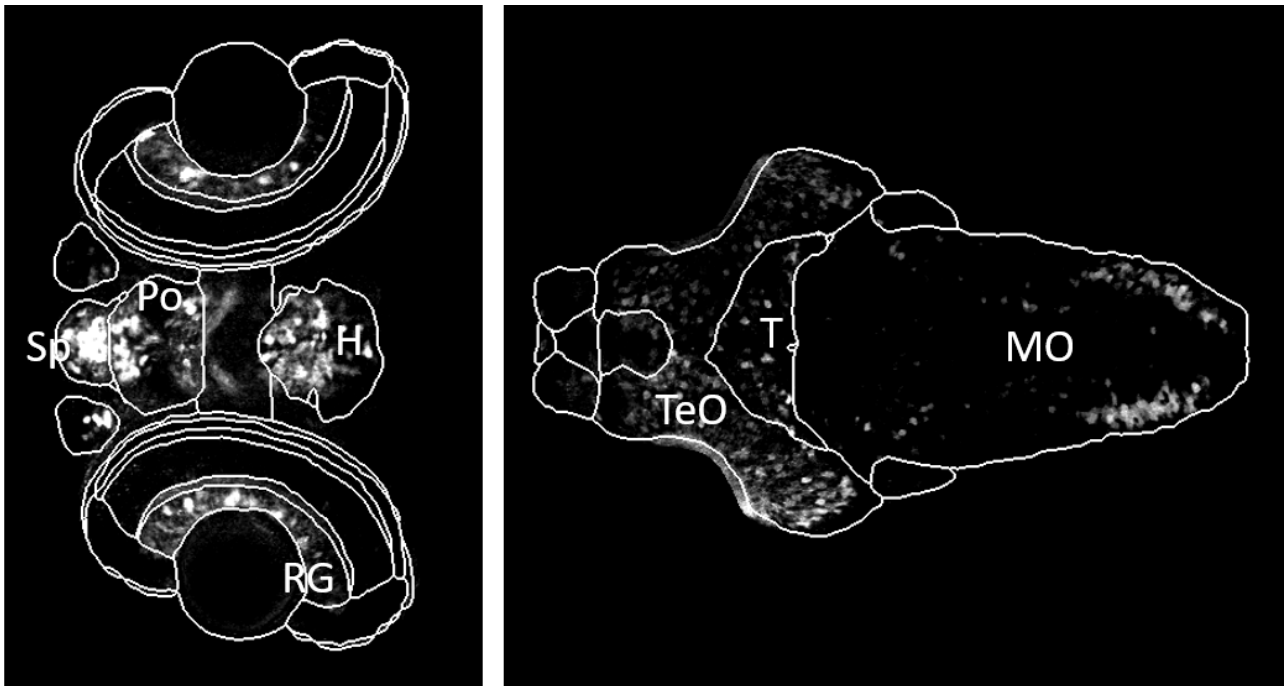


Fig. S1: Stat3 reporter is expressed in erythroid progenitor cells. A-A'': Confocal lateral view of haematopoietic tissue of double transgenic embryos obtained crossing the *Tg(7xStat3:EGFP)* (A) with *Tg(gata1a:DsRed)* (A'').



- 77 Tectum opticum (TeO)
- 97 Medulla oblongata (MO)
- 81 tegmentum (T)
- 7 subpallium (Sp)
- 45 preoptic region (Po)
- 48 hypothalamus (H)
- 59 retinal layer (RG)

Fig. S2: *Tg(7xStat3:EGFP)* reporter expression in the brain of 72-hpf larvae. A-B: Single planes of *Tg(7xStat3:EGFP)* brain at 72 hpf. Images have been obtained with VIBEZ-Z software. H= hypothalamus, Sp= subpallium, Po= preoptic region, RG= retinal layer, TeO= tectum opticum, MO=medulla oblongata, T= tegumentum.

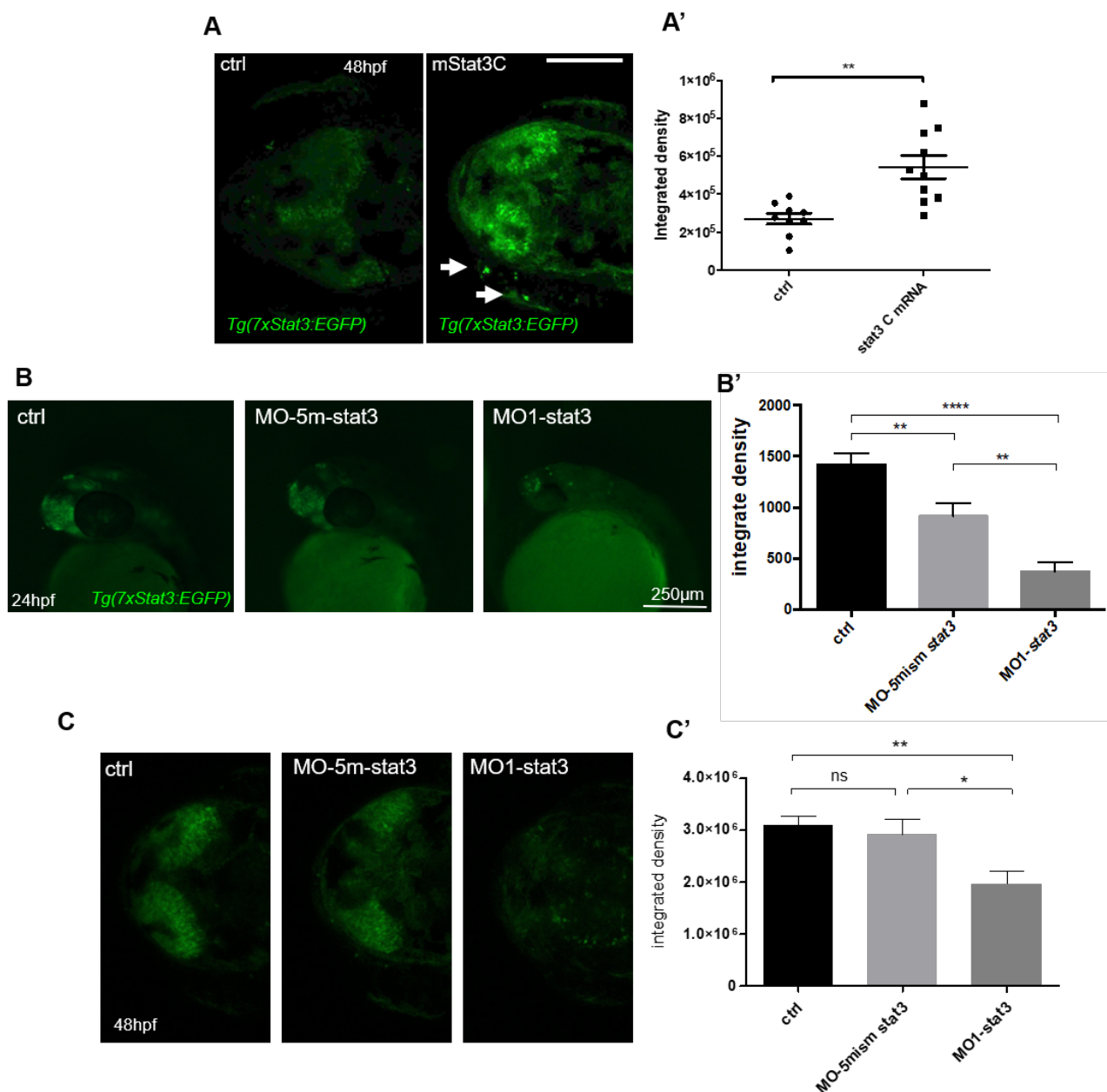


Fig. S3: *Tg(7xStat3:EGFP)* reporter line respond to silencing and overexpression of *stat3*. A: Dorsal view live image of the head of *Tg(7xStat3:EGFP)* controls (left) or *mStat3C* mRNA injected embryos (right) at 48hpf. Arrowheads highlight ectopic fluorescent signal. Scale bar: 100µm. A': EGFP fluorescence quantification in the TeO of *mStat3C* injected and control larvae at 48 hpf. Statistical analysis was performed by unpaired t-test (n=20); **p<0.01 B: representative pictures of 24-hpf *Tg(7xStat3:EGFP)* injected with *stat3*-MO1 (Liu *et al.*, 2017; Miyagi *et al.*, 2004; Yamashita *et al.*, 2002) 5-mismatch morpholino B': Quantification of EGFP fluorescence in 24-hpf embryos injected with *stat3*-MO1 morpholino and 5-mismatch morpholino on the reporter line. Scale bar=250µm, statistical analysis was performed by unpaired t-test on 3 biological replicates. **p<0.01, ****p<0.0001; error bars=SEM. C: dorsal view of 48-hpf *Tg(7xStat3:EGFP)* injected with *stat3*-MO1

and 5-mismatch morpholino. C': Quantification of EGFP fluorescence in 48-hpf *Tg(7xStat3:EGFP)* embryos injected with stat3-MO1 morpholino and 5-mismatch morpholino. * $p < 0.05$, ** $p < 0.01$; error bars=SEM

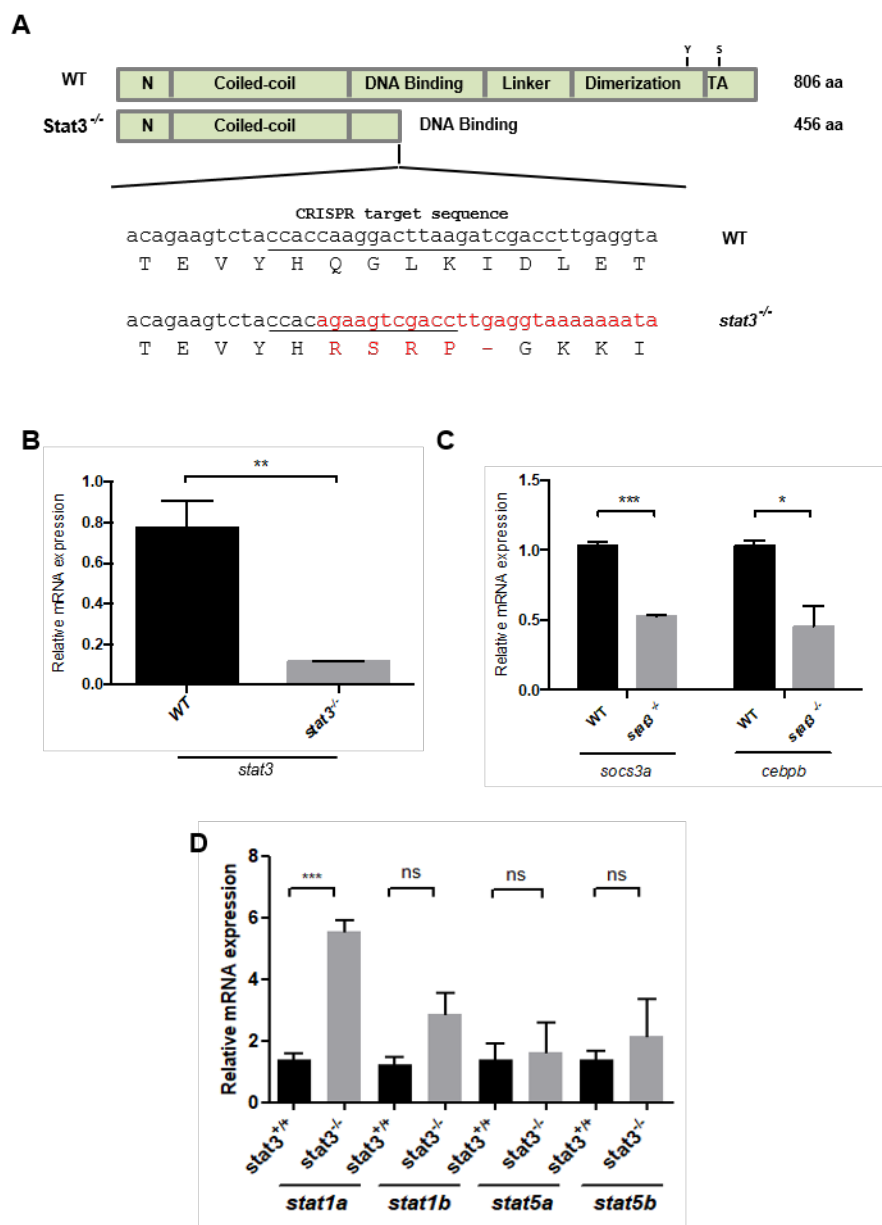


Fig. S4: *stat3^{ia23}* mutant validation. A: Schematic representation of *stat3^{-/-}* mutant allele in comparison with WT. B: qRT-PCR analysis of *stat3* mRNA expression normalized on *gapdh* from WT and *stat3^{-/-}* siblings at 6 dpf (p-value= 0,0073). C: qRT-PCR analysis of *socs3a* and *cebpb* Stat3 targets expression normalized on *gapdh* from WT and *stat3^{-/-}* siblings at 6 dpf, *zgapdh* was used as internal control (p-values= 0,001; 0,0205); Statistical analysis was performed by unpaired t-test on 3 independent biological samples. **p<0,01; error bars=SEM. D: qRT-PCR analysis of *stat1a*, *stat1b*, *stat5a* and *stat5b* mRNA expression levels normalized on *gapdh* from *stat3^{+/+}* and *stat3^{-/-}* siblings at 6 dpf (**p<0.001)

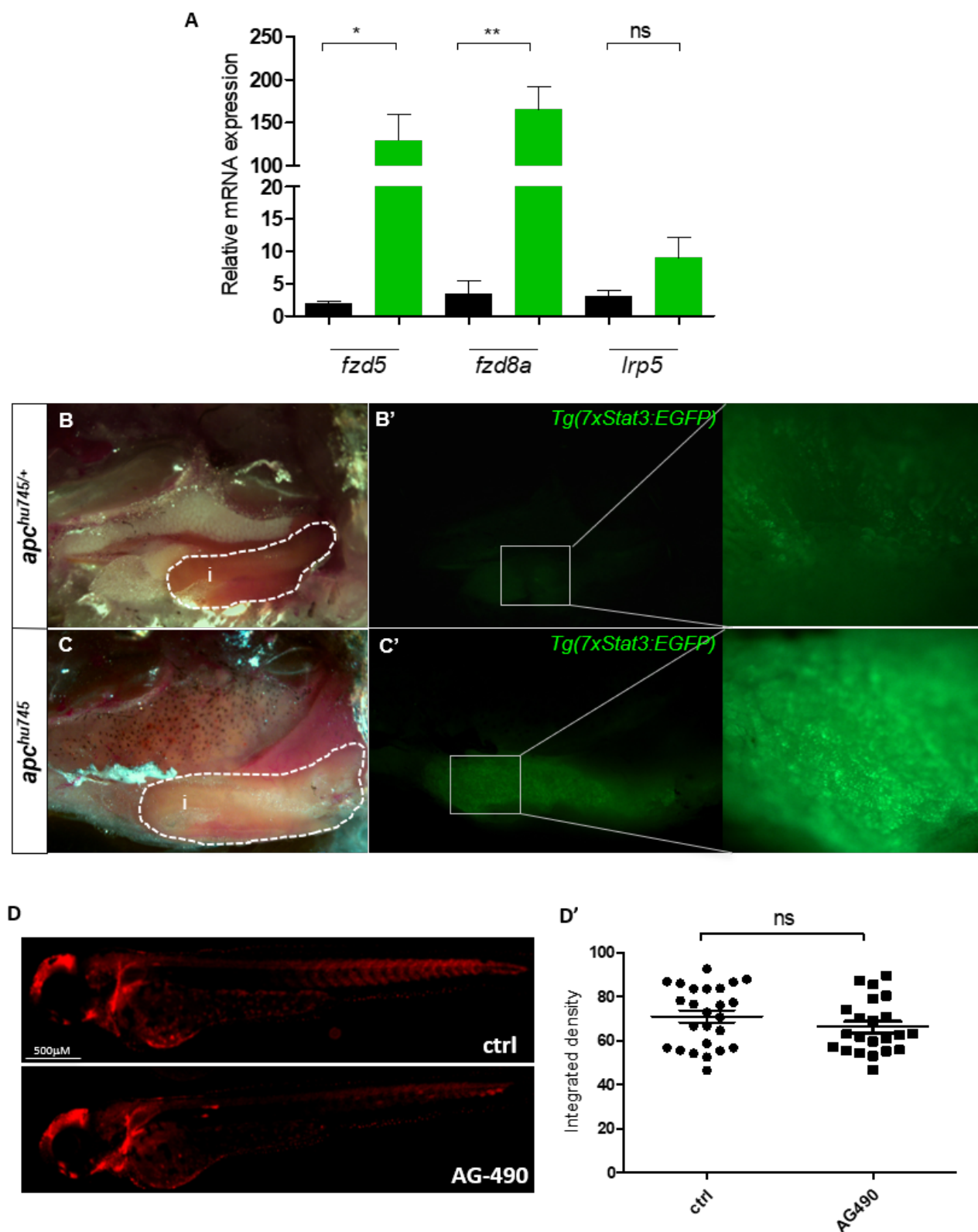


Fig. S5: Stat3 pathway is strongly activated in zebrafish *apc*-driven tumor. A: qRT-PCR analysis of *fzd5*, *fzd8a* and *lrp5* on EGFP-positive and EGFP-negative cells sorted from adult intestines. B-C'': *In vivo* EGFP expression is ectopic in 12 mpf *Tg(7xStat3:EGFP)/apc^{hu745}* hyperplastic intestine (B-B') with respect to *Tg(7xStat3:EGFP)/apc^{hu745/+}* siblings (B-B'). C-C': representative pictures of 3-dpf

Tg(7xTCF-Xla.Siam:nlsmCherry)^{ia5} reporter larvae treated with DMSO and 80 μ M AG-490 from 8-72 hpf (D); quantification of *Tg(7xTCF-Xla.Siam:nlsmCherry)^{ia5}* reporter larvae fluorescence (D'). Statistical analysis was performed by unpaired t-test. ns=not significant.

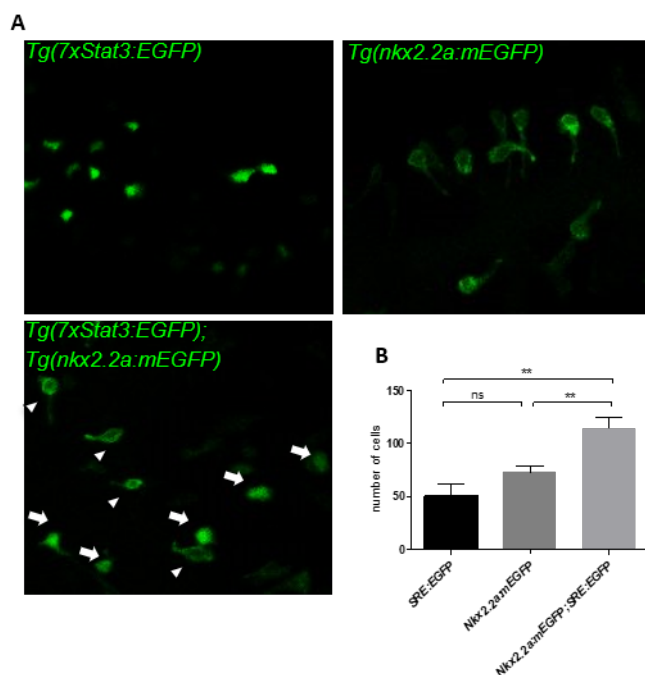


Fig. S6: EGFP positive cells of *Tg(7xStat3:EGFP)* zebrafish line are not secretory cells. A: representative pictures of *Tg(7xStat3:EGFP)*, *Tg(nkx2.2a:mEGFP)* and *Tg(7xStat3:EGFP)/Tg(nkx2.2a:mEGFP)* 6-dpf larvae intestine. B: number of EGFP-positive cells measured in intestines of *Tg(7xStat3:EGFP)*, *Tg(nkx2.2a:mEGFP)* and *Tg(7xStat3:EGFP)/Tg(nkx2.2a:mEGFP)* 6-dpf. Arrows indicate *Tg(nkx2.2a:mEGFP)* positive cells; arrowheads indicate *Tg(7xStat3:EGFP)* positive cells. ** $p < 0.01$, ns=not significant; error bars=SEM.

Gene	Forward primer sequence	Reverse primer sequence
<i>zstat3</i>	TGCCACCAACATCCTAGTGT	GCTTGTTTGCACCTTTTGACTGA
<i>zgapdh</i>	GTGGAGTCTACTGGTGTCTTC	GTGCAGGAGGCATTGCTTACA
<i>zcebpb</i>	CCAAAAGTAACGGGCGACAC	ATCTTCCCTTACCTGACGGC
<i>zsocs3</i>	GGAAGACAAGAGCCGAGACT	GCGATACACACCAAACCCTG
<i>egfp</i>	ACGTAAACGGCCACAAGTTC	AAGTCGTGCTGCTTCATGTG
<i>zstat1a</i>	GCAGCTCAAGAACTCCTGG	AAAGGTCTCTGCAGTTGGGT
<i>zstat1b</i>	CGAGTGGAAGAAGAGACAGC	GCTGGCCCCTTCTAGATTT
<i>zstat5a</i>	TGACCCGAGAAGCTAACACC	GTATGTCCAGTCCTCCCT
<i>zstat5b</i>	TGAGGAAACAGCAAACCGTG	GCTGCTGAGTCAAGTGTTCA
<i>zil6</i>	CGTAAAGAGTCTCCTTGGCG	GGTTTGAGGAGAGGAGTGCT
<i>zgp130</i>	TGCTGGAGTGGGTGAATGAA	GGCTTGGTTACTGGTGTTC
<i>zjak2a</i>	CTTCGAGAGTCAGGAGCCC	CTGAAGCTTCTTCACCGCC
<i>zjak2b</i>	ACGTATTGTGATTCGCGGA	ACAAAAGACAAGGCCTGCAT
<i>zsox9b</i>	CTCGGCAAACCTCTGGAGACT	GCGCATTGGTGGAGATCTG
<i>zagr2</i>	GCACAGACATACGAGGAAGC	GGAGACAAGTGCTTATCTGTG
<i>zpept1</i>	GATTGCTTTGGGAACAGGAGG	GATGGGTGTGATGAGAGTGG
<i>zfabp2</i>	GCTGCCCATGACAACCTG	CGTGTCTCCCTCTATGACC
<i>znotch2</i>	GACGAATGCATCTCCAGTGC	GCAGCAGCCACAGCAACC
<i>zpcna</i>	CCTTGCCACTGGTCTTTGAA	GGCACACGAGATCATGACAG
<i>zcyclinD1</i>	CCAACTTCTCTCGCAAGTC	TGGTCTCTGTGGAGATGTGC
<i>zfzd5</i>	CCTAACTGTGCACTGCCTTG	ATTTGAAGCGCTCCATGTGC
<i>zfzd8a</i>	TGCAATCGGGAGTATGACGT	CTCGTTTCCCCACTTCATGC
<i>zlrp5</i>	TTCTCGGAGGGCCTGATTTT	TTGTCTCCGAGTCAGTCCAG

Tab. 1: list of primer used for Real Time qPCR.