



**UMR\_S 1155**

Unité mixte Inserm-Université

**MALADIES RENALES FREQUENTES ET RARES:  
DES MECANISMES MOLECULAIRES A LA MEDECINE PERSONNALISEE**

**Directeur : Dr CHRISTOS CHATZIANTONIOU**

Paris, the 8<sup>th</sup> of April 2020,

**Object: Review for PONE-D-19-36071**

Dear Prof. Dussaule,

The manuscript PONE-D-19-36071, entitled ‘Sodium–calcium exchanger 1 is the key molecule for urinary potassium excretion against acute hyperkalemia’, by Shoda and co-authors, describes the characterisation of the crucial role played by the Na-Ca exchanger NCX1 in the regulation of the activity of the Na-Cl cotransporter NCC in the distal nephron during a K-load.

The paper is well written. The data are solid and support the hypotheses and conclusions drawn. I have a few comments, which you will find below. Therefore, I recommend that the paper should be revised, with major revisions.

Best regards,

Juliette Hadchouel, VetMD, PhD

## Review for PONE-D-19-36071

The manuscript PONE-D-19-36071, entitled ‘Sodium–calcium exchanger 1 is the key molecule for urinary potassium excretion against acute hyperkalemia’, by Shoda and co-authors, describes the characterisation of the crucial role played by the Na-Ca exchanger NCX1 in the regulation of the activity of the Na-Cl cotransporter NCC in the distal nephron during a K-load.

Overall, the results described here support the hypotheses and the conclusions drawn. However, I have the following comments.

### Major comments:

**Material and Methods** – Animal experiment : why were the kidneys collected 15 minutes after the oral K gavage for protein extraction? The metabolic data were collected every 30 minutes after the oral gavage. Why is it different?

### Results :

(1) Figure 1 : Calcineurin is essential for K<sup>+</sup>-induced NCC dephosphorylation

- I don't understand the result and relevance of the experiment describing the consequence of the overexpression of CaN-A on NCC phosphorylation. How could CaN-A work without CaN-B, which is barely expressed in HEK293 cells, as explained by the authors in the next paragraph?

Conversely, why did the authors overexpress CaN-A when transfecting CaN-B is sufficient to induce NCC dephosphorylation (Fig 1B)?

(2) Figure 2D: why did the authors quantify pNCC-S71 in kidney slices and mice rather than NCC-p53, used *in vitro*?

In addition, the figures would be easier to read if the system in which the experiments are conducted were indicated on the figure.

(3) Figure S11: why did the authors quantify the pNCC/tNCC ratio rather than pNCC and tNCC separately like in the other experiments?

(4) Table 2: ‘Blood K<sup>+</sup> level at 120 min following a high-K<sup>+</sup> load was significantly higher in SEA0400-treated mice than in vehicle-treated counterparts (Table 2)’.

In their previous study, using tacrolimus to inhibit calcineurin, the authors did not observe a change in plasma potassium in tacrolimus-treated animals even though NCC dephosphorylation was inhibited. Could the authors comment on the difference in plasma K between tacrolimus- and SEA0400-treated animals?

(5) Have the authors studied the role of NCX1 after a longer exposure to high K ?

**Minor comments:**

- Abstract : the authors should remove the following sentence, which is not at the correct place :  
'The mice were housed in metabolic cages for urine sample collection'.

- Introduction, page 6 : the reference's number in the following sentence is incorrect: 'In our previous study, we observed that the CaN inhibitor, tacrolimus, ...urinary K<sup>+</sup> excretion in the acute phase[22]'. The authors should check all the other references' number.

- Results: Figures 3 and 5 should be merged.