

Peer Review Information

Journal: NSMB

Manuscript Title: A helical assembly of human ESCRT-I scaffolds reverse-topology membrane scission

Corresponding author name(s): James H. Hurley

Reviewer Comments & Decisions:

Decision Letter, initial version:
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Dear Jim,

Thank you again for submitting your manuscript "A helical assembly of human ESCRT-I scaffolds reverse-topology membrane scission". We now have comments (below) from 2 reviewers who evaluated your paper; both are experts in ESCRT systems. In light of those reports, we remain interested in your study and would like to see your response to the comments of the referees, in the form of a revised manuscript.

You will see that reviewer 1 is quite positive about the interest of the work, and has requests for clarifications on reproducibility of some of the data presented. Reviewer 2 is concerned about the physiological relevance of the helical assembly, and that the mutations that disrupt such assembly could affect interaction of ESCRT-I with other factors. We would like to know the availability (or feasibility) of cellular data that could address such concerns. We'd be happy to discuss a revision, in which case a point-by-point response (for editorial assessment) would be very useful.

We are committed to providing a fair and constructive peer-review process. Do not hesitate to contact us if there are specific requests from the reviewers that you believe are technically impossible or unlikely to yield a meaningful outcome.

We expect to see your response within 1 week; you may send that to me directly by e-mail.

As you already know, we put great emphasis on ensuring that the methods and statistics reported in our papers are correct and accurate. As such, if there are any changes that should be reported, please

submit an updated version of the Reporting Summary along with your revision.

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SOURCE DATA: we urge authors to provide, in tabular form, the data underlying the graphical representations used in figures. This is to further increase transparency in data reporting, as detailed in this editorial (<http://www.nature.com/nsmb/journal/v22/n10/full/nsmb.3110.html>). Spreadsheets can be submitted in excel format. Only one (1) file per figure is permitted; thus, for multi-paneled figures, the source data for each panel should be clearly labeled in the Excel file; alternately the data can be provided as multiple, clearly labeled sheets in an Excel file. When submitting files, the title field should indicate which figure the source data pertains to. We encourage our authors to provide source data at the revision stage, so that they are part of the peer-review process.

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We require deposition of coordinates (and, in the case of crystal structures, structure factors) into the Protein Data Bank with the designation of immediate release upon publication (HPUB). Electron microscopy-derived density maps and coordinate data must be deposited in EMDB and released upon publication. Deposition and immediate release of NMR chemical shift assignments are highly encouraged. Deposition of deep sequencing and microarray data is mandatory, and the datasets must

be released prior to or upon publication. To avoid delays in publication, dataset accession numbers must be supplied with the final accepted manuscript and appropriate release dates must be indicated at the galley proof stage.

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We look forward to seeing the revised manuscript and thank you for the opportunity to review your work.

Sincerely,
Ines

Ines Chen, Ph.D.
Chief Editor
Nature Structural & Molecular Biology

ORCID 0000-0002-1405-9703

Reviewers' Comments:

Reviewer #1:

Remarks to the Author:

Flower et al reports the structure of the headpiece of human ESCRT-I. The subcomplex serendipitously crystallized as a helical assembly. The authors propose that this assembly recapitulates a key functional feature of ESCRT-I during its role in vesicle budding. Evidence that the helical arrangement observed in the crystals include showing that ESCRT-I subcomplexes form helical assemblies in solution and that mutagenesis of helical interface residues affect assembly in vitro and abrogate function during membrane fission in cells. The authors then used molecular simulations of modeled full-length ESCRT-I helices in context of HIV-1 budding. They conclude that ESCRT-I may have a scaffolding and mechanical role during vesicle formation. Whereas the structure of the equivalent complex from yeast has been previously reported by the Hurley group, this paper is still quite novel and substantially extends those studies. The finding that ESCRT-I forms helical assemblies is the most impactful aspect of the paper as it significantly changes understanding of its role in membrane fission and will inform future studies in this area. The paper also clarifies the relationship between yeast Mvb12 and human MVB12A which is not apparent from sequence, the role of MVB12A (VPF motif) in assembly of the headpiece, and the relationship between the UEV domain and PTAP motifs of TSG101 – while these aspects are of more specialized interest, they remain an important contribution and fill in gaps in understanding of ESCRT-I function.

The paper is very well written throughout; concepts are presented clearly and logically. The biochemical and structural data are all of high quality. I have no changes to suggest, except that for the biochemical and functional experiments the figure legends 2d, 2f, 3e, 3f, 5 and S1 should indicate the number of biological/technical repeats if any.

Reviewer #2:

Remarks to the Author:

Flower et al define the structure of the human ESCRT-I headpiece, a fragment of the full ESCRT-I complex that is composed of ~80 amino acids of Tsg101, ~70 amino acids of Vps37B, ~120 amino acids of Vps28, and ~60 amino acids of Mvb12A (small domains of each subunit). Despite lacking sequence identity with yeast Mvb12, human Mvb12A occupies a similar location on the ESCRT-I headpiece. Overall, the complex solved exhibits a highly similar conformation as compared to the previously published yeast ESCRT-I headpiece. The structure shown is well characterized, and on its own, is publishable at another more appropriate venue, given its relatively modest novelty. During analysis, the authors find that the headpiece is capable of oligomerization and filament formation in vitro, mediated largely by electrostatic interactions between Vps28 subunits within each headpiece. Although they are able to disrupt this interface with a series of significant charge reversals (K54D and K58D) and charge neutralization (D59A) mutations, there is no evidence that these mutations do not also impact other

contacts needed for Vps28 to function in cells. Unfortunately, all assays performed in cells leverage these mutations, making them inconclusive. Ultimately, the data do not directly support a putative role for ESCRT-I oligomerization in autophagy or HIV-1 budding. In its present form, the work is not suitable for publication in NSMB, as the findings seem overinterpreted and remain ambiguous.

Major comments:

1. Currently there is no evidence that ESCRT-I forms anything larger than a heterotetramer in cells. Specifically, the authors fail to provide data using biochemical methods or imaging that directly demonstrate ESCRT-I oligomerizes during autophagosome closure or HIV-1 budding, which is absolutely necessary to make their model plausible.
2. Could the K54D, K58D, D59A mutations in Vps28 not impair its other associations in cells? The authors seem to overinterpret the impact of these mutations, as they may act in other ways beyond specifically interfering with Vps28-Vps28 interactions, which may only be an artifact of crystallization.
3. The indirect assay examining Vps37 accumulation following CHMP2A depletion is insufficient evidence that "...the mutations within the helical interface of VPS28 disrupt the higher order oligomerization of ESCRT-I." To make any strong conclusions regarding the ability of ESCRT-I to oligomerize or not, the authors need to examine wild-type ESCRT-I assembly in a manner similar to that of Teis and Kirchhausen (at the single molecule level; Adell et al., 2017).

Author Rebuttal to Initial comments

Response to Reviewers' Comments:

Reviewer #1:

Remarks to the Author:

[We greatly appreciate Reviewer 1's thoughtful comments and enthusiasm for the work.](#)

The paper is very well written throughout; concepts are presented clearly and logically. The biochemical and structural data are all of high quality. I have no changes to suggest, except that for the biochemical and functional experiments the figure legends 2d, 2f, 3e, 3f, 5 and S1 should indicate the number of biological/technical repeats if any.

[The pull down experiments shown in figure 2d and f were repeated twice with the same results. Gel filtration experiments shown in figure 3 were repeated three times. The experiment in figure 5 was repeated three times and the Western blot is representative of the outcome on all occasions. The gel in figure S1 represents two biological replicates. All of this information is now noted in the respective figure legends.](#)

Reviewer #2:

Remarks to the Author:

We appreciate Reviewer 2's comments on the high quality of the structural work. However, the reviewer's concerns about the cellular function are overstated, as detailed below.

Major comments:

1. Currently there is no evidence that ESCRT-I forms anything larger than a heterotetramer in cells. Specifically, the authors fail to provide data using biochemical methods or imaging that directly demonstrate ESCRT-I oligomerizes during autophagosome closure or HIV-1 budding, which is absolutely necessary to make their model plausible.

We would like to call the referee's attention to a recent publication demonstrating that 3-11 copies of ESCRT-I mediate HIV-1 release, consistent with our findings. Please see Hoffman et al. <https://www.jbc.org/content/294/44/16266.long>. This publication is cited on pg. 17 as ref. 56. We have expanded the discussion of the Hoffman et al. findings.

2. Could the K54D, K58D, D59A mutations in Vps28 not impair its other associations in cells? The authors seem to overinterpret the impact of these mutations, as they may act in other ways beyond specifically interfering with Vps28-Vps28 interactions, which may only be an artifact of crystallization.

We are not aware of any other interactors reported with the N-terminal domain of VPS28. With respect to the possibility of a crystallization artifact, this is ruled out by the EM of filaments assembled in solution, which we provide in Fig. 3.

3. The indirect assay examining Vps37 accumulation following CHMP2A depletion is insufficient evidence that "...the mutations within the helical interface of VPS28 disrupt the higher order oligomerization of ESCRT-I." To make any strong conclusions regarding the ability of ESCRT-I to oligomerize or not, the authors need to examine wild-type ESCRT-I assembly in a manner similar to that of Teis and Kirchhausen (at the single molecule level; Adell et al., 2017).

The suggested wild-type ESCRT assembly imaging was already published by Hoffman et al. <https://www.jbc.org/content/294/44/16266.long> and the results are consistent with our

conclusions. Out of respect to the reviewer's concern we have however rephrased the quoted sentence and now state that the mutations "inhibit the accumulation" of ESCRT-I instead of "disrupt the higher order oligomerization of ESCRT-I."

Decision Letter, first revision:

Dear Jim,

Thank you again for submitting your manuscript "A helical assembly of human ESCRT-I scaffolds reverse-topology membrane scission". As previously communicated, after editorial assessment of the revision and response, we are happy to accept your paper, in principle, for publication as an Article in Nature Structural & Molecular Biology, on the condition that you revise your manuscript in response to our editorial requirements.

The text and figures require revisions. Note that, within a few days, we will send you detailed instructions for the final revision, along with information on editorial and formatting requirements. We recommend that you do not start revising the manuscript until you receive this additional information.

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Sincerely,
Ines

Ines Chen, Ph.D.
Chief Editor
Nature Structural & Molecular Biology

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Final Decision Letter:

Dear Jim,

Hope all is well with you and your team.

We are now happy to accept your revised paper "A helical assembly of human ESCRT-I scaffolds reverse-topology membrane scission" for publication as a Article in Nature Structural & Molecular Biology.

Acceptance is conditional on the manuscript's not being published elsewhere and on there being no announcement of this work to the newspapers, magazines, radio or television until the publication date in Nature Structural & Molecular Biology.

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Sincerely,
Ines

Ines Chen, Ph.D.
Chief Editor
Nature Structural & Molecular Biology

ORCID 0000-0002-1405-9703