

Supplementary Methods

1.1. RNA extraction and quantitative real-time PCR (qPCR)

Total RNA from cells was isolated using peqGOLD RNAPure reagent (PEQLAB Biotechnologie GmbH, Erlangen, Germany) and transcribed by utilizing Maxima first-strand cDNA synthesis kit (Thermo Scientific, Dreieich, Germany), each according to the manufacturer's protocol. qPCR assays were performed using IQ SYBR® Green Supermix (Bio-Rad Laboratories GmbH, Munich, Germany) on the CFX96 Touch™ Real-Time PCR (Bio-Rad) cycler. Data were analyzed via CFX Manager™ 3.1 software (Bio-Rad) and $\Delta\Delta C_t$ were calculated. 18S rRNA was used as internal control. Primer sequences are listed in Supplementary Tab. 1. Inducible nitric oxide synthase (*iNOS*) mRNA expression (Mm_LOC673161_1_SG) was determined using QuantiTect Primer Assay (QT01547980) from Qiagen (Hilden, Germany).

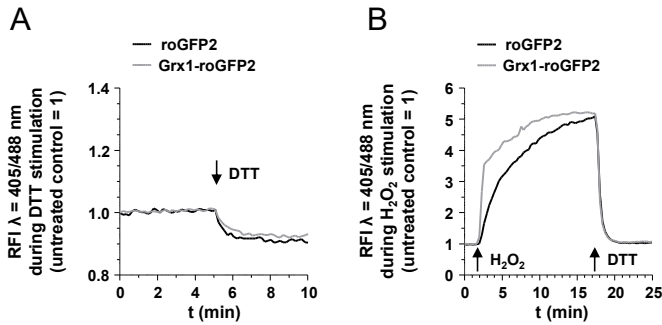
Supplementary Tab. 1 qPCR primers used in this study.

Target	Forward primer (5' - 3')	Reverse Primer (5' - 3')
18S	GTAACCCGTTGAACCCATT	CCATCCAATCGGTAGTAGCG
<i>Arg1</i>	GTGAAGAACCACGGTCTGT	CTGGTTGTCAGGGGAGTGT

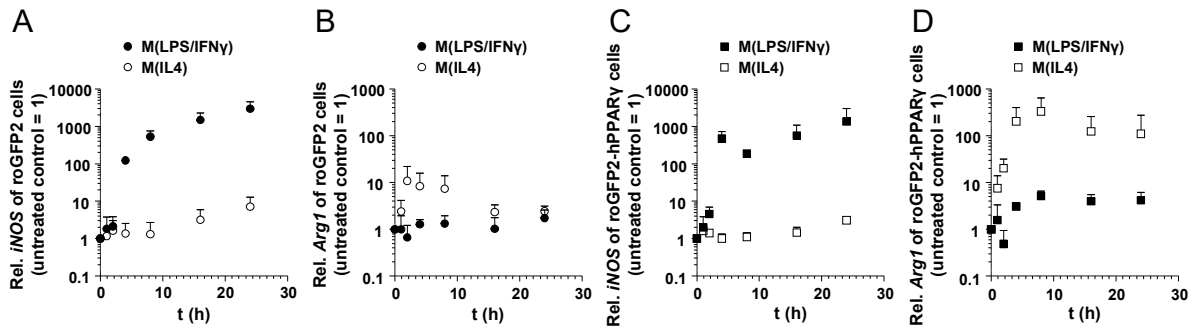
1.2. BIAM switch assay

Cellular available reduced cysteines of total protein content of HEK293T stably expressing HA-tagged hPPAR γ were labeled by NEM (20 mM in DMEM) for 10 min on ice, before cells were washed 3 times with PBS and harvested afterwards. Further sample preparation started with cell lyses, followed by labeling of cellular oxidized cysteines by biotin-iodoacetamide (BIAM) and finally biotin-immunoprecipitation as described elsewhere⁷⁵. Samples were analyzed by Western blot studies, using detection of peroxiredoxin 3 (Prx3) as control for cellular redox stress.

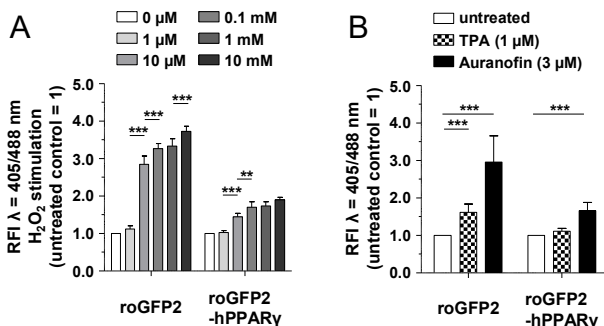
Supplementary Figures



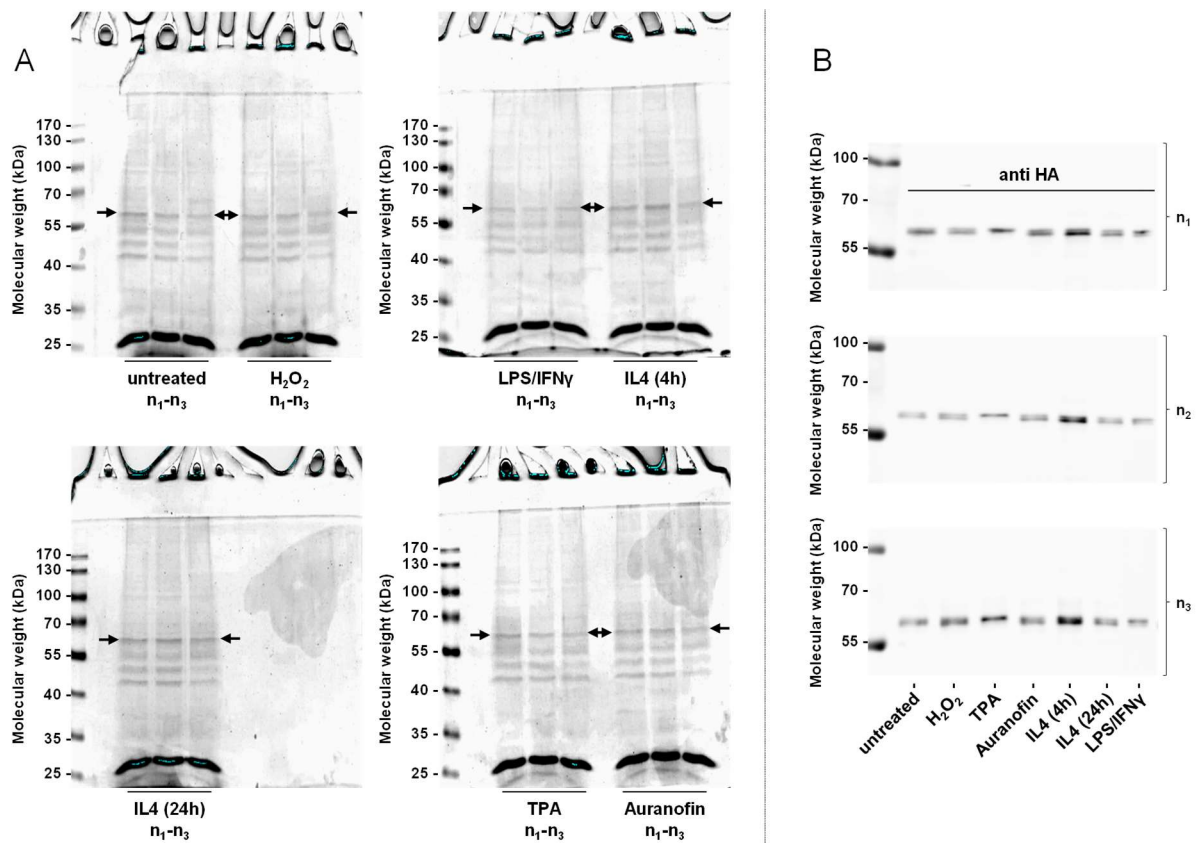
Supplementary Fig. 1. Validation of roGFP2 readout system to determine the redox status of MΦ. J774A.1 cells were transduced with vectors encoding the redox sensitive marker proteins roGFP2 or Grx1-roGFP2. To standardize the experimental setup, the fluorescence of untreated cells was initially monitored at 405 nm and 488 nm. To inflict changes in cellular redox milieu, cells were treated either by 10 mM DTT to cause reducing (A) or 10 mM H₂O₂ to induce oxidizing conditions (B). Fluorometric changes were followed till a constant value was reached. Untreated cells were set as 1.



Supplementary Fig. 2. Effective macrophage polarization in response to LPS/IFN γ and IL4 was verified by expression of corresponding target genes. J774A.1 cells were stably transduced with vectors encoding the redox sensitive marker proteins roGFP2 (A, B) or roGFP2-hPPAR γ (C, D). Cells were harvested at the indicated time points. mRNA expression of target genes was normalized to 18S rRNA. Mean values \pm SD are provided. Untreated control cells were set as 1.

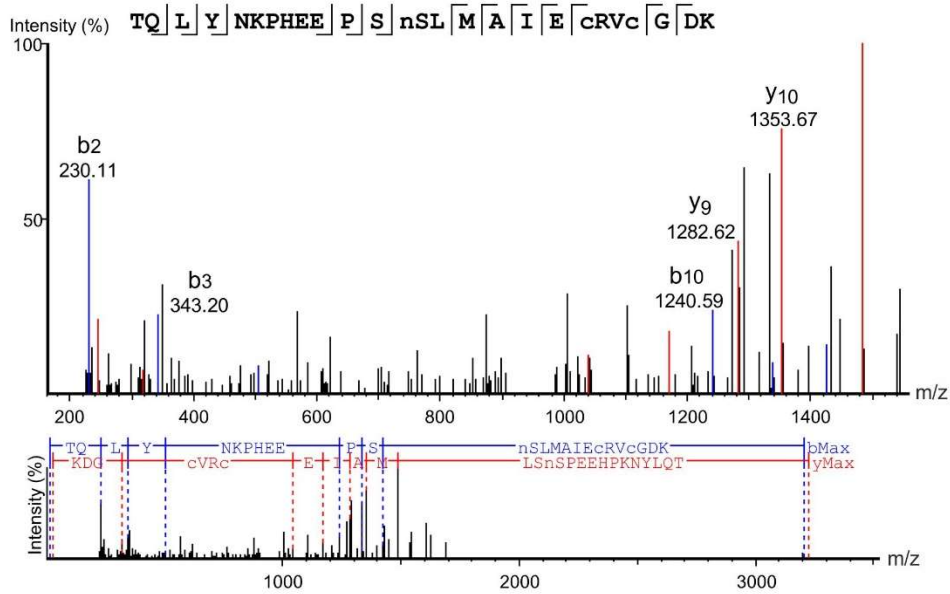


Supplementary Fig. 3. Impact of short-term redox stress on redox status of cytosolic and nuclear proteins. Lentiviral transduced J774A.1 MΦ expression cytosolic roGFP2 and nuclear roGFP2-hPPAR γ were treated with the different redox stress inducers H₂O₂ (1 μ M – 10 mM, 15 min) (A), auranofin (3 μ M, 40 min) and TPA (1 μ M, 30 min) (B). The effect of the stimulation was monitored by FACS-analysis at 405 nm and 488 nm. Quantification (Mean values \pm SD) of flow cytometric data is provided in (A, B). All experiments were performed at least three times. Untreated control cells were set as 1. (**p \leq 0.01, ***p \leq 0.001).

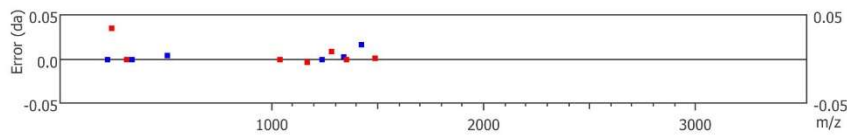


Supplementary Fig. 4. Protein input of carried out LC/MS. J774A.1 cells were transduced using lentiviral particles with integrated N- and C-terminal HA-labelled hPPAR γ . To induce redox stress, M Φ were treated by 1 μ g/ml LPS combined with 10 U/ml IFN γ for 4 h, 10 ng/ml IL4 for 4 h and 24 h, H_2O_2 (100 μ M, 15 min), auranofin (3 μ M, 40 min), or TPA (1 μ M, 30 min). Untreated cells were used as controls. The redox states of reduced cysteines were fixed by NEM before cell harvest on ice. HA-tagged hPPAR γ was HA-immunoprecipitated and passed SDS-PAGE before LC/MS. For this, three SDS gel pieces (n_1-n_3) containing HA-tagged hPPAR γ of each sample treatment were separately cut out of the coomassie stained SDS gel at the indicated areas and later on used as LC/MS samples (**A**, arrows). Western blot studies were performed by using an anti HA antibody to determine the specific size of HA-tagged hPPAR γ (**B**).

A



#	Immonium	b	b-H2O	Seq	y	y-H2O	#
1	74.06	102.06	84.04	T			26
2	101.07	230.11	212.10	Q	3122.48	3104.47	25
3	86.10	343.20	325.19	L	2994.42	2976.41	24
4	136.08	506.26	488.25	Y	2881.34	2863.33	23
5	87.06	620.30	602.29	N	2718.27	2700.26	22
6	101.11	748.40	730.39	K	2604.23	2586.22	21
7	70.07	845.45	827.44	P	2476.14	2458.13	20
8	110.07	982.51	964.50	H	2379.08	2361.07	19
9	102.06	1111.55	1093.54	E	2242.02	2224.01	18
10	102.06	1240.59	1222.59	E	2112.98	2094.97	17
11	70.07	1337.64	1319.64	P	1983.94	1965.93	16
12	60.04	1424.66	1406.67	S	1886.89	1868.88	15
13	88.04	1539.71	1521.70	N(+.98)	1799.85	1781.84	14
14	60.04	1626.74	1608.73	S	1684.83	1666.82	13
15	86.10	1739.82	1721.81	L	1597.80	1579.79	12
16	104.05	1870.86	1852.85	M	1484.71	1466.70	11
17	44.05	1941.90	1923.89	A	1353.67	1335.66	10
18	86.10	2054.99	2036.98	I	1282.62	1264.62	9
19	102.06	2184.03	2166.02	E	1169.55	1151.54	8
20	206.10	2417.12	2399.11	C(+130.08)	1040.51	1022.50	7
21	129.11	2573.22	2555.21	R	807.42	789.41	6
22	72.08	2672.29	2654.28	V	651.32	633.31	5
23	206.10	2905.37	2887.36	C(+130.08)	552.25	534.24	4
24	30.03	2962.40	2944.39	G	319.16	301.15	3
25	88.04	3077.42	3059.41	D	262.14	244.09	2
26	101.11			K	147.11	129.10	1

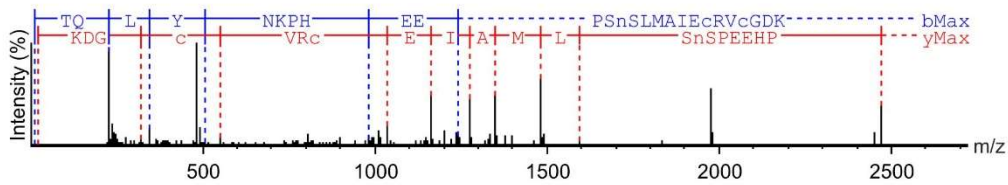
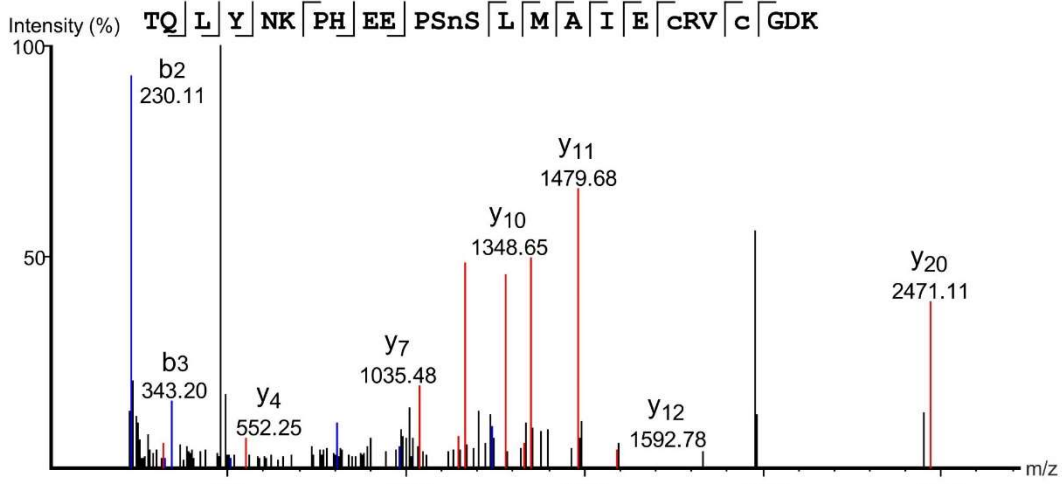


B

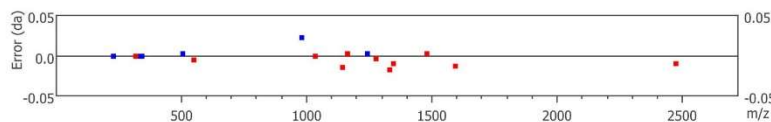
T QLYNKPHEEP SNSLMAIECR VCGDK



- d Deamidation (NQ) (+0.98)
- c D5 N-ethylmaleimide on cysteines (+130.08)
- c Disulfide Bridge unpaired fragmentation (-2.02)
- n N-ethylmaleimide on cysteines (+125.05)



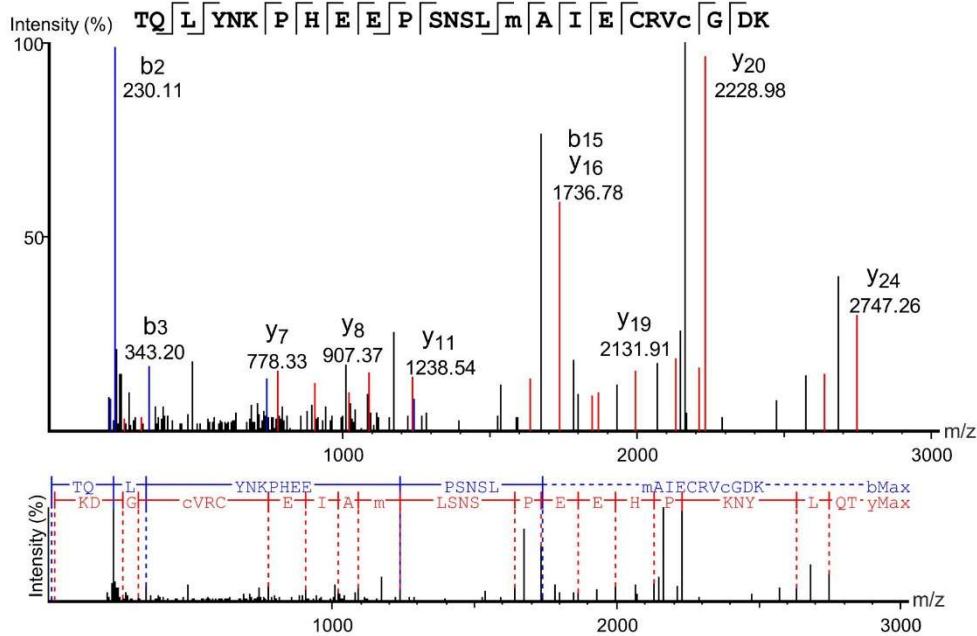
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4	136.08	506.26	488.25	Y	2876.31	2858.30	23
5	87.06	620.30	602.29	N	2713.24	2695.23	22
6	101.11	748.40	730.39	K	2599.20	2581.19	21
7	70.07	845.45	827.44	P	2471.11	2453.09	20
8	110.07	982.49	964.50	H	2374.05	2356.04	19
9	102.06	1111.55	1093.54	E	2236.99	2218.98	18
10	102.06	1240.59	1222.59	E	2107.95	2089.94	17
11	70.07	1337.65	1319.64	P	1978.91	1960.90	16
12	60.04	1424.68	1406.67	S	1881.86	1863.84	15
13	88.04	1539.71	1521.70	N(+.98)	1794.82	1776.81	14
14	60.04	1626.74	1608.73	S	1679.80	1661.79	13
15	86.10	1739.82	1721.81	L	1592.78	1574.75	12
16	104.05	1870.86	1852.85	M	1479.68	1461.67	11
17	44.05	1941.90	1923.89	A	1348.65	1330.65	10
18	86.10	2054.99	2036.98	I	1277.61	1259.59	9
19	102.06	2184.03	2166.02	E	1164.51	1146.52	8
20	201.07	2412.09	2394.07	C(+125.05)	1035.48	1017.47	7
21	129.11	2568.19	2550.18	R	807.42	789.41	6
22	72.08	2667.25	2649.24	V	651.32	633.31	5
23	206.10	2900.34	2882.33	C(+130.08)	552.25	534.24	4
24	30.03	2957.36	2939.35	G	319.16	301.15	3
25	88.04	3072.39	3054.38	D	262.14	244.13	2
26	101.11			K	147.11	129.10	1



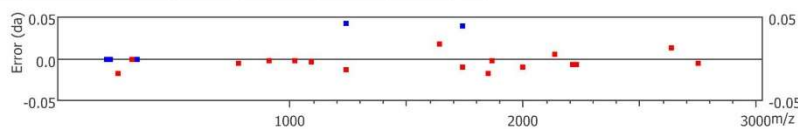
C

T QLYNKPHEEP SNSLMAIECR VCGDK

Disulfide Bridge unpaired fragmentation (-2.02)
Oxidation (M) (+15.99)



#	Immonium	b	b-H2O	Seq	y	y-H2O	#
1	74.06	102.06	84.04	T			26
2	101.07	230.11	212.10	Q	2875.31	2857.30	25
3	86.10	343.20	325.19	L	2747.26	2729.24	24
4	136.08	506.26	488.25	Y	2634.16	2616.16	23
5	87.06	620.30	602.29	N	2471.11	2453.10	22
6	101.11	748.40	730.39	K	2357.06	2339.05	21
7	70.07	845.45	827.44	P	2228.98	2210.97	20
8	110.07	982.51	964.50	H	2131.91	2113.91	19
9	102.06	1111.55	1093.54	E	1994.87	1976.85	18
10	102.06	1240.55	1222.59	E	1865.82	1847.82	17
11	70.07	1337.65	1319.64	P	1736.78	1718.76	16
12	60.04	1424.68	1406.67	S	1639.70	1621.71	15
13	87.06	1538.72	1520.71	N	1552.69	1534.68	14
14	60.04	1625.76	1607.75	S	1438.64	1420.63	13
15	86.10	1738.80	1720.83	L	1351.61	1333.60	12
16	120.05	1885.88	1867.86	M(+15.99)	1238.54	1220.52	11
17	44.05	1956.91	1938.90	A	1091.50	1073.48	10
18	86.10	2070.00	2051.99	I	1020.46	1002.45	9
19	102.06	2199.04	2181.03	E	907.37	889.36	8
20	76.02	2302.05	2284.04	C	778.33	760.32	7
21	129.11	2458.15	2440.14	R	675.32	657.31	6
22	72.08	2557.22	2539.21	V	519.22	501.21	5
23	74.00	2658.21	2640.20	C(-2.02)	420.15	402.14	4
24	30.03	2715.23	2697.22	G	319.16	301.15	3
25	88.04	2830.26	2812.24	D	262.16	244.13	2
26	101.11			K	147.11	129.10	1



Supplementary Fig. 5. Representative MS2 spectra of a peptide at AA position 90-115. This peptide contains C109 and C112, shown in (A) with D5-NEM ($m/z=645.5149$, $z=5$, $\text{ppm}=5.5$, $-10\lg P=20.61$), in (B) with D5-NEM, NEM, and disulfide ($m/z=805.3776$, $z=4$, $\text{ppm}=-2.4$, $-10\lg P=34.15$), and in (C) with disulfide only ($m/z=744.8454$, $z=4$, $\text{ppm}=-0.3$, $-10\lg P=57.03$). Ion tables shows fragment coverage by b- and y-ions.

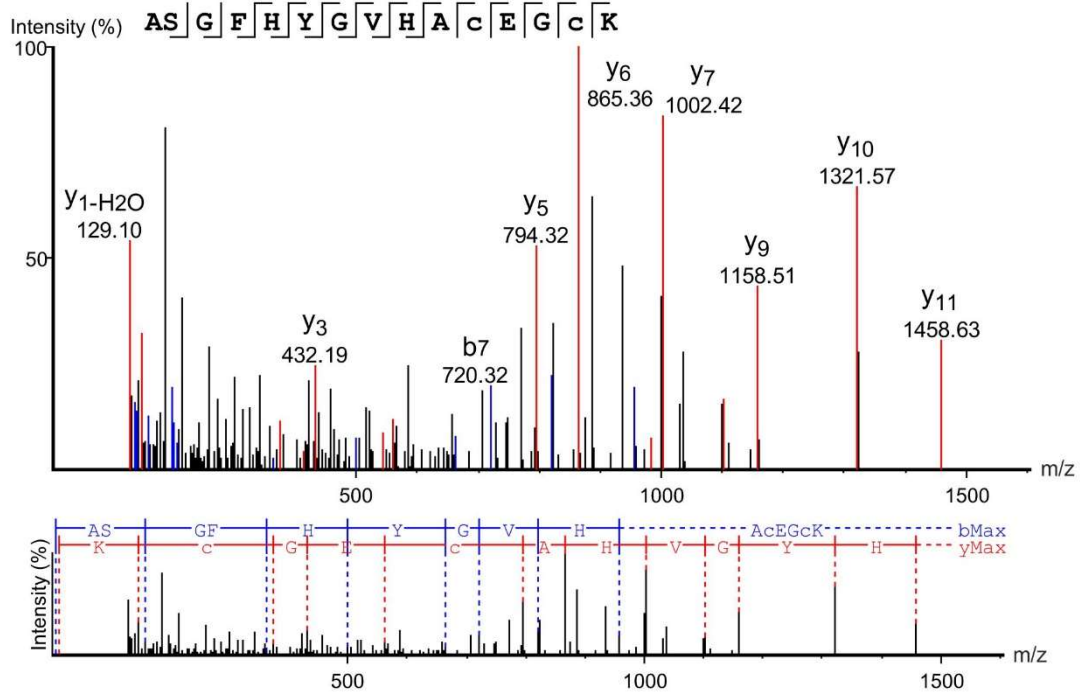
A

ASGFH YGVHACEGCK

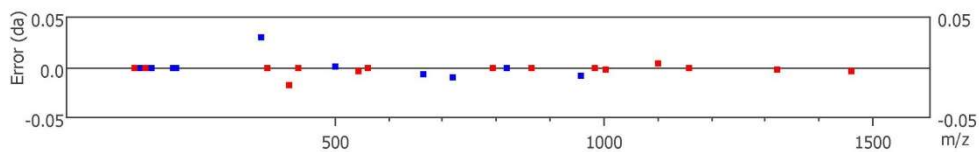


d D5 N-ethylmaleimide on cysteines (+130.08)

n N-ethylmaleimide on cysteines (+125.05)



#	Immonium	b	b-H2O	Seq	y	y-H2O	#
1	44.05	72.04	54.03	A			15
2	60.04	159.08	141.07	S	1749.75	1731.74	14
3	30.03	216.10	198.09	G	1662.72	1644.71	13
4	120.08	363.14	345.16	F	1605.70	1587.69	12
5	110.07	500.22	482.22	H	1458.63	1440.62	11
6	136.08	663.30	645.28	Y	1321.57	1303.56	10
7	30.03	720.32	702.30	G	1158.51	1140.50	9
8	72.08	819.38	801.37	V	1101.48	1083.48	8
9	110.07	956.45	938.43	H	1002.42	984.41	7
10	44.05	1027.47	1009.46	A	865.36	847.35	6
11	206.10	1260.56	1242.55	C(+130.08)	794.32	776.31	5
12	102.06	1389.61	1371.60	E	561.23	543.23	4
13	30.03	1446.63	1428.62	G	432.19	414.20	3
14	201.07	1674.68	1656.67	C(+125.05)	375.17	357.16	2
15	101.11			K	147.11	129.10	1

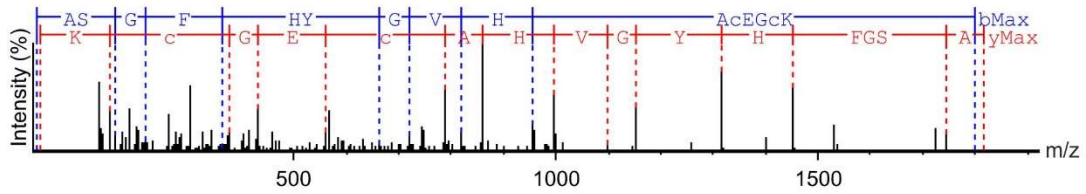
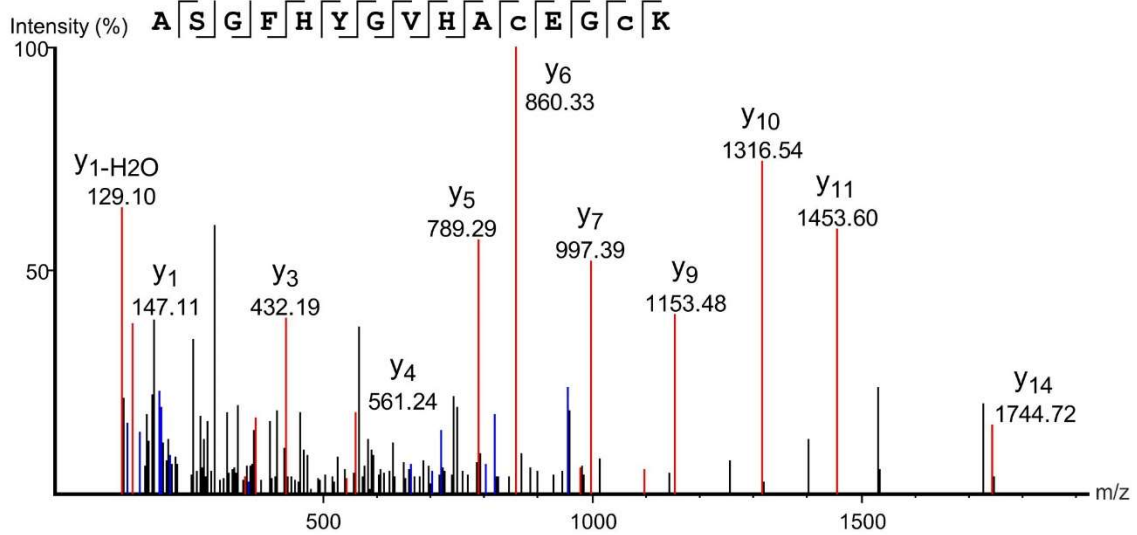


B

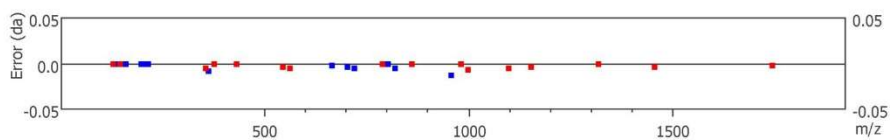
ASGFH YGVHACEGCK



n N-ethylmaleimide on cysteines (+125.05)



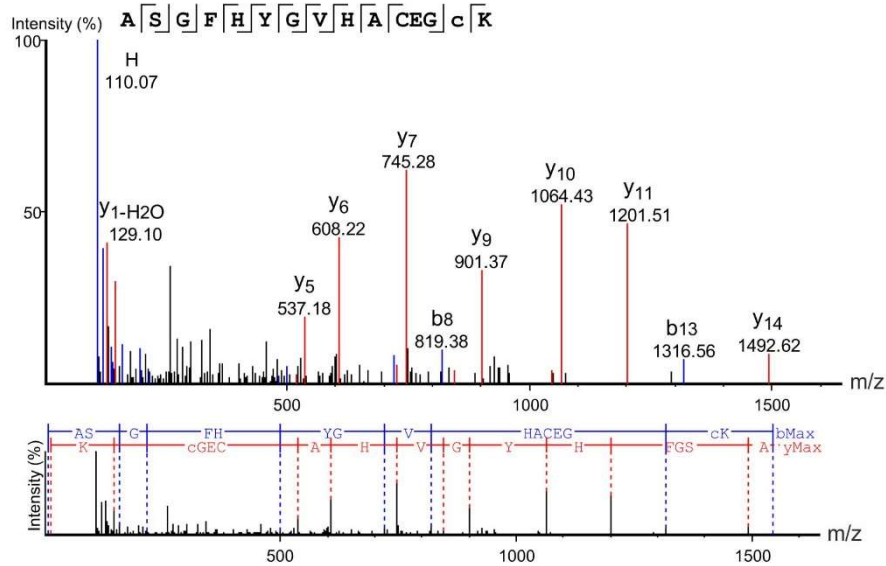
#	Immonium	b	b-H2O	Seq	y	y-H2O	#
1	44.05	72.04	54.03	A			15
2	60.04	159.08	141.07	S	1744.72	1726.71	14
3	30.03	216.10	198.09	G	1657.69	1639.68	13
4	120.08	363.18	345.16	F	1600.67	1582.66	12
5	110.07	500.23	482.22	H	1453.60	1435.59	11
6	136.08	663.29	645.28	Y	1316.54	1298.53	10
7	30.03	720.32	702.30	G	1153.48	1135.47	9
8	72.08	819.38	801.37	V	1096.46	1078.44	8
9	110.07	956.45	938.43	H	997.39	979.38	7
10	44.05	1027.47	1009.46	A	860.33	842.32	6
11	201.07	1255.53	1237.52	C(+125.05)	789.29	771.28	5
12	102.06	1384.57	1366.56	E	561.24	543.23	4
13	30.03	1441.60	1423.59	G	432.19	414.18	3
14	201.07	1669.65	1651.64	C(+125.05)	375.17	357.17	2
15	101.11			K	147.11	129.10	1



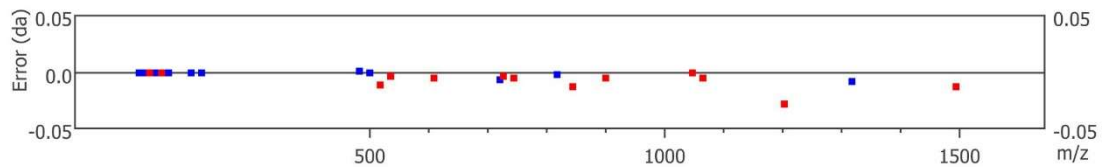
C

ASGFH YGVHACEGCK

d Disulfide Bridge unpaired fragmentation (-2.02)

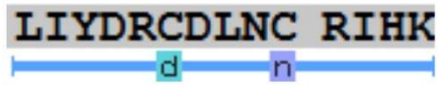


#	Immonium	b	b-H2O	Seq	y	y-H2O	#
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2	60.04	159.08	141.07	S	1492.62	1474.59	14
3	30.03	216.10	198.09	G	1405.57	1387.56	13
4	120.08	363.17	345.16	F	1348.55	1330.54	12
5	110.07	500.23	482.21	H	1201.51	1183.47	11
6	136.08	663.29	645.28	Y	1064.43	1046.41	10
7	30.03	720.32	702.30	G	901.37	883.35	9
8	72.08	819.38	801.37	V	844.35	826.33	8
9	110.07	956.44	938.43	H	745.28	727.27	7
10	44.05	1027.47	1009.46	A	608.22	590.20	6
11	76.02	1130.48	1112.47	C	537.18	519.18	5
12	102.06	1259.53	1241.52	E	434.17	416.16	4
13	30.03	1316.56	1298.54	G	305.12	287.11	3
14	74.00	1417.54	1399.53	C(-2.02)	248.10	230.09	2
15	101.11			K	147.11	129.10	1

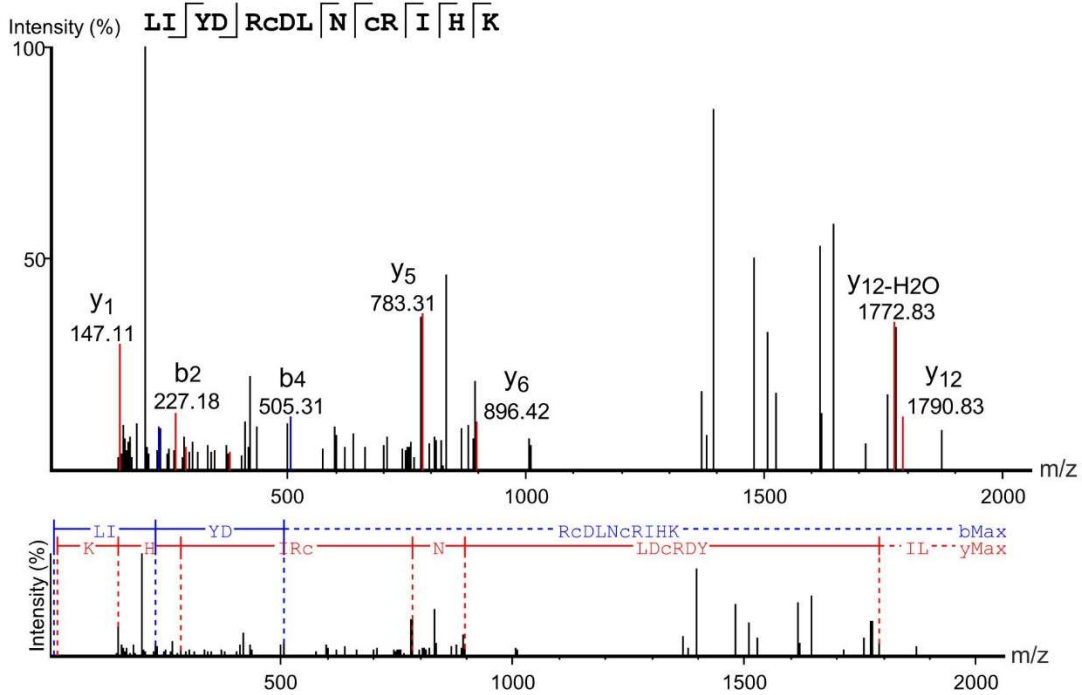


Supplementary Fig. 6. Representative MS2 spectra of a peptide at AA position 116-130. This peptide contains C126 and C129, shown in (A) with D5-NEM and NEM ($m/z=607.6014$, $z=3$, $\text{ppm}=0.3$, $-\log P=48.60$), in (B) with NEM ($m/z=605.9240$, $z=3$, $\text{ppm}=-0.3$, $-\log P=59.65$), and in (C) with a disulfide ($m/z=521.8882$, $z=3$, $\text{ppm}=4.9$, $-\log P=50.31$). Ion tables shows fragment coverage by b- and y-ions.

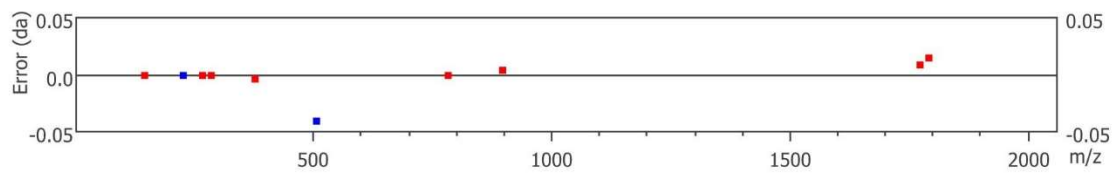
A



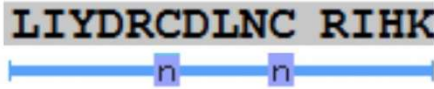
d D5 N-ethylmaleimide on cysteines (+130.08)
n N-ethylmaleimide on cysteines (+125.05)



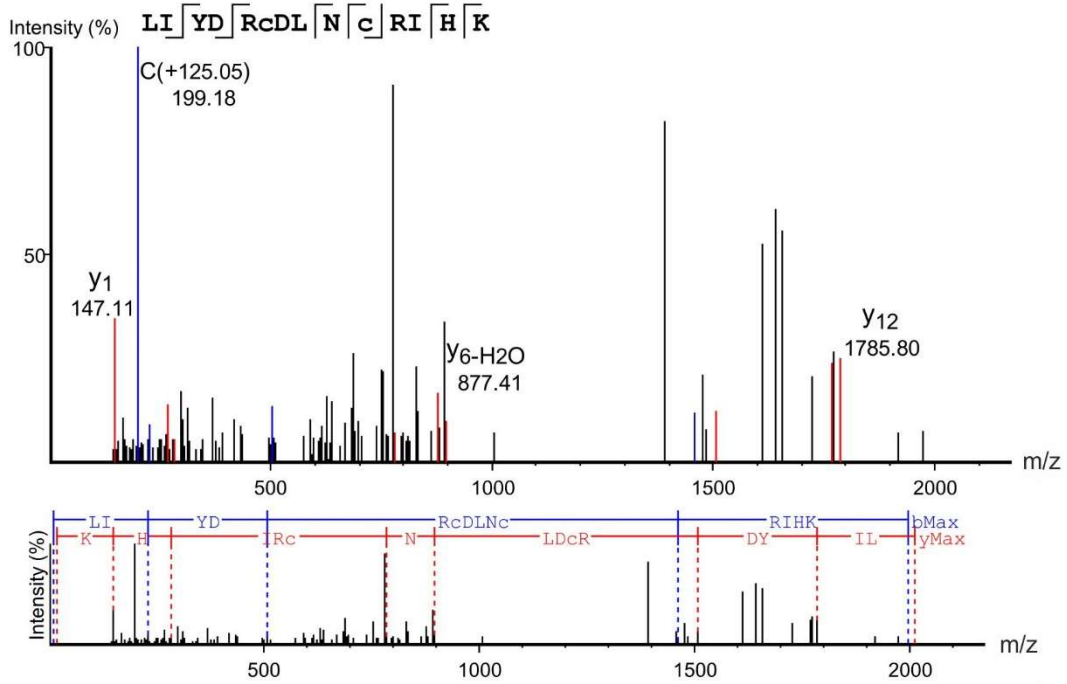
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2	86.10	227.18	209.17	I	1903.93	1885.92	13
3	136.08	390.24	372.23	Y	1790.83	1772.83	12
4	88.04	505.31	487.26	D	1627.78	1609.77	11
5	129.11	661.37	643.36	R	1512.76	1494.75	10
6	206.10	894.46	876.45	C(+130.08)	1356.66	1338.65	9
7	88.04	1009.48	991.47	D	1123.57	1105.56	8
8	86.10	1122.57	1104.56	L	1008.54	990.53	7
9	87.06	1236.61	1218.60	N	895.45	877.45	6
10	201.07	1464.67	1446.66	C(+125.05)	781.41	763.40	5
11	129.11	1620.77	1602.76	R	553.36	535.35	4
12	86.10	1733.85	1715.84	I	397.26	379.25	3
13	110.07	1870.91	1852.90	H	284.17	266.16	2
14	101.11			K	147.11	129.10	1



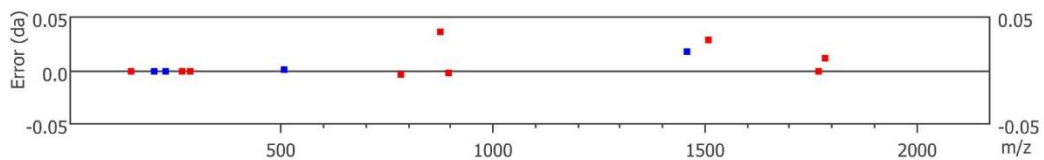
B



n N-ethylmaleimide on cysteines (+125.05)

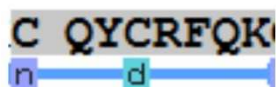


#	Immonium	b	b-H2O	Seq	y	y-H2O	#
1	86.10	114.09	96.08	L			14
2	86.10	227.18	209.17	I	1898.90	1880.89	13
3	136.08	390.24	372.23	Y	1785.80	1767.81	12
4	88.04	505.26	487.26	D	1622.75	1604.74	11
5	129.11	661.37	643.36	R	1507.70	1489.72	10
6	201.07	889.42	871.41	C(+125.05)	1351.62	1333.61	9
7	88.04	1004.45	986.44	D	1123.57	1105.56	8
8	86.10	1117.54	1099.52	L	1008.54	990.53	7
9	87.06	1231.58	1213.57	N	895.46	877.41	6
10	201.07	1459.62	1441.62	C(+125.05)	781.42	763.40	5
11	129.11	1615.74	1597.73	R	553.36	535.35	4
12	86.10	1728.82	1710.81	I	397.26	379.25	3
13	110.07	1865.88	1847.87	H	284.17	266.16	2
14	101.11			K	147.11	129.10	1



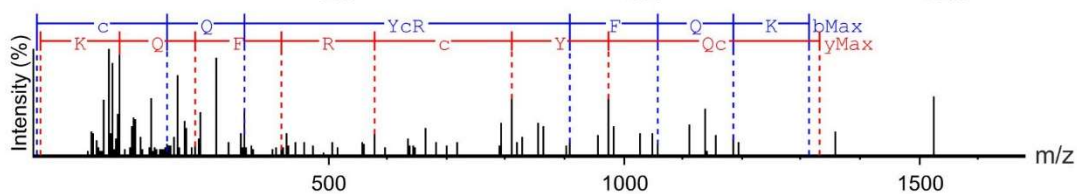
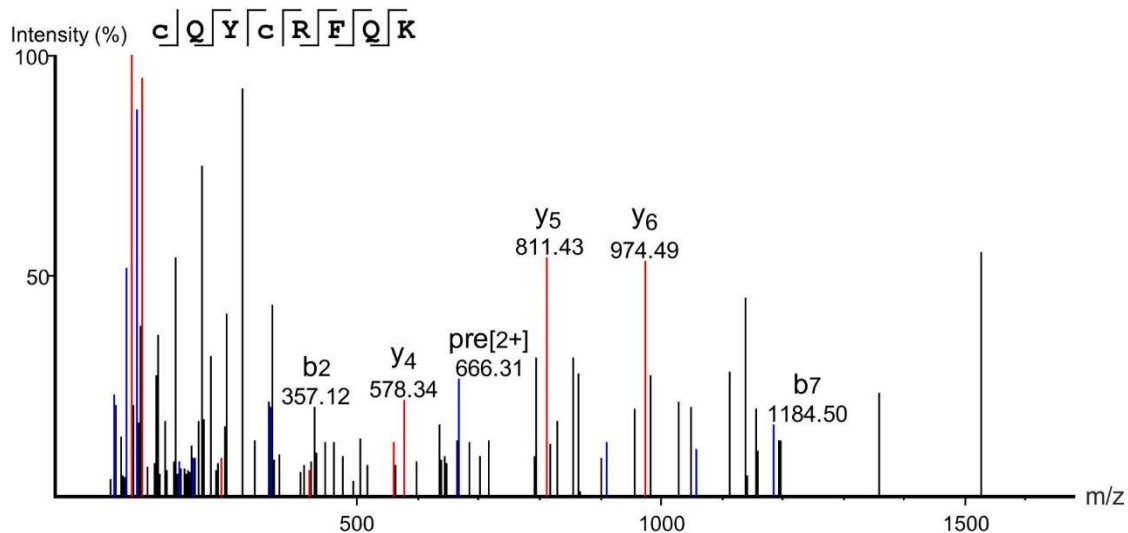
Supplementary Fig. 7. Representative MS2 spectra of a peptide at AA position 141-154. This peptide contains C146 and C150, shown in (A) with D5-NEM and NEM ($m/z=505.0099$, $z=4$, $\text{ppm}=1.2$, $-\log P=24.48$), and in (B) with NEM ($m/z=503.7509$, $z=3$, $\text{ppm}=-1.1$, $-\log P=25.98$). Ion tables shows fragment coverage by b- and y-ions.

A

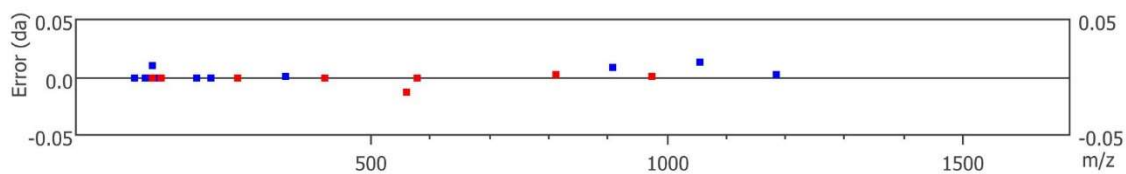


d D5 N-ethylmaleimide on cysteines (+130.08)

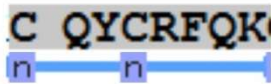
n N-ethylmaleimide on cysteines (+125.05)



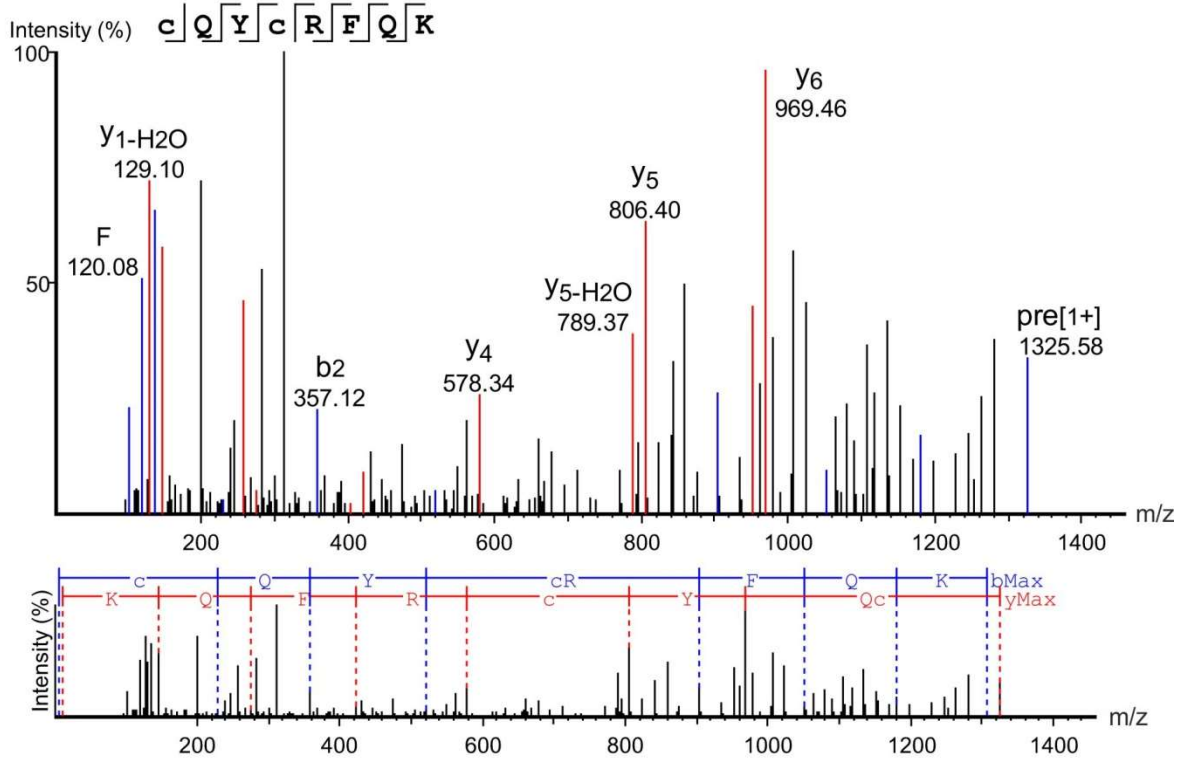
#	Immonium	b	b-H2O	Seq	y	y-H2O	#
1	201.07	229.06	211.05	C(+125.05)			8
2	101.07	357.12	339.11	Q	1102.55	1084.54	7
3	136.08	520.19	502.18	Y	974.49	956.48	6
4	206.10	753.27	735.26	C(+130.08)	811.43	793.42	5
5	129.10	909.37	891.37	R	578.34	560.34	4
6	120.08	1056.43	1038.43	F	422.24	404.23	3
7	101.07	1184.50	1166.49	Q	275.17	257.16	2
8	101.11			K	147.11	129.10	1



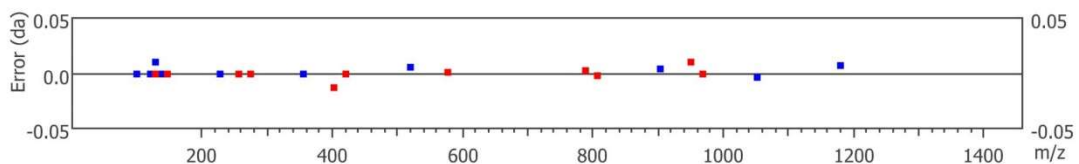
B



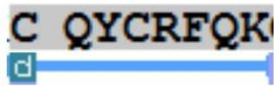
n N-ethylmaleimide on cysteines (+125.05)



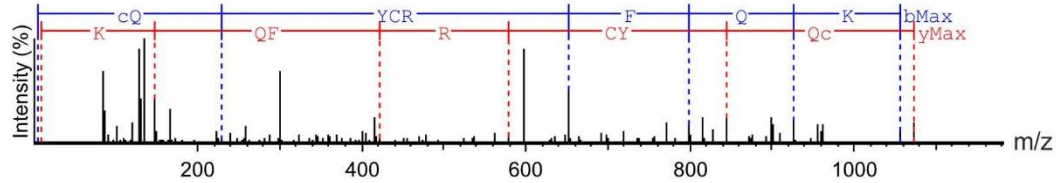
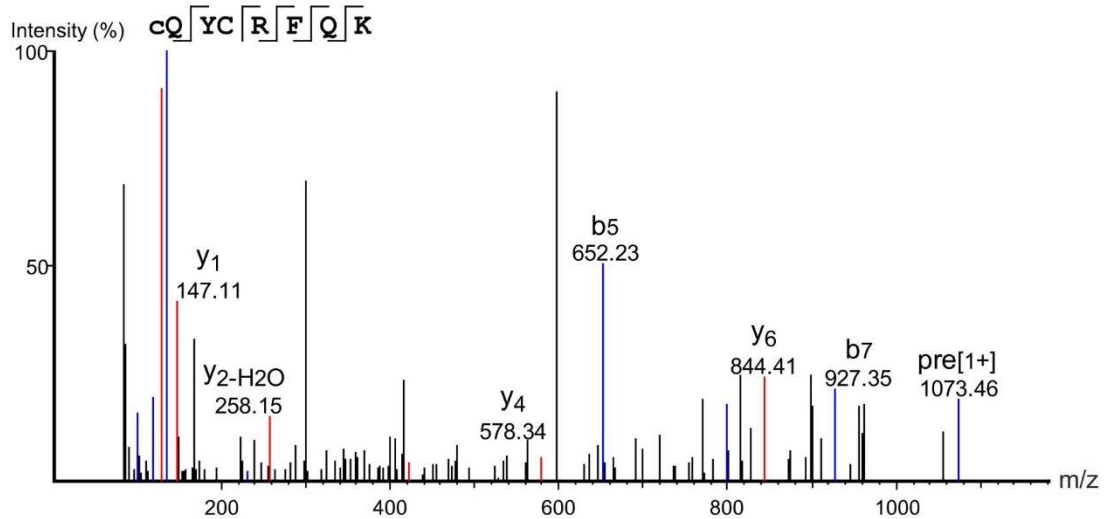
#	Immonium	b	b-H2O	Seq	y	y-H2O	#
1	201.07	229.06	211.05	C(+125.05)			8
2	101.07	357.12	339.11	Q	1097.52	1079.51	7
3	136.08	520.18	502.18	Y	969.46	951.44	6
4	201.07	748.24	730.23	C(+125.05)	806.40	788.38	5
5	129.10	904.34	886.33	R	578.34	560.33	4
6	120.08	1051.42	1033.40	F	422.24	404.24	3
7	101.07	1179.46	1161.46	Q	275.17	257.16	2
8	101.11			K	147.11	129.10	1



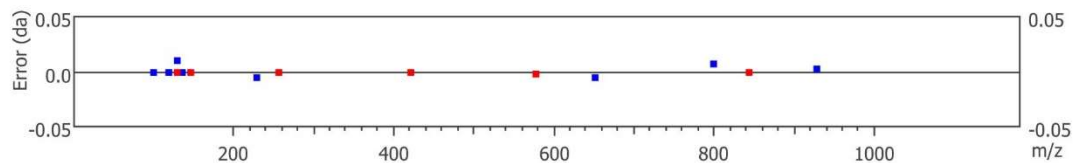
C



Disulfide Bridge unpaired fragmentation (-2.02)



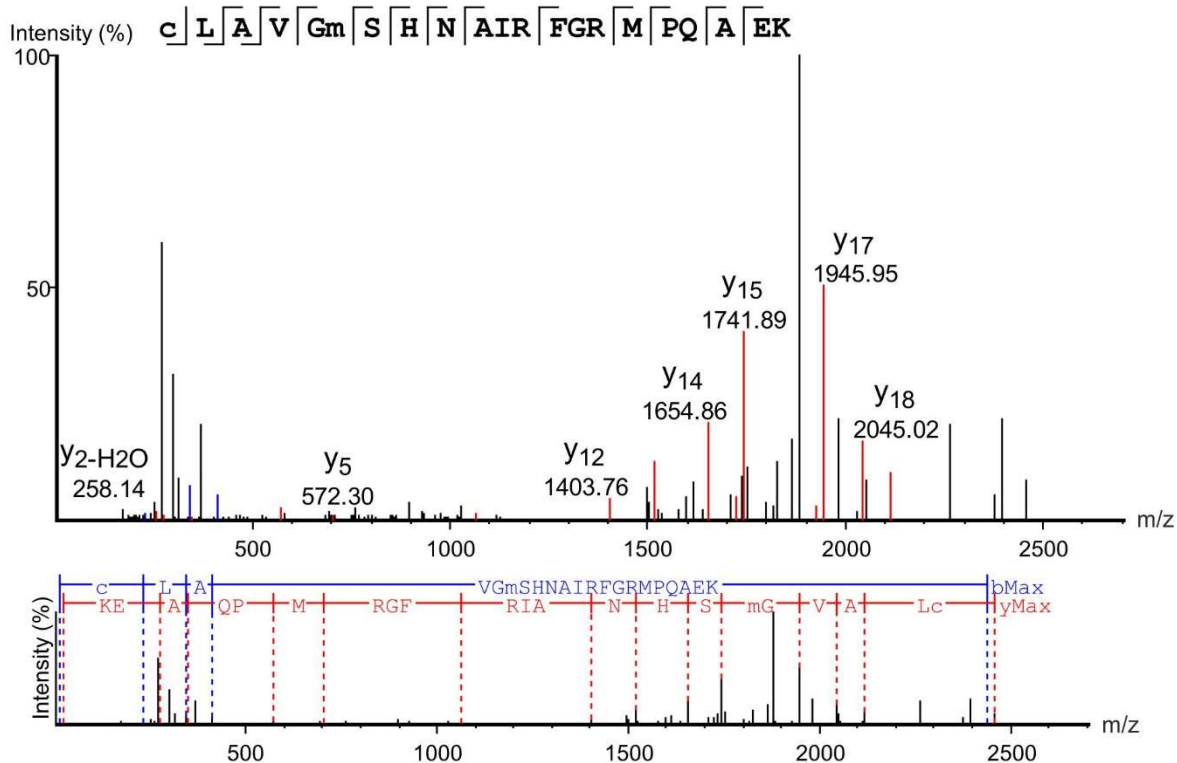
#	Immonium	b	b-H2O	Seq	y	y-H2O	#
1	74.00	102.00	83.99	C(-2.02)			8
2	101.07	230.06	212.05	Q	972.47	954.46	7
3	136.08	393.12	375.11	Y	844.41	826.40	6
4	76.02	496.13	478.12	C	681.35	663.34	5
5	129.10	652.23	634.22	R	578.34	560.33	4
6	120.08	799.29	781.29	F	422.24	404.23	3
7	101.07	927.35	909.35	Q	275.17	257.16	2
8	101.11			K	147.11	129.10	1



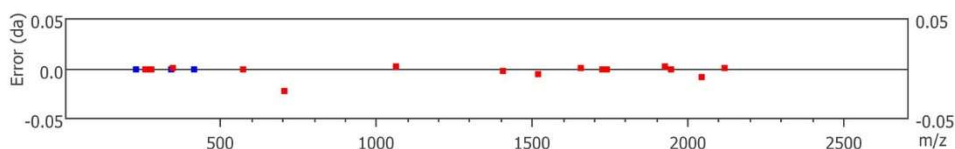
Supplementary Fig. 8. Representative MS2 spectra of a peptide at AA position 160-167. This peptide contains C160 and C163, shown in (A) with D5-NEM and NEM ($m/z=665.8071$, $z=2$, $\text{ppm}=-0.7$, $-\log P=27.36$), in (B) with NEM ($m/z=663.2920$, $z=2$, $\text{ppm}=0.2$, $-\log P=37.87$), and in (C) with a disulfide ($m/z=537.2368$, $z=2$, $\text{ppm}=4.9$, $-\log P=27.52$). Ion tables shows fragment coverage by b- and y-ions.

CLA VGMSHNAIRF GRMPQAEK

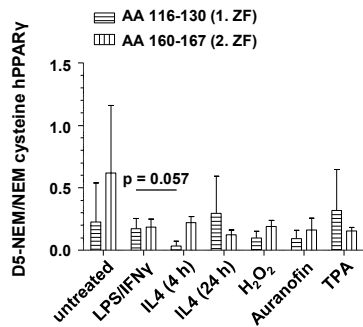
n N-ethylmaleimide on cysteines (+125.05)
 □ Oxidation (M) (+15.99)



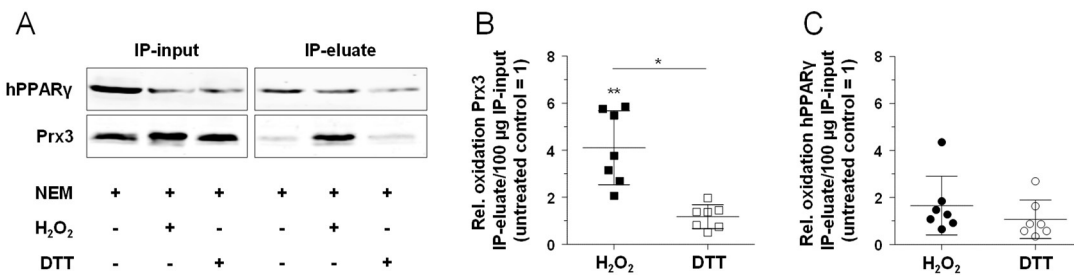
#	Immonium	b	b-H2O	Seq	y	y-H2O	#
1	201.07	229.06	211.05	C(+125.05)			21
2	86.10	342.15	324.14	L	2229.14	2211.13	20
3	44.05	413.19	395.18	A	2116.05	2098.04	19
4	72.08	512.25	494.24	V	2045.02	2027.01	18
5	30.03	569.28	551.27	G	1945.95	1927.93	17
6	120.05	716.31	698.30	M(+15.99)	1888.93	1870.92	16
7	60.04	803.34	785.33	S	1741.89	1723.88	15
8	110.07	940.40	922.39	H	1654.86	1636.85	14
9	87.06	1054.45	1036.43	N	1517.81	1499.79	13
10	44.05	1125.48	1107.47	A	1403.76	1385.75	12
11	86.10	1238.57	1220.56	I	1332.72	1314.71	11
12	129.11	1394.67	1376.66	R	1219.64	1201.63	10
13	120.08	1541.74	1523.73	F	1063.53	1045.52	9
14	30.03	1598.76	1580.75	G	916.47	898.46	8
15	129.11	1754.86	1736.85	R	859.45	841.43	7
16	104.05	1885.90	1867.89	M	703.37	685.33	6
17	70.07	1982.95	1964.94	P	572.30	554.29	5
18	101.07	2111.01	2093.00	Q	475.25	457.24	4
19	44.05	2182.05	2164.04	A	347.19	329.18	3
20	102.06	2311.09	2293.08	E	276.16	258.14	2
21	101.11			K	147.11	129.10	1



Supplementary Fig. 9. Representative MS2 spectra of a peptide at AA position 168-188. This peptide contains C168, shown with NEM ($m/z=615.0542$, $z=0.1$, $\text{ppm}=0.2$, $-10\lg P=48.30$). Ion tables shows fragment coverage by b- and y-ions.



Supplementary Fig. 10. Heavy (D5-NEM) to light (NEM) ratios of hPPAR γ cysteines after redox stress. J774A.1 cells were transduced using lentiviral particles with integrated N- and C-terminal HA-labelled hPPAR γ . To induce redox stress, M Φ were treated by 1 μ g/ml LPS combined with 10 U/ml IFN γ for 4 h, 10 ng/ml IL4 for 4 h and 24 h, H₂O₂ (100 μ M, 15 min), auranofin (3 μ M, 40 min), or TPA (1 μ M, 30 min). Untreated cells were used as controls. The redox states of reduced cysteines were fixed by NEM before cell harvest on ice. HA-tagged hPPAR γ was HA-immunoprecipitated, existing oxidized cysteines denatured and labelled by D5-NEM. Peptides were analysed by LC/MS after LysC digestion. Quantification of the ratio of cellular oxidized (D5-NEM) and reduced (NEM) cysteine are mapped. Peptides of different hPPAR γ amino acid sequences (AA) are depicted in Table 1. Cysteines affiliations to the first and second zinc finger (ZF) are indicated. All experiments were performed at least three times. Mean values \pm SD are provided.



Supplementary Fig. 11. Validation of H₂O₂ induced PTMs at the cysteines of proteins via BIAM switch assay. HEK293T cells were transduced using lentiviral particles with integrated N- and C-terminal HA-labelled hPPAR γ . To induce redox stress, cells were treated by 100 μ M H₂O₂ as oxidizing or 100 μ M DTT as reducing condition. Untreated cells were used as controls. The redox states of present reduced cysteines were fixed by NEM before cell harvest on ice. Remaining oxidized cysteines were reduced with 2.5 mM DTT, respectively, biotin labelled after cell lysis and protein-TCA precipitation. Immunoprecipitation was carried out against biotin-labelled cysteines (oxidized cysteines) and samples were analysed via Western blot. (A) shows a representative Western blot with total lysate (IP-input, each lane 50 μ g) as loading control and the biotinylated IP-eluate. Positive induced redox stress was verified with protein intensities of Prx3 as control protein on each blot in parallel to hPPAR γ . Quantification with mean values \pm SD out of 7 separate experiments of Prx3 (B) and hPPAR γ (C) is provided. Untreated control cells were set as 1 (* \triangleq $p \leq 0.05$, ** \triangleq $p \leq 0.01$).