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## **Supplemental Information**

**METTL3 Induces AAA Development and Progression**

**by Modulating N6-Methyladenosine-Dependent**

**Primary miR34a Processing**

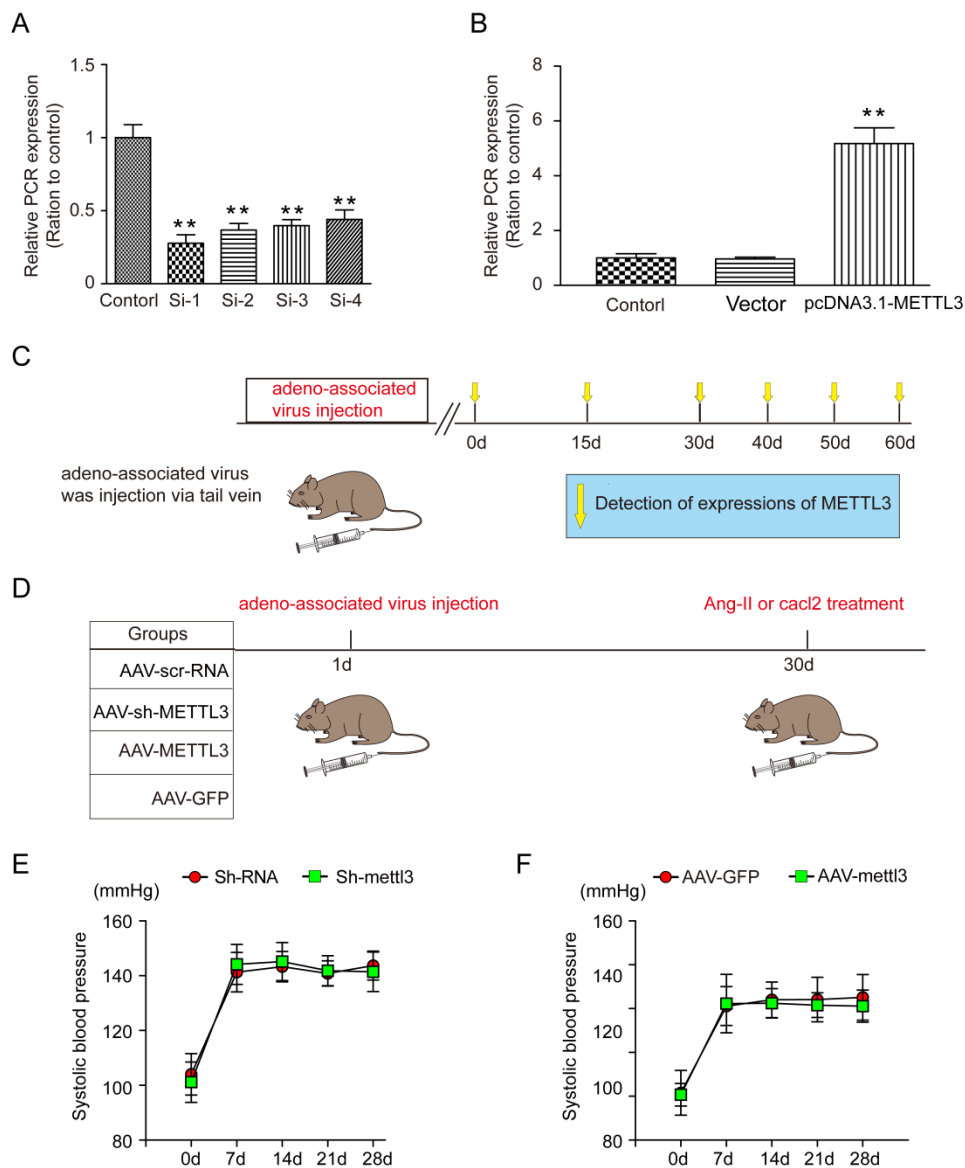
**Lintao Zhong, Xiang He, Haoyu Song, Yili Sun, Guojun Chen, Xiaoyun Si, Jie Sun, Xiaoqiang Chen, Wangjun Liao, Yulin Liao, and Jianping Bin**

**METTL3 induces abdominal aortic aneurysm development and progression by modulating N6-methyladenosine-dependent primary miR34a processing**

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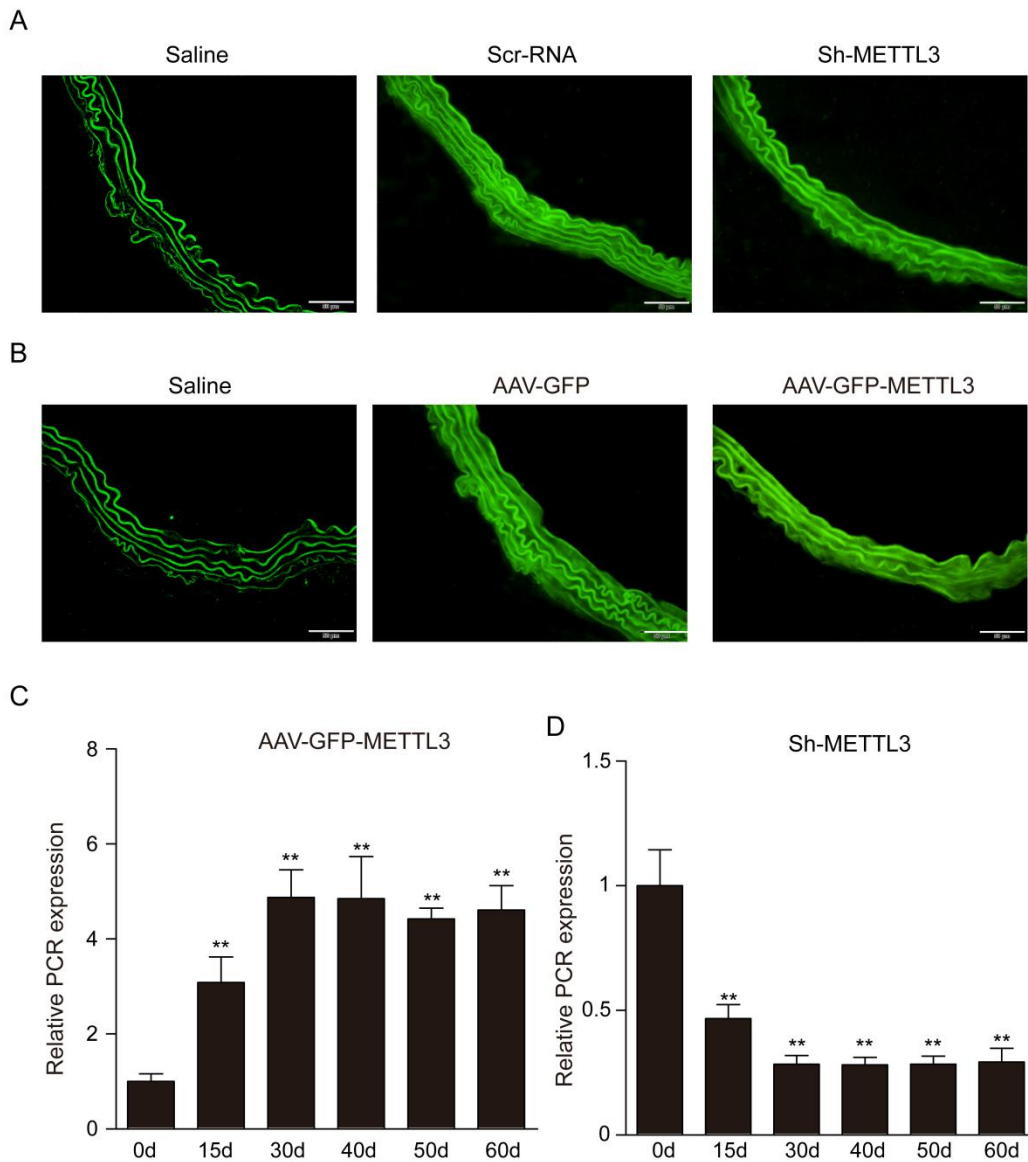
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Online Figure 1

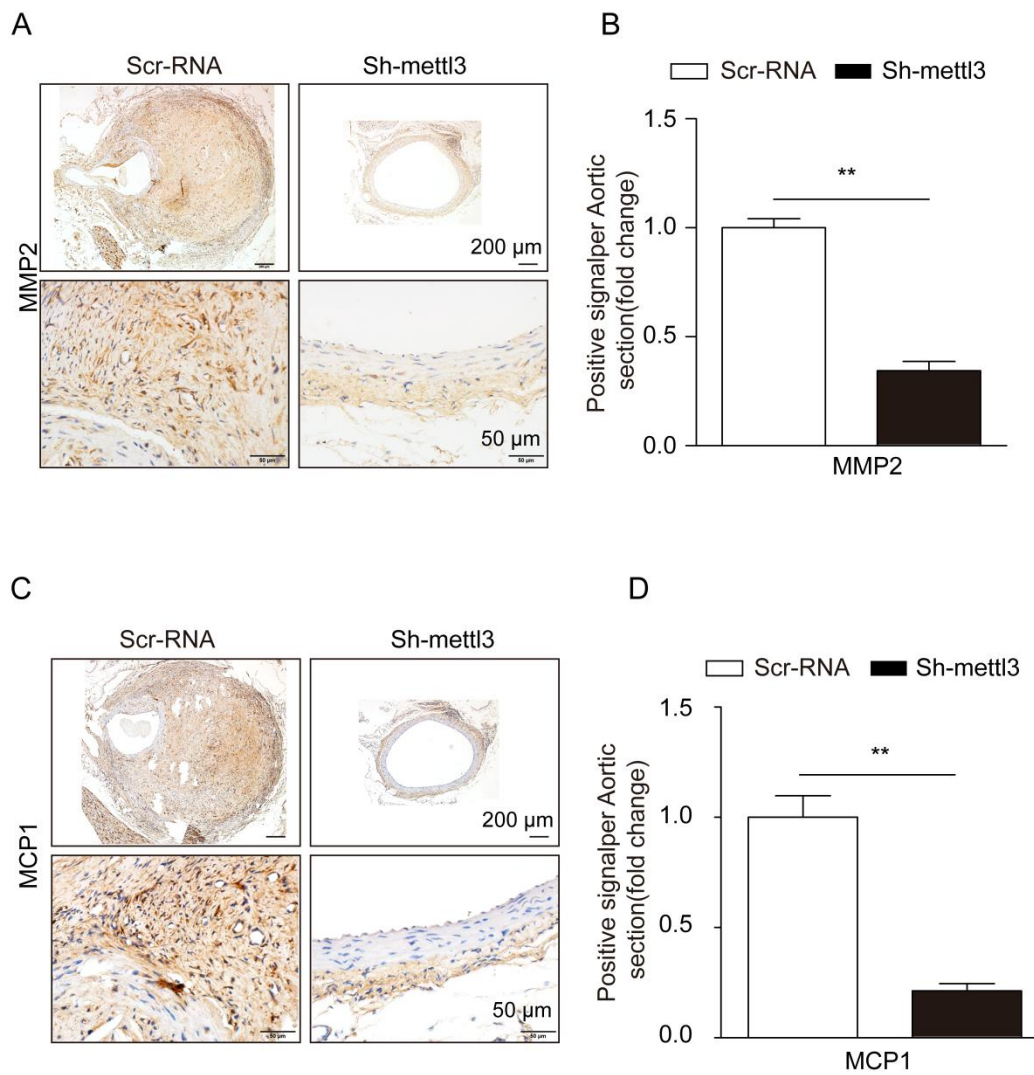
**Online Figure 1.** Selection of a potent adeno-associated virus (AAV) carrying METTL3 siRNA and overexpression plasmids and a diagram of the virus-related experimental flow. A, Inhibitory effects of four small interfering RNAs (siRNAs) against METTL3 in VSMCs as assessed by qRT-PCR analysis. B, Overexpression effects of the constructed pcDNA3.1-METTL3 plasmid in VSMCs as assessed by qRT-PCR analysis. C, Protocol for the in vivo AAV-mediated METTL3 knockdown and overexpression experiment. A predetermined number of male mice were injected with Scr-RNA, sh-METTL3, AAV-GFP or AAV-METTL3. Before injection and at 15 days, 30 days, 40 days, 50 days, and 60 days after the initial injection, a predetermined number of mice were sacrificed, and aortic samples were collected to detect aortic METTL3 expression (n=3). D, Thirty days after the initial AAV transfection, mice were treated with Ang II via minipump for four weeks or were

subjected to CaCl<sub>2</sub>-treatment surgery and sacrificed after 3 or six weeks. The data are presented as the mean  $\pm$  SD. \*P<0.05, \*\*P<0.01.



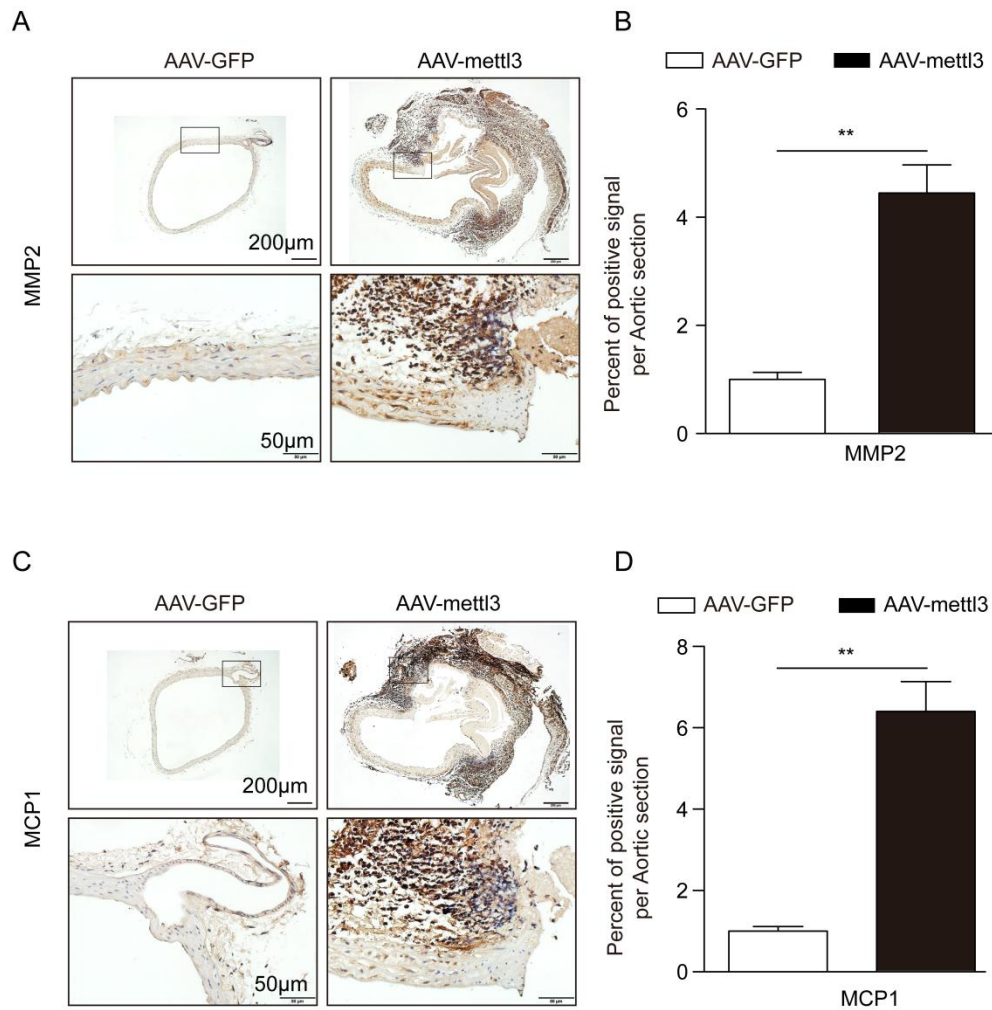
OnlineFigure 2

**Online Figure 2.** Confirmation of the viral infection efficiency in the suprarenal aortas of mice. A and B, Representative immunofluorescent staining of virus-borne green fluorescent protein (GFP) in the aortas of mice from the different virus-mediated groups and the saline group (scale bar, 50  $\mu$ m). C and D, qRT-PCR analysis of the mRNA expression of METTL3 over time after sh-METTL3 or AAV-METTL3 transfection in the suprarenal aortas of mice (n=3). The data are presented as the mean  $\pm$  SD. \*P<0.05.



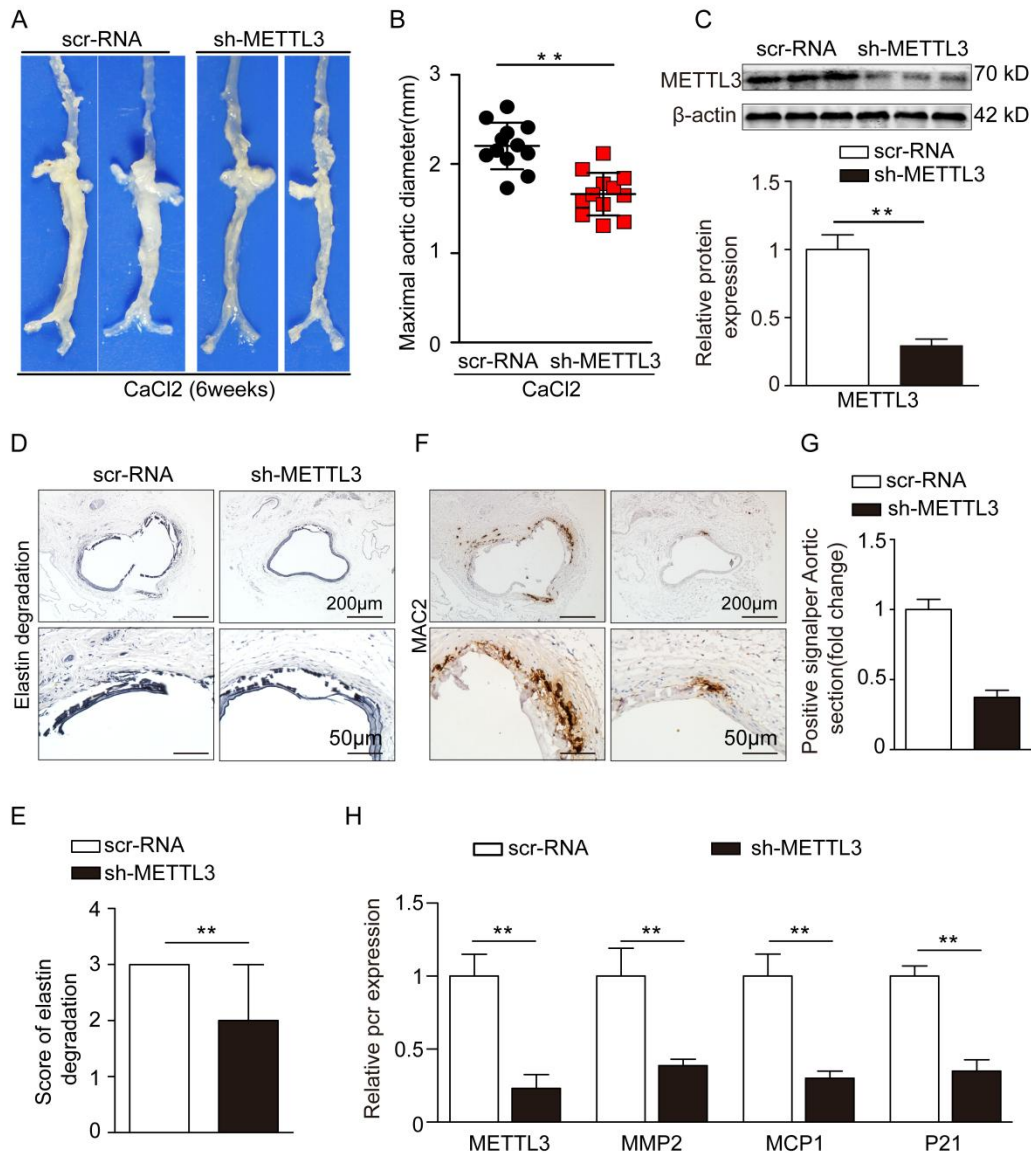
Online Figure 3

**Online Figure 3.** METTL3 knockdown attenuates MMP2 and MCP1 expression in Ang II-infused ApoE<sup>-/-</sup> mice. A to D, Immunohistochemical staining of abdominal aortic MMP2 (A,B) and MCP1 (C,D) in Ang II-infused male ApoE<sup>-/-</sup> mice (n=3; scale bars, 200 μm (upper) and 50 μm (lower)). The data are presented as the mean ± SD. \*P<0.05.



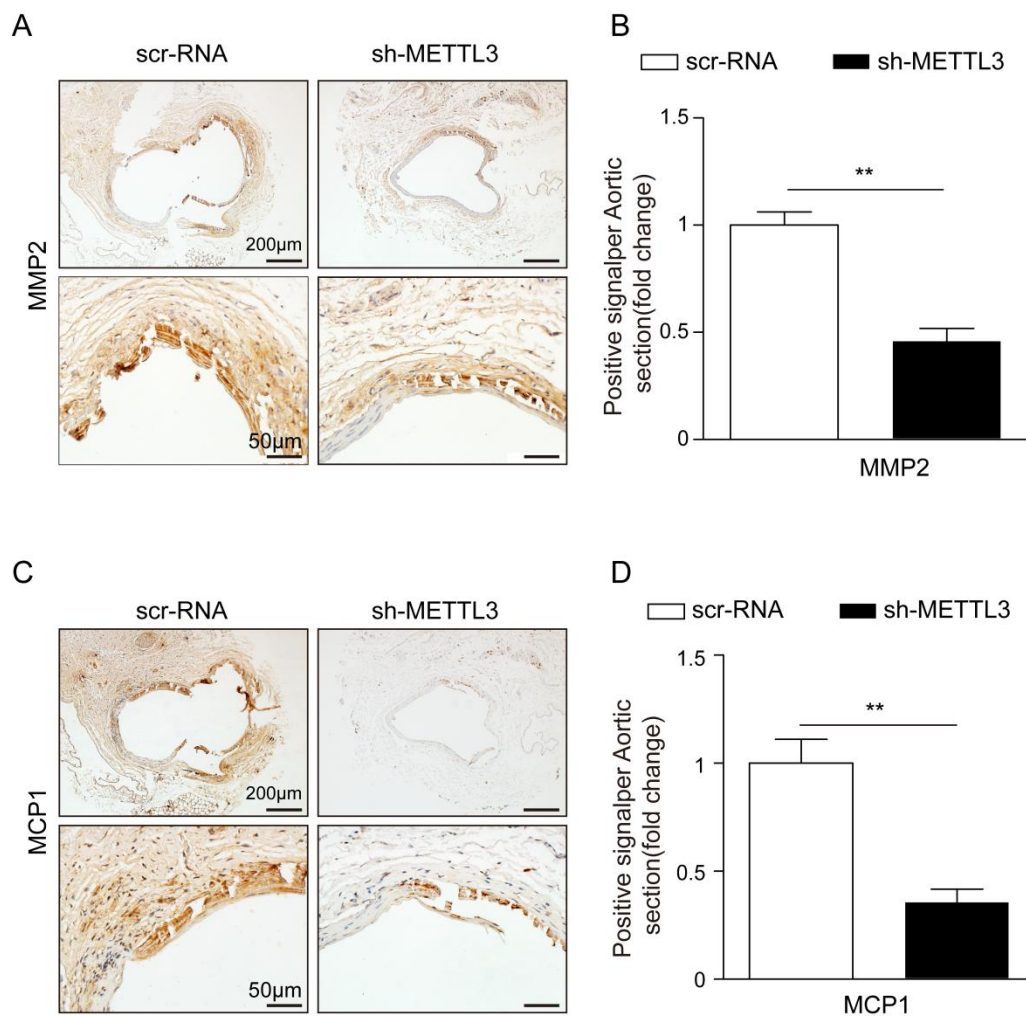
Online Figure 4

**Online Figure 4.** Overexpression of METTL3 increases the expression of MMP2 and MCP1 in Ang II-infused C57BL/6J mice. A to D, Immunohistochemical staining of abdominal aortic MMP2 (A,B) and MCP1 (C,D) in Ang II-infused C57BL/6J mice (n=3; scale bars, 200 µm (upper) and 50 µm (lower)). The data are presented as the mean  $\pm$ SD. \*P<0.05.



Online Figure 5

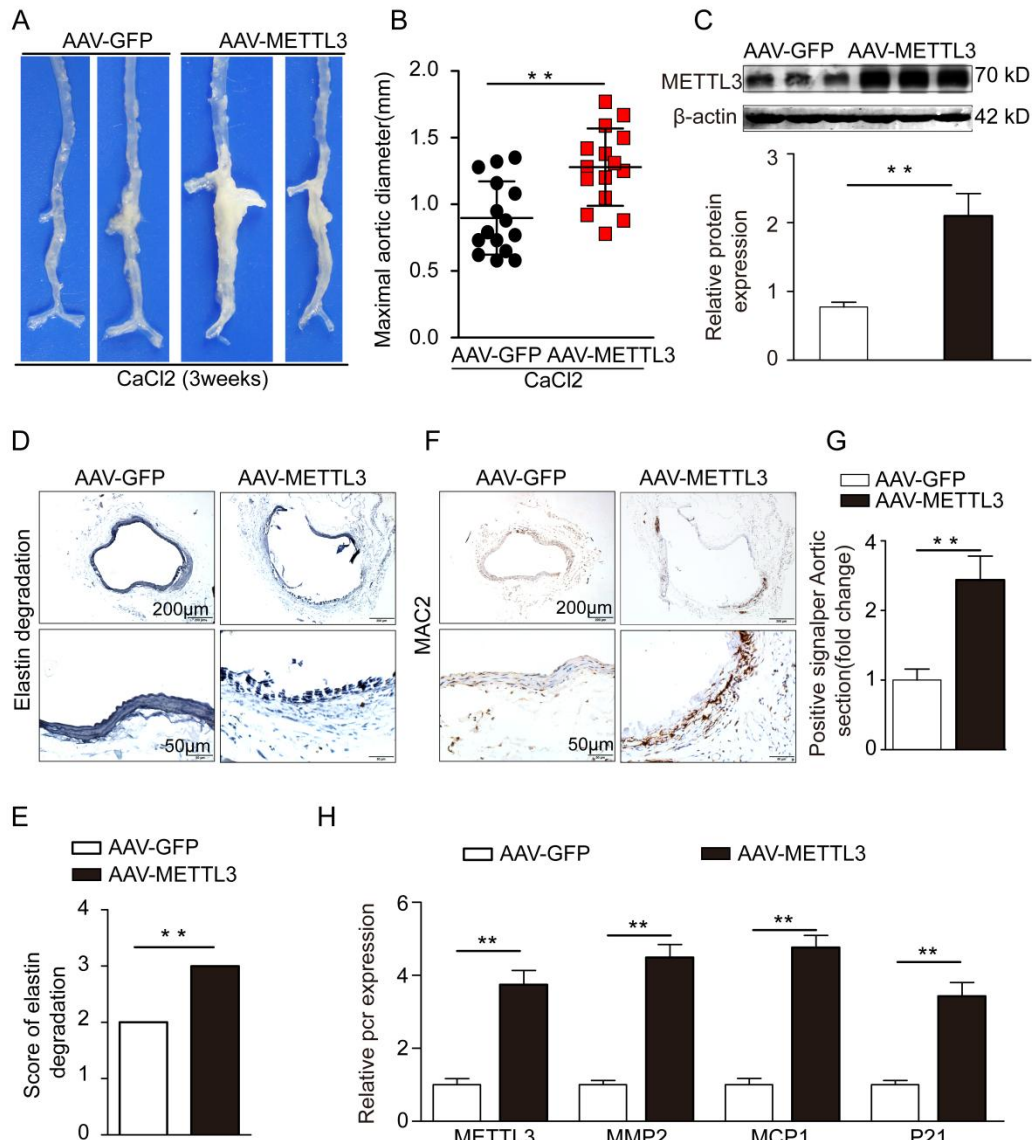
**Online Figure 5.** METTL3 suppression inhibits CaCl<sub>2</sub>-induced AAA formation. C57BL/6J mice were injected with Scr-RNA or sh-METTL3, and their infrarenal aortas were then treated with CaCl<sub>2</sub> for 15 min. After six weeks, the mice were sacrificed and used for analysis. A, Representative images showing C57BL/6J mouse infrarenal aortas treated with CaCl<sub>2</sub>. B, Maximal diameters of infrarenal aortas from CaCl<sub>2</sub>-induced C57BL/6J mice. C, Representative western blots and statistical analysis of aortic METTL3 in CaCl<sub>2</sub>-treated C57BL/6J mice. D and E, Representative elastin staining and statistical analysis of elastin degradation scores in the two groups of mice. F and G, Representative immunohistochemical staining (F) and statistical analysis (G) of MAC2 in aortas from the two groups of mice (n=4 per group). G and H, mRNA levels of aortic METTL3, MCP-1, MMP-2 and P21 in CaCl<sub>2</sub>-treated C57BL/6J mice. The data are presented as the mean ± SD. \*P<0.05, \*\*P<0.01.



Online Figure 6

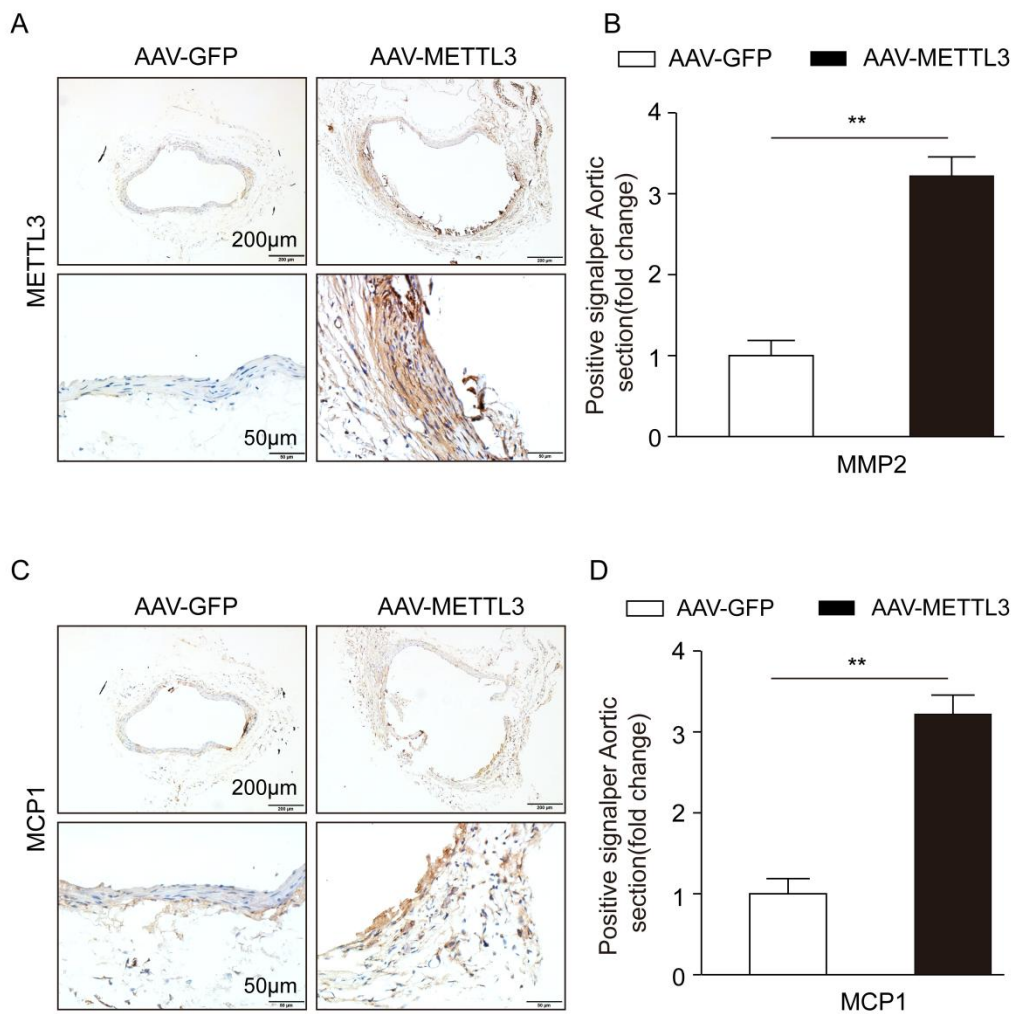
**Online Figure 6.** METTL3 suppression inhibits vascular MMP2 and MCP1 expression. A to C, Immunofluorescent staining for MMP2 (A,B) and MCP1 (C,D) (scale bars, 200  $\mu\text{m}$  (upper) and 50  $\mu\text{m}$  (lower)). The data are presented as the mean  $\pm$  SD. \* $P < 0.05$ .





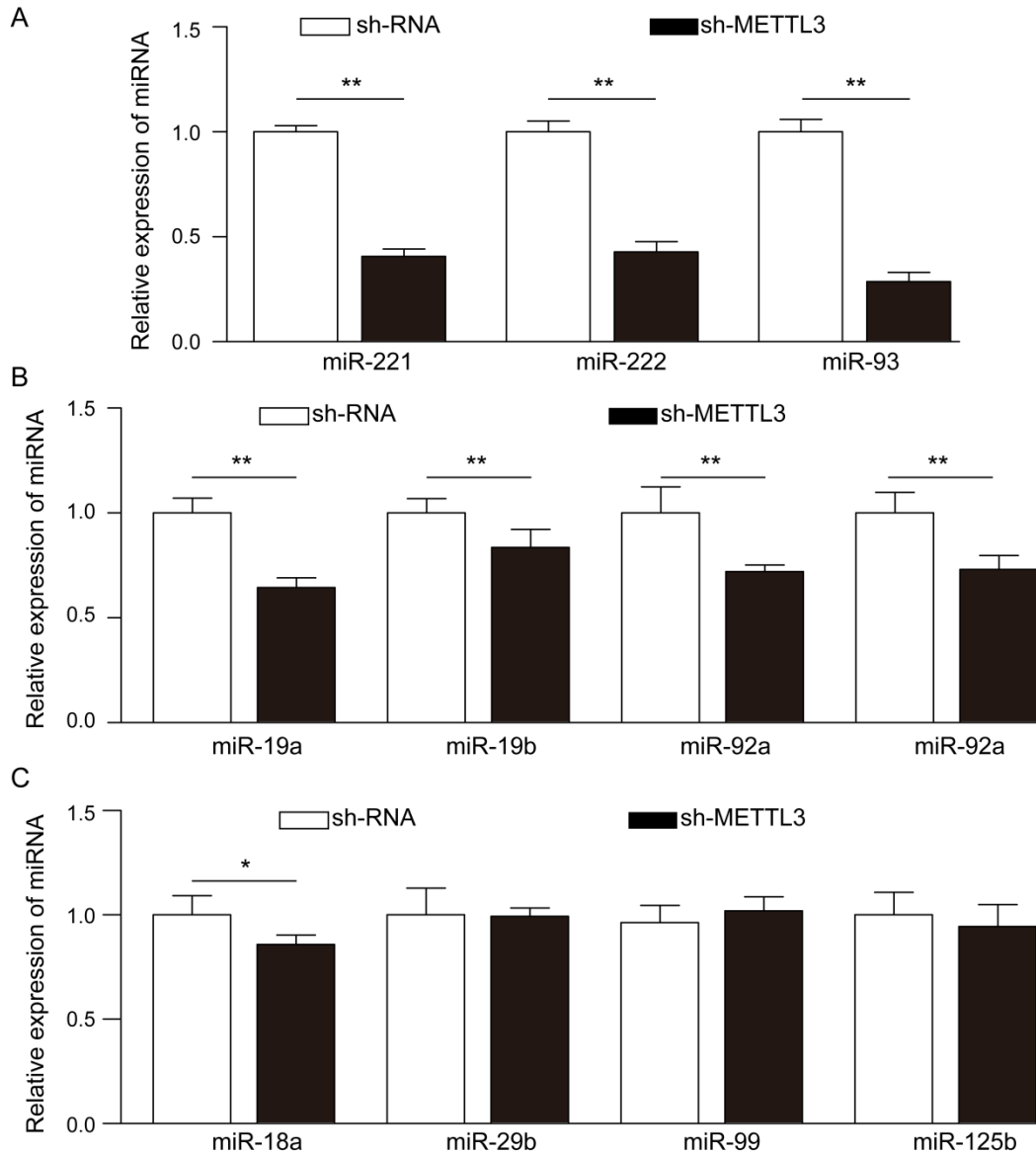
Online Figure 7

**Online Figure 7.** METTL3 overexpression exacerbates  $\text{CaCl}_2$ -induced AAA formation. C57BL/6J mice were injected with AAV-GFP or AAV-METTL3, and their infrarenal aortas were then treated with  $\text{CaCl}_2$  for 15 min. After three weeks, the mice were sacrificed and used for analysis. A, Representative images showing C57BL/6J mouse infrarenal aortas treated with  $\text{CaCl}_2$ . B, Maximal diameters of infrarenal aortas from  $\text{CaCl}_2$ -induced C57BL/6J mice. C, Representative western blots and statistical analysis of aortic METTL3 in  $\text{CaCl}_2$ -treated C57BL/6J mice. D and E, Representative elastin staining and statistical analysis of elastin degradation scores in the two groups of mice. F and G, Representative immunohistochemical staining (F) and statistical analysis (G) of MAC2 in aortas from the two groups of mice (n=4 per group). G and H, mRNA levels of aortic METTL3, MCP-1, MMP-2 and P21 in  $\text{CaCl}_2$ -treated C57BL/6J mice. The data are presented as the mean  $\pm$  SD. \* $P < 0.05$ , \*\* $P < 0.01$ .



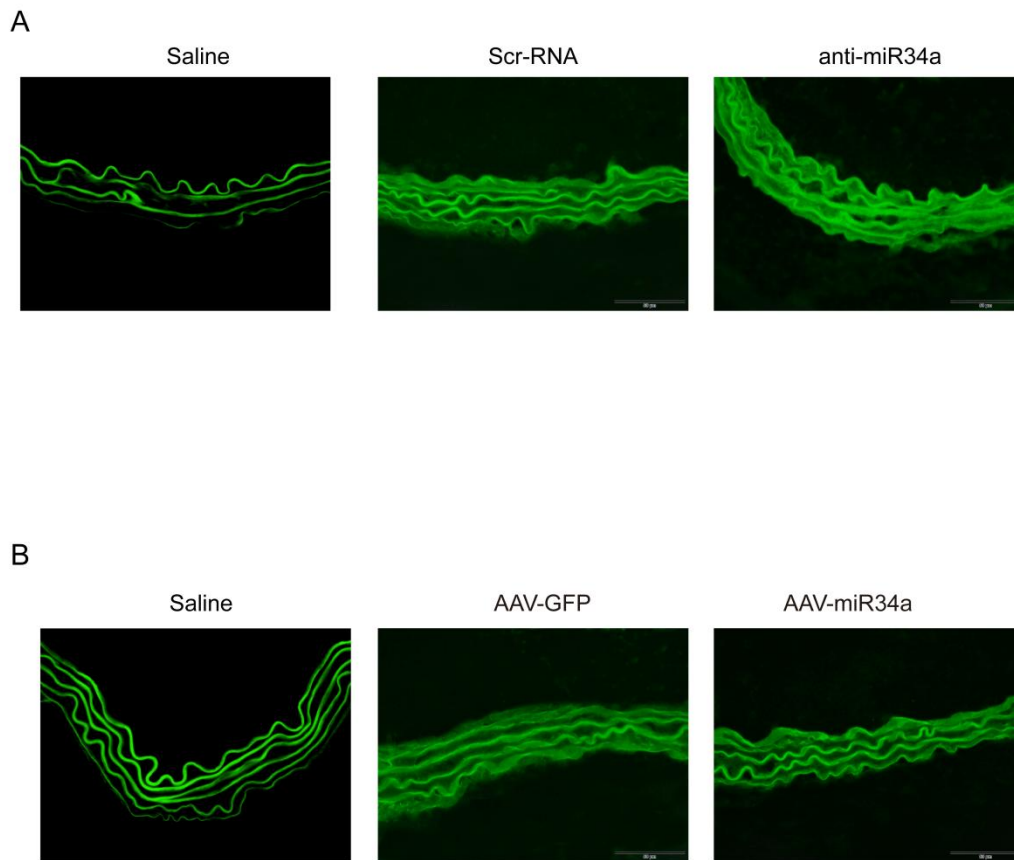
Online Figure 8

**Online Figure 8.** METTL3 overexpression increases vascular MMP2 and MCP1 expression. A to D, Immunofluorescent staining for MMP2 (A,B) and MCP1 (C,D) (scale bars, 200 µm (upper) and 50 µm (lower)). The data are presented as the mean ± SD. \* $P < 0.05$ , \*\* $P < 0.01$ .



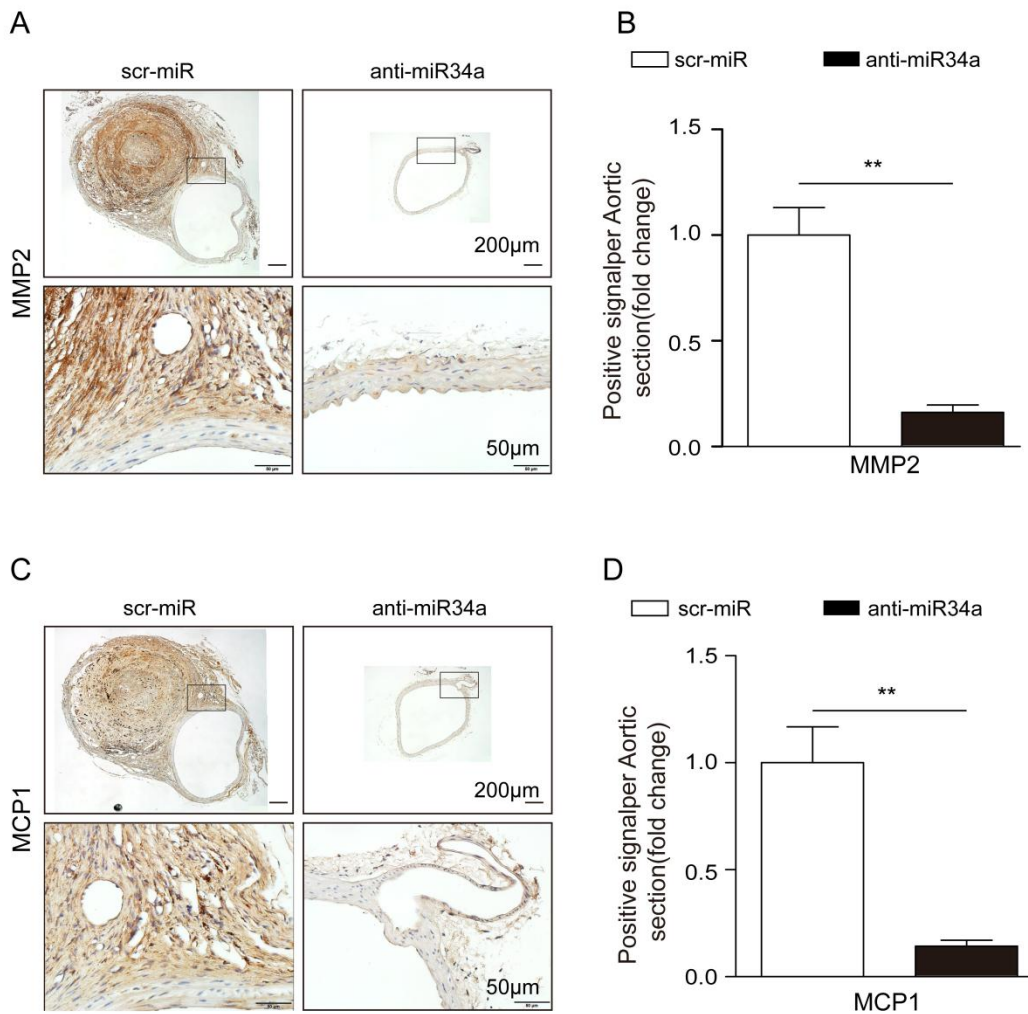
Online Figure 9

**Online Figure 9.** Expression of miRNAs in METTL3-depleted SMCs. A, miR-34a, miR-221, miR-222 and miR-93 were quantified by qRT-PCR upon METTL3 depletion in SMCs. B, miR-19a, miR-19b, miR-92a and miR-20a were quantified by qRT-PCR upon METTL3 depletion in SMCs. C, miR-18a, miR-29b, miR-99 and miR-125b were quantified by qRT-PCR upon METTL3 depletion in SMCs. Abbreviations: siRNA, small interfering fragment control; sh-METTL3, METTL3 knockdown. The data are presented as the mean  $\pm$  SD. \* $P < 0.05$ , \*\* $P < 0.01$ .



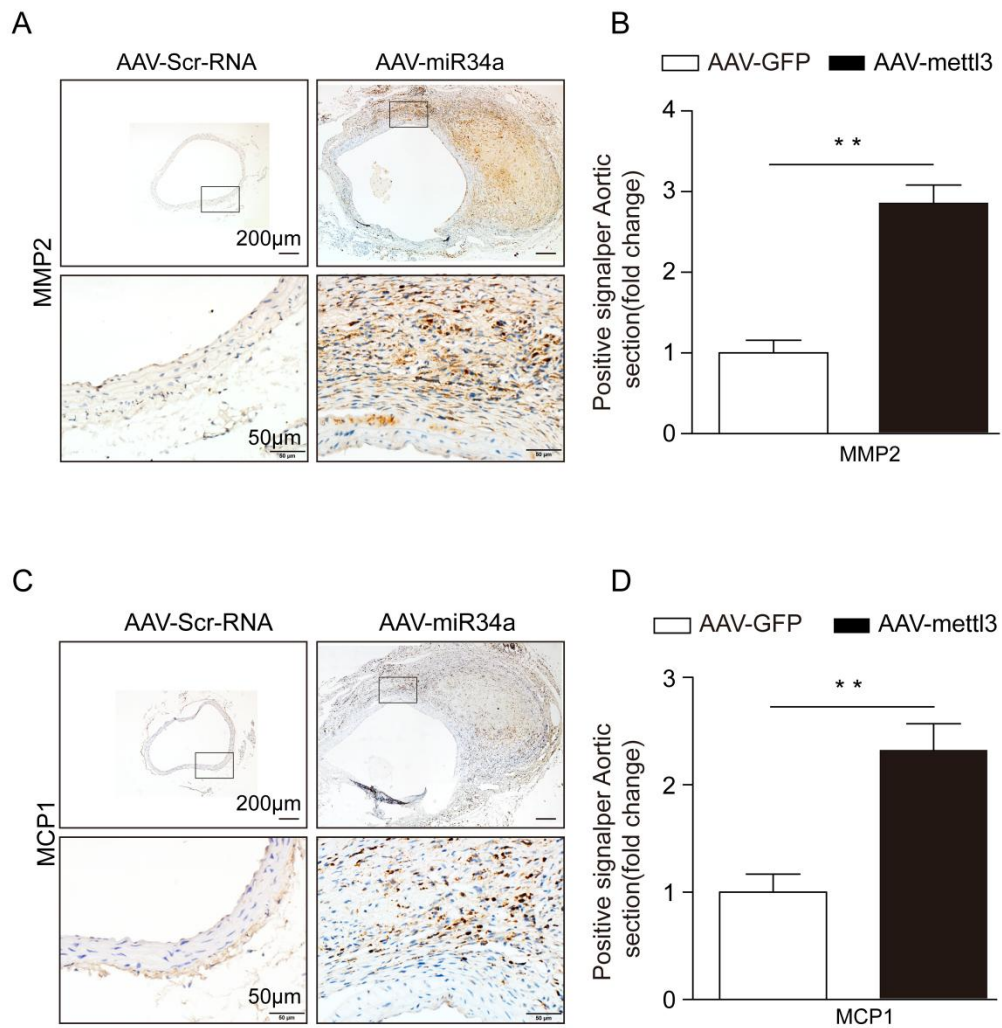
Online Figure 10

**Online Figure 10.** Confirmation of the transfection efficiency of anti-miR34a, AAV-miR34a, and control sequences in the supracrenal aortas of mice. A and B, Representative immunofluorescent staining of virus-borne green fluorescent protein (GFP) in the aortas of mice from the different virus-mediated groups and the saline group (scale bar, 50  $\mu$ m).



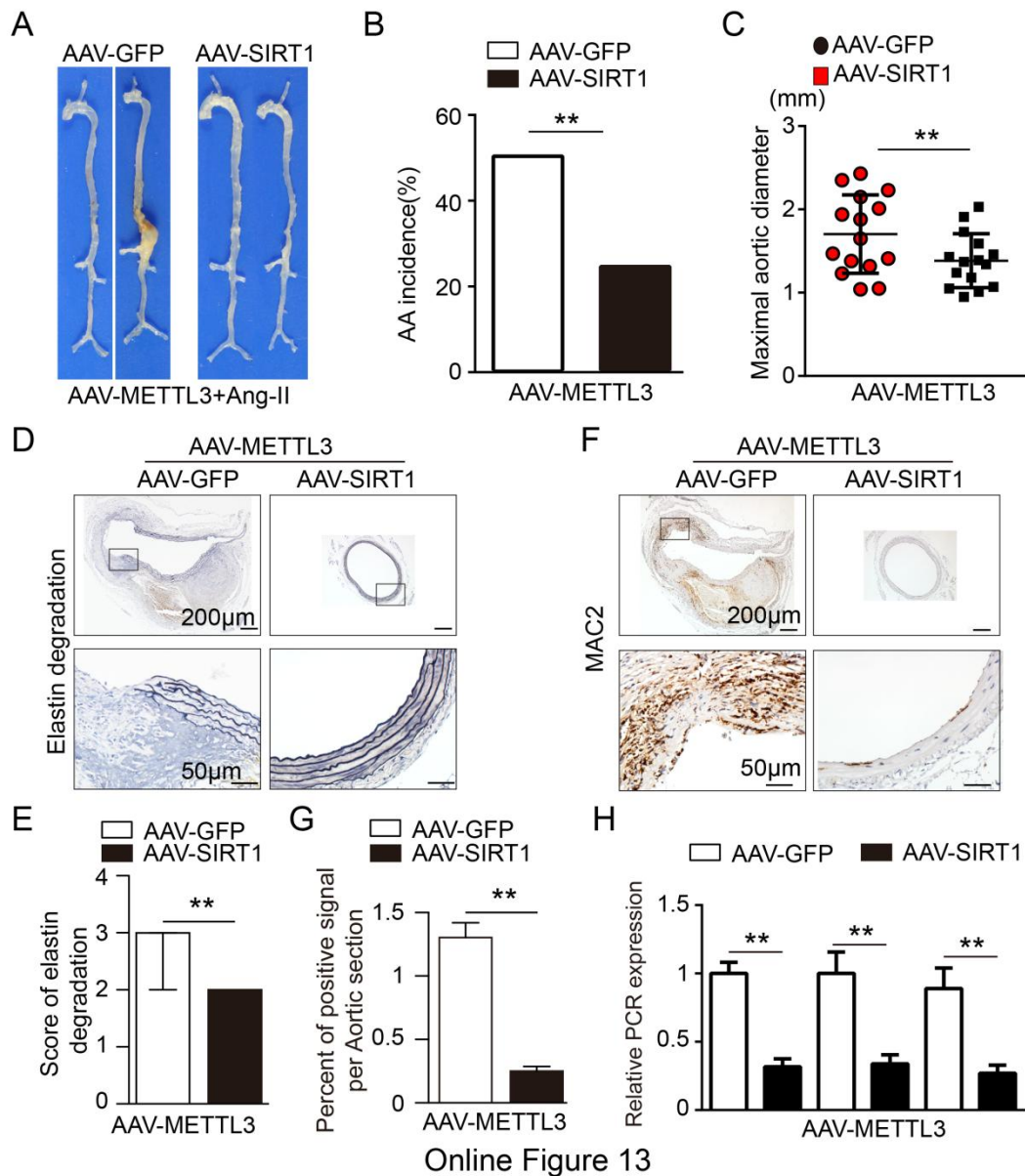
Online Figure 11

**Online Figure 11.** miR34a knockdown inhibits vascular MMP2 and MCP1 expression. A to C, Immunofluorescent staining for MMP2 (A,B) and MCP1 (C,D) (scale bars, 200  $\mu\text{m}$  (upper) and 50  $\mu\text{m}$  (lower)). The data are presented as the mean  $\pm$  SD. \* $P < 0.05$ , \*\* $P < 0.01$ .

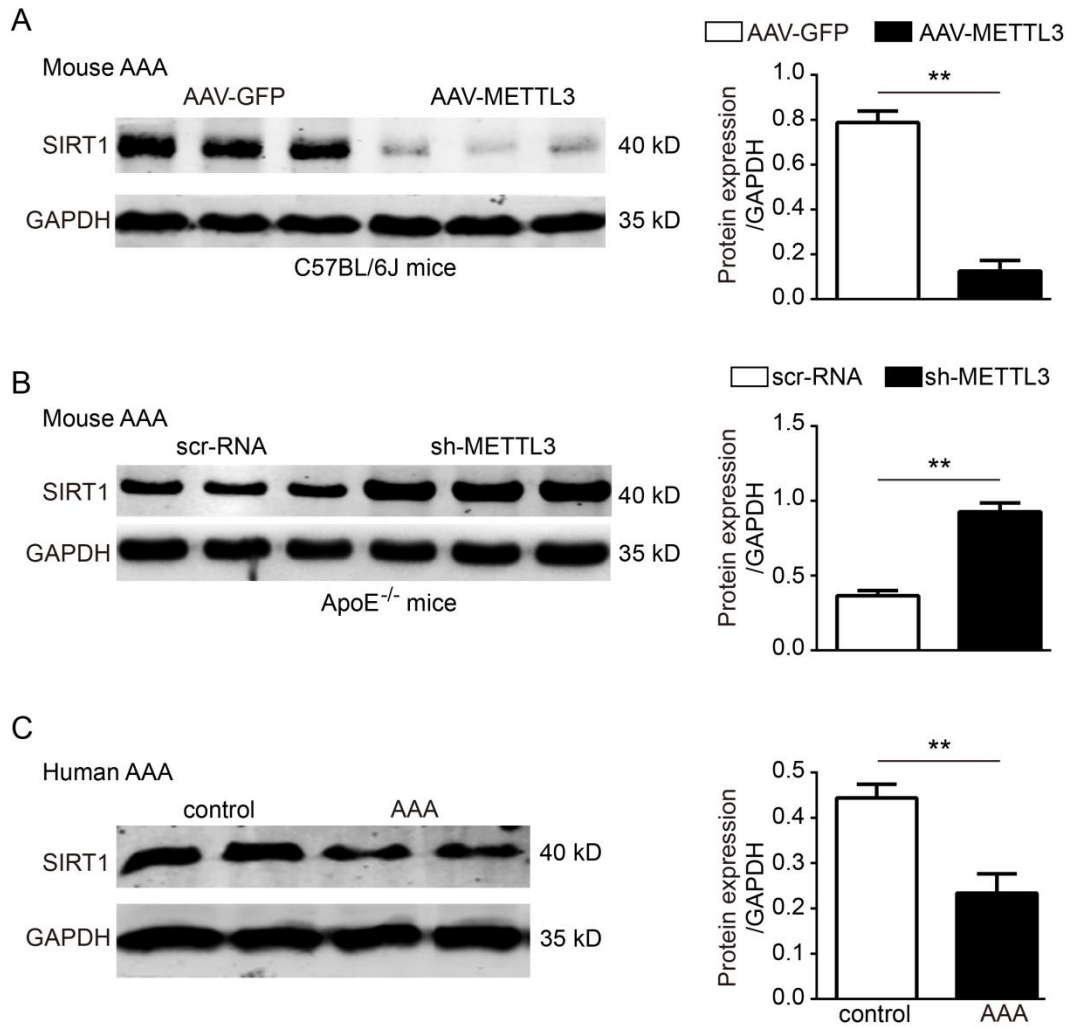


Online Figure 12

**Online Figure 12.** miR34a overexpression increases vascular MMP2 and MCP1 expression. A to D, Immunofluorescent staining for MMP2 (A,B) and MCP1 (C,D) (scale bars, 200 µm (upper) and 50 µm (lower)). The data are presented as the mean ± SD. \* $P < 0.05$ , \*\* $P < 0.01$ .

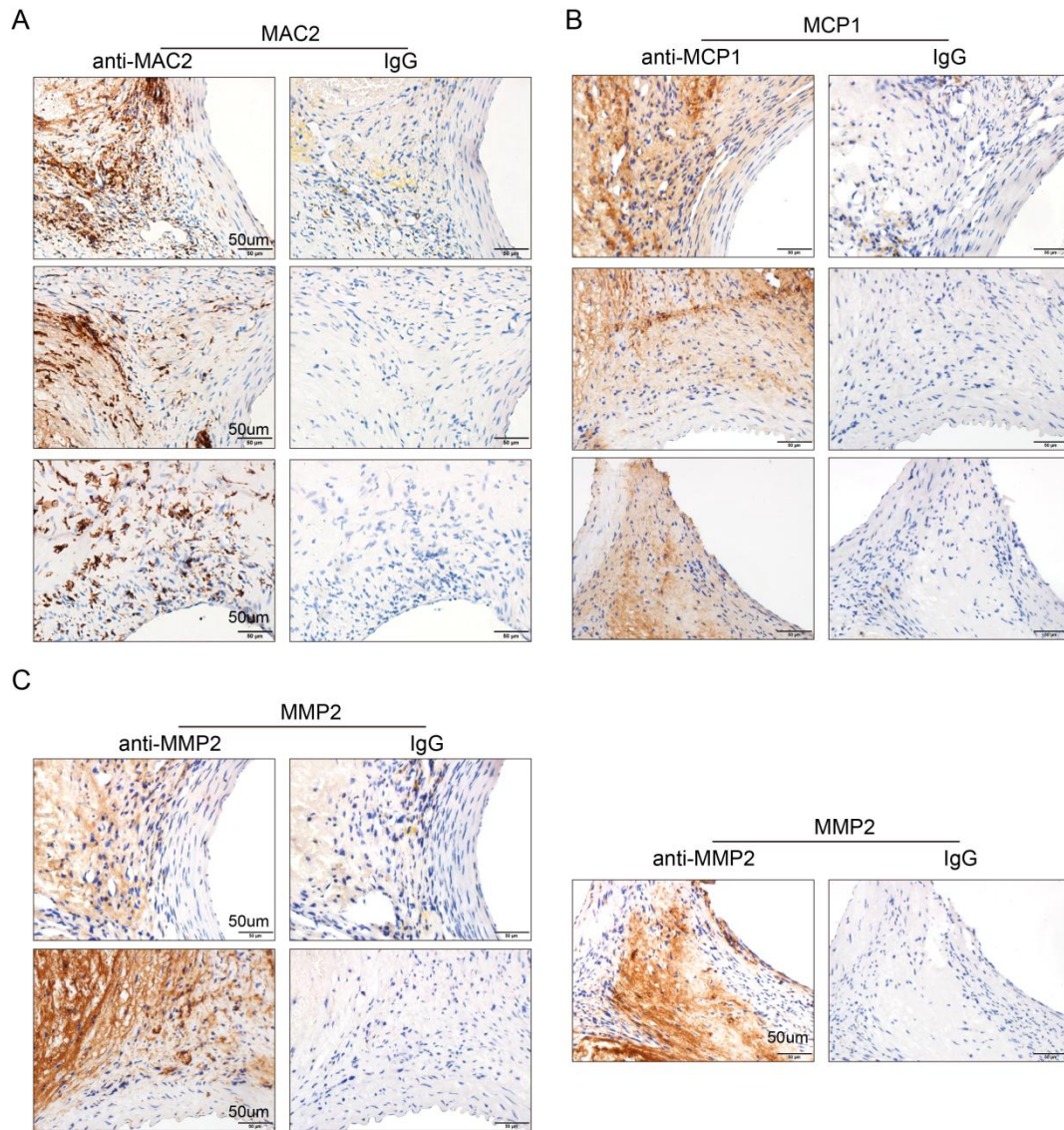


**Online Figure 13.** METTL3 overexpression promotes AAA via miR34a/SIRT1. **A**, Representative photographs of the macroscopic features of AAAs in AAV-METTL3 transfected Ang II-infused C57BL/6J mice in the AAV-GFP group or the AAV-SIRT1 group. **B**, Statistical analysis of AAA incidence in AAV-METTL3 transfected Ang II-infused C57BL/6J mice. **C**, Maximal aortic diameters in the AAV-METTL3-transfected Ang II-infused C57BL/6J mice in the two groups. **D** and **E**, Representative elastin staining and elastin degradation scores in suprarenal aortas from AAV-METTL3 transfected Ang II-infused C57BL/6J mice. The data are presented as the medians and quartiles. **\*\*** $P < 0.01$ . **F** and **G**, Representative immunostaining for MAC2 (scale bars, 200 and 50  $\mu$ m) and the corresponding densitometric analysis ( $n=3$ ). **H**, Relative mRNA expression of MMP2, MCP1 and P21 in AAV-METTL3 transfected Ang II-infused C57BL/6J mouse aortas ( $n=4$ ). The data are presented as the mean  $\pm$  SD. **\*** $P < 0.05$ , **\*\*** $P < 0.01$ .



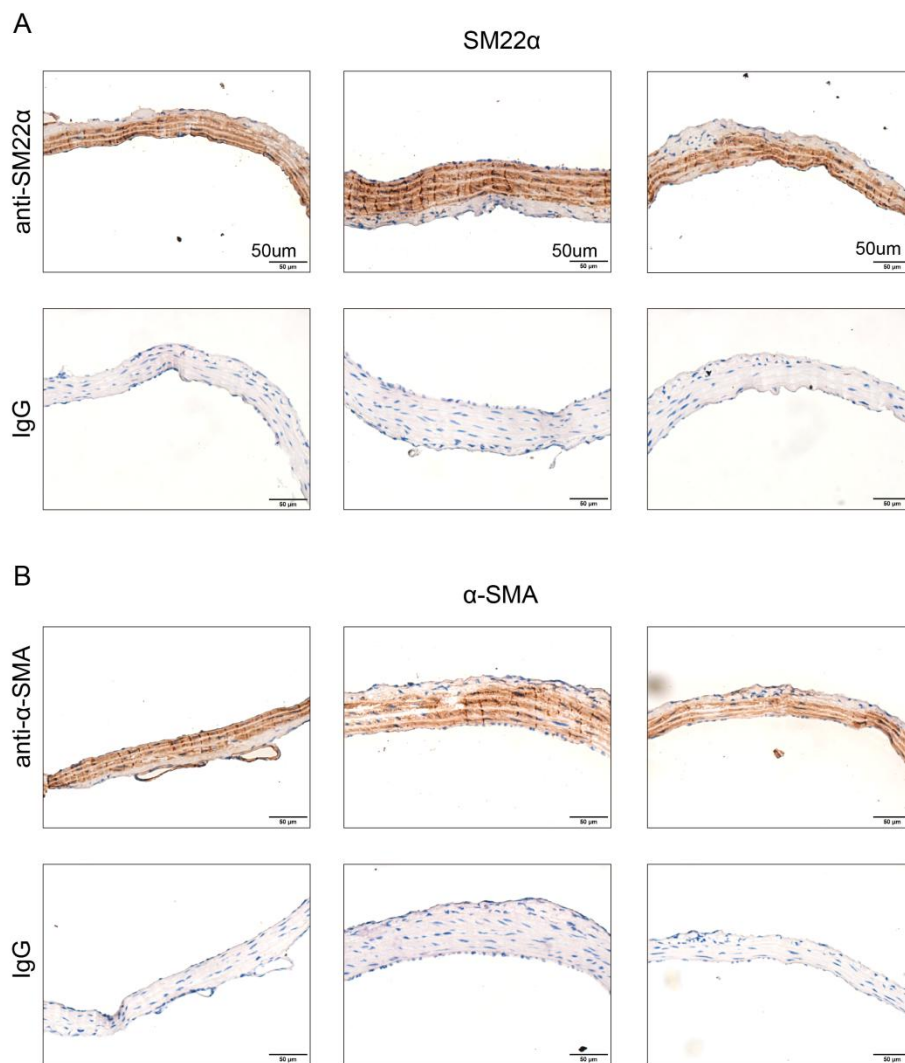
**Online Figure 14.** SIRT1 was substantially upregulated in METTL3 knockdown samples but downregulated in METTL3 overexpression samples. A, Western blot analysis of SIRT1 in Ang II-infused C57BL/6J mice in the AAV-GFP group or the AAV-METTL3 group (n=4). B, Western blot analysis of SIRT1 in Ang II-infused male ApoE<sup>-/-</sup> mice in the scr-RNA group or the sh-METTL3 group (n=4). C, Western blot analysis of SIRT1 in human AAA and adjacent nonaneurysmal aortic samples. \*\*P<0.01.





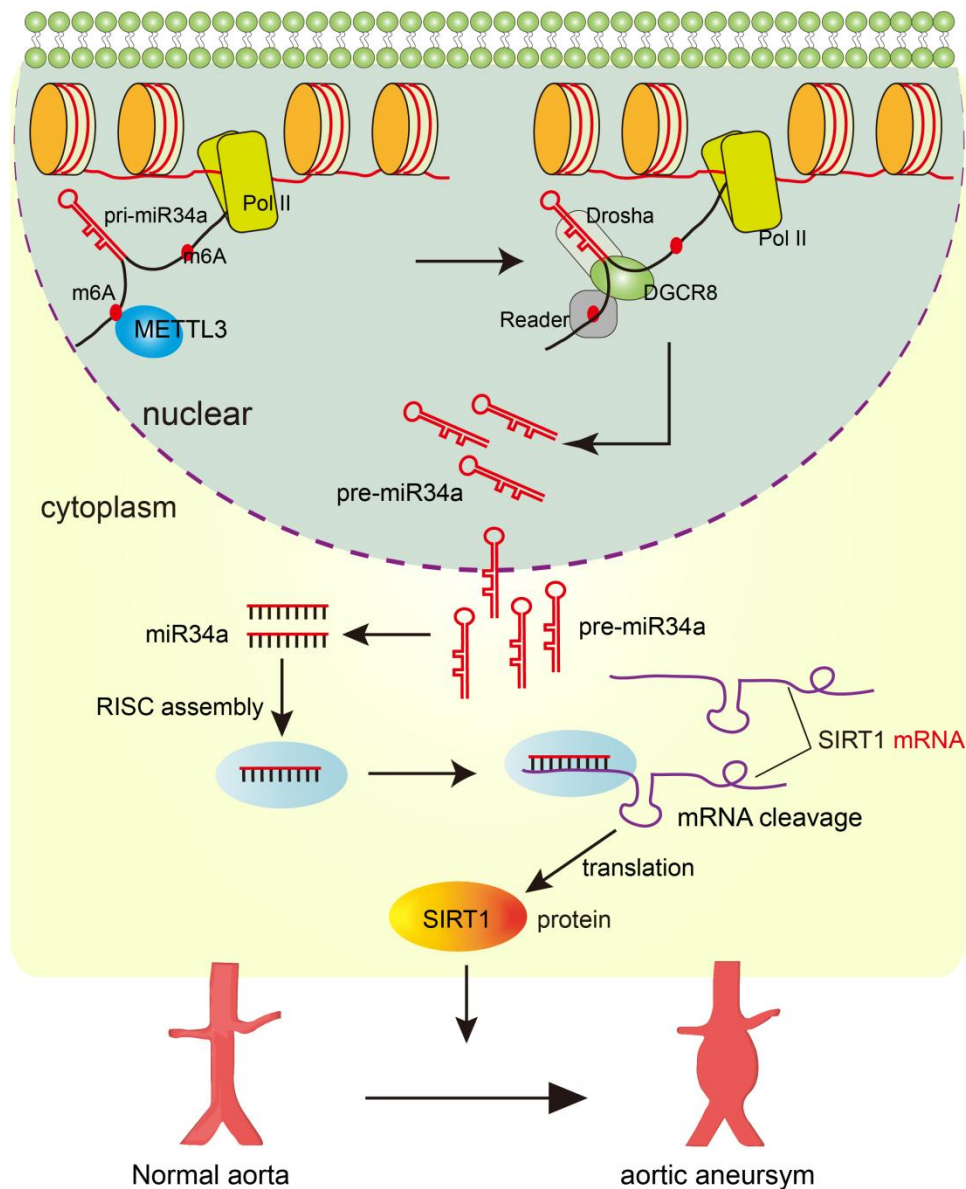
Online Figure 15

**Online Figure 15.** Negative control experiments confirming the specificity of antibody binding in the immunohistochemistry results. A to C, Representative images of immunohistochemical staining for MAC2 (A), MCP1 (B), and MMP2 (C) (scale bars, 50 μm).



Online Figure 16

**Online Figure 16.** Negative control experiments confirming the specificity of antibody binding in the immunohistochemistry results. A and B, Representative images of immunohistochemical staining for SM22 $\alpha$  (A) and  $\alpha$ -SMA (B) (scale bars, 50  $\mu$ m).



Online Figure 17

**Online Figure 17.** Working model of the role of METTL3 in AAA formation. METTL3 increases m<sup>6</sup>A modification of pri-miR34a, which favors the binding of pri-miR34a to DGCR8. METTL3 promotes mature miR34a expression in a DGCR8-dependent manner, which leads to AAA formation through inhibition of SIRT1 expression.

## Major Resources Tables

**Table 1**

**Patient clinical information ( n=5 )**

characteristics	AAA
Ever-smoker	100%
hypertensive	100%
hyperlipidemia	100%
coronary artery disease	80%
gender	male
average age	64.2±4.44 years

**Table 2**

**Specific siRNAs against METTL3, miR34a and their nonspecific controls (NCs)**

siMETTL3,sense:GCAUUGGUGCUGUGUUA AATTUUUAACACAGCACCAAU GCTT
siNC,sense: UUCUCCGAACGUGUCACGUTTACGUGACACGU UCGGAGAATT
Anti-miR-34a,sense: ACAACCAGCTAAGACACTGCCA
Scr-miR, sense: TTCTCCGAACGTGTCACGT

**Table 3**

Antibodies for immunohistochemistry analysis

name	Vendor or Source	Catalog #
anti-SM22 $\alpha$	Abcam	Ab170902
anti- $\alpha$ SMA	Abcam	ab32575
anti-MMP2	Abcam	ab37150
anti-MCP1	Thermo Fisher	PA5-34505
anti-MAC2	Abcam	ab76245
anti-IgG	Abcam	ab172730

**Table 4**

Antibodies for immunofluorescent analysis

name	Vendor or Source	Catalog #
anti- METTL3	Abcam	ab195352
anti- SM22 $\alpha$	Abcam	ab10135
Alexa Fluor 488	Abcam	ab150129
Alexa Fluor 594	Abcam	ab150088

**Table 5**

Antibodies for western blots

name	Vendor or Source	Catalog #
anti- METTL3	Abcam	ab195352
anti- MCP1	Thermo Fisher	PA5-34505
anti- MMP2	Abcam	ab37150
anti- P21	Abcam	Ab109119
anti- SM22 $\alpha$	Abcam	ab155272
anti- SIRT1	Abcam	ab110304
anti- $\beta$ -actin	Abcam	ab5694
anti-GAPDH	Abcam	Ab9485

**Table 6**

Quantitative real-time PCR

Primer	Sequence (5'-3')
miR34AHG _forward	TGGCAGTGTCTTAGCTGGTTGT
miR34AHG _reverse	TGGCGTCTCCCCTGGTCT
miR34a _forward	TGGCAGTGTCTTAGCTGGTTGT
miR34a _reverse	AGTGCAGGGTCCGAGGTATT
U6 _forward	CTCGCTTCGGCAGCACA
U6 _reverse	AACGCTTCACGAATTTGCGT
METTL3 _forward	TTCATCTTGGCTCTATCCGGC
METTL3 _reverse	GCACGGGACTATCACTACGG
METTL14 _forward	CCATAATGATTACTGCCAAC
METTL14 _reverse	GTCAAAGGCTTCTATGTCTG
WTAP _forward	GCAACCAAAGAGCAGGAGAT
WTAP _reverse	CTTCCAGGCACTCAGTTCAT
YTHDF2 _forward	TAGCCAGCTACAAGCACACC
YTHDF2 _reverse	TTTCCCACGACCTTGACGTT
FTO _forward	GAGCAGCCTACAACGTGACT
FTO _reverse	GAAGCTGGACTCGTCCTCAC
KIAA1429 _forward	GCTGATGACTGCAATCTGCG
KIAA1429 _reverse	CTCCACAACAGCCCATAGCA
METTL4 _forward	TTCGAAGTTAATCCAAGAAGG T
METTL4 _reverse	CGTTTGAAGCTCCATTCAT
ALKBH5 _forward	TGTGCTCAGTGGGTATGCTG
ALKBH5 _reverse	CTGACAGGCGATCTGAAGCA
MMP2 _forward	ACCAACACTGGGACCTGTAC
MMP2 _reverse	CGAAGAACACAGCCTTCTCCT
MMP9 _forward	GCGTGTCTGGAGATTCGACTTG
MMP9 _reverse	ACTGCAGGAGGTCTAGGTCAC
MCP1 _forward	ACCTGCTGCTACTCATTAC
MCP1 _reverse	CATTCAAAGGTGCTGAAGAC
$\beta$ -actin _forward	GGCTGTATTCCTCCATCG
$\beta$ -actin _reverse	CCAGTTGGTAACAATGCCATGT

**Table 7**

**(DGCR8) RIP-specific primer pairs for the miR34AHG gene**

5'- TGGCAGTGTCTTAGCTGGTTGT -3' (forward)
5'- TGGCGTCTCCCACTGGTCT -3' (reverse)

**Table 8**

**(m<sup>6</sup>A) RIP-specific primer pairs for the miR34AHG gene**

Site 1: 5'- ATGCCAACTTTGAGGCCA-3' (forward)
Site 1: 5'- CTCTCCATCCTCCGGTGA-3' (reverse)
Site 2 : 5'- ATGCCAACTTTGAGGCCA-3' (forward)
Site 2 : 5'- AAGACCTGGGGAAGCCAC-3' (reverse)
Site 3 : 5'-AAGAGGTGACGCCAAACG-3' (forward)
Site 3 : 5'-CCTGGCCTGTGTGAAAGG-3' (reverse)