTAAGCTTTCGGATTC
IbBT4-R	TCATGTTGCTTCAACTGAGAAAAAT
<i>IbBT4</i> -DW-F (<i>Pac</i> I)	CCTTAATTAAATGGGTAAGCTTTCGGATTC
<i>IbBT4</i> -DW-R (<i>Asc</i> I)	<u>GGCGCGCC</u> ATGTTGCTTCAACTGAGAAAAAT
<i>IbBT4</i> -OS-F(<i>Xba</i> I)	GCTCTAGAAATGGGTAAGCTTTCGGATTC
<i>IbBT4</i> -OS-R(<i>Pst</i> I)	AACTGCAGTGTTGCTTCAACTGAGAAAAAT
pBD-F- <i>Nde</i> I	<u>GGAATTC</u> ATGGGTAAGCTTTCGGATTC
pBD-R- <i>Sal</i> II	AACTGCAGTCATGTTGCTTCAACTGAGAAAAAT
<i>IbBEE</i> -AD-F(<i>Nde</i> I)	GGAATTC <u>GAAATTC</u> ATGCTTCGCTGCGCGC
<i>IbBEE</i> -AD-F(<i>Bam</i> H I)	CGGGATCCTCACTTTCCCAACCTTGCAGC
<i>AtBEE</i> -AD-F(<i>Nde</i> I)	GGAATTC <u>GAAATTC</u> ATGGACTTGTCTGTACTTGATA
<i>AtBEE</i> -AD-F(<i>Bam</i> H I)	CGGGATCCTTACTTGAGGCTGAAGAAATTGG
CE-IbBEE-F(<i>Asc</i> I)	<u>GGCGCGCC</u> ATGCTTCGCTGCGCGC
CE-IbBEE-R(<i>Kpn</i> I)	GGGGTACCCTTTCCCAACCTTGCAGC
CE-AtBEE-F(<i>Asc</i> I)	<u>GGCGCGCC</u> ATGGACTTGTCTGTACTTGATAGG
CE-AtBEE-R(<i>Kpn</i> I)	GGGGTACCCTTGAGGCTGAAGAAATTGG
NE-IbBT4-F(<i>Asc</i> I)	<u>GGCGCGCC</u> ATGGGTAAGCTTTCGGATTC
NE-IbBT4-R(<i>Kpn</i> I)	GGGGTACCCTGTTGCTTCAACTGAGAAAAAT
Primers for transformant identification	
pSuper-1300-F	GACGCCATTTCGCTTTTCA

pSuper-1300-R

TGAACTTGTGGCCGTTTACGTC

Primers for qRT-PCR

<i>Ibactin</i> -F	AGCAGCATGAAGATTAAGGTTGTAGCAC
<i>Ibactin</i> -R	TGGAAAATTAGAAGCACTTCCTGTGAAC
<i>IbBT4</i> -F	CCGATTATGAAAGCCATGTTGAG
<i>IbBT4</i> -R	TACGAATGCGACAGCACCAGTAA
<i>Atactin</i> -F	GCACCCTGTTCTTCTTACCGA
<i>Atactin</i> -R	AGTAAGGTCACGTCCAGCAAGG
<i>AtDWF4</i> -F	CCGTACACCGCCACAA
<i>AtDWF4</i> -R	GAATCTATTAAGTCCAGCATCAG
<i>AtCPD</i> -F	GCTGATCGGAGCTTACAAAAC
<i>AtCPD</i> -R	AAATCGTCGGTTCACCAAAA
<i>AtDET2</i> -F	CACCAACCGCCGTCCTT
<i>AtDET2</i> -R	CGGTGGAGATACGGTGGGAC
<i>AtROT3</i> -F	AACTTCATCGCTTGTGGTTATT
<i>AtROT3</i> -R	TTGGTGTCCCTATTATGTTCGT
<i>AtCYP90D1</i> -F	TTTATCATCATCGTCATCTCA
<i>AtCYP90D1</i> -R	TTTGGTCCGTGACTCTGG
<i>AtBR6ox1</i> -F	AAACCAAAGACTCCGATACGG
<i>AtBR6ox1</i> -R	CGATTGTGGGTAACCAGGAA
<i>AtBR6ox2</i> -F	ACCAAAGACTAAGATATGGGAGT
<i>AtBR6ox2</i> -R	AAGCATAGATTGCGGGTAA
<i>AtBR11</i> -F	CTCTCCTGTCTCTCACCGGA
<i>AtBR11</i> -R	GCACCGGAGATTGAATTCGC
<i>AtBIN2</i> -F	AGATGCCTGCTGCTGTAGTTG
<i>AtBIN2</i> -R	CCACGGTTTCTCCAGTCTCC
<i>AtBZR1</i> -F	ATGGTGGCATTCTTCTTCTC
<i>AtBZR1</i> -R	GCAACGGTTTCGGGTTCTT
<i>AtBES1</i> -F	CCCAAACCATTGCCTACTTG
<i>AtBES1</i> -R	GGTGCAGACACCGCATAAAA
<i>AtBR6ox1</i> -F	AAACCAAAGACTCCGATACGG
<i>AtBR6ox1</i> -R	CGATTGTGGGTAACCAGGAA
<i>AtBR6ox2</i> -F	ACCAAAGACTAAGATATGGGAGT

<i>AtBR6ox2</i> -R	AAGCATAGATTGCGGGTAA
<i>AtSOD</i> -F	ATGAGAAGTTCTATGAAGAG
<i>AtSOD</i> -R	GTCTTTATGTAATCTGGT
<i>AtGPX</i> -F	ATGGCGACGAAGGAACCAG
<i>AtGPX</i> -R	ATCGCCGAAGATTCCCCATT
<i>AtPOD</i> -F	TCCGGGAGCACACCATTGG
<i>AtPOD</i> -R	TGGTCGGAATTCAACAG
<i>AtCAT</i> -F	GCAACTACCCCCGAGTGGAAA
<i>AtCAT</i> -R	TG TTCAGAACCAAGCGACCA
<i>AtNCED</i> -F	AGAAGCAGGGCAAATAACAAG
<i>AtNCED</i> -R	CCGTCGCCGTACCTAAACTC
<i>AtABA1</i> -F	TACTTGGGGTAAAGGGCGTG
<i>AtABA1</i> -R	CAAAGCACCCCTGCAATAACT
<i>IbBEE</i> -F	GAGGAAGATAAGAAATGGGAAGGAGA
<i>IbBEE</i> -R	ATGGCTGTCGGTGGCTTGG
