Tumor microenvironment remodeling enables bypass of oncogenic KRAS dependency in pancreatic cancer

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Supplementary Figures S1-S9

Supplementary Figure 1



Supplementary Figure 1. Epigenetic library screening to bypass KRAS* dependency. A, KRAS*-independent escaper tumor numbers generated from 10 mice with orthotopically transplanted iKPC cells for each pool. Fifteen red color highlighted pools promoted the generation of more than 5 escapers and were further validated for enriched ORF expression. **B**, Validation of ORF enrichment in escaper tumors. Gene expression in escapers from 15 highlighted pools in (A) were analyzed by qRT-PCR. ORFs with gene expression levels higher in escapers than in "input" iKPC cells were considered "enriched". The top 10 ORFs enriched in more than 5 escaper tumors are highlighted. **C**, Validation of the overexpressed top 10 gene candidates in iKPC-1 cells by western blot analysis. **D**, Distribution of the top 10 ORFs in screening sub-pools. **E**, BLI imaging

of nude mice orthotopically transplanted with GFP-, HDAC5- or HDAC5D-overexpressed (OE) iKPC-3 cells with luciferase reporter. **F**, Mutation of *HDAC5* (HDAC5D) interrupted HDAC5-HDAC3 interaction to form functional repressive complex by co-IP analysis. **G**, The capability of HDAC family members to bypass KRAS* dependency extracted from ORF screening.



Supplementary Figure 2. Characterization of *HDAC5* escapers. A, Validation of gene expression of endogenous *Kras*, transgenic KRAS*, *Yap1*, and transgenic *HDAC5* (Tg-HDAC5) by qRT-PCR in *HDAC5* escaper cells. KRAS*-reactivated escaper cells and *Yap1*-amplified escaper cells were served as positive controls. Data are represented as mean \pm SD. **B**,

Determination of RAS activity in primary iKPC cells, *HDAC5* escapers, KRAS* on and off iKPC-5 cells, and iKPC-5 cells after inhibition of KRAS* downstream pathways. Active RAS was pulled down by agarose beads crosslinked with Ras-binding domain (RBD) of Raf1, and detected by (H+K) RAS antibody. **C**, Validation of KRAS* downstream signaling pathways in *HDAC5* escaper cells by western blot analysis. **D-E**, The 3-D colony formation assay of GFP-, HDAC5or HDAC5D-OE iKPC-1 cells after KRAS* extinction in MethoCult (D) or soft agar (E) culture under normoxia or hypoxia conditions. KRAS*-expressing cells were used as positive control. **F**, Cell cycle analysis of iKPC-1 cells with or without KRAS* expression in Matrigel culture *in vitro*. The iKPC-1 cells overexpressing GFP, HDAC5 or HDAC5D were seeded in Matrigel with or without DOX treatment, and collected after 4 days for propidium iodide staining. Three independent experiments were performed for statistical analysis. Two-tailed unpaired t tests were performed to calculate the p values.



Supplementary Figure 3. Activation of the TGFβ pathway promotes pancreatic cancer cells to bypass KRAS* dependency. A, Summary of 18 candidate receptors, corresponding ligands and small molecule activators/cytokines. **B,** Experimental design and summary of the screening results of the 13 small molecule activators or cytokines to bypass KRAS* dependency in iKPC-3

cells. Colonies were counted at Day 9 after KRAS* extinction. C, TGF β 1 concentration in 5 mouse plasma samples. D, Validation of *Hdac5* knockout in iKPC-5 cells by western blot analysis. E, TGFβ1 (0.5 ng/ml) drove KRAS*-independent colony growth after KRAS* extinction in Hdac5 wildtype and *Hdac5* knockout iKPC cells. Images were taken at Day 8 after KRAS* extinction. F, TGF^{β1} (0.5 ng/ml) drove KRAS*-independent colony growth after KRAS* extinction in iKPC-1 cells. Images were taken at Day 5 after KRAS* extinction. G, TGF^β treatment attenuated colony growth of KRAS*-expressing iKPC-5 cells in 3-D culture. H, Activation of pSMAD2/3 in iKPC cells after TGF^{β1} treatment at indicated concentrations by western blot analysis. I, TGF^{β2} and TGF β 3 drove iKPC-3 cells to bypass KRAS* dependency in Matrigel culture (n = 3). Colonies were counted at Day 9 after KRAS* extinction. J, TGF^{β1} promoted MEK inhibition (Trametinib, 50 nM)-resistant iKPC-3 colony growth at Day 15 (n = 3). Representative images and colony number quantification are shown. K, Another independent experiment showing that neutralization of TGF_β impaired KRAS*-independent tumor growth of HDAC5-OE iKPC-5 cells subcutaneously transplanted in nude mice (n = 3). L, IHC staining of pSMAD3 in tumors from isotype control group and TGF β neutralizing antibody treatment group related to Fig. 2F. For I. J and K, data are represented as mean \pm SEM, and two-tailed unpaired t tests were performed to calculate the p values.

Supplementary Figure 4



Supplementary Figure 4. Necessity of the canonical TGF β pathway to promote pancreatic cancer cells to bypass KRAS* dependency. A and B, Validation of knockdown efficiency of *Smad2*, *Smad3* and *Smad4* shRNAs in iKPC-1 cells by qRT-PCR (A) and western blot (B) analysis. C, Representative images of TGF β 1-driven KRAS*-independent colony formation comparing scramble control and knockdown of *Smad2*, *Smad3* and *Smad4*. D, Gene signatures enriched in iKPC cells treated with TGF β 1 versus cells treated with vehicle control 5 days after KRAS* extinction in 3-D culture by GSEA analysis of RNA-seq data (n = 3 for each group). E, TGF β promotes MIA PaCa-2 cells to get resistant to KRAS^{G12C} inhibitor ARS-1620 treatment *in vitro*. Data are represented as mean \pm SD. F, Validation of SMAD4 knockout in human MIA PaCa-2 cells by western blot analysis. For A and E, data are represented as mean \pm SD, and two-tailed unpaired t tests were performed to calculate the p values.



Supplementary Figure 5. Characterization of tumor-infiltrated immune cells. A, Expression distribution of lineage marker genes by tSNE plot analysis of CyTOF data related to Fig. 3A. B and C, Gates of macrophages, monocytes and neutrophils in a representative iKPC tumor and a HDAC5-driven escaper by CyTOF (B) and FACS (C) analysis. Data are displayed by FlowJo. CD45⁺CD11b⁺F4/80⁺Ly6C⁻ represented macrophages; CD45⁺CD11b⁺F4/80⁻ cells Ly6G^{high}Ly6C^{low} cells represented neutrophils. CD45⁺CD11b⁺F4/80⁻Ly6G^{low}Ly6C^{high} cells represented monocytes. D-E, Percentage of S100A8⁺ cells in myeloid cells comparing iKPC tumors and HDAC5 escapers by CyTOF (D) and FACS (E) analysis. F and G, Percentage of different S100A8⁺ myeloid cell types in iKPC tumors and HDAC5 escapers by CyTOF (F) and FACS (G) analysis. H, Comparison of F4/80⁺ cell numbers by IHC staining in different HDAC5 escaper tumors (left) or iKPC primary tumors (middle) generated from subcutaneous and orthotopic allograft models; comparison of S100A8⁺ cell numbers after IHC staining in different HDAC5 escapers (right) generated from subcutaneous and orthotopic allograft models. Two-tailed unpaired t tests between the orthotopic and subcutaneous (subQ) groups including all the samples were performed to calculate the p values. I, RNA expression of Csf1 and Csf2 in iKPC cells (n = 3) and HDAC5 escaper cells (n = 5) by qRT-PCR analysis. J, Gates of the MHC II⁺ cells in bone marrow (BM) myeloid cells and in macrophages from an iKPC-3 tumor and a HDAC5 escaper by FlowJo analysis of the FACS data. K, Gates of tissue-resident and HSC-derived macrophages in a representative iKPC-5 tumor and a HDAC5 escaper by FACS analysis. Data are displayed by FlowJo. CXCR4⁺CCR2⁺ macrophages represented tissue-resident TAMs; CXCR4⁻CCR2⁺ macrophages cells represented HSC-derived TAMs. L, Quantification of the tissue-resident, HSCderived and other tumor associated macrophages (TAMs) in iKPC-5 primary tumors and HDAC5 escapers from orthotopic allograft mouse model in nude mice. M, Representative overlaid histograms of TGFB1-164Dy intensity distribution in S100A8⁺ and S100A8⁻ macrophages from HDAC5 escapers and primary tumors, which were generated from subcutaneous allograft models in nude mice. B cells expressed low TGFB1 in our models, so we used the histogram of TGFB1-164Dy intensity distribution in B cells from spleen as the control for low TGFB1 expressing cells. N, The quantification of TGFB1-164Dy median intensities in S100A8⁺ and S100A8⁻ macrophages from HDAC5 escapers and primary tumors, which were generated from subcutaneous allograft models in nude mice. O, Representative overlaid histograms of TGFB1-PE intensity distribution in S100A8⁺ and S100A8⁻ macrophages from HDAC5 escapers and primary tumors, which were generated from orthotopic allograft models in nude mice. Unstained cells were used as negative

TGFB1 control. **P**, The quantification of TGFB1-PE median intensities in S100A8⁺ and S100A8⁺ macrophages from HDAC5 escapers and primary tumors, which were generated from orthotopic allograft models in nude mice. For **D**, **E** and **L**, data are represented as mean \pm SEM. For **I**, data are represented as mean \pm SD. For **D**-**G**, **I**, **N** and **P**, two-tailed unpaired t tests were performed to calculate the p values.

Supplementary Figure 6



Supplementary Figure 6. Overexpression of *HDAC5* in iKPC cells promotes macrophage infiltration via CCL2/CCR2 axis. A, Representative images of migrated macrophages in transwell assay quantified in Fig. 4C and 4D. **B**, Tumor growth analysis comparing iKPC-1 cells overexpressing GFP w/o Doxy feeding and *Ccl2* w/o Doxy feeding in subcutaneous allograft model in nude mice (n=5 for each group). Two-tailed unpaired t tests were performed to calculate the p values. C, Validation of gene expression of endogenous *Kras*, transgenic KRAS*, *Yap1*, and *Ccl2* by qRT-PCR in *Ccl2* escapers. The KRAS*-expressing iKPC-1 tumor and iKPC-1 tumor after KRAS* extinction for 3 days were served as positive and negative controls, respectively. Data are represented as mean \pm SD. **D**, Validation of KRAS* downstream signaling pathways in *Ccl2* escaper cells by western blot analysis. **E**, Related to Fig. 4I, CCR2 inhibitor RS 504393 (RS) and CCL2 neutralizing antibody (CCL2 Ab) attenuated macrophage infiltration into tumors by IHC analysis of F4/80 compared to vehicle control (VEH); TGFBR inhibitor Galunisertib (GAL) inhibited TGF β pathway activation in tumor cells by IHC analysis of pSMAD3 compared to vehicle control (VEH).

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Supplementary Figure 7. *HDAC5* regulates expression of macrophage-recruiting chemokines through *Socs3*. A, Exploration of *HDAC5* targets by overlapping 3 profiling datasets as described in Fig. 5A. Seventeen candidate genes were filtered out and ranked by p-values in the 2 RNA-seq datasets from low to high. **B**, Gene expression of neutrophil- and macrophage-attracted chemokines after knockdown of *Zfp36* in iKPC cells. Data are represented as mean \pm SD, and two-tailed unpaired t tests were performed to calculate the p values. **C**, Identification of potential HDAC5 interactors by co-IP/MS analysis. **D**, Schematic model of the potential HDAC5 co-repressor complex. **E** and **F**, Validation of knockdown efficiency of *Nfix* (E) and *Mef2d* (F) using CRISPR/Cas9 in HDAC5 escaper cells by western blot analysis.



Supplementary Figure 8. HDAC5 is upregulated after inhibition of the KRAS*/MAPK pathway in *de novo* generated escapers and human PDAC lines. A, Hdac expression in iKPC tumors, *de novo* generated KRAS* negative escapers and KRAS* reactivated escapers. "1", "2" and "3" are representing "iKPC cells", "KRAS*-negative escapers" and "KRAS*-positive escapers", representatively. **B**, Hdac expression in KRAS*-expressing iKPC tumor samples and samples 24 hours after KRAS* extinction. **C**, *Hdac5* is upregulated in surviving cells. Two-tailed unpaired t tests were performed to calculate the p values. **D**, Dosage titration and on-target effect determination of MEK inhibitor Trametinib, mTOR inhibitor Rapamycin, and PI3Ka inhibitor LY294002 in iKPC-3 cells by western blot analysis. The red- highlighted concentrations were chosen for molecular and functional analysis *in vitro*. **E**, HDAC5 is upregulated in transplanted iKPC tumors after Trametinib (TRA) treatment. **F**, RNA expression of chemokines and cytokines that chemoattract macrophages and neutrophils in iKPC cells", "KRAS*-negative escapers" and KRAS*-positive escapers. "1", "2" and "3" represent "iKPC cells", "KRAS*-negative escapers"

and "KRAS*-positive escapers", representatively. **G**, Comparison of tumor growth between vehicle control, MEK inhibitor Trametinib only and dual inhibition of MEK by Trametinib and PI3K α by Alpelisib in transplanted syngeneic iKPC tumors (n = 7). Tumor sizes were measured at Day 7 and Day 11 post-treatment. Data are represented as mean ± SEM. **H**, Western blot analysis shows regulation of HDAC5 expression by KRAS* downstream signaling pathways in human PDAC cell lines. **I**, Western blot analysis shows upregulation of HDAC5 expression by KRAS^{G12C} inhibitor ARS-1620 in human NSCLC cell lines with KRAS^{G12C} mutation. **J**, Related to Fig. 6J, pharmacodynamic determination of KRAS^{G12C} inhibitor ARS-1620 alone and in combination with MEK inhibitor Trametinib in MIA PaCa-2 xenograft tumors in nude mice.



Supplementary Figure 9. Characterization of *HDAC5* escapers generated in syngeneic mouse models. **A**, MRI images to examine the tumor burden of orthotopically transplanted GFP-, *HDAC5-*, HDAC5D and *Ccl2-*OE iKPC-5 cells in C57BL/6 mice after KRAS* extinction at indicated days. MRI images of KRAS*-expressing GFP-OE iKPC-5 tumors were used as positive control. **B**, Transcriptional expression of KRAS*, endogenous *Kras*, endogenous *Yap1*, transgenic HDAC5 and total *Ccl2* in *HDAC5* and *Ccl2* escapers by qRT-PCR analysis. The iKPC tumors were used as control. **C**, Activation of KRAS* signaling pathway in *HDAC5* and *Ccl2* escapers by western blot analysis. The iKPC tumors were used as control. **D**, Gates of macrophages, monocytes and neutrophils in a representative iKPC tumor and a *HDAC5* escaper tumor generated from orthotopic allograft models in C57BL/6 mice by FACS analysis. Data are displayed by FlowJo. **E**,

CyTOF analysis of immune cell subtypes in iKPC-5 primary tumors (n = 5) and *HDAC5* escaper tumors (n = 4) generated from orthotopic allograft models in C57BL/6 mice. Two-tailed unpaired t tests were performed to calculate the p values. **F**, Validation of SMAD4 knockout in iKPC-5 cells by western blot analysis. For **B** and **E**, data are represented as mean ± SEM.