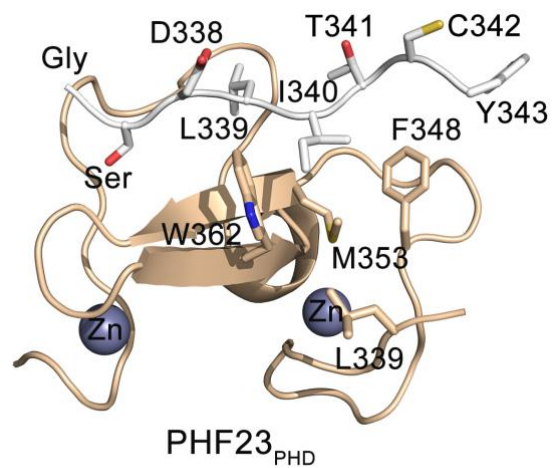


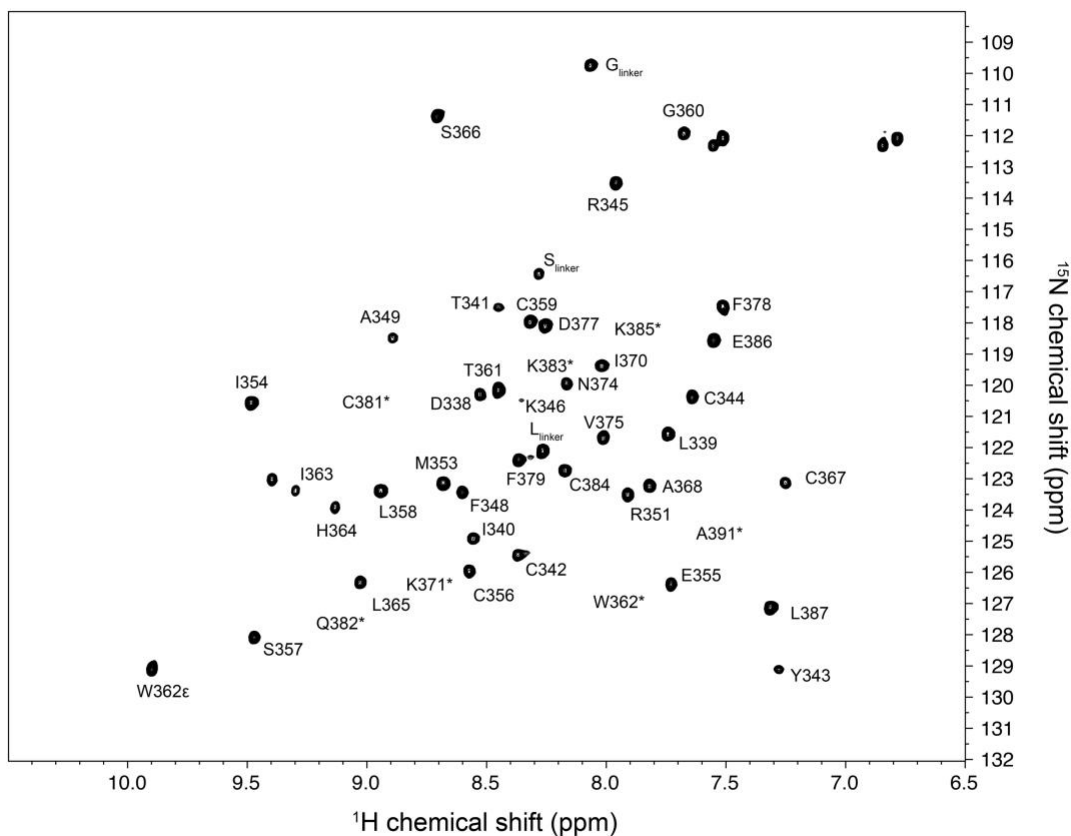
## **Supplementary Information**

### **Mechanistic insights into chromatin targeting by leukemic NUP98-PHF23 fusion**

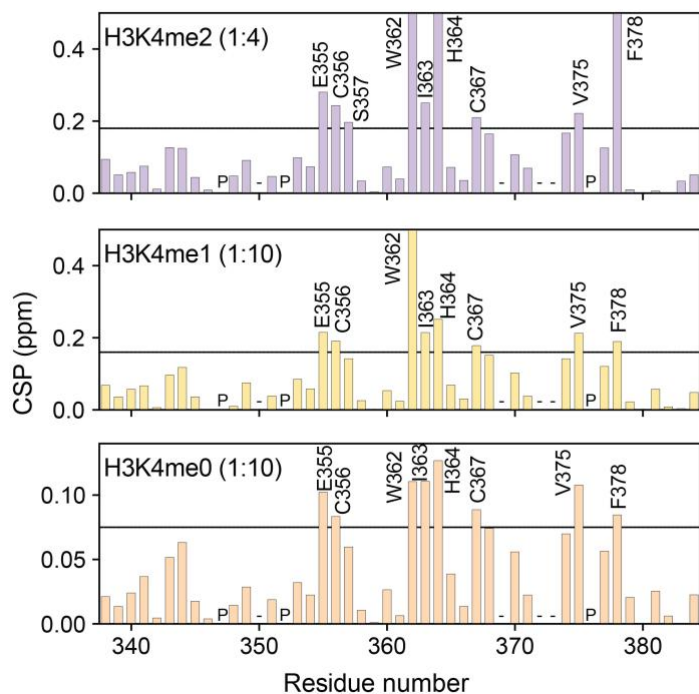
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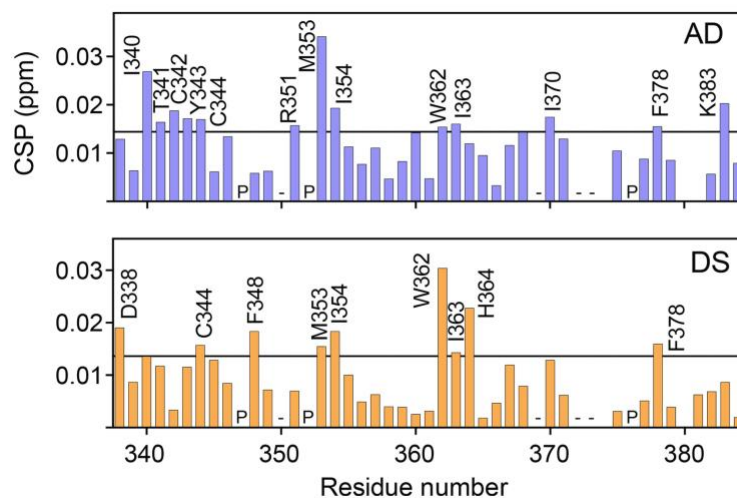
**Supplementary Figure 1.** Structure of PHF23<sub>PHD</sub>. The N-terminus of another PHF23<sub>PHD</sub> molecule (white; labeled) binds in the histone-binding site of the domain. The side chain of I340 occupies the aromatic cage. Gly and Ser residues derived from a vector are shown and labeled. Related to Figure 2.



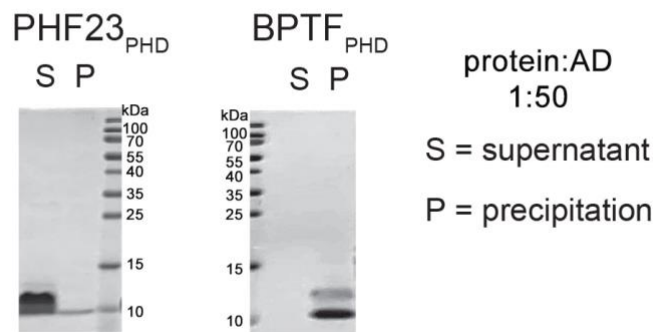
**Supplementary Figure 2. Backbone assignments of PHF23<sub>PHD</sub>.**  $^1\text{H},^{15}\text{N}$  HSQC spectrum of PHF23<sub>PHD</sub>. Backbone assignment of cross-peaks is shown using the one-letter code for amino acids. ‘\*’ indicates cross-peak invisible at the plotting level. ‘linker’ indicates cross-peaks from residues (GPLGS) derived from the vector. Related to Figures 1, 2 and 3.



**Supplementary Figure 3. Recognition of histone H3 peptides by PHF23<sup>PHD</sup>.** Chemical shift perturbation (CSP) analysis of PHF23<sup>PHD</sup> upon binding of H3K4me2 (1:4, top), H3K4me1 (1:10, middle) and H3K4me0 (1:10, bottom) peptides. 'P' indicates a proline residue. '-' indicates unassigned residue. Bars reaching the maximum of y-axis indicate disappeared cross-peaks. Related to Figures 1 and 2.



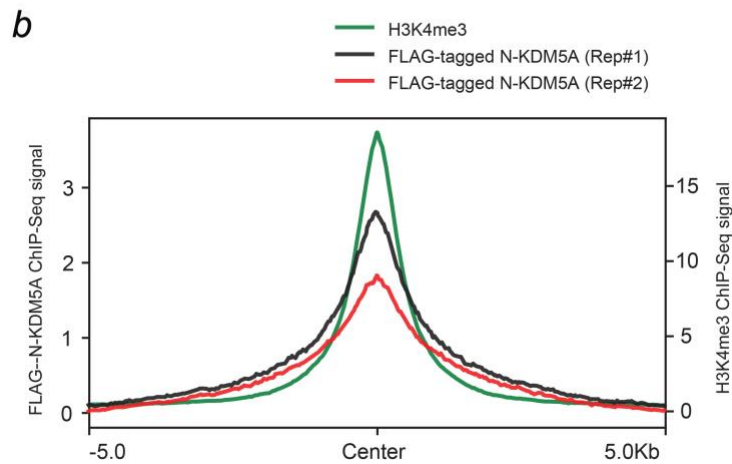
**Supplementary Figure 4. Characterization of AD and DS inhibitory mechanisms.** Chemical shift perturbation (CSP) analysis of the  $^1\text{H},^{15}\text{N}$  HSQC spectra of PHF23<sub>PHD</sub> at a 1:20 ratio of AD (purple) and 1:2 ratio of DS (orange). 'P' indicates a proline residue. '-' indicates an unassigned residue. Related to Figure 3.



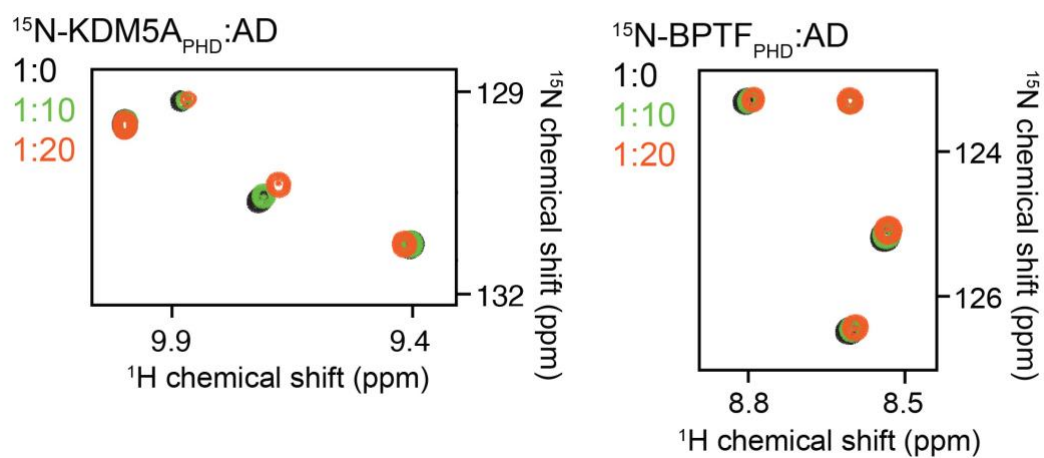
**Supplementary Figure 5.** Samples containing 0.1 mM PHF23<sub>PHD</sub> (left) and BPTF<sub>PHD</sub> (right) were incubated with AD at a protein:compound ratio of 1:50 and resolved by SDS-PAGE under non-reducing condition. 'S' and 'P' indicate supernatant and precipitation, respectively, after spinning the samples down. Experiment was repeated independently two times with similar results. Related to Figure 3.

**a**

Sample	Read # (raw)
INPUT	9860281
H3K4me3	8651824
H3K27me3	13855403
NUP98-KDM5A (FLAG tagged) FLAG Rep#1	15101034
FLAG Rep#2	21301507

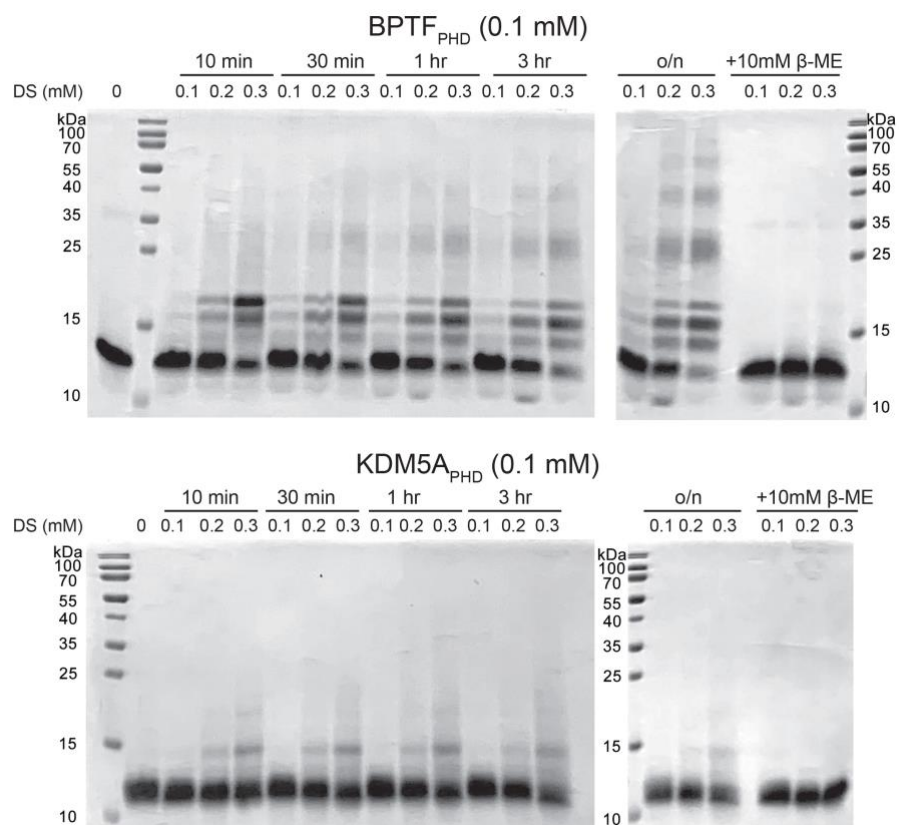


**Supplementary Figure 6. NUP98-KDM5A binding sites overlap with H3K4me3-loci.** (a). ChIP-seq read depths for the indicated sample. (b) Averaged H3K4me3 and NUP98-KDM5A ChIP-seq signals centered on H3K4me3 binding sites in a +/- 5.0-kb window in murine leukemia cells expressing FLAG-tagged NUP98-KDM5A. ChIP-seq signals were normalized to input. Related to Figures 5 and 6.



**Supplementary Figure 7. AD binds to KDM5A<sub>PHD</sub> and BPTF<sub>PHD</sub>.** Superimposed  $^1\text{H},^{15}\text{N}$  HSQC spectra of KDM5A<sub>PHD</sub> (left) and BPTF<sub>PHD</sub> (right) collected upon titration with AD. Spectra are color coded according to the protein:compound molar ratio. Related to Figures 5 and 6.





**Supplementary Figure 8.** Samples containing 0.1 mM BPTF<sub>PHD</sub> (top) and KDM5A<sub>PHD</sub> (bottom) were incubated with indicated amounts of DS for 10 min, 30 min, 1 hr, 3 hr, and overnight (o/n). All samples were flash-frozen and resolved by SDS-PAGE under non-reducing and reducing (10 mM β-ME) conditions. Experiment was repeated independently two times with similar results. Related to Figures 5 and 6.

**Supplementary Table 1.** Data collection and refinement statistics for PHF23<sub>PHD</sub>

PHF23 PHD domain	
<b>Data Collection</b>	
Space group	P 4 <sub>1</sub> 2 <sub>1</sub> 2
Wavelength (Å)	1.54
Resolution (Å)	50.0- 2.9 (3.0-2.9)*
Unit-cell dimensions	
a, b, c (Å)	66.92, 66.92, 151.05
α, β, γ (°)	90, 90, 90
No. of measured reflections	27524
No. of unique reflections	8025
Multiplicity	3.4 (3.0)
I/σ	13.4 (1.9)
Completeness (%)	97.8 (97.1)
R <sub>sym</sub> # (%)	8.5 (49.4)
R <sub>pim</sub> # (%)	5.1 (32.2)
No. of molecules in ASU	5
Matthews coefficient (Å <sup>3</sup> Da <sup>-1</sup> )	2.28
Solvent content (%)	46.1
<b>Refinement</b>	
R <sub>work</sub> /R <sub>free</sub> (%)	24.2/29.4
No. of atoms	
Protein	1953
Ligand/ion	10
Water	0
B-factors (Å <sup>2</sup> )	
Protein	58.71
Ligand/ion	58.58
R.M.S.D	84.36
Bond lengths (Å)	
Bond angles (°)	0.007
Ramachandran favored (%)	0.989
Ramachandran allowed (%)	95.0
Ramachandran outliers	5.0
Clashscore	0
Clashscore	8.36

\*Values in parentheses are for the highest resolution shell (Å).

#R<sub>sym</sub> =  $\sum |I_{\text{obs}} - I_{\text{avg}}| / I_{\text{avg}}$ , where I<sub>obs</sub> is intensity of any given reflection and I<sub>avg</sub> is the weighted mean I.

**Supplementary Table 2. List of primers used in this study.**

P1	CGCGAATTCGATCTGATCACATGTTACTGTCTG
P2	CCGCTCGAGTTACAGTTCCTTGCATTTTC
P3	CTGGGATCCGATCTGATCGCATGTTACTGTCTGAAAG
P4	CTTTTCGACAGTAACATGCGATCAGATCGGATCCCAG
P5	ATCACATGTTACTGTCTAAAGCCCTTTGCAGGGCGG
P6	CCGCCCTGCAAAGGGCTTTAGACAGTAACATGTGAT
P7	GTTACTGTCTGAAAGCCCGCTGCAGGGCGGCCCATGAT
P8	ATCATGGGCCGCCCTGCAGCGGGCTTTTCGACAGTAAC
P9	CTTTGCAGGGCGGCCCGTGATTGAGTGCAGCCTGTGT
P10	ACACAGGCTGCACTCAATCACGGGCCGCCCTGCAAAG
P11	GCCCATGATTGAGTGCAGCATGTGTGGGACGTGGATTC
P12	GAATCCACGTCCCACACATGCTGCACTCAATCATGGGC
P13	CAGCCTGTGTGGGACGGCGATTACCTCTCCTGTGCT
P14	AGCACAGGAGAGGTGAATCGCCGTCCCACACAGGCTG