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Supplementary Information

A nongenomic mechanism for “metalloestrogenic” effects of cadmium in human uterine leiomyoma cells through G protein-coupled estrogen receptor

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Content:**Part 1: Methods**

The text includes details of immunohistochemistry and real time-PCR analysis for GPER.

Part 2: 1 Table

Table S1. List of antibodies used for western blotting.

Part 3: 6 Figures

Figure S1. Expression of GPER, ER α 36, and phospho-EGFR (P-EGFR) in patient-matched myometrial (M) and uterine leiomyoma (L) tissues.

Figure S2. Percent of ht-UtLM cells positive for GPER by immunofluorescence.

Figure S3. Effects of Cd on cell proliferation in ht-UtLM cells.

Figure S4. Percent of ht-UtLM cells positive for phospho-MAPK by immunofluorescence.

Figure S5. Activation of different tyrosine (Tyr) residues of EGFR in Cd-treated ht-UtLM cells for 10 min. Figure

S6. Mechanism of EGFR transactivation by GPER.

Methods

Immunohistochemistry

Human uterine leiomyoma and myometrial tissue samples were embedded in paraffin and stained for GPER by immunohistochemistry. Sections were deparaffinized and rehydrated. Antigen retrieval was performed with proteinase K retrieval (Agilent Technologies, Santa Clara, CA) at room temperature for 2 minutes. Endogenous peroxidase was blocked with IHC/ISH peroxidase block (Enzo Life Sciences, Farmingdale, NY) for 5 minutes. The sections were incubated with Antibody Blocker/Diluent (Enzo Life Sciences, Farmingdale, NY) for 10 minutes, followed by incubation with GPER (Catalog no. ab39742, abcam, Cambridge, MA) polyclonal IgG primary antibody and an equivalent dilution of rabbit IgG, polyclonal – isotype control (negative control; EMD Millipore, Billerica, MA) for 60 minutes at 1:100 dilution. Sections were then incubated with Polyview plus HRP (Enzo Life Sciences, Farmingdale, NY) for 30 minutes. Antigen-antibody complexes were visualized using HDDAB (Enzo Life Sciences, Farmingdale, NY) for 5 minutes and counterstained with ENZO hematoxylin for 30 seconds, dehydrated, cleared, and coverslipped.

Real Time-PCR analysis

Ht-UtLM cells treated with Cd (0.1 μ M and 10 μ M) were collected at 24 h, 48 h and 72 h for extraction of total RNA using a TRIzolTM Reagent (catalog no. 15596026, Thermo Fisher Scientific). After DNase treatment and RNA purification, one microgram of total RNA was used to prepare cDNA and primed with GPER and GAPDH (house-keeping gene used as a control) and reverse transcribed with High-Capacity cDNA Reverse Transcription Kit (catalog no. 4368814, Thermo Fisher Scientific). The primer sequence for GPER is: forward primer 5'-TGCACCTTCATGTCGCTCTT-3' and reverse primer 5'-GCGGTCGAAGCTCATCCA-3'. The primer sequence for GAPDH is: forward primer 5'-AATCCCATCACCATCTTCCA-3' and reverse primer 5'-TGGACTCCACGACGTACTCA-3'. Real time-PCR was performed with Applied Biosystems Power SYBR Green PCR Mix on an AB cycler (Quantstudio 7 Flex, Thermo Fisher Scientific). The data analysis was based on the $\Delta\Delta$ Ct method with normalization to GAPDH.

Table S1. List of antibodies used for western blotting.

| Name of antibody | Dilution | Catalog No. | Company |
|-------------------------|-----------------|--------------------|---------------------------|
| phospho-EGFR (Y845) | 1:400 | AF3394 | R&D |
| EGFR | 1:1000 | 2232 | Cell signaling technology |
| phospho-p44/42 MAPK | 1:1000 | 9101 | Cell signaling technology |
| p44/42 MAPK | 1:1000 | 9102 | Cell signaling technology |
| phospho-Src | 1:1000 | 2101 | Cell signaling technology |
| Src | 1:1000 | 2123 | Cell signaling technology |
| MMP2 | 1:1000 | 4022 | Cell signaling technology |
| MMP9 | 1:1000 | 3852 | Cell signaling technology |
| TIMP2 | 1:1000 | 5738 | Cell signaling technology |
| TIMP1 | 1:1000 | 8946 | Cell signaling technology |
| HB-EGF | 1:4000 | 185555 | Abcam |
| HPRT | 1:1000 | 20975 | Santa cruz |

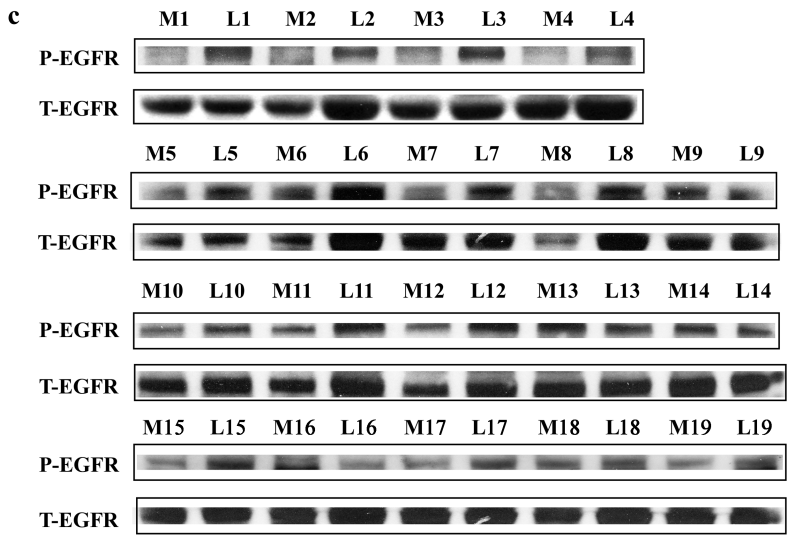
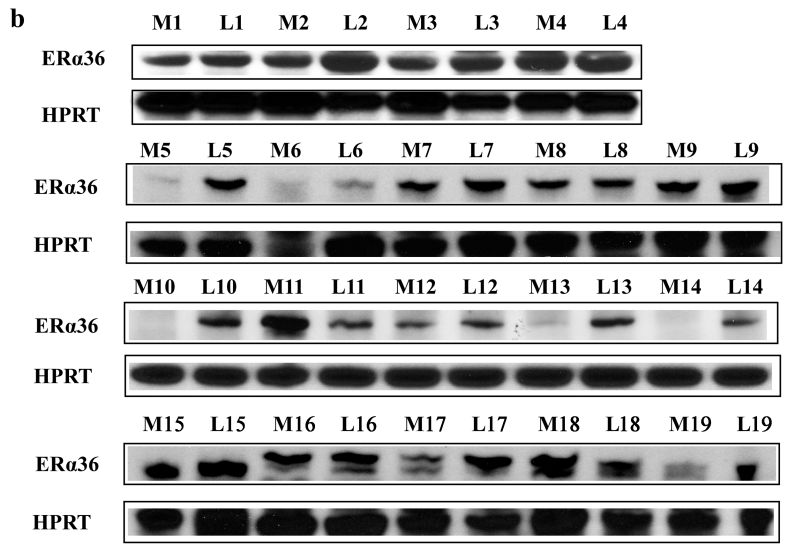
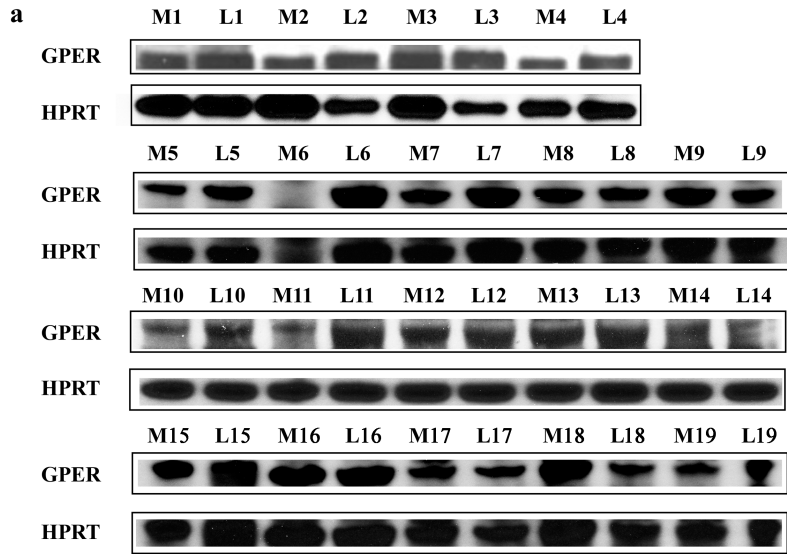


Fig. S1 Expression of GPER, ER α 36, and phospho-EGFR (P-EGFR) in patient-matched myometrial (M) and uterine leiomyoma (L) tissues. (a) GPER (b) ER α 36 and (c) phospho-EGFR by western blotting. n = 15 pairs of human tissues.

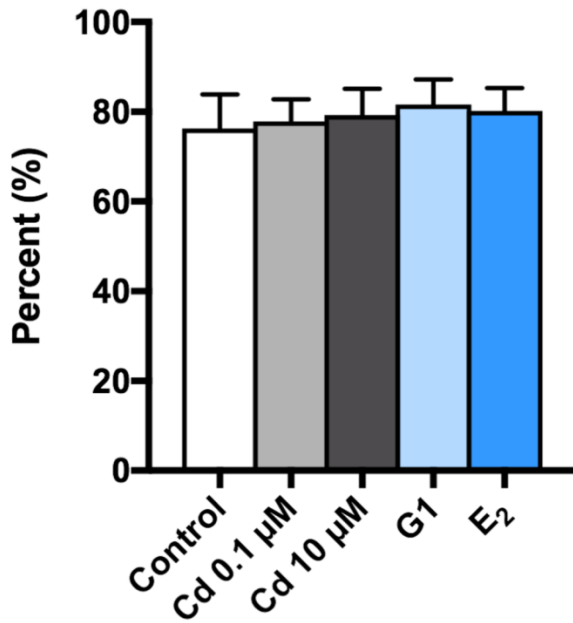


Fig. S2 Percent of ht-UtLM cells positive for GPER by immunofluorescence.

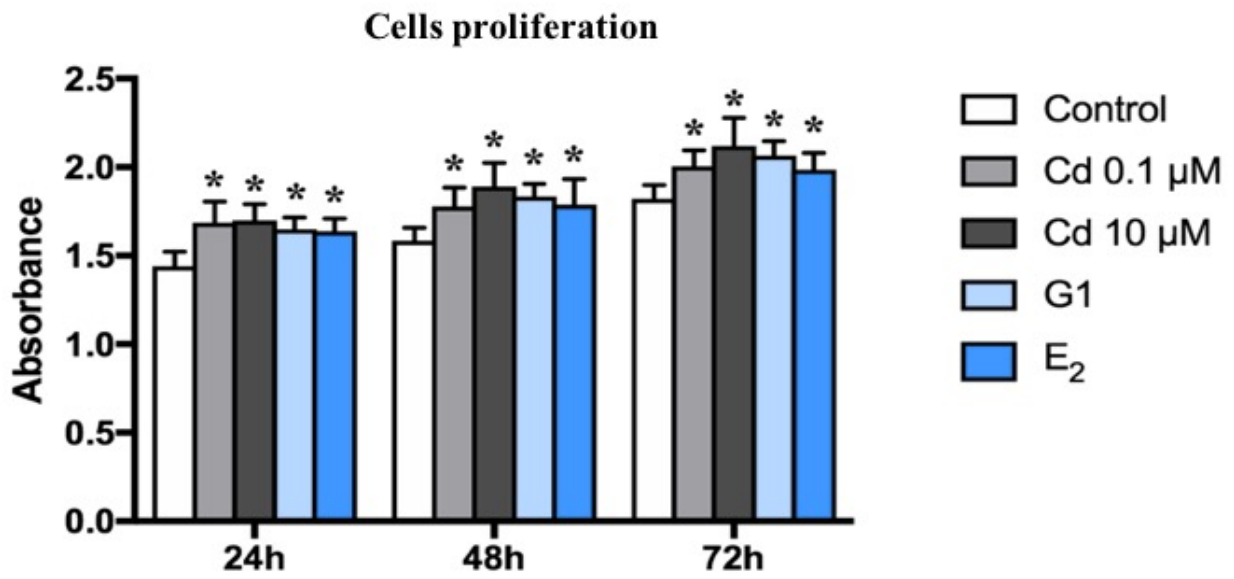


Fig. S3 Effects of Cd on cell proliferation in ht-UtLM cells. * $P < 0.05$ compared to control.

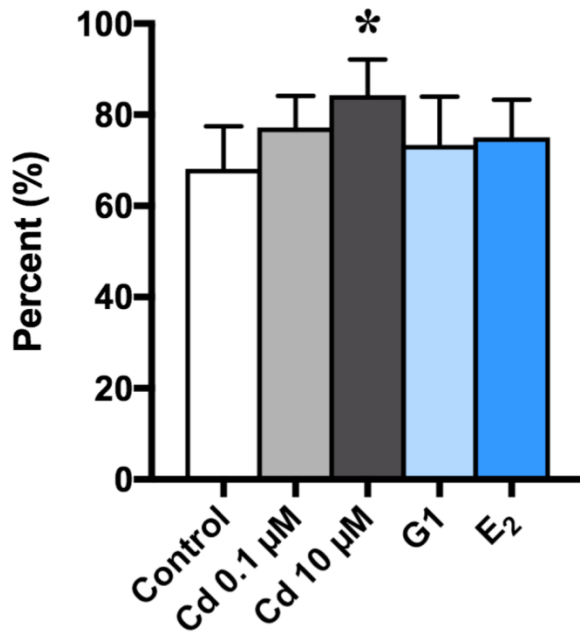


Fig. S4 Percent of ht-UtLM cells positive for phospho-MAPK by immunofluorescence. * $P < 0.05$ compared to control.

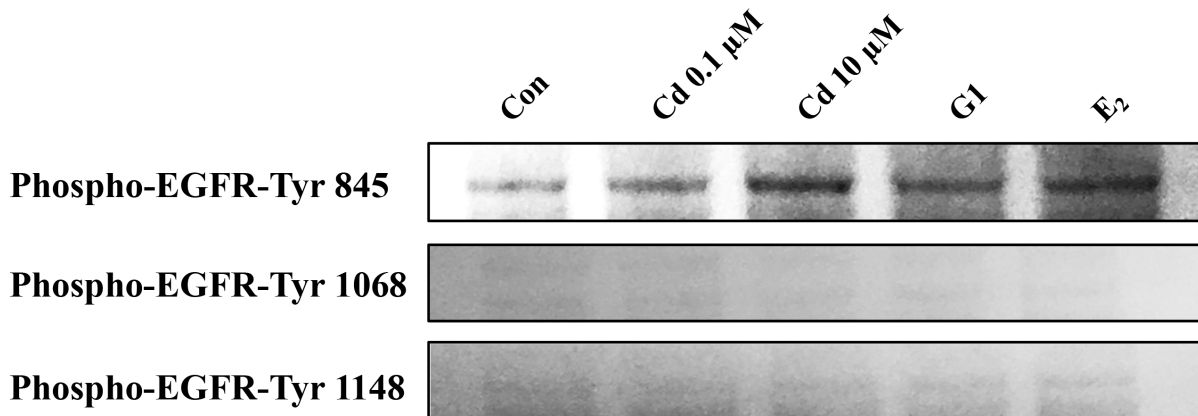


Fig. S5 Activation of different tyrosine (Tyr) residues of EGFR in Cd-treated ht-UtLM cells for 10 min.

Representative immunoblots by western blotting.

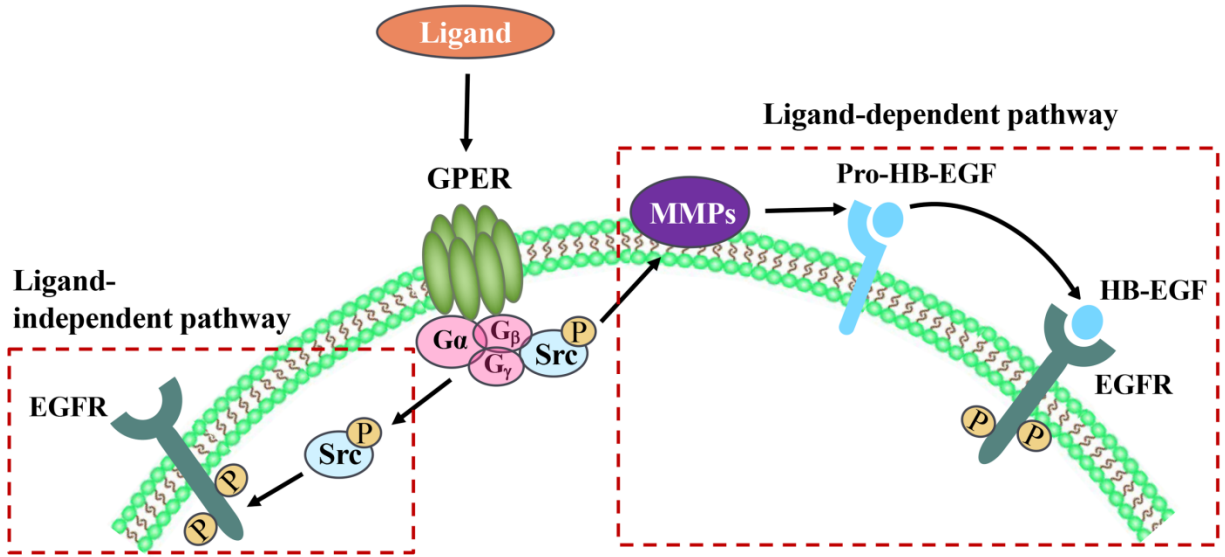


Fig. S6 Mechanisms of EGFR transactivation by GPER.