Strain or plasmid	Genotype, characteristic(s) and/or use	Reference or source
Streptomyces coelicolor A	(3(2) strains	
M145	SCP1 ⁻ SCP2 ⁻	(1)
E117	∆ <i>rpfA-E</i> (<i>rpf</i> null)	(2)
J3385	3×∆PASTA	(3)
Escherichia coli strains		
DH5a	Routine cloning	
ET12567(pUZ8002)	<i>dam dcm</i> ; with transmobilizing plasmid pUZ8002	(4,5)
Rosetta 2	Protein overexpression	Novagen
Plasmids		
pET15b	Overexpression of His ₆ tagged proteins	Novagen
pMC177	pET15b carrying <i>rpfA</i>	(6)
pMC200	pET15b carrying <i>rpfA∆LysM</i>	This study
pMC178	pET15b carrying <i>rpfB</i>	(2)
pMC179	pET15b carrying <i>rpfC</i>	(2)
pMC181	pET15b carrying <i>rpfD</i>	(2)
pMC201	pET15b carrying <i>rpfD</i> Δ <i>LytM</i> Δ <i>LysM</i>	This study
pMC202	pET15b carrying <i>rpfD</i> Δ <i>LysM</i>	This study
pMC203	pET15b carrying <i>rpfD</i> Δ <i>LytM</i>	This study
pMC182	pET15b carrying <i>rpfE</i>	(2)

Table S1: Plasmids and Streptomyces coelicolor and Escherichia coli strains

Table S2: Oligonucleotides used in this study

Primer Name	Sequence (5′- 3′) ¹	Use
0974 PP 5'	CAGTAC <u>CATATG</u> GCCGACGCGCGACCTGGGAC	Overexpression of RpfD
0974 PP 3'	CAGTAC <u>GGATCC</u> TCAGATCCTGACGCCGCCGGC	Overexpression of RpfD
0974∆lytM PP 3'	CATCAT <u>GGATCC</u> CCGGGTGGTCCCCTGCCCGC	Overexpression of RpfD∆LytM
SCO0974∆lysM rev	TGCTCTTGCTTCTGCTCTTTCAGTCCGGCCCGCTCCGAGCA	RpfD∆LysM overexpression
SCO0974∆lysM fwd	AAAGAGCAGAAGCAAGAGCA	RpfDALysM overexpression
0974∆lytM∆lysM PP3'	CATCAT <u>GGATCC</u> TCCGGCCCGCTCCGAGCACA	Overexpression of RpfDALytMALysM
3097 PP 5'	CAGTAC <u>CATATG</u> GCCACCGCGTCCG	Overexpression of RpfA
3097 PP 3'	CGAAGT <u>GGATCC</u> TTACTTCAGGTGCAGCTGCTG	Overexpression of RpfA
3097∆lysM PP 3'	CATCAT <u>GGATCC</u> TCAGCCGGTGCCGCA	Overexpression of RpfA∆LysM
T7 promoter	TAATACGACTCACTATAGGG	Sequencing
T7 terminator	GCTAGTTATTGCTCAGCGG	Sequencing

¹Restriction enzyme sites are underlined

Table S3:	Conditions for	protein	overexpression

	Induction	[IPTG]	Induction time	Induction	Molecular
	OD ₆₀₀	(mM)	(hr)	temperature (°C)	weight (kDa) ¹
RpfA	0.8	1	2.5	30	22.8
RpfA∆LysM	0.6	1	2.5	30	10.0
RpfB	0.8	0.25	16	16	36.6
RpfC	0.8	1	16	16	40.0
RpfD	0.8	1	4.5	30	42.6
RpfD∆LysM	0.8	1	16	16	30.3
RpfD∆LysM∆LytM	0.8	1	2.5	30	10.9
RpfE	0.8	1	2.5	30	10.2

¹Calculated without the SignalP predicted signal peptide and with the 6×His tag

NlpD	-NKGIDIAGSKGQAIIATADGRVVYAGNALRG-YGNLIIIKHNDDYLSAYAHNDTMLVRE	58
RpfD	-HTGVDFPVPTGTSVKSVADGRVVSAGWGGSYGYQV-V-VRHGDGRYSQYAHLSAISVKS	57
LytM	AHYGVDYAMPENSPVYSLTDGTVVQAGWSNYGGGNQVTIKEANSNNYQWYMHNNRLTVSA	60
	· *:* : : : ** ** · · · · · * * · : *	
NlpD	QQEVKAGQKIATMGSTGTSS-TRLHFEIRYKGKSV 92	
RpfD	GQSVGVGQRLGRSGSTGNVTGPHLHFEVRTGPGFGSDVDP 97	
LytM	GDKVKAGDQIAYSGSTGNSTAPHVHFQRMSGG 92	
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Figure S1: Catalytic residues in the LytM domain of *S. aureus* LytM are conserved in **RpfD.** Alignment of the amino acid sequence of LytM domains from *Escherichia coli* NlpD, *Staphylococcus aureus* LytM, and *Streptomyces coelicolor* RpfD, using Clustal Omega. LytM catalytic residues are highlighted in grey. Asterisks denote residues that are conserved, colons denote conservation of residues with strongly similar properties, and periods denote conservation of residues with weakly similar properties at a specific position.



Figure S2: Peptidoglycan binding assays for RpfA and RpfD variants. Peptidoglycan binding by the different Rpf proteins was assessed by incubating approximately two nanomoles of each Rpf protein with peptidoglycan isolated from *Streptomyces coelicolor*. Following separation of bound (+PG, left) and unbound (Supernatant, centre) protein by centrifugation, the fractionated samples were separated on a tricine polyacrylamide gel (PAG) and stained using Coomassie Brilliant Blue. –PG (right) = input protein (positive control). Protein molecular weight is shown on the far right (kDa), with molecular weight relative to protein marker after separation on a PAG, shown in brackets (~kDa on gel).

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