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Reporting Checklist For Nature Communications Life Sciences Articles






This checklist is used to ensure good reporting standards and to improve the reproducibility of published results. For more information, please read [Reporting Life Sciences Research](#).

Figure legends

- Check here to confirm that the following information is available in all relevant figure legends (or Methods section if too long):
 - the **exact sample size (n)** for each experimental group/condition, given as a number, not a range;
 - a **description of the sample collection** allowing the reader to understand whether the samples represent **technical or biological replicates** (including how many animals, litters, culture, etc.);
 - a **statement of how many times the experiment shown was replicated in the laboratory**;
 - **definitions of statistical methods and measures**: (For small sample sizes (n<5) descriptive statistics are not appropriate, instead plot individual data points)
 - very common tests, such as *t*-test, simple χ^2 tests, Wilcoxon and Mann-Whitney tests, can be unambiguously identified by name only, but more complex techniques should be described in the methods section;
 - are tests one-sided or two-sided?
 - are there adjustments for multiple comparisons?
 - **statistical test results**, e.g., **P values**;
 - definition of '**center values**' as **median** or **mean**;
 - definition of **error bars** as **s.d.** or **s.e.m.** or **c.i.**

This checklist will not be published. Please ensure that the answers to the following questions are reported in the manuscript itself. We encourage you to include a specific subsection in the Methods section for statistics, reagents and animal models. Below, provide the page number or section and paragraph number (e.g. "Page 5" or "Methods, 'reagents' subsection, paragraph 2").

▶ <u>Statistics and general methods</u>	Reported in section/paragraph or page #:
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<div style="display: flex; align-items: flex-start;"> <div style="margin-right: 10px;"></div> <div> <p>How was the sample size chosen to ensure adequate power to detect a pre-specified effect size? (Give section/paragraph or page #)</p> <p>For animal studies, include a statement about sample size estimate even if no statistical methods were used.</p> </div> </div>	<div style="background-color: #f0f0f0; padding: 5px;"> <p>Methods section "Animals", page 18. Sample sizes chosen based on typical sample sizes in the literature and previous studies.</p> </div> <div style="background-color: #f0f0f0; padding: 5px; margin-top: 5px;"> <p>Methods section "Animals", page 18. No power calculations were performed to calculate sample sizes.</p> </div>
<div style="display: flex; align-items: flex-start;"> <div style="margin-right: 10px;"></div> <div> <p>Describe inclusion/exclusion criteria if samples or animals were excluded from the analysis. Were the criteria pre-established? (Give section/paragraph or page #)</p> </div> </div>	<div style="background-color: #f0f0f0; padding: 5px;"> <p>Methods section "Surgical procedures and intrinsic imaging", page 16. Methods section "Behavioral training", page 20. Mice with surgical complications or that did not accommodate to head-fixation were excluded from the study.</p> </div>
<div style="display: flex; align-items: flex-start;"> <div style="margin-right: 10px;"></div> <div> <p>If a method of randomization was used to determine how samples/animals were allocated to experimental groups and processed, describe it. (Give section/paragraph or page #)</p> <p>For animal studies, include a statement about randomization even if no randomization was used.</p> </div> </div>	<div style="background-color: #f0f0f0; padding: 5px;"> <p>Methods section "Animals", page 18. No randomization of mice into different experimental groups were performed. Our groups consisted of different genotypes, and not the same genotype subjected to different experimental conditions. Group randomization was hence not relevant.</p> </div> <div style="background-color: #f0f0f0; padding: 5px; margin-top: 5px;"> <p>There was no randomization, "Animals", p18</p> </div>
<div style="display: flex; align-items: flex-start;"> <div style="margin-right: 10px;"></div> <div> <p>If the investigator was blinded to the group allocation during the experiment and/or when assessing the outcome, state the extent of blinding. (Give section/paragraph or page #)</p> <p>For animal studies, include a statement about blinding even if no blinding was done.</p> </div> </div>	<div style="background-color: #f0f0f0; padding: 5px;"> <p>Methods section "Animals". No blinding was performed.</p> </div> <div style="background-color: #f0f0f0; padding: 5px; margin-top: 5px;"> <p>Methods section "Animals". No blinding was performed.</p> </div>
<div style="display: flex; align-items: flex-start;"> <div style="margin-right: 10px;"></div> <div> <p>For every figure, are statistical tests justified as appropriate?</p> <p>Do the data meet the assumptions of the tests (e.g., normal distribution)?</p> <p>Is there an estimate of variation within each group of data?</p> <p>Is the variance similar between the groups that are being statistically compared? (Give section/paragraph or page #)</p> </div> </div>	<div style="background-color: #f0f0f0; padding: 5px;"> <p>Methods section "Statistical analyses", Page 30. Due to the hierarchical structure of the data, linear mixed effects statistics on log-transformed data was used. P-values from two-sided test.</p> </div> <div style="background-color: #f0f0f0; padding: 5px; margin-top: 5px;"> <p>Methods section "Statistical analyses", Page 30</p> </div> <div style="background-color: #f0f0f0; padding: 5px; margin-top: 5px;"> <p>Methods section "Statistical analyses", Page 30</p> </div> <div style="background-color: #f0f0f0; padding: 5px; margin-top: 5px;"> <p>Methods section "Statistical analyses", Page 30</p> </div>

► **Reagents**



To show that antibodies were profiled for use in the system under study (assay and species), provide a citation, catalog number and/or clone number, supplementary information or reference to an antibody validation profile (e.g., [Antibodypedia](#), [1DegreeBio](#)).

7. Cell line identity:
 - a. Are any cell lines used in this paper listed in the database of commonly misidentified cell lines maintained by [ICLAC](#) (also available in [NCBI Biosample](#))?
 - b. If yes, include in the Methods section a scientific justification of their use – indicate here on which page (or section and paragraph) the justification can be found.
 - c. For each cell line, include in the Methods section a statement that specifies:
 - the source of the cell lines
 - have the cell lines been authenticated? If so, by which method?
 - have the cell lines been tested for mycoplasma contamination? In this checklist, indicate on which page (or section and paragraph) the information can be found.

Reported in section/paragraph or page #:

Methods, "Immunohistochemistry", page 23. All antibodies are commercial (see URLs for validation data). Primary antibodies: polyclonal chicken anti-GFP (1:3000, Abcam Cat#ab13970: <https://www.abcam.com/gfp-antibody-ab13970.html>), rabbit anti-GFP (1:4000, Abcam Cat#ab6556: <https://www.abcam.com/gfp-antibody-ab6556.html>), mouse anti-NeuN (1:1000, Merck Cat#MAB377), mouse anti-GFAP (1:1000, Merck/Sigma-Aldrich Cat#MAB360: <https://www.sigmaaldrich.com/catalog/product/mm/mab377?lang=en®ion=NO>), rabbit anti-Iba1 (1:1000, Wako Cat#019-19741: <https://labchem-wako.fujifilm.com/us/category/01213.html>). Secondary antibodies (all diluted 1:200, Jackson Laboratories): Cy5-coupled anti-rabbit (Cat#111-175-144), Cy5-coupled anti-mouse (Cat#115-175-146), FITC-coupled anti-rabbit (Cat#111-095-144) and FITC-coupled anti-chicken (Cat#703-095-155).

N/A

N/A

N/A

N/A

N/A

► **Animal Models**



Report species, strain, sex and age of animals



For experiments involving live vertebrates, include a statement of compliance with ethical regulations and identify the committee(s) approving the experiments.

10. We recommend consulting the ARRIVE guidelines ([PLoS Biol. 8\(6\), e1000412,2010](#)) to ensure that other relevant aspects of animal studies are adequately reported.

Reported in section/paragraph or page #:

Methods, "Animals". Male C57BL/6J, *Itpr2^{-/-}* and GLT1-eGFP mice of 8–10 weeks

Methods, section "Animals". All procedures were approved by the Norwegian Food Safety Authority (FOTS 11983).

► **Human subjects**

11. Identify the committee(s) approving the study protocol.
12. Include a statement confirming that informed consent was obtained from all subjects.
13. For publication of patient photos, include a statement confirming that consent to publish was obtained.
14. Report the clinical trial registration number (at [ClinicalTrials.gov](#) or equivalent).
15. For phase II and III randomized controlled trials, please refer to the [CONSORT statement](#) and submit the CONSORT checklist with your submission.
16. For tumor marker prognostic studies, we recommend that you follow the [REMARK reporting guidelines](#).

Reported in section/paragraph or page #:

N/A

N/A

N/A

N/A

N/A

N/A

► **Data Availability**

Reported in section/paragraph or page #



Provide a Data availability statement in the Methods section under "**Data availability**", which should include, where applicable:

- Accession codes for deposited data
- Other unique identifiers (such as DOIs and hyperlinks for any other datasets)
- At a minimum, a statement confirming that all relevant data are available from the authors
- Formal citations of datasets that are assigned DOIs
- A statement regarding data available with restrictions

See our [data availability and data citations policy page](#) for more information.

Data deposition in a public repository is mandatory for:

- a. Protein, DNA and RNA sequences
- b. Macromolecular structures
- c. Crystallographic data for small molecules
- d. Microarray data

Deposition is strongly recommended for many other datasets for which structured public repositories exist; more details on our data policy are available [here](#). We encourage the provision of other source data in supplementary information or in unstructured repositories such as [Figshare](#) and [Dryad](#). We encourage publication of Data Descriptors (see [Scientific Data](#)) to maximize data reuse



3. If computer code was used to generate results that are central to the paper's conclusions, include a statement in the Methods section under "**Code availability**" to indicate whether and how the code can be accessed. Include version information as necessary and any restrictions on availability.

Section "Data Availability", page 31:
 "The raw data that support the findings of this study are available from the corresponding author upon reasonable request. The source data underlying Figs. 2c–d, 3b, 4b–c, 5e–g, 6b, e, 7b–e, 8a–d, f, h, 9a, c, and Supplementary Figs. 2a–d, 4b–c, 5e–f, 8g–i, 9, 10c–h, 11, 14a–e, 15 a–c, are provided as Source Data files."

Furthermore a small dataset to illustrate the workflow is available with the Supplementary Software.

Methods, section "Software Availability", page 30. The code and a small dataset i provided as Supplementary Software.