

Table S1. Comparison of the extent of RNA editing of chloroplast sites between the wild type and *yl* mutant by RNA-seq.

Gene	Position ^a	Wild type		<i>yl</i>	
		Coverage [reads]	Editing (%)	Coverage [reads]	Editing (%)
<i>accD</i>	617	696	91.24	258	84.50
<i>atpF</i>	92	3531	97.73	2260	96.59
<i>clpP</i>	559	2338	94.65	1990	55.73
<i>ndhA</i>	341	689	91.29	450	94.00
<i>ndhB</i>	1073	2277	90.65	1553	90.79
	9	28	100.00	20	100.00
	149	50	96.00	28	96.43
	542	290	100.00	192	100.00
	586	95	91.58	74	20.27
	737	126	98.41	147	6.12
	746	175	99.43	153	43.79
	830	266	99.62	134	46.27
	836	267	100.00	78	100.00
	1112	189	96.83	71	100.00
	1255	727	99.72	209	98.56
	1481	138	100.00	50	100.00
	<i>ndhC</i>	323	1231	95.69	636
<i>ndhD</i>	2	1686	55.28	1488	55.85
	383	1896	91.59	1532	92.95
	674	2597	89.37	1836	4.52
	878	3442	84.02	2354	31.61
	1298	2026	87.76	1376	94.40
<i>ndhE</i>	233	2342	89.75	1136	84.86
<i>ndhF</i>	290	645	58.29	192	73.44
<i>petB</i>	611	1260	99.05	3557	18.11
<i>psaI</i>	79	2311	94.50	884	87.78
<i>psbE</i>	214	1658	99.70	1132	99.65
<i>psbF</i>	6	1654	85.31	1289	92.47
	77	2694	94.51	1523	93.11
<i>psbL</i>	2	1239	89.99	700	46.14
<i>rpoA</i>	200	1430	73.01	1091	72.23
<i>rpoB</i>	338	132	68.18	187	66.84
	551	173	39.88	120	0.00
	566	160	37.50	121	57.02
	2000	145	86.90	161	37.27
	41	556	96.22	497	97.99
<i>rpoC1</i>	488	1054	47.25	1152	51.48
	134	1566	96.87	817	35.50
<i>rps2</i>	248	1875	98.56	967	57.81
<i>rps12</i>	554 ^b	139	94.96	48	81.25
<i>rps14</i>	80	1159	86.11	2641	89.10
<i>rps16</i>	499 ^b	350	73.14	260	11.92
	212	2662	95.79	643	91.91
<i>rps18</i>	221	1349	84.36	516	73.84

The coverage of each editing site is presented in reads. Editing (%) = $U/(C + U) \times 100$.

^a Position is given with respect to the initiation codon of each chloroplast transcript.

^b The RNA editing site is in the intron of the plastid gene. The position here is given with respect to the initiation codon of the DNA sequence of each gene.

Table S2. Verification of the extent of chloroplast RNA editing by Sanger sequencing.

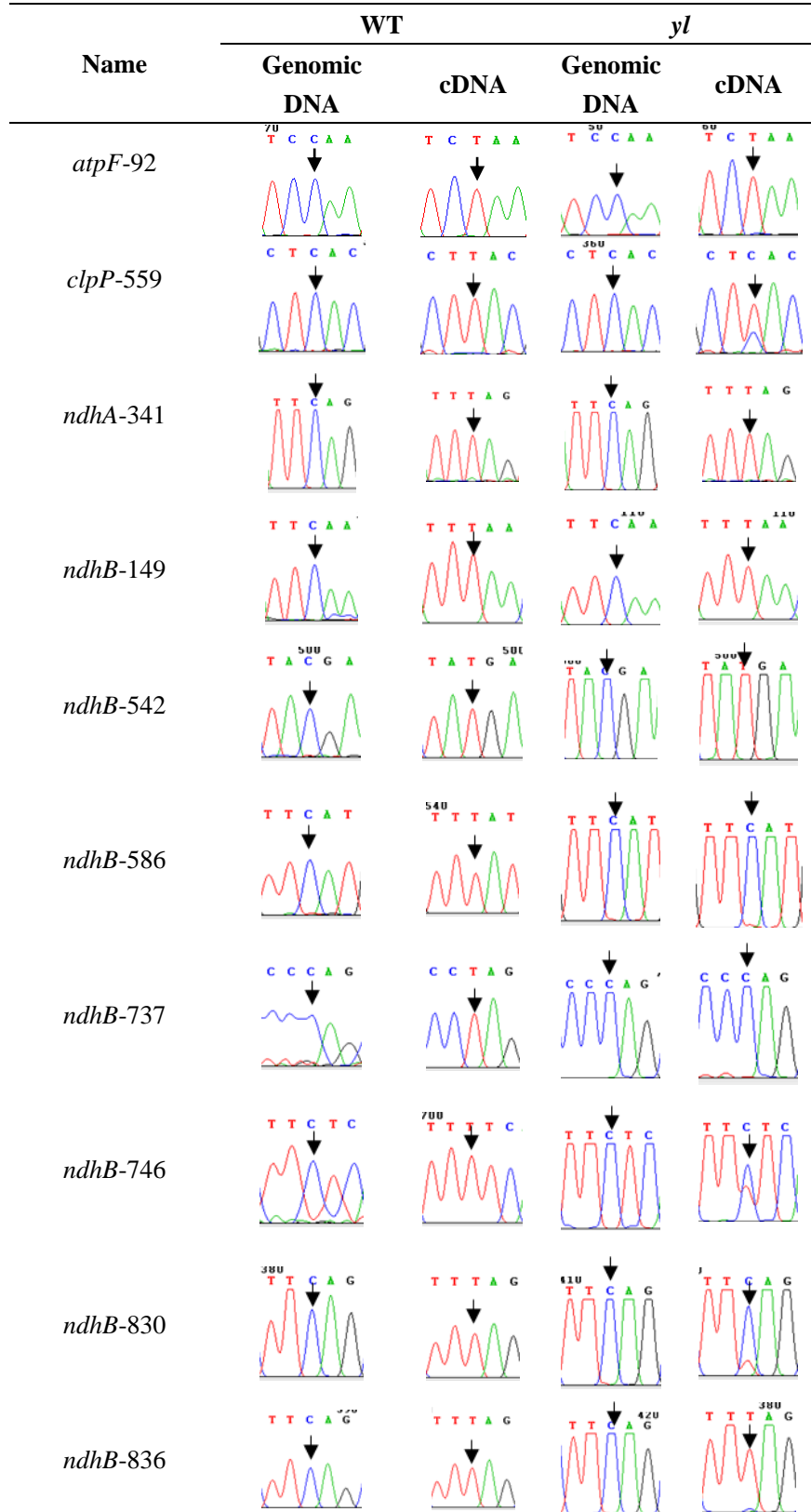


Table S2. (continued).

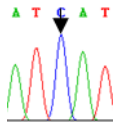
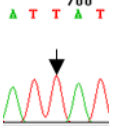
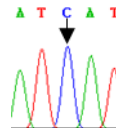
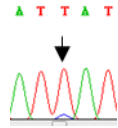
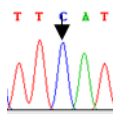
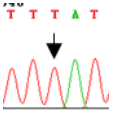
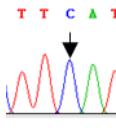
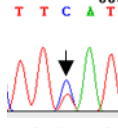
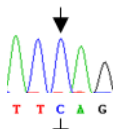
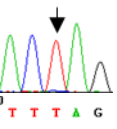
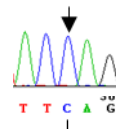
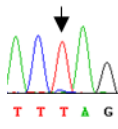
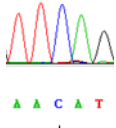
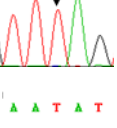
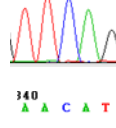
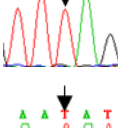
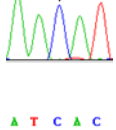
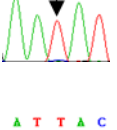
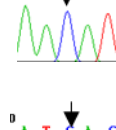
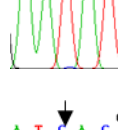
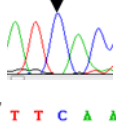
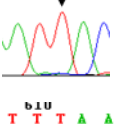
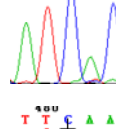
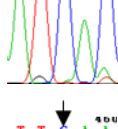
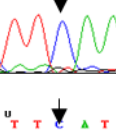
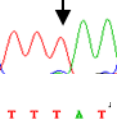
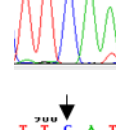
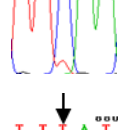
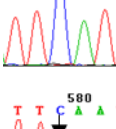
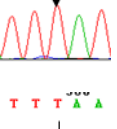
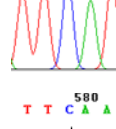
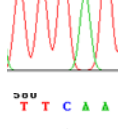
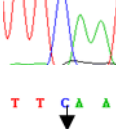
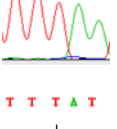
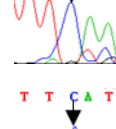
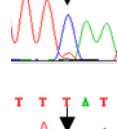
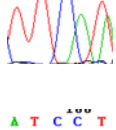
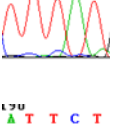
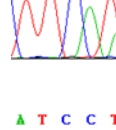
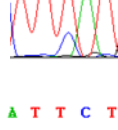
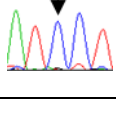
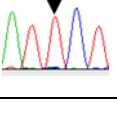
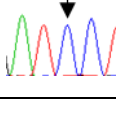
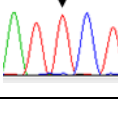
Name	WT		<i>yl</i>	
	Genomic DNA	cDNA	Genomic DNA	cDNA
<i>ndhB</i> -1112				
<i>ndhB</i> -1255				
<i>ndhB</i> -1481				
<i>ndhC</i> -323				
<i>ndhD</i> -383				
<i>ndhD</i> -674				
<i>ndhD</i> -878				
<i>ndhD</i> -1298				
<i>petB</i> -611				
<i>psaI</i> -79				
<i>psbE</i> -214				

Table S2. (continued).

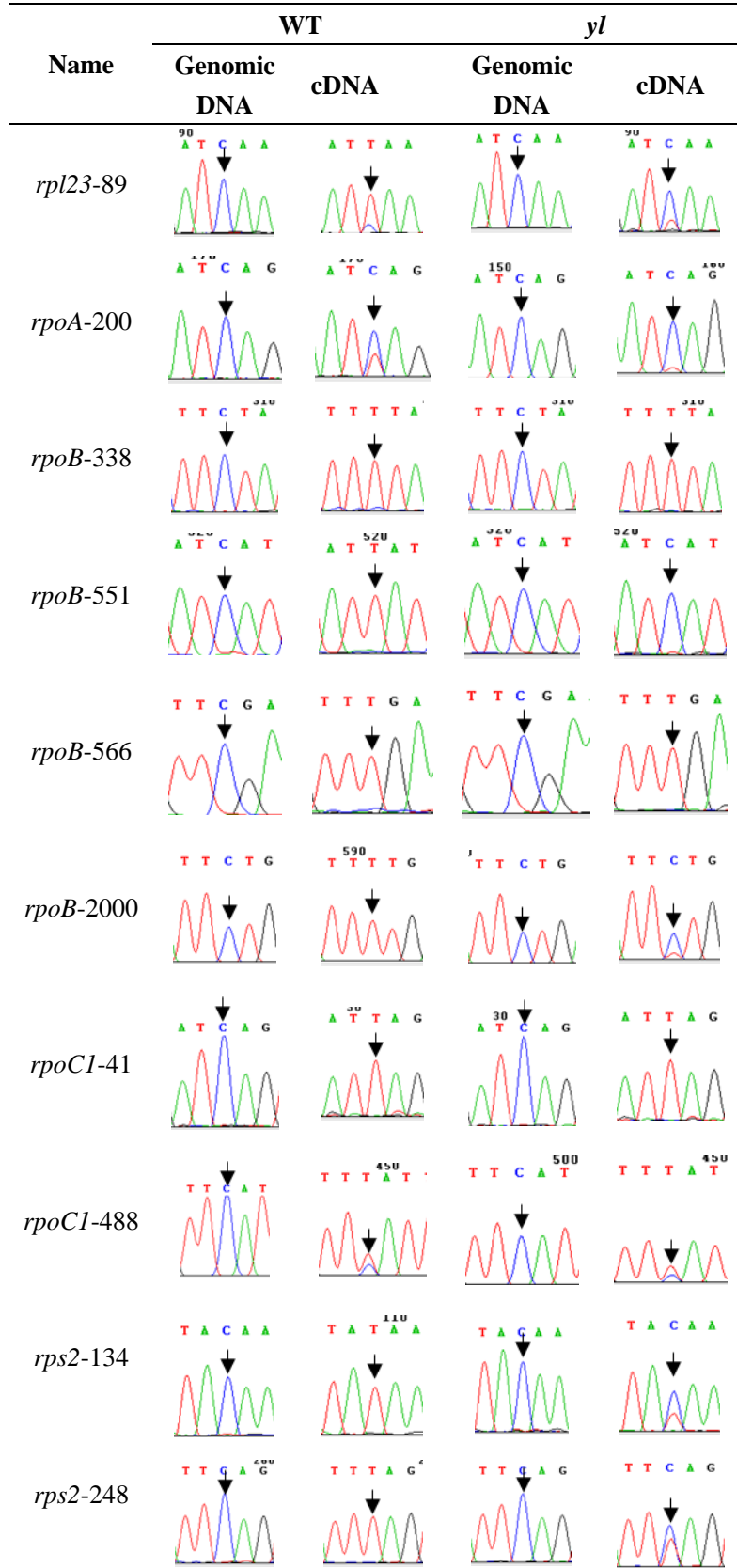


Table S2. (continued).

Name	WT		<i>yl</i>	
	Genomic DNA	cDNA	Genomic DNA	cDNA
<i>rps14-80</i>				
<i>rps16-212</i>				
<i>rps18-221</i>				

The sequence results were derived from PCR or RT-PCR products amplified from DNA or cDNA, separately. The editing sites are indicated by arrows.

Table S3. Primers used in this study.

Primers used for fine mapping of <i>YL</i>		
Name	Forward primers (5'-3')	Reverse primers (5'-3')
at009	AATCCATTCACTCTCCCAA	TCATCGCCAACATTTCACTA
att002	TTGCCCAAGGACTTCCACAT	GAACAAGCCCTTCGGAGATT
at015	AATAATGCACACGACTTAGG GT	TAATCAGCATCAGCACATACAA
at022	CGGAGCAAGTTGGTTTTGAC TG	ATTATTTGGGGAATTGGGGTGT
S3	GTGATGCTGTTTCTTGCAG TT	ATGAGTTATCCTGCTGCCTTGA
S7-3	AATACGATGGCGATAAGAA C	ACAGGAACAGGAGGAAGTGA
Primers used for determining the <i>YL</i> gene		
Name	Forward primers (5'-3')	Reverse primers (5'-3')
Seq28k-1	CGCAACTTGTTTCATCACTG TA	TGAAGGGAATAGGGAGAATCT C
Seq28k-2	GAGATTCTCCCTATTCCCTT	TACTCACATTTGCACCTTCG
Seq28k-3	AATACGATGGCGATAAGAA C	ACAGGAACAGGAGGAAGTGA
Seq28k-4	AAATACGTAATCGCATTTCGA GT	GCAAAGCAGGAAATCAGATAC A
Seq28k-5	CATGGCTACTTTCACCTTTA	TTCACCTTCTCTCCTTCTA
Seq28k-6	ACTTTGTAGTTCTGTGCCTCT TG	AACATGCTGAGCTGGAATCTTA A
Seq28k-7	ATGACTTTAAGATTCCAGCT CAG	GGACATTTCCCTTCACTACCAC C
Seq28k-8	CTCATAGATAAACTGGATA GGGAAG	TTTTAGGCACAAGGGCAAATAG AAC
Seq28k-9	CATGGCTACTTTCACCTTTA	ATTAAGATGCCATGCTGACA
Seq28k-10	CTTATCAAGTCACATGGACA TTGTG	CTACAAGGCTAACCAGCAAGCT CTT
Seq28k-11	AGGTCATGGGAAAGTGAGG ACAGTG	TGATTCCTGTGGTATCTTGGTG GGT
Seq28k-12	TGTTGAATACACCACAGAGG AGGCT	CAATAGGAACAACCTCGAAGGG ACTG
Seq28k-13	AATATTGTAGCCTCTATAGG AATCTGGC	GTCTTTTGTGTTAGATCAACC CAAATA
Seq28k-14	TGAGATGGTCCCTGATGGAA AATAT	TCTGAAAGCTGACCCAGTAGTG GAA
Seq28k-15	ATTACTGGTCCTTTCTGCTTC AC	TAATAGCGTAAGTCTGCACAAC AAG
Seq28k-16	GCCTCTTTTGTATCATCTAT GTCCTTG	TAGTTCAGACCCTACTTACTTT ACTCTG

Seq28k-17	TCCCACTATTTAATGTGATA TTTGC	AAACAACAAGACAAGAAGACT ACCC
Seq28k-18	TGCCTACTCGTGGAAATGTTG	TGAAGGTTTCGGGTTTGT
Seq28k-19	TTTTCTTATAGCGACTTGCC	CATCAGGGACCATCTCATTC
Seq28k-20	ATGTCATACTCCGACTTGTT	CATAGTTTCTGATATTTCCA
Seq28k-21	ACATTTTCGCTCTATTCTGTTC	GACGGTFACTTGACTTCTTTCT
Seq28k-22	TTCATATTTGCGTTAATGAT TTCTC	TCATTTCAAAGTGTTTCATTGGA TAC
Seq28k-23	ACTGCAATGAGTAATGCTGA GCCACC	GTCGTCGTCAAACCAAACCAAA TCCTAT
Seq28k-24	GGCGGCATAGCATCGTCATC AT	TGGCGAAGGAGACTCTAGGAA CT
Seq28k-25	GCTAAATGAGAAGCTAAAC TTACCCTAA	CAAGACAAGAAGACTACCCTA AACTA
Seq28k-26	TCAAGTCTCAAGATACGCTT ATTATCAT	CGTCAAACTTTAATTTTATTCT TTCTG
Seq28k-27	AAAAGTATAGGGAAAGGGT GGGAGA	TTGGCATTGCTAAGTGAGACA GAC
Seq28k-28	TTGAATGCTGTTGTCGTAAT TTTCT	CCATAGGCAAAGCTTATACCGT ATT
Seq28k-29	AACCTCCACGATACTTCCAG AACCT	TCAATCATCCAGAACAGGGGAAT CAG
Seq28k-30	AAATTGGTAGGAAGGTTACT TATGT	CAATGATTATCTTTTCTAGGAT TTG
Seq28k-31	GTGGATCTTGGAGATAGAA ATA	TTTCATTACTIONGTACCGATTAC
Seq28k-32	GGGTCAAGGATGTGACAAC GAT	CTGATGCCACTCAACCATGACA
Seq28k-33	ATTCACCAGGGCTTAACAAA GT	AAAAGAACGAAGGGAGTATCA T
Seq28k-34	CACCGCCTCTAATACTACGC TA	GGCGAAGGAGACTCTAGGAAC T

Primers used for vector construction

Name	Forward primers (5'-3')	Reverse primers (5'-3')
<i>YL-GFP</i>	TGCCATGGCAATGGAAGTTC TCTCTGTGTCTGTCT	GGACTAGTTCAGCATGATTCC TGTGGTATCTT

Primers used for real-time PCR

Name	Forward primers (5'-3')	Reverse primers (5'-3')
<i>Actin</i>	ATCCGGAATCAAACGCACCC T	AGAGCGTCAGCGAAGAAGGAA G
<i>ATP synthase</i>	CGATTTCCGCATCCGCATCG AG	GAGCCCTTCCCGGTGAACTCAT
<i>YL</i>	TACCCCTCCTTCAATCCTTCC AC	TGAGCATCTTTCTCACTGCCAA G

Primers used for verifying RNA editing sites by Sanger sequencing		
Name	Forward primers (5'-3')	Reverse primers (5'-3')
<i>accD</i>	ATGGA AAAATGGTGGTTTAA TTCTAT	CTATGCCTTATCATT TTTTAGTCA AAG
<i>atpF</i>	ATGAAAAATATAACCGATTC TTTCC	CTAATCAGTTAT TTTTCTTTCA TCGTC
<i>clpP</i>	TTATTCAACGGCTACAAGAT CTACA	ATGCCCAT TGGTGTCCAAGAG TAC
<i>ndhA</i>	ATGATAATTTATTTAACAGA GATAC	TTACAGTGAAAACAGTTGAGAC GAA
<i>ndhB</i>	ATGATCTGGCATGTACAGAA TGAAA	CTAAAAAAGGGTATCCTGAGC AATT
<i>ndhC</i>	ATGTTTCTTCTTTACGAATAT GATA	TTAAGACCATTCTAATGCCCCC TTT
<i>ndhD</i>	ACGAATTATTTTCCTTGGTT AACAAACAG	CTATAAAAAATTAGATAGAATA GCTTCA
<i>petB</i>	ATGAGTGTGCGGCTTGTTTT AATGGATC	CTATAAAGGACCAGAAATGCCT TGCTTA
<i>psaI</i>	ATGATAAACTTTCCCTCCAT TT	CTAAAAAATCTTGTTTTTTTGA AC
<i>psbE</i>	CTAAAAAGATCTACTAAATT CATCAAGT	ATGTCTGGAAGCACAGGAGAA CGTT
<i>rpl23</i>	ATGAATGGAATAAAATATG CAGTAT	TTAAGTTTTTTTCTTTCTAAGAG GT
<i>rpoA</i>	TTATGCAAAATGCTTTTCTA TTT	ATGGTTCGAGAGAAAATAAGA GT
<i>rpoB</i>	ATGCTTGGGGATGGAAATG AAG	TTAAGCTTCCTTCCTATGAATC CGG
<i>rpoC1</i>	ATGATTGATCAGTATAAACA TCAACAAC	TTAGATATCATATGAATAGGCC CGG
<i>rps2</i>	ATGACAAAAAGATATTGGA ACATAA	TCAAGAATTTTCGTATATAGCTA GAA
<i>rps14</i>	ATGGCAAGGAAAAGTGTGA TTCAGA	TTACCAACTGGATCTTGTTGCA CCG
<i>rps16</i>	ATGGTAAAACCTTCGTTTGAA ACGAT	TTAATGGAATTTTCGTTTGTTGA TTC
<i>rps18</i>	ATGGA AAAATCTAAGCGAC TCTTTATGA	TTAATTTCTATAT TTTT TTTTC TTAAA