

SUPPLEMENTARY MATERIAL

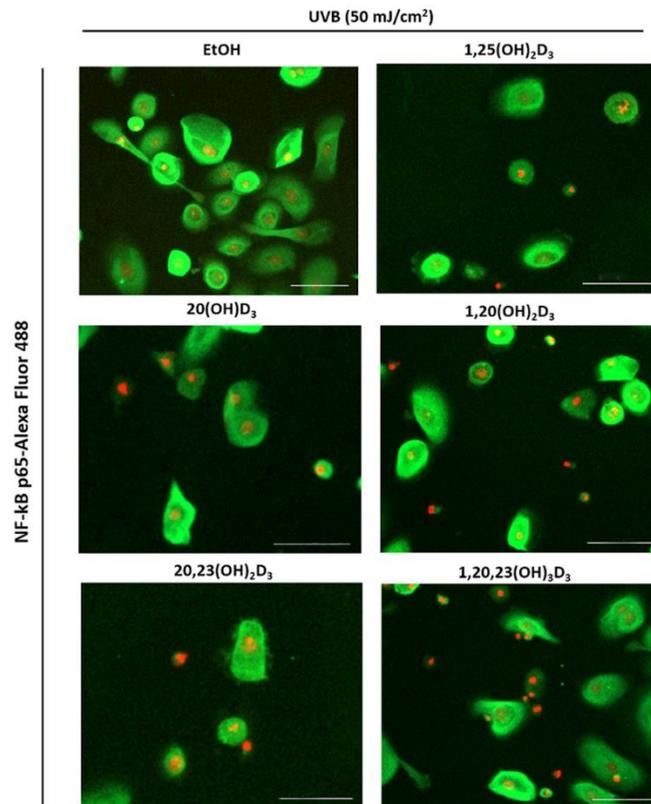
CYP11A1-derived vitamin D₃ products protect against UVB-induced inflammation and promote keratinocytes differentiation

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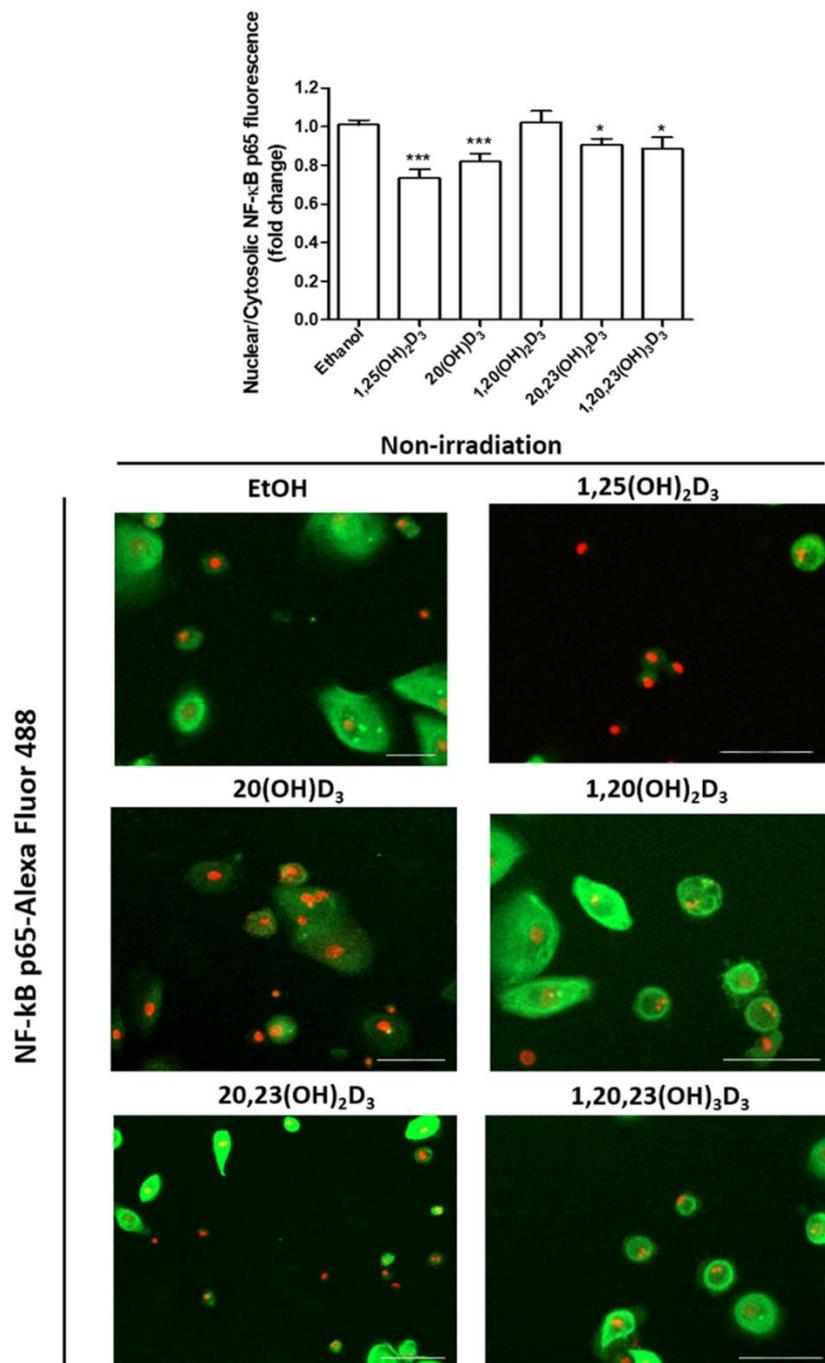
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Supplementary Figure S1. Immunofluorescent images of UVB-irradiated cells treated with vitamin D₃ hydroxyderivatives: nuclear/cytosolic NF-κB p65 detection

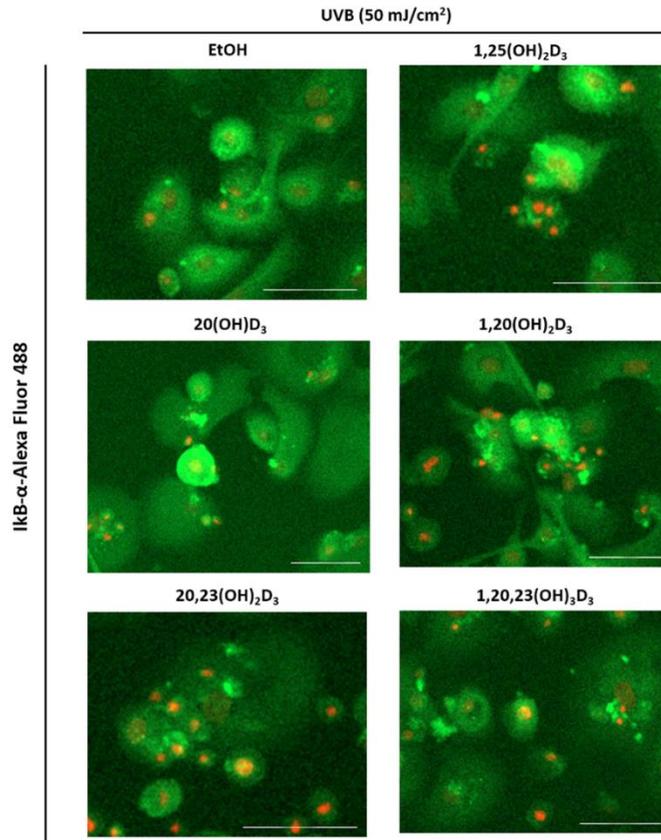
Keratinocytes were pretreated individually with 100 nM of each of the hydroxyderivatives (or ethanol vehicle), for 24 h before UVB irradiation (50 mJ/cm²) then further incubated with the same hydroxyderivative for an additional 24 h. Cells were fixed and stained with anti-NF-κB p65 antibody and imaged with a fluorescence microscope. Immunofluorescent cells show the NF-κB p65 levels in UVB-irradiated cells treated with the indicated secosteroid. Keratinocytes stained with NF-κB p65 antibody exhibited green fluorescence while nuclear staining with propidium iodide caused red fluorescence. Immunofluorescent positive cells were captured using the Cytation™ 5 cell imaging, n ≥ 100 cells for each condition. Scale bar = 100 μm. The immunofluorescent images are representative pictures for the immunofluorescent graph analysis.



Supplementary Figure S2. Immunofluorescent images of non-irradiated cells treated with hydroxyderivatives of vitamin D₃: nuclear/cytosolic NF-κB p65 detection

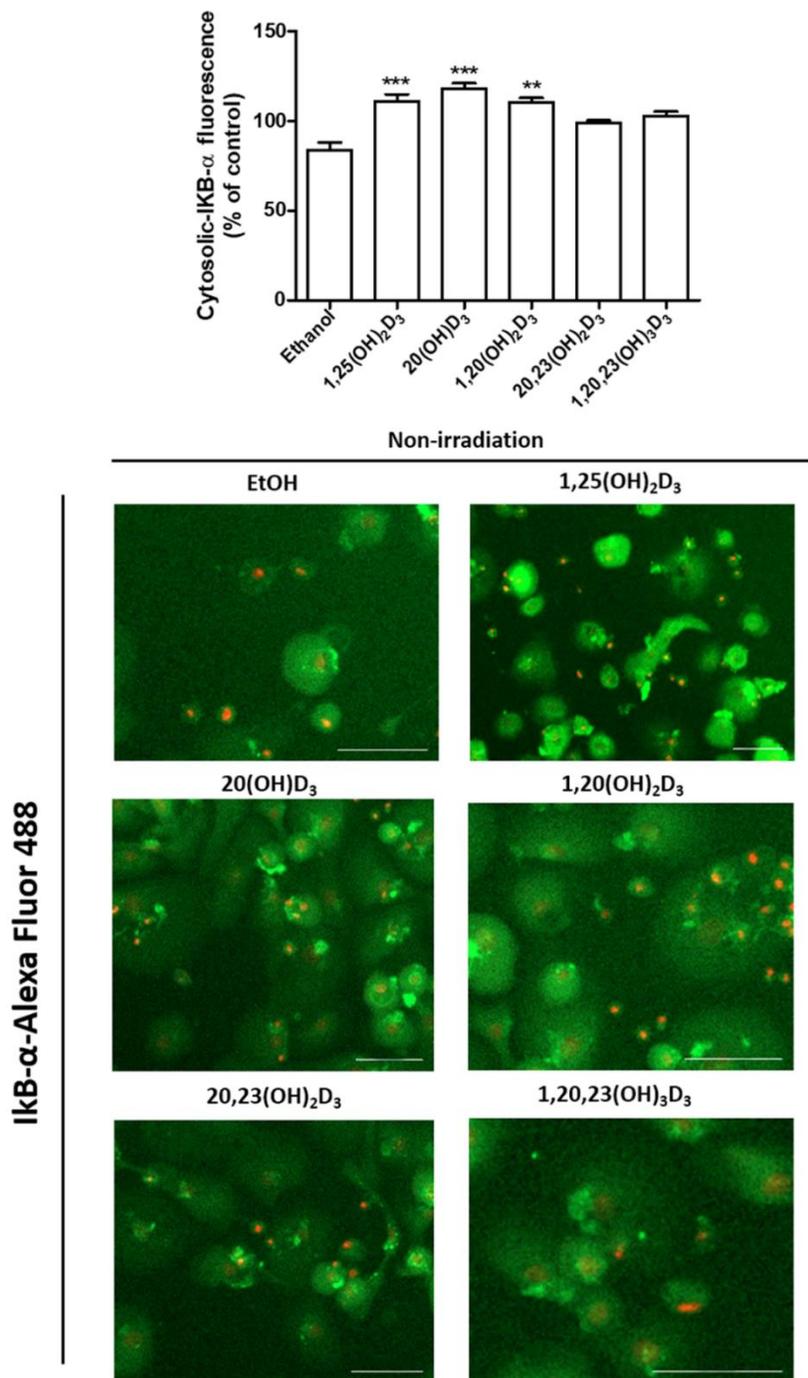
Keratinocytes were incubated with the indicated hydroxyderivative (100 nM), or ethanol vehicle, for 24 h. Cells were fixed and stained with anti-NF-κB p65 antibody and imaged with a

fluorescence microscope. Fluorescence intensity was measured using the Cytation 5 reader and data were analyzed using Graph Pad Prism. Images and graphs show the levels of fluorescence intensities in non-irradiated cells with the effects of the hydroxyderivatives shown in the graph (upper panel) and images in the lower panels. Keratinocytes stained with NF- κ B p65 antibody exhibited green fluorescence while nuclear staining with propidium iodide caused red fluorescence. Data are presented as the ratio (mean \pm SD) nuclear/cytoplasmic staining calculated from fluorescence intensities. Immunofluorescent positive cells were captured using the CytationTM 5 cell imaging, $n \geq 100$ cells for each condition. Scale bar = 100 μ m. The statistical significance of differences was evaluated using the student t-test, * $P < 0.05$ and *** $P < 0.001$, for all conditions. The immunofluorescent images are representative pictures for the immunofluorescent graph analysis.



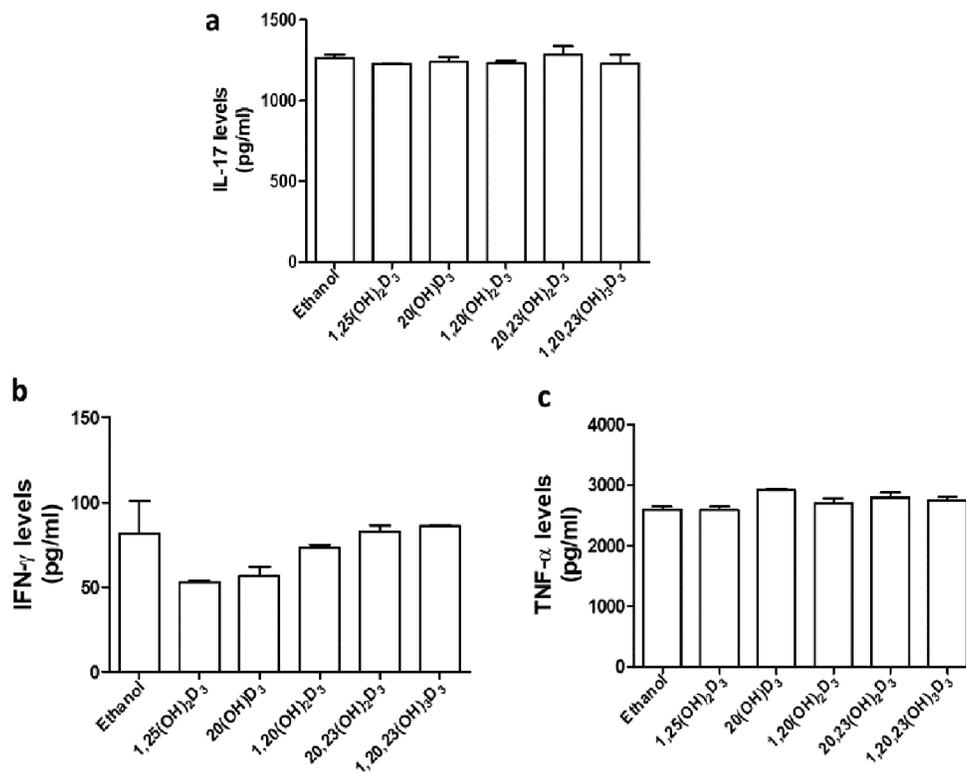
Supplementary Figure S3. Immunofluorescent images of UVB-irradiated cells treated with hydroxyderivatives of vitamin D₃: cytosolic-IκB-α detection

Keratinocytes were pretreated individually with each of the indicated vitamin D₃ hydroxyderivatives (100 nM) or ethanol vehicle, for 24 h before UVB irradiation (50 mJ/cm²) then further incubated with the same hydroxyderivative for an additional 24 h. Cells were fixed and stained with anti-IκB-α antibody and imaged with a fluorescence microscope. Immunofluorescent cells show the IκB-α levels in UVB-irradiated cells treated with the indicated secosteroid. Keratinocytes stained with IκB-α antibody exhibited green fluorescence while nuclear staining with propidium iodide caused red fluorescence. Immunofluorescent positive cells were captured using the Cytation™ 5 cell imaging, n ≥ 100 cells for each condition. Scale bar = 100 μm. The immunofluorescent images are representative pictures for the immunofluorescent graph analysis.



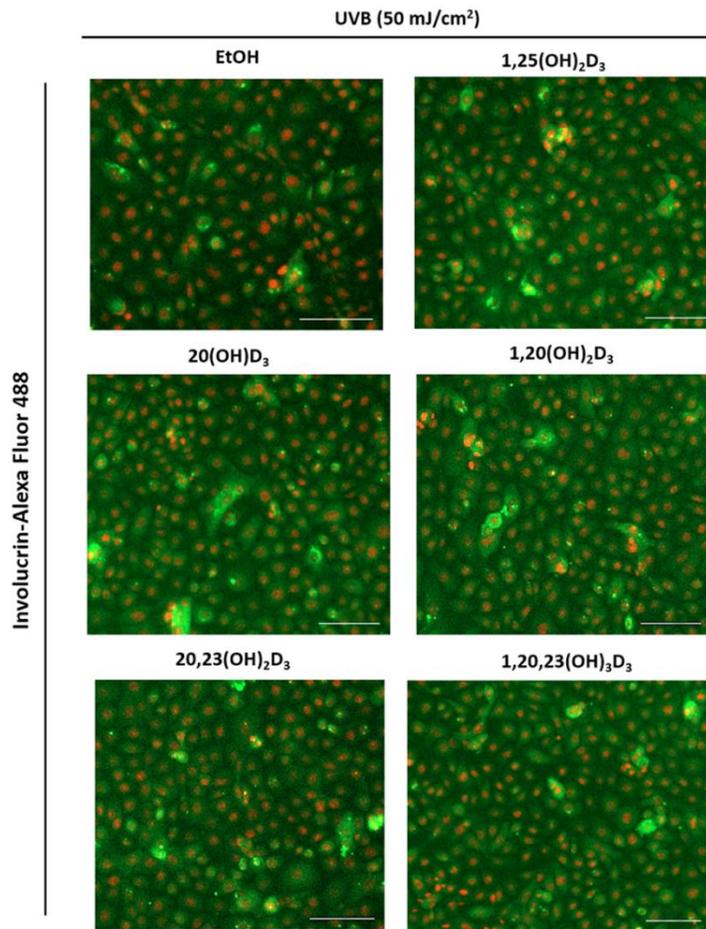
Supplementary Figure S4. Immunofluorescent images of non-irradiated cells treated with hydroxyderivatives of vitamin D₃: cytosolic-IκB-α detection

Keratinocytes were incubated with the indicated hydroxyderivative (100 nM) or ethanol vehicle, for 24 h. Cells were fixed and stained with anti-I κ B- α antibody and imaged with a fluorescence microscope. Fluorescence intensity was measured using the Cytation 5 reader and data were analyzed using Graph Pad Prism. Images and graphs show the levels of fluorescence intensities in non-irradiated cells with the effects of the hydroxyderivatives shown in the graph (upper panel) and images in the lower panels. Keratinocytes stained with I κ B- α antibody exhibited green fluorescence while nuclear staining with propidium iodine caused red fluorescence. Immunofluorescent positive cells were captured using the CytationTM 5 cell imaging, $n \geq 100$ cells for each condition. Scale bar = 100 μ m. Data are presented as the ratio (mean \pm SD) nuclear/cytoplasmic staining calculated from fluorescence intensities. The statistical significance of differences was evaluated using the student t-test, ** $P < 0.01$ and *** $P < 0.001$, for all conditions. The immunofluorescent images are representative pictures for the immunofluorescent graph analysis.



Supplementary Figure S5. The levels of IL-17, IFN- γ , and TNF- α in non-irradiated cells treated with hydroxyderivatives of vitamin D₃

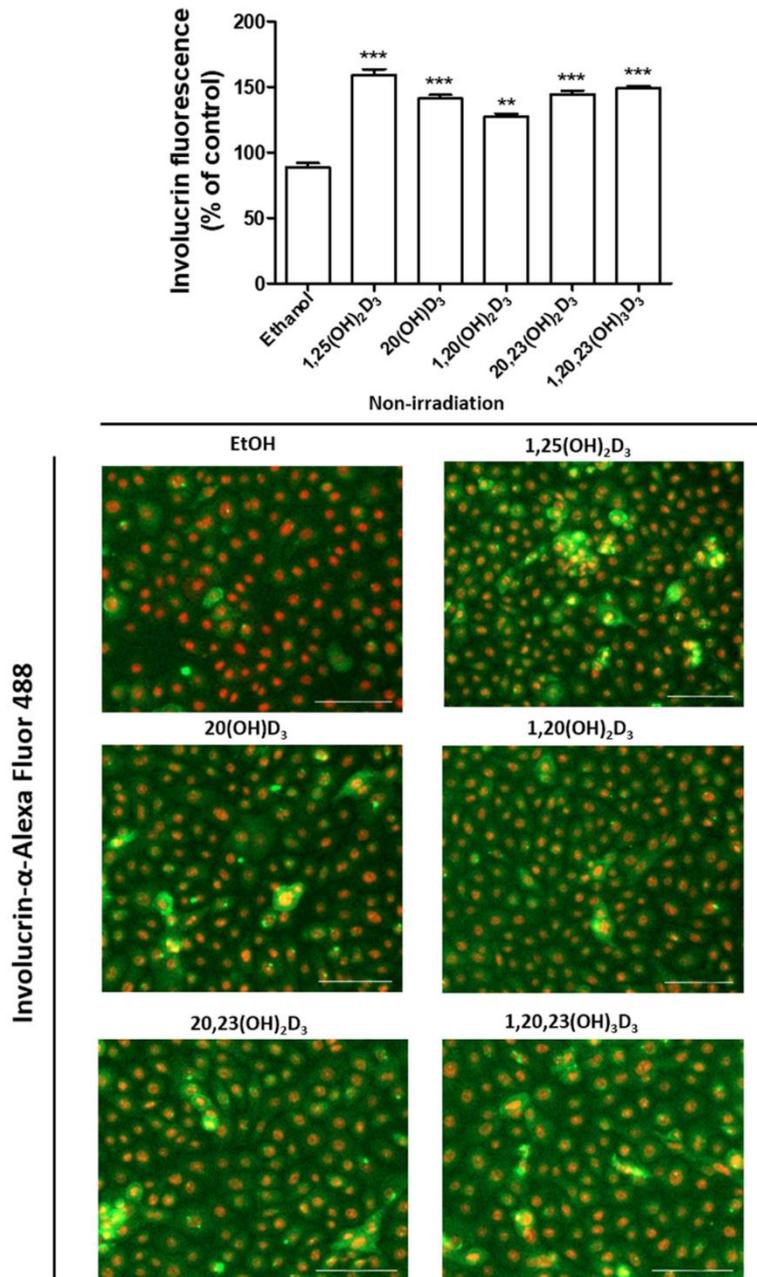
Keratinocytes were incubated with the indicated hydroxyderivative (100 nM) or ethanol vehicle, for 24 h prior to UVB irradiation (50 mJ/cm²). Cell supernatants were plated using an ELISA assay kit tagged with (a) IL-17, (b) IFN- γ , and (c) TNF- α antibody. The cytokine levels of secosteroid-treated cells compared to vehicle-treated cells are presented as bar graphs. The levels of inflammatory cytokines (pg/ml) were calculated from protein standards. The statistical significance of differences was evaluated by the student t-test, for all conditions.



Supplementary Figure S6. Immunofluorescent images of UVB-irradiated cells treated with hydroxyderivatives of vitamin D₃: involucrin (IVL) detection

Keratinocyte were pretreated individually with 100 nM of each of the indicated hydroxyderivatives (or ethanol vehicle) for 24 h before UVB irradiation (50 mJ/cm²), then further incubated with the same hydroxyderivative for an additional 24 h. Cells were fixed and stained with anti-IVL antibody and imaged with a fluorescence microscope. Immunofluorescent cells show the IVL levels in UVB-irradiated cells treated with the indicated secosteroid. Keratinocytes stained with IVL antibody exhibited green fluorescence while nuclear staining with propidium iodine caused red fluorescence. Immunofluorescent positive cells were captured using the Cytation™ 5 cell imaging,

$n \geq 100$ cells for each condition. Scale bar = 100 μm . The immunofluorescent images are representative pictures for the immunofluorescent graph analysis.



Supplementary Figure S7. Immunofluorescent images of non-irradiated cells treated with hydroxyderivatives of vitamin D₃: involucrin (IVL) detection

Keratinocytes were incubated with the indicated hydroxyderivative (100 nM) or with ethanol vehicle, for 24 h. Cells were fixed and stained with anti-IVL antibody and imaged with a

fluorescence microscope. Fluorescence intensity was measured using the Cytation 5 reader and data were analyzed using Graph Pad Prism. Images and graphs show the levels of fluorescence intensities in non-irradiated cells with the effects of the hydroxyderivatives shown in the graph (upper panel) and images in the lower panels. Keratinocytes stained with IVL antibody exhibited green fluorescence while nuclear staining with propidium iodine caused red fluorescence. Immunofluorescent positive cells were captured using the Cytation™ 5 cell imaging, $n \geq 100$ cells for each condition. Scale bar = 100 μm . Data are presented as the ratio (mean \pm SD) nuclear/cytoplasmic staining calculated from fluorescence intensities. The statistical significance of differences was evaluated using the student t-test, * $P < 0.05$ and *** $P < 0.001$, for all conditions. The immunofluorescent images are representative pictures for the immunofluorescent graph analysis.

Supplementary Table S1. Sequences of primers used in the heat map analysis

Additional details are in [1-6].

Function	Gene	Description	Sequence (L and R)
The biological actions of vitamin D ₃	<i>VDR</i>	Vitamin D ₃ receptor	CTTACCTGCCCCCTGCTC AGGGTCAGGCAGGGAAGT
Inflammation	<i>TLR4</i>	Toll-like receptor 4	CAG GTG GAA TTG TAT CGC CT CGA GGC TTT TCC ATC CAA TA
	<i>COX2</i>	Cyclooxygenase-2	GAATGGGGTGATGAGCAGTT CAGAAGGGCAGGATACAGC
	<i>ICAM</i>	Intracellular adhesion molecule	CCTTCCTCACCGTGTACTGG AGCGTAGGGTAAGGTTCTTGC
	<i>IL-6</i>	Interleukin 6	GAAGCTCTATCTCGCCTCCA AGCAGGCAACACCAGGAG
	<i>IL-8</i>	Interleukin 8	AGACAGCAGAGCACACAAGC ATGGTTCCTTCCGGTGGT
	<i>IL-17</i>	Interleukin 17	TGGGAAGACCTCATTGGTGT GGATTTTCGTGGGATTGTGAT
	<i>IL-33</i>	Interleukin 33	TGTCAACAGCAGTCTACTGTGGAGTGCT GCAAAAGTAATGGATTGATCATTGTA

	<i>IL-1α</i>	Interleukin 1 alpha	GGTTGAGTTTAAGCCAATCCA TGCTGACCTAGGCTTGATGA
	<i>IL-1β</i>	Interleukin 1 beta	CRGTCCTGCGTGTTGAAAGA TTGGGTAATTTTTGGGATCTACA
	<i>IL-10</i>	Interleukin 10	TGGGGGAGAACCTGAAGAC CCTTGCTCTTGTTTTACAGG
	<i>CD14</i>	Cluster of differentiation 14	GTTCGGAAGACTTATCGACCAT ACAAGGTTCTGGCGTGGT
	<i>NFκB</i> <i>p50</i>	nuclear factor kappa B p50	ACCCTGACCTGCCTATTTG AGCTCTTTTTCCCGATCTCC
	<i>NFκB</i> <i>p65</i> (<i>RelA</i>)	nuclear factor kappa B p65 (RelA)	CGGGATGGCTTCTATGAGG CTCCAGGTCCCGCTTCTT
	<i>IκB-α</i>	Inhibitory-kappa B- alpha	GTCAAGGAGCTGCAGGAGAT GATGGCCAAGTGCAGGAA
	<i>bcl2</i>	B-cell lymphoma 2	AGTACCTGAACCGGCACCT GGCCGTACAGTTCCACAAA
	<i>BNIP</i>	BCL2 adenovirus E1B interacting	CCGGGATGCAGGAGGAGAG TTATAAATAGAAACCGAGGCTGGAAC
Differentiation	<i>IVL</i>	Involucrin	TGCCTCAGCCTTACTGTGAGT TCATTTGCTCCTGATGGGTA

	<i>LOR</i>	Loricrin	GTGGGAGCGTCAAGTACTCC AGAGTAGCCGCAGACAGAGC
	<i>FLG</i>	Filaggrin	GGCACTCATCATGCAGAGAA ATGGTGTCTGACCCTCTTG
	<i>TGM1</i>	Transglutaminase	TCTGTGGGTCCTGTCCCATCCATCCTGACC CCCCAACGGCCACATCGGAACGTGGCCCATCCATCATGC
	<i>KRT1</i>	Cytokeratin 1	GTTCCAGCGTGAGGTTTGT TAAGGCTGGGACAAATCGAC
	<i>KRT10</i>	Cytokeratin 10	GGCTCTGGAAGAATCAAACCTATGAGC GGATGTTGGCATTATCAGTTGTTAGG
	<i>KRT14</i>	Cytokeratin 14	CTGTCTCCCGCACCAGCTTCACCTCC CTCCACAAGCACCCGCAAGGCTGACC
	<i>FGF23</i>	Fibroblast growth factor 23	CAGCATGAGCGTCCTCAGAG GCCAGCATCCTCTGATCTGATC
House-keeping genes	<i>β-actin</i>	β-actin	CCAACCGCGAGAAGATGA CCAGAGGCGTACAGGGATAG
	<i>CYCB</i>	cyclophilin B	TGTGGTGTGGCAAAGTTC GTTTATCCCGGCTGTCTGTC
	<i>GAPDH</i>	Glyceraldehyde-3-Phosphate Dehydrogenase	AGCCACATCGCTCAGACAC GCCCAATACGACCAAATCCC

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