



**Supplemental Figure 1. Generation of the *Aifm1* floxed mice strain and genotypic analysis of the AIF<sup>-Y</sup> MEFs produced by tamoxifen treatment. (A)** Schematic representation of the wild-type *Aifm1* allele, targeting vector, and the resulting floxed *Aifm1* locus. The targeting vector was generated by classical recombination. Briefly, exon 11 of *Aifm1* was flanked by LoxP sequences in direct orientation along with a NEO cassette used for selection. Then, Sv129 ES cells were electroporated with this vector, transfected with a plasmid containing FLP recombinase to eliminate the NEO cassette and, after selecting positive clones, injected into C57BL/6J blastocysts to obtain the chimeric mice. After at least fifteen backcrosses in the C56BL/6J background, heterozygous *Aifm1*-floxed males (*Aifm1*<sup>fl/Y</sup>) were crossed to PGK-Cre females. This crossing induced the excision of exon 11 in *Aifm1* that resulted in a frameshift mutation and the creation of a stop codon in exon 12. **(B)** Representative southern blot analysis of WT (Co) and AIF<sup>-Y</sup> MEFs at 4 days post-tamoxifen treatment. **(C)** Schematic representation of the PCR genotyping strategy used to identify AIF transgenic mice and AIF<sup>-Y</sup> MEFs. The location, name and expected size of the different PCR products is indicated. The table specifies the sequence of the two primers used in the screening. Representative genomic PCR assessment performed in MEFs generated as described in **Figure 1** and left untreated (0) or treated with tamoxifen (4-OHT) at the time indicated.