











Days of surgery



















Days after viral microinjection

Supplementary Figure 1 (a) Expression of FTO in the ipsilateral (Ipsi) L4 DRG and contralateral (Con) L5 DRG after SNL. n = 6 rats/time point. One-way ANOVA with repeated measures followed by *post hoc* Tukey test. (b) Expression of FTO in the ipsilateral L4/5 DRGs after injection of complete Freund's adjuvant (CFA) into unilateral hind paw. n = 6 rats/time point. One-way ANOVA with repeated measures followed by *post hoc* Tukey test.

6 Supplementary Figure 2 Effect of pre-microinjection of Runnx1 siRNA or scrambled siRNA 7 (Scr) into the ipsilateral L5 DRG of the rats on the development of SNL-induced mechanical 8 allodynia (a), heat hyperalgesia (b) and cold allodynia (c) on days 3 and 5 post-surgery. (a,d) 9 Paw withdrawal thresholds (PWT) in response mechanical stimulation on the ipsilateral (a) and 10 contralateral (d) sides. (b,c,e) Paw withdrawal latencies in response to heat stimulation in the 11 ipsilateral (b) and contralateral (e) and to cold stimulation on the ipsilateral side (c). n = 8rats/group. *P < 0.05, **P < 0.01 versus the Scr plus SNL group at the corresponding time 12 13 points by two-way ANOVA with repeated measures followed by *post hoc* Tukey test.

Supplementary Figure 3 Effect of microinjection of AAV5-*Runx1* or AAV5-*Gfp* into unilateral L4/5 DRGs of the rats on paw withdrawal thresholds (PWT) in response to mechanical stimulation (**a**) and on paw withdrawal latencies (PWL) in response to heat (**b**) and cold (**c**) stimuli on the ipsilateral (Ipsi) and contralateral (Con) sides at time points as shown. BL: baseline. n = 10 rats/group. *P < 0.05, **P < 0.01 versus the AAV5-*Runx1*-treated group on the ipsilateral side at the corresponding time points by two-way ANOVA with repeated measures followed by *post hoc* Tukey test.

Supplementary Figure 4 (a) Effect of pre-microinjection of *Fto* siRNA (siFTO), scrambled siRNA (Scr) or PBS into the ipsilateral L5 DRG of the rats on the expression of FTO, DNMT1, DNMT3a and Kv1.4 in the ipsilateral L5 DRG on day 7 after SNL or sham surgery. n = 6

1 rats/group. One-way ANOVA with repeated measures followed by *post hoc* Tukey test. **P <2 0.01 versus the corresponding naive group. #P < 0.01 versus the corresponding PBS-treated SNL group. (b,c) Effect of pre-microinjection of *Fto* siRNA (siFTO), scrambled siRNA (Scr) or 3 4 PBS into the ipsilateral L5 DRG of the rats on basal paw withdrawal responses to mechanical (b) 5 and heat (c) stimuli on the contralateral side at days indicated post-SNL or sham surgery. n = 66 rats/group. Two-way ANOVA with repeated measures followed by *post hoc* Tukey test. (d,e) 7 Effect of post-microinjection of Fto siRNA (siFTO), scrambled siRNA (Scr) or PBS into 8 ipsilateral L5 DRG of the rats on basal paw withdrawal responses to mechanical (d) and heat (e) 9 stimuli on the contralateral side at days indicated post-SNL or sham surgery. n = 6 rats/group. 10 Two-way ANOVA with repeated measures followed by *post hoc* Tukey test.

11 Supplementary Figure 5 (a,b) Levels of FTO protein (a) and mRNA (b) in PC12 cells 12 transfected with the *Fto* guide RNA (gRNA) or negative control sequence (NC). n = 3 biological repeats *P < 0.05, **P < 0.01 versus the NC-treated group by two-tailed unpaired Student's t-13 14 test. (c,d) Levels of FTO protein (c) and mRNA (d) in the cultured rat DRG neurons transduced 15 with AAV5-*Fto* guide RNA (gRNA) or AAV5-negative control sequence (NC). n = 3 biological repeats *P < 0.05, **P < 0.01 versus the AAV5-negative control sequence-treated group by two-16 17 tailed unpaired Student's t-test. (e) Expression of FTO protein in the ipsilateral L5 DRG on day 7 18 post-SNL or sham surgery from the rats microinjected with AAV5-Fto guide RNA (gRNA), PBS 19 or AAV5-negative control sequence (NC). n = 3 biological repeats (6 rats)/group. Two-way ANOVA with repeated measures followed by post hoc Tukey test. **P < 0.01 versus the PBS-20 21 treated sham group. #P < 0.01 versus the PBS-treated SNL group. (f) Levels of p-ERK1/2, ERK1/2 and GFAP in the ipsilateral L5 dorsal horn 7 days after SNL or sham surgery from the 22 23 rats microinjected with AAV5-Fto guide RNA (gRNA), PBS or AAV5-negative control

1 sequence (NC). n = 3 biological repeats (3 rats)/group. Two-way ANOVA with repeated measures followed by post hoc Tukey test. **P < 0.01 versus the corresponding PBS-treated 2 3 sham group. #P < 0.01 versus the corresponding PBS-treated SNL group. (g,h) Effect of pre-4 microinjection of AAV5-Fto guide RNA (gRNA), PBS or AAV5-negative control sequence 5 (NC) into ipsilateral L5 DRG on basal paw withdrawal responses to mechanical (g) and heat (h) 6 stimuli on the contralateral side at days indicated post-SNL or sham surgery. PWT: paw 7 withdrawal threshold. PWL: paw withdrawal latency. Two-way ANOVA with repeated measures 8 followed by post hoc turkey test. n = 5 rats/group. (i-l) Effect of pre-microinjection of AAV5-*Fto* 9 guide RNA (gRNA), PBS or AAV5-negative control sequence (NC) into ipsilateral L4/5 DRGs 10 on paw withdrawal thresholds (PWT) in response to mechanical stimulation (i, j) and paw 11 withdrawal latencies (PWL) in response to heat stimulation (**k**, **l**) stimuli on the ipsilateral (**i**, **k**) and contralateral (j, l) sides at days indicated post-CCI or sham surgery. Two-way ANOVA with 12 repeated measures followed by post hoc turkey test. n = 6 rats/group. **P < 0.01 versus the 13 14 corresponding PBS-treated sham group. #P < 0.01 versus the corresponding PBS-treated CCI 15 group.

Supplementary Figure 6 (a) FTO protein expression in the ipsilateral L4 DRG, spinal cord 16 (SC) and brain 7 weeks after microinjection of AAV5-Gfp (Gfp) or AAV5-Cre (Cre) into 17 unilateral L4 DRG from the $Fto^{fl/fl}$ mice. n = 3 biological repeats (6 mice). **P < 0.01 versus the 18 19 corresponding AAV5-Gfp-treated group by two-tailed unpaired Student's t-test. (b) FTO protein expression in the ipsilateral L4 DRG on day 14 post-SNL or sham surgery from the *Fto*^{fl/fl} mice 20 21 microinjected with AAV5-*Cre* or AAV5-*Gfp*. n = 3 biological repeats (12 mice)/group. One-way ANOVA followed by post hoc Tukey test. **P < 0.01 versus the PBS-treated sham group. ##P <22 23 0.01 versus the AAV5-Gfp-treated SNL group. (c, d) Effect of pre-microinjection of AAV5-Gfp

(*Gfp*) or AAV5-*Cre* (*Cre*) into ipsilateral L4 DRG of the *Fto*^{fl/fl} mice on basal paw withdrawal
responses to mechanical (c) and heat (d) stimuli on the contralateral side at days indicated post SNL or sham surgery. PWF: paw withdrawal frequency. PWL: paw withdrawal latency. Two way ANOVA with repeated measures followed by post hoc turkey test. n = 10 mice/group.

5 Supplementary Figure 7 (a) FTO protein expression in the DRGs, spinal cord (SC) and cortex from the *Fto*^{fl/fl} mice (fl/fl) and the conditional *Fto* knockout mice (KO). (b) Level of FTO 6 protein in the ipsilateral L4 DRG on day 14 post-SNL or sham surgery from the *Fto*^{fl/fl} mice 7 8 (fl/fl) and the conditional *Fto* knockout mice (KO). n = 3 biological repeats (12 mice)/group. One-way ANOVA with repeated measures followed by post hoc Tukey test. *P < 0.05, **P < 0.059 0.01 versus the sham $Fto^{\text{fl/fl}}$ mice. ##P < 0.01 versus the SNL conditional *Fto* knockout mice. (**c**-10 11 i) Paw withdrawal frequency (PWF) to low (0.07 g; c and g) and medium (0.4 g; d and h) force 12 von Frey filament stimuli and paw withdrawal latency (PWL) to heat (e and i) and cold (f) stimuli on the ipsilateral (c-f) and contralateral (g-i) sides from $Fto^{fl/fl}$ mice (fl/fl) or conditional 13 14 *Fto* knockout mice (KO) after SNL or sham surgery. n = 10 mice/group. **P < 0.01 versus the SNL *Fto*^{fl/fl} mice at the corresponding time points by two-way ANOVA with repeated measures 15 followed by post hoc Tukey test. 16

Supplementary Figure 8 (a) FTO protein expression in the cultured rat DRG neurons transduced with AAV5-*Fto* (*Fto*) or AAV5-*Gfp* (*Gfp*). n = 3 biological repeats/group. ***P* < 0.01 versus the AAV5-*Gfp*-treated group by two-tailed unpaired Student's t-test. (b) FTO protein expression in the ipsilateral L4/5 DRGs 8 weeks after microinjection of AAV5-*Fto* (*Fto*) or AAV5-*Gfp* (*Gfp*) into unilateral L4/5 DRGs of the rats. n = 3 biological repeats (6 rats)/group. ***P* < 0.01 versus the AAV5-*Gfp*-treated group by two-tailed unpaired Student's t-test. (c, d) Basal paw withdrawal response to mechanical (c) and heat (d) stimuli on the contralateral side on weeks as indicated after microinjection of AAV5-*Fto* (*Fto*) or AAV5-*Gfp* (*Gfp*) into
unilateral L4/5 DRGs of the rats. BL: baseline. PWT: paw withdrawal threshold. PWL: paw
withdrawal latency. n = 12 rats/group. Two-way ANOVA with repeated measures followed by
post hoc Tukey test.

Supplementary Figure 9 (a) Dynamic changes of m⁶A sites across transcripts from the 5 6 ipsilateral L5 DRG on day 7 after SNL or sham surgery. This chart is based on clustering relative 7 tag numbers of peak-associated genes. The intragenic regions of peak-associated genes are 8 classified into four categories: intron, CDS (coding sequence region), 5'-UTR and 3'-UTR. 9 Peaks are categorized into each of the intragenic regions. Thus, each gene has four values 10 according to the relative tag numbers in the four regions. The relative tag numbers are log2_ratio 11 of different tag numbers ((tag number in the SNL group) – (tag number in the sham group)) in 12 peaks in each region. If no peaks were detected in one gene locus in both sham and SNL groups, 13 this gene was discarded for analysis. (b,c) GO enrichment analyses of the transcripts with 14 increased (b) and decreased (c) m^6A sites from the ipsilateral L5 DRG on day 7 after SNL or 15 sham surgery. (d) FTO protein expression in cultured rat DRG neurons transduced with the viruses as indicated. Gfp: AAV5-Gfp. Fto: AAV5-Fto. gRNA: AAV5-Fto guide RNA. NC: 16 17 AAV5-control negative sequence. n = 3 biological repeats/group. One-way ANOVA with 18 repeated measures followed by post hoc Tukey test. **P < 0.01 versus the *Gfp* plus NC-treated 19 group. #P < 0.01 versus the *Fto* plus NC-treated group. (e,f) Levels of FTO and G9a proteins 20 (e) and *Fto* mRNA (f) in PC12 cells transduced by the vectors as shown. *Gfp*: vector expressing 21 *Gfp. Fto*: vector expressing full-length *Fto*. Scr: vector expressing scrambled shRNA. shRNA: 22 vector expressing Fto shRNA. n = 3 biological repeats. One-way ANOVA with repeated

1 measures followed by *post hoc* Tukey test. *P < 0.05 versus corresponding naive group. #P <2 0.05, ##P < 0.01 versus the corresponding *Fto* plus Scr-treated group.

3 Supplementary Figure 10 (a) Levels of G9a and MOR protein in the ipsilateral L5 DRG on day 4 7 post-SNL or sham surgery from the rats microinjected with AAV5-Fto guide RNA (gRNA), 5 PBS or AAV5-negative control sequence (NC). n = 3 biological repeats (6 rats)/group. One-way ANOVA with repeated measures followed by post hoc Tukey test. **P < 0.01 versus the 6 7 corresponding PBS-treated sham group. #P < 0.01 versus the corresponding PBS-treated SNL 8 group. (b) Levels of FTO, G9a and MOR proteins in the cultured rat DRG neurons transduced 9 with AAV5-Fto (Fto) or AAV5-Gfp (Gfp). n = 3 biological repeats. **P < 0.01 versus the 10 corresponding AAV5-Gfp-treated group by two-tailed unpaired Student's t-test. 11 Supplementary Figure 11 Effect of microinjection of *Ehmt2* siRNA (*siEhmt2*) or scrambled 12 siRNA (Scram) into ipsilateral L4/5 DRGs on contralateral paw withdrawal responses to

13 mechanical (**a**) and heat (**b**) stimuli at time points as shown in the rats pre-microinjected with 14 AAV5-*Fto* (*Fto*) or AAV5-*Gfp* (*Gfp*) into unilateral L4/5 DRGs. PWT: paw withdrawal 15 threshold. PWL: paw withdrawal latency. n = 10 rats/group. Two-way ANOVA with repeated 16 measures followed by *post hoc* Tukey test.

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Treatments/genotyping	Functional test		
	Placing	Grasping	Righting
<i>Fto</i> siRNA + Sham	5 (0)	5 (0)	5 (0)
Fto siRNA + SNL	5 (0)	5 (0)	5 (0)
<i>Fto</i> scramble + SNL	5 (0)	5 (0)	5 (0)
PBS + SNL	5 (0)	5 (0)	5 (0)
PBS + Sham	5 (0)	5 (0)	5 (0)
sg3+Sham	5 (0)	5 (0)	5 (0)
sg3+ SNL	5 (0)	5 (0)	5 (0)
NC + Sham	5 (0)	5 (0)	5 (0)
NC + SNL	5 (0)	5 (0)	5 (0)
<i>Fto^{fl/fl}</i> + AAV5-Cre + Sham	5 (0)	5 (0)	5 (0)
<i>Fto^{fl/fl}</i> + AAV5-Cre + SNL	5 (0)	5 (0)	5 (0)
<i>Fto^{fl/fl}</i> + AAV5-GFP + Sham	5 (0)	5 (0)	5 (0)
$Fto^{fl/fl} + AAV5-GFP + SNL$	5 (0)	5 (0)	5 (0)
AAV5-GFP	5 (0)	5 (0)	5 (0)
AAV5-FTO	5 (0)	5 (0)	5 (0)
$Fto^{fl/fl}$ + Sham	5 (0)	5 (0)	5 (0)
$Fto^{fl/fl} + SNL$	5 (0)	5 (0)	5 (0)
$Adv^{cre} Fto^{fl/fl} + Sham$	5 (0)	5 (0)	5 (0)
$Adv^{cre} Fto^{fl/fl} + SNL$	5 (0)	5 (0)	5 (0)

Supplementary Table 1: Locomotor functions

Scores for placing, grasping and righting reflexes were based on counts of each normal reflex

exhibited in five trials. All values are Mean (SEM). n = 10 rats or mice/group.

PCR or Real-RT-PCR
Fto-F 5'- TGGATCCGTGCATCTGTAAAGCTCAGG -3'
Fto-R 5'- CCTGACAAATAACGACAATCGAGATG -3'
Ehmt2-F 5'-TGCCTATGTGGTCAGCTCAG-3'
Ehmt2-R 5'-GGTTCTTGCAGCTTCTCCAG-3'
Cre-F 5'-TGC CAC GAC CAA GTG ACA GCA ATG-3'
Cre-R 5'-ACC AGA GAC GGA AAT CCA TCG CTC-3'
Rat-Fto-F 5'- TCTTACAACGCTGCCAGTTG -3'
Rat-Fto-R 5'- GAAACCAGAACTGCCTCAGC -3'
Mouse-Fto-F 5'- GCGGGAAGCTAAGAAACTGA -3'
Mouse-Fto-R 5'- ATGCAGCTCCTCTGGTATGC -3'
Rat-Runx1-F 5'- CGAAGACATCGGCAGAAACT -3'
Rat-Runx1-R 5'- GCTGAGGGTTAAAGGCAGTG -3'
Rat-Ehmt2-F 5'- GATCATCTGCCGGGATGTAG -3'
Rat-Ehmt2-R 5'- AGTGCTGCAGGTGAGTGATG -3'
Mouse-Ehmt2-F 5'- AAATTGGGAACTTGGAAATGG-3'
Mouse-Ehmt2-R 5'- CACTACCCGTGAAGGAGGC-3'
Gapdh-F 5'-TCG GTG TGA ACG GAT TTG GC-3'
Gapdh-R 5'-TCC CAT TCT CGG CCT TGA CT-3'
Rat-Oprm1-F 5'- TTCCTGGTCATGTATGTGATTGT-3'
Rat-Oprm1-R 5'- GGGCAGTGTACTGGTCGCTAA-3'
Rat-Oprk1-F 5'- TTTGTGGTGGGCTTAGTGGG -3'
Rat-Oprk1-R 5'- CTCTGGAAGGGCATAGTGGT -3'
Kcna2-F 5'- CTGCAAGGGCAACGTCACAC -3'
Kcna2-R 5'-GGGACAGTGAGATGCTTGGC-3'
ChIP-PCR
Fto-ChIP-F 5'- GAGTGGCACCACACCCTACT -3'
<i>Fto-</i> ChIP-R 5'- GACAGGGTAGGGGGGCGCGTAT-3'
RIP-PCR
Rat Ehmt2-RIP-F 5'- GATCATCTGCCGGGATGTAG-3'
Rat Ehmt2-RIP-R 5'-AGTGCTGCAGGTGAGTGATG-3'
Fto Guide RNA
GATGAGGATGCGAGACACCGGG (PAM: GTGAG, targeting exon 3)
Fto shRNAs
<i>Fto</i> siRNA: ACGUGACUUUGCUAAACUUTT
<i>Fto</i> siRNA: AAGUUUAGCAAAGUCACGUTG
Scramble siRNA: AAGGCUCUAUGAAGAGGCUTG
Cloning
FTO shRNA sense:
GATCCGATGAAGTGGACCTTAAGAGAAGCTTGTCTTAAGGTCCACTTCATCTTT
TTTT
FTO shRNA antisense:
CTAGAAAAAAAGATGAAGTGGACCTTAAGACAAGCTTCTCTTAAGGTCCACTTC

Supplementary Table 2. All primers and other sequences used.

ATCG Runx1 shRNA sense: GATCCGCAGGCTCCTACCAATTCTGAAGCTTGAGAATTGGTAGGAGCCTGCTTT TTTT Runx1 shRNA antisense: CTAGAAAAAAGCAGGCTCCTACCAATTCTCAAGCTTCAGAATTGGTAGGAGC CTGCG Scramble shRNA sense: GATCCGGTTCAGATGTGCGGCGAGTGAAGCTTGACTCGCCGCACATCTGAACCT TTTTTT Scramble shRNA antisense: CTAGAAAAAAGGTTCAGATGTGCGGCGAGTCAAGCTTCACTCGCCGCACATCT GAACCG Runx1 RT F: TAACCCTGCCTGGGTGTAAG Runx1 RT R: AATAACGACCACCCAGATGC Runx1 nested F: CGCTCTAGAGCCACCATGCGTATCCCCGTAGA Runx1 nested R: ATATAGCGGCCGCTCAGTAGGGCCGCCAGACA Fto RT F: ATGAAGCGCGTCCAGACCGC Fto RT R: GGGAGAAAAGCCAAGGACAT Fto nested F: ATATCCGGAGCCACCATGAAGCGCGTCCAGACC Fto nested R: GCTAGCGGCCGCCTAGGATCTTGCTTCCAG FTO-luc-F: ATAGGTACCTAAGGGAAGCCTATGCAAGC FTO-luc-R: ATCAAGCTTACTCAGGCCTGCATCACAG m⁶A-eCLIP-seq 3'-RNA adaptor 1: ATTGCTT AGATCGGAAGAGCACACGTCT 3'-RNA adaptor 2: ACAAGCCAGATCGGAAGAGCACACGTCT 3'-RNA adaptor 3: AACTTGT AGATCGGAAGAGCACACGTCT nested specific primer: AGACGTGTGCTCTTCCGA

RT: reverse transcription. F: forward. R: reverse.