

1 Supplemental material for
2 Potential of Using cell-free DNA and miRNA in breast milk to Study Breast Cancer
3 Risk
4 Qinghe Song, Yingying Zhang, Hongtao Liu, Yuguang Du

5 **Supplemental Text**

6 **Human donor breast milk samples collection.** Each donor was asked to provide at least 100
7 mL of breast milk (BM) by pumping or hand expressing. Breast milk was processed for BM
8 cells isolation within 2 h of collection using the protocol described behind. Whole BM for
9 stability experiments were packed into 15 ml centrifuge tubes (JetBiofil) and stored at room
10 temperature.

11 **Processing of milk samples and Isolation of BM cells and BM supernatant.** To separate
12 the BM cells, fresh breast milk samples were centrifuged immediately on arrival at the
13 laboratory at 1,500 G for 10 min in 15 mL plastic centrifuge tubes. After removal the milk fat
14 by aspiration, the supernatant was transferred to new tubes, the cell pellet was washed in ~5
15 mL PBS one time, then used for isolation of RNA via traditional TRIZol (Ambion) method,
16 and extraction of genomic DNA (gDNA) via the QIAamp DNA mini kit (Qiagen). The newly
17 transferred supernatant was centrifuged again at 10,000g at RT for 10 min, removed milk fat
18 and collected BM supernatant for stability experiments and future extraction of cell-free
19 nucleic acid. BM cfDNA was extracted using a cfDNA purification Kit (SummerBio). BM
20 cfRNA isolation was carried out using a cfRNA purification Kit (SummerBio). All nucleic
21 acid extraction processes are carried out following the manufacturer's procedure.

22 **Stability experiments of BM cells, cell-free DNA and miRNA.** For BM cells stability test,
23 whole BM from donor 3 and 4 was packed by 10ml each into centrifuge tubes. After
24 incubation for designed time at RT, BM cells were separated to isolate gDNA for detection.
25 For cfDNA stability test, whole BM and BM supernatant from donor 1, 2, 3, 4 and 5 were
26 packed by 2ml each into centrifuge tubes and incubation for designed time at RT, then
27 isolated BM supernatant was used to extract cfDNA for detection of abundance by qRT-PCR.
28 For cell-free miRNA stability test, 2ml of whole BM from donor 3 was incubated 7 days at RT,
29 then isolated BM supernatant to extract cfRNA for generating NGS libraries of miRNA. All
30 samples for incubation at RT were added with sodium azide at 0.05% final concentration as
31 antimicrobial.

32 **Nucleic acid Extraction Procedures.** Isolation of cellular RNA via traditional TRIZol
33 (Ambion) method, extraction of genomic DNA (gDNA) via the QIAamp DNA mini kit
34 (Qiagen). BM cfDNA was extracted using a cfDNA purification Kit (SummerBio). BM
35 cfRNA isolation was carried out using a cfRNA purification Kit (SummerBio). All nucleic
36 acid extraction processes are carried out following the manufacturer's procedure.

37 **Measurement of cfDNA abundance in BM by Using qRT-PCR Assays.** To accurately
38 measure abundance of cfDNA, quantitative real-time PCR method is designed. We choose
39 human long interspersed nuclear elements (LINE) as target amplification region and designed
40 forward primer (5' CAAGTCCTGAGTGACCTAC 3') and reverse primer (5'
41 GTCTCGTTGATCTGTCTAATG 3'). Human DNA Standard (Thermo) was used to generate
42 standard curve. cfDNA samples and diluted DNA standards were subjected to qRT-PCR as

43 follows: initial denaturation at 95 °C for 10 min, then (95 °C for 10 s, 60 °C for 20 s) x 40
44 cycles, and default melt curve setting. Ct values of gradient standards were used to generate
45 standard curve, to which concentrations of cfDNA samples were calculated according. Then
46 concentration of cfDNA was normalized into ng cfDNA per milliliter of BM by converting
47 sample volume of BM for extracting cfDNA and final elution volume

48 **Measurement of RNA difference between BM cellular RNA and BM cfRNA By Using**
49 **RT-qPCR Assays.** RNA samples from different donors were subjected to RT-qPCR Assays to
50 determine the difference of expression levels of LALBA, K-CSN, B-CSN, ELF5, TACSTD2
51 and PPBP, CD99, ITGB3, RGS18, CD45. Reverse transcription was performed using
52 SumScript 1st strand cDNA synthesis kit (SummerBio). qPCR analysis was carried out at a
53 CFX connect Real-time PCR instrument (BioRad Laboratories) using SYBR QPCR Mixture
54 (SummerBio). Forward and reverse primers listed in Table S1. Thermal cycling was set up as
55 the following condition: 95 °C for 10 min, followed by 40 cycles of 10 s at 95 °C, 20 s at 60 °C,
56 and default melt curve setting. The comparative Ct method ($-\Delta Ct$) was used to determine the
57 relative expression levels of each gene. $-\Delta Ct$ was calculated by formula of $-\left[Ct_{(\text{Target gene})}-\right.$
58 $Ct_{(\text{GAPDH})}]$.

59 **Whole Genome Bisulfite Sequencing (WGBS) Library Generation and NGS data**
60 **analysis** were performed by LC Sciences (Houston, TX, USA). The brief process is as
61 follows.

62 ***WGBS Library construction and sequencing.*** cfDNA and sonication-fragmented DNA
63 samples were subjected to bisulfite conversion. The Accel-NGS Methyl-Seq DNA Library Kit
64 (Swift, MI, USA) was utilized for attaching adapters to single-stranded DNA fragments.
65 Briefly, the Adaptase step is a highly efficient, proprietary reaction that simultaneously
66 performs end repair, tailing of 3' ends, and ligation of the first truncated adapter complement
67 to 3' ends. The Extension step is used to incorporate truncated adapter 1 by a primer extension
68 reaction. The Ligation step is used to add the second truncated adapter to the bottom strand
69 only. The Indexing PCR step increases yield and incorporates full length adapters. Bead-based
70 SPRI clean-ups are used to remove both oligonucleotides and small fragments, as well as to
71 change enzymatic buffer composition. Finally, the pair-end 2×150bp sequencing was
72 performed on an illumina Hiseq 4000 platform.

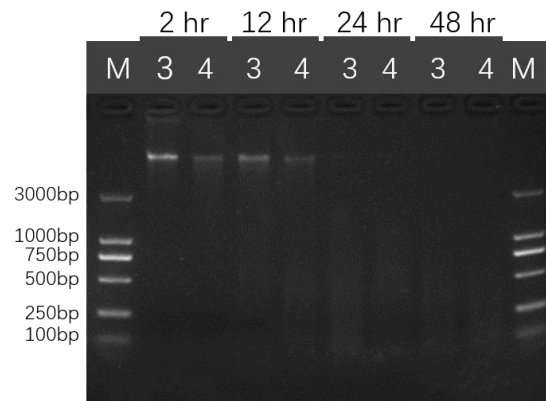
73 ***WGBS Bioinformatics analysis.*** Firstly, Cutadapt [1] and perl scripts in house were used to
74 remove the reads that contained adapter contamination, low quality bases and undetermined
75 bases. Then sequence quality was verified using FastQC (<http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>). Reads that passed quality control were mapped to reference
76 genome using WALT [2]. After alignment, reads were further deduplicated using samtool [3].
77 For each cytosine site (or guanine corresponding to a cytosine on the opposite strand) in the
78 reference genome sequence, the DNA methylation level was determined by the ratio of the
79 number of reads supporting C (methylated) to that of total reads (methylated and
80 unmethylated) using per scripts in house and MethPipe [4]. Differentially methylated regions
81 (DMRs) were calculated by R package-MethylKit [5] with default parameters (1000 bp slide
82 windows, 500 bp overlap, p value < 0.05).

84 **Whole Genome Sequencing(WGS) Library Generation and NGS data analysis for**
85 **cfDNA.** cfDNA samples were directly used for WGS library generation by utilizing NEBNext
86 Ultra DNA Library Prep Kit for Illumina (NEB #E7370) according to manufacturer's

87 recommended protocol. After passing the quality control, libraries were subjected to the
88 pair-end 2×150bp sequencing on an illumina Hiseq 4000 platform. Alignment analysis for
89 raw reads were processed by the Burrows-Wheeler Aligner software to simply determine the
90 chromosomal origin of each sequenced DNA fragments. we counted only sequences that
91 could be mapped to just one location in the hg19 reference human genome with no mismatch
92 [6]. To reduce GC bias introduced by the PCR process, we applied weight to each sequenced
93 read based on the local genomic GC content of large regions of the human genome with
94 relatively homogeneous GC content [7]. We first got the read count for bins of size of 500kb,
95 then averaged the counts in each 0.5% of GC content bin. Counts in each GC content bin is
96 weighted by $W_i = \bar{M} / M_i$, where \bar{M} is the average counts across all GC content bins, and
97 M_i is the average count in bin i. Counts ratio is normalized by *500kb bin counts/ all counts*,
98 and a reference baseline of mean of counts ratios based on a set of normal diploid cfDNAs
99 was created. Copy number of cfDNA samples was calculated by *sample counts ratios/*
100 *reference mean of counts ratios*, which was further plotted into genome-wide copy number for
101 data visualization using ggplot2 [8]. Z-score was calculated based on the same reference data
102 set by referring to previous reports [9].

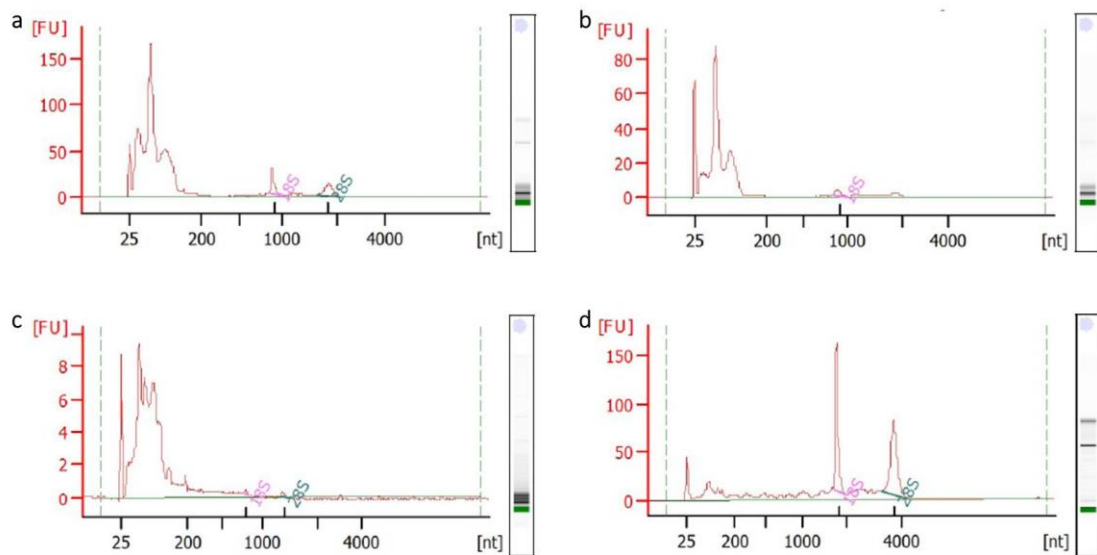
103 **miRNA Sequencing and Bioinformatics analysis.** Service of miRNA library generation and
104 NGS data analysis was provided by LC Sciences (Houston, Texas, USA). In brief, miRNA
105 library was prepared by TruSeq Small RNA Sample Prep Kits (Illumina) and single-end
106 sequencing (36 bp or 50 bp) was performed on an Illumina Hiseq 2500 following the vendor's
107 recommended protocol. Raw reads were subjected to the Illumina pipeline filter (Solexa 0.3),
108 and then the dataset was further processed with an in-house program, ACGT101-miR (LC
109 Sciences) to remove adapter dimers, junk, low complexity, common RNA families (rRNA,
110 tRNA, snRNA, snoRNA) and repeats. Subsequently, unique sequences with length in 18~26
111 nucleotide were mapped to human species precursors in miRBase 22.0 by BLAST search to
112 identify known miRNAs and novel 3p- and 5p- derived miRNAs. Length variation at both 3.
113 and 5. ends and one mismatch inside of the sequence were allowed in the alignment. The
114 unique sequences mapping to human species mature miRNAs in hairpin arms were identified
115 as known miRNAs. The unique sequences mapping to the other arm of known human species
116 precursor hairpin opposite to the annotated mature miRNA-containing arm were considered to
117 be novel 5p- or 3p- derived miRNA candidates. The remaining sequences were mapped to
118 other selected species precursors (with the exclusion of human species) in miRBase 22.0 by
119 BLAST search, and the mapped pre-miRNAs were further BLASTed against the human
120 species genomes to determine their genomic locations. The above two we defined as known
121 miRNAs.

122 **Supplementary Figures**



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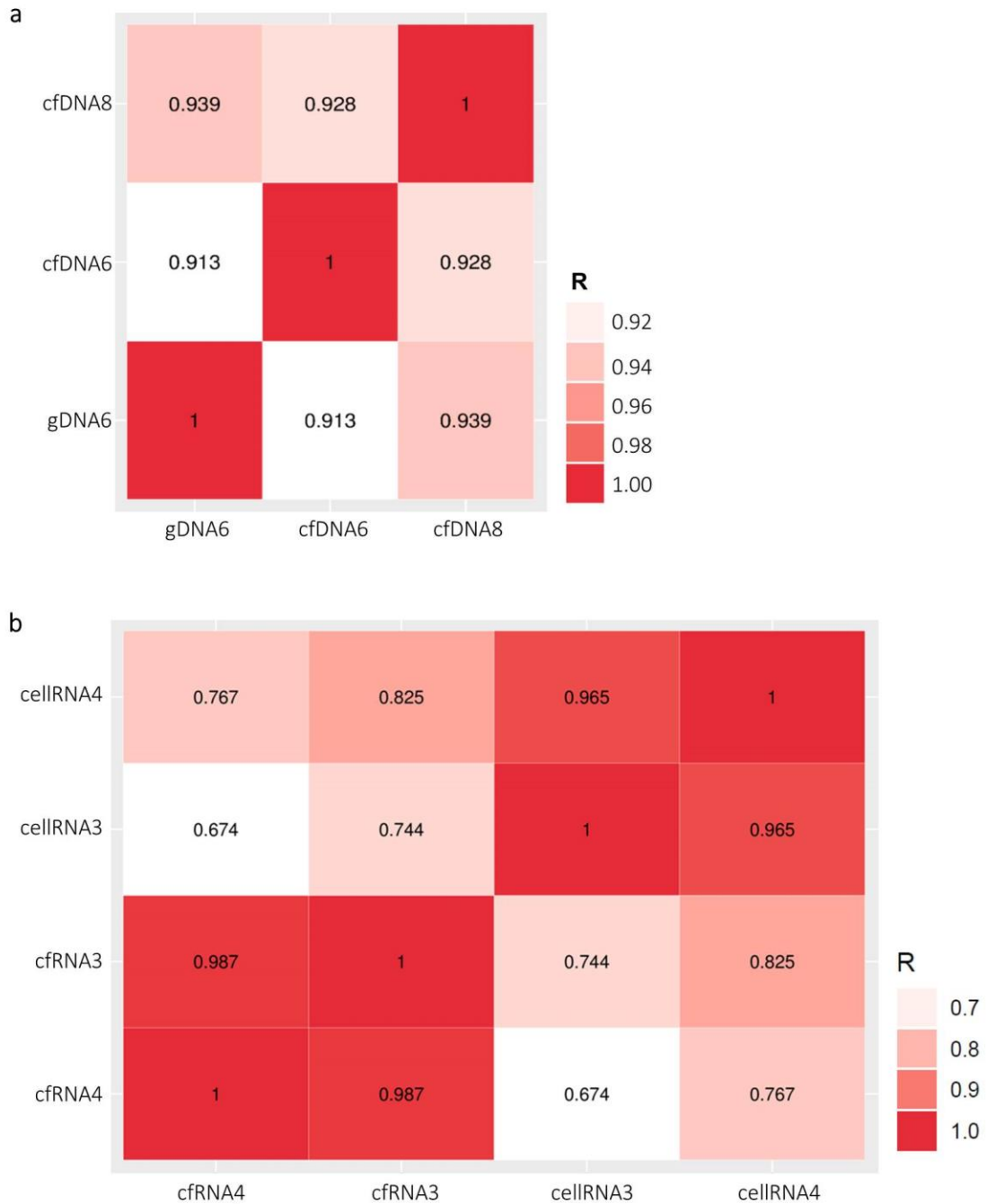
124 **Figure S1:** genomic DNA (gDNA) of BM cells was easily degraded when BM was stored at
125 room temperature. 10mL of Whole BM samples (from donor 3 and 4) were incubated at room
126 temperature for indicated time, then BM cells was isolated and gDNA was extracted detected
127 by agarose gel electrophoresis.



128

129 **Figure S2:** Identification of existence and stability of cfRNA in human BM. (a) cfRNA
130 extracted from fresh BM samples; (b) cfRNA extracted from RT-2-days BM sample; (c)
131 cfRNA extracted from RT-7-days BM sample; (d) Total RNA isolated from fresh BM cells.
132 RNA and cfRNA were detected by Agilent 2100 Bioanalyzer. The BM samples used in this
133 experiment are from donor 3.

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135

136 **Figure S3:** Pearson correlation between samples. (A) Pearson correlation of methylation
 137 between indicated samples according to data of Whole Genome Bisulfite Sequencing
 138 (WGBS). (B) Pearson correlation of miRNA abundance profiles between indicated samples
 139 according to the normalized reads of 413 identical known human miRNAs detectable in all
 140 samples.

141 **Supplementary Tables**

142 **Table S1:** Forward and reverse primers used in real-time PCR.

Genes	Forward primers	Reverse primers
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LALBA	CCTGGATGATGACATTACTGA	ACACTCACAACCTTCTCACAA
B-CSN	GAGGATGAACACCAGGATAA	TGAGCAAGAGGCAGAATG
K-CSN	ATTCTCAGCGGCAATAC	GTTCAACAGTAGCAATGG
ELF5	ACTTCCACAGACATCCTTC	ATGCCAGCCATCAGTTAG
TACSTD2	CACGCTTCCTGATTCCTC	AATGCTTCCTTCAAGACAGA
PPBP	TAGACAGTGACTTGTATGCT	ATCCTTCAGTGTGGCTATC
RGS18	GACACATTCATCTCTACAGAAC	CGGTGATAAGGTAGTCTACTT
ITGB3	TTAGCAGCCAGTCTTGAAG	AGAGAATCTTAACCGCACAT
CD99	ATAGAAGATTGTCCGGCAGAA	CAAAGCACATCGCAAGATT
CD45	TGCTCAGAATGGACAAGTAA	ACCTTCAGCCTGTTCCTT
GAPDH	TCTCTGCTCCTCCTGTTC	GACTCCGACCTTCACCTT

143 LALBA: lactalbumin, alpha; B-CSN: casein beta; K-CSN: casein kappa; ELF5: E74-like factor 5;
144 TACSTD2: tumor-associated calcium signal transducer 2; SPP1: secreted phosphoprotein 1; ELF5:
145 E74-like factor 5; PPBP: pro-platelet basic protein; RGS18: regulator of G-protein signaling 18; ITGB3:
146 integrin, beta 3; CD99: Cluster of differentiation 99; CD45: Cluster of differentiation 45; GAPDH:
147 glyceraldehyde-3-phosphate dehydrogenase.

148 **Table S2: Partial data of normalized reads of detectable miRNAs in all samples.**

miRNA name	miRNA sequence	Normalized reads			
		cfRNA4	cfRNA3	cellRNA3	cellRNA4
hsa-miR-1-3p	TGGAATGTAAAGAAGTATGTAT	13.84	40.95	32.43	15.32
hsa-let-7f-5p	TGAGGTAGTAGATTGTATAGTT	73829.24	39642.02	169680.29	251397.85
hsa-let-7f-1-3p_R-1_1ss21CT	CTATACAATCTATTGCCTTCT	142.51	127.52	245.32	132.35
hsa-let-7d-5p	AGAGGTAGTAGTTGCATAGTT	23649.91	16202.40	15598.37	19948.15
hsa-let-7d-3p_R-2	CTATACGACCTGCTGCCTTT	10422.37	12390.31	4978.73	3429.99
hsa-let-7e-5p	TGAGGTAGGAGTTGTATAGTT	11059.61	3558.48	3586.36	10139.19
hsa-let-7e-3p_R-2	CTATACGGCCTCCTAGCTTT	89.93	108.80	91.17	89.16
hsa-let-7a-5p	TGAGGTAGTAGTTGTATAGTT	69874.08	57908.58	83430.84	129086.21
hsa-let-7a-3p_R-1	CTATACAATCTACTGTCTTT	253.42	427.21	932.40	470.43
hsa-miR-7-5p	TGGAAGACTAGTGATTTGTGTGT	303.00	168.85	732.19	2653.07
hsa-let-7i-5p	TGAGGTAGTAGTTTGTGCTGTT	10651.35	8548.39	36036.83	72137.46
hsa-let-7i-3p	CTGCGCAAGCTACTGCCTTGCT	318.22	445.73	1184.29	759.98
hsa-let-7c-5p	TGAGGTAGTAGTTGTATGGTT	2273.01	2554.75	2570.07	2421.30
hsa-let-7g-5p	TGAGGTAGTAGTTGTACAGTT	95427.35	76654.95	254574.04	418970.55
hsa-let-7g-3p_R-1	CTGTACAGCCACTGCCTTG	193.70	63.17	276.34	149.07
hsa-let-7b-5p	TGAGGTAGTAGTTGTGTGGTT	82701.24	116853.47	81207.83	48586.97
hsa-let-7b-3p_R-1_1ss21CT	CTATACAACCTACTGCCTTCT	3748.07	2484.85	1170.19	601.85
hsa-miR-7-1-3p	CAACAAATCACAGTCTGCCATA	135.59	166.12	166.37	216.64
hsa-let-7f-2-3p_1ss22CT	CTATACAGTCTACTGTCTTCT	5.07	1.95	142.87	85.91
hsa-miR-9-5p	TCTTTGGTTATCTAGCTGTATGA	271.64	224.62	318.00	137.23
hsa-miR-10a-5p_R-1	TACCCTGTAGATCCGAATTTGT	18065.16	11759.74	4595.25	8784.65
hsa-miR-10b-5p_R-1	TACCCTGTAGAACCGAATTTGT	182.63	122.84	69.55	87.77
hsa-miR-15a-5p	TAGCAGCACATAATGGTTTGTG	135.59	272.58	1843.17	1779.78
hsa-miR-15b-5p	TAGCAGCACATCATGGTTTACA	1285.33	827.11	2492.66	2148.97

hsa-miR-15b-3p_R-1	CGAATCATTATTTGCTGCTCT	59.49	63.17	216.18	376.85
hsa-miR-16-5p	TAGCAGCACGTAAATATTGGCG	7283.76	7890.33	12244.75	13131.01
hsa-miR-16-2-3p_L+1R-1	ACCAATATTACTGTGCTGCTTT	221.37	391.91	563.95	693.80
hsa-miR-17-5p	CAAAGTGCTTACAGTGCAGGTAG	1691.40	1989.98	7665.48	6633.94
hsa-miR-17-3p	ACTGCAGTGAAGGCACTTGTAG	134.21	116.99	531.05	475.07
hsa-miR-18a-5p	TAAGTGTCATCTAGTGCAGATAG	81.63	91.25	794.23	714.00
hsa-miR-19b-3p_R-1	TGTGCAAATCCATGCAAAACTG	549.97	624.14	3133.21	3018.31
hsa-miR-19a-3p	TGTGCAAATCTATGCAAACTGA	196.47	291.30	1393.89	1409.19
hsa-miR-20a-5p	TAAAGTGCTTATAGTGCAGGTAG	4964.22	3870.00	31801.10	31943.72
hsa-miR-20a-3p	ACTGCATTATGAGCACTTAAAG	16.60	24.57	155.09	113.54
hsa-miR-21-5p	TAGCTTATCAGACTGATGTTGA	167597.73	225676.62	1216907.49	773411.75
hsa-miR-21-3p_R+1	CAACACCAGTCGATGGGCTGTC	123.14	188.35	2935.36	2381.63
hsa-miR-22-5p_R-1	AGTTCTTCAGTGGCAAGCTTT	3782.66	5124.12	3181.61	2292.47
hsa-miR-22-3p	AAGCTGCCAGTTGAAGAAGTGT	161231.96	171429.96	128999.63	88835.69
hsa-miR-23a-5p_R-1	GGGGTTCCTGGGGATGGGATT	19.37	104.12	9.40	9.75
hsa-miR-23a-3p_R-1	ATCACATTGCCAGGGATTTC	3745.99	4037.88	32975.05	25369.68
hsa-miR-23b-3p_R-5	ATCACATTGCCAGGGATT	729.83	697.84	3237.07	1712.21
hsa-miR-24-3p_R-2	TGGCTCAGTTCAGCAGGAAC	3255.52	4550.88	15252.95	7701.11
hsa-miR-24-2-5p_L+1R-1	GTGCCTACTGAGCTGAAACACA	224.14	331.08	775.90	807.34
hsa-miR-25-5p_R+2	AGGCGGAGACTTGGCAATTGCT	34.59	42.12	28.20	43.19
hsa-miR-25-3p	CATTGCACTGTCTCGGTCTGA	24937.31	34049.67	24623.90	23471.49
hsa-miR-26b-5p_R+1	TTCAAGTAATTCAGGATAGGTT	37110.81	43039.89	98969.64	81495.06
hsa-miR-26b-3p	CCTGTTCTCCATTACTTGGCT	114.84	70.19	112.79	137.23
hsa-miR-26a-5p	TTCAAGTAATTCAGGATAGGCT	60712.99	45029.88	117750.56	109287.12
hsa-miR-26a-2-3p_R-1	CCTATTCTTGATTACTTGTIT	34.59	24.57	179.52	94.04
hsa-miR-27a-5p	AGGGCTTAGCTGCTTGTGAGCA	85.78	104.12	578.05	822.67
hsa-miR-27a-3p_R-1	TTCACAGTGGCTAAGTTCGG	4813.64	12190.85	79125.52	29570.33
hsa-miR-27b-3p_R-1	TTCACAGTGGCTAAGTTCGT	2902.94	3878.77	17108.50	7536.25
hsa-miR-28-5p_R-1	AAGGAGCTCACAGTCTATTGA	2992.18	2159.03	5951.70	4937.64
hsa-miR-28-3p	CACTAGATTGTGAGCTCCTGGA	6856.93	4332.11	1423.03	5482.83
hsa-miR-29a-3p_R-1	TAGCACCATCTGAAATCGGTT	65680.20	92681.09	207835.42	98081.72
hsa-miR-29b-3p_R-3	TAGCACCATTTGAAATCAGT	2256.36	3032.36	5154.10	2840.22
hsa-miR-29c-5p_R-1	TGACCGATTTCCTCGGTGTT	672.41	541.66	1325.28	1090.85
hsa-miR-29c-3p_R-1	TAGCACCATTTGAAATCGGTT	2998.87	5932.52	15753.37	7486.56
hsa-miR-29b-2-5p_R+1	CTGGTTTCACATGGTGGCTTAGA	358.34	604.83	568.65	1151.46
hsa-miR-30b-5p	TGTAAACATCCTACACTCAGCT	383302.82	264658.95	372313.79	346934.96
hsa-miR-30b-3p_R-2	CTGGGAGGTGGATGTTTACT	478.71	377.87	203.96	172.06
hsa-miR-30c-5p_R+1	TGTAAACATCCTACACTCAGCT	54995.42	46195.28	42790.74	47149.16
hsa-miR-30c-2-3p_R-2	CTGGGAGAAGGCTGTTTACT	80.25	46.80	146.63	242.41
hsa-miR-30a-5p	TGTAAACATCCTCGACTGGAAG	143946.44	97103.28	178924.64	183650.88
hsa-miR-30a-3p_1ss22CT	CTTTCAGTCGATGTTTGAGT	3262.44	3162.80	3578.73	5698.78
hsa-miR-30d-5p	TGTAAACATCCCCGACTGGAAG	3694759.6	2510709.18	1131940.55	1230804.8
hsa-miR-30d-3p_R-2	CTTTCAGTCAGATGTTTGCT	1120.68	1056.41	1885.94	1538.41
hsa-miR-30e-5p_R+2	TGTAAACATCCTTACTGGAAGCT	63270.96	44668.19	92393.68	156105.41

hsa-miR-30e-3p_1ss22CT	CTTTCAGTCGGATGTTTACAGT	2883.34	4249.63	4055.73	9630.66
hsa-miR-32-5p_R-2	TATTGCACATTACTAAGTTG	408.15	1109.06	3934.49	1496.27
hsa-miR-32-3p_R-1	CAATTTAGTGTGTGTGATATT	42.89	64.34	143.81	128.17
hsa-miR-33a-3p_R-1	CAATGTTTTCCACAGTGCATCA	15.22	26.91	121.25	107.27
hsa-miR-33b-3p_R-1	CAGTGCCTCGGCAGTGCAGCC	304.38	214.09	86.47	71.75
hsa-miR-34a-5p_R+1	TGGCAGTGTCTTAGCTGGTTGTT	95.47	113.48	1263.25	1480.94
hsa-miR-92b-3p_R-2	TATTGCACTCGTCCCGGCT	3241.68	2805.40	802.06	544.03
hsa-miR-92a-1-5p	AGGTTGGGATCGGTTGCAATGCT	24.90	65.51	25.38	34.83
hsa-miR-92a-3p	TATTGCACTTGTCCCGGCTGT	71133.05	52695.39	36107.63	24183.40
hsa-miR-93-5p	CAAAGTGCTTTCGTGCAGGTAG	7586.07	6588.83	12813.87	14362.22
hsa-miR-93-3p_R+1	ACTGCTGAGCTAGCACTCCCGA	166.03	277.26	158.85	137.92
hsa-miR-96-5p_R-2	TTTGGCACTAGCACATTTTTG	12768.89	12150.49	16593.27	26239.02
hsa-miR-98-5p	TGAGGTAGTAAGTTGTATTGTT	2425.25	1655.57	5080.95	7802.61
hsa-miR-98-3p_L-1R+1_1ss22CT	TATACAACCTACTACTTTCCCTT	52.58	93.59	89.29	60.60
hsa-miR-99a-5p_R-1	AACCCGTAGATCCGATCTTGT	161476.85	114593.15	62097.86	45214.04
hsa-miR-99a-3p_R+1	CAAGCTCGCTTCTATGGGTCTGA	98.23	87.74	112.79	80.80
hsa-miR-99b-5p	CACCCGTAGAACCAGCTTGCG	220722.34	112993.91	19611.33	48173.84
hsa-miR-99b-3p_R-2	CAAGCTCGTGTCTGTGGGTC	864.73	765.11	423.90	803.86
hsa-miR-100-5p	AACCCGTAGATCCGAACCTGTG	11176.41	5719.60	2669.83	4109.16
hsa-miR-101-5p_L+1R-2	TCAGTTATCACAGTGTGATG	8.30	8.19	55.46	20.20
hsa-miR-101-3p_R+1	TACAGTACTGTGATAACTGAAG	16836.56	29211.00	136941.91	79067.46
hsa-miR-103a-2-5p	AGCTTCTTTACAGTGTGCCTTG	35.97	7.02	52.64	43.19
hsa-miR-103a-3p	AGCAGCATTGTACAGGGCTATGA	38637.80	22561.93	35255.44	76849.76
hsa-miR-106b-5p	TAAAGTGCTGACAGTGCAGAT	909.00	2044.97	8091.73	5302.42
hsa-miR-106b-3p_L+2R-2	TACCGCACTGTGGTACTTGCT	2252.44	2146.75	1731.32	2544.63
hsa-miR-106a-5p_1ss1AC	CAAAGTGCTTACAGTGCAGGTAG	1691.40	1989.98	7665.48	6633.94
hsa-miR-107_R-2	AGCAGCATTGTACAGGGCTAT	2940.53	3355.25	6352.57	5061.63
hsa-miR-125b-5p_R-1	TCCCTGAGACCCTAACTTGTTG	29145.42	29130.87	18317.07	8758.18
hsa-miR-125b-2-3p_L-2	ACAAGTCAGGCTCTTGGGAC	936.67	560.38	443.64	335.75
hsa-miR-125a-5p_R-1	TCCCTGAGACCCTTAACTGTG	142400.31	113799.96	35452.67	40911.92
hsa-miR-125a-3p_R-1	ACAGGTGAGGTTCTTGGGAGC	799.70	659.82	62.97	93.34
hsa-miR-125b-1-3p	ACGGGTTAGGCTCTTGGGAGCT	9.68	24.57	10.34	9.75

149 **Table S3:** Partial data of normalized reads of miRNAs that only detectable in partial samples.

miRNA name	miRNA sequence	Normalized reads			
		cfRNA4	cfRNA3	cellRNA3	cellRNA4
hsa-miR-15a-3p_1ss22AT	CAGGCCATATTGTGCTGCCTCT	0	0	4.70	0
hsa-miR-18b-5p_R-4	TAAGGTGCATCTAGTGCAG	0	0	3.76	0
hsa-miR-23b-5p_L-1R-3	GGTTCTCGCATGCTGA	0	23.40	0	0
hsa-miR-26a-1-3p_R+1	CCTATTCTTGTTACTTGCACGA	0	9.36	4.70	3.48
hsa-miR-29b-1-5p_L-1	CTGGTTTCATATGGTGGTTAGA	0	16.38	0	13.24
hsa-miR-31-3p_R+1	TGCTATGCCAACATAATGCCATC	0	0	12.22	17.41
hsa-miR-33b-5p_R-1	GTGCATTGCTGTTGCATTG	0	14.04	136.29	27.17

hsa-miR-34c-3p	AATCACTAACCACACGGCCAGG	0	5.85	0	0
hsa-mir-96-p3	TGTGTCTCTCCGCTCTGAG	0	8.19	0	3.48
hsa-mir-106a-p3	ACTGCAATGTAAGCACTTCT	0	0	0	9.06
hsa-miR-129-5p	CTTTTTGCGGTCTGGGCTTGC	0	4.09	0	0
hsa-miR-139-5p	TCTACAGTGACGTGTCTCCAGT	0	9.36	21.62	10.45
hsa-miR-146a-3p_L+2R-3	GACCTCTGAAATTCAGTTCTT	0	0	15.04	16.02
hsa-miR-147b-5p	TGGAAACATTTCTGCACAAACT	0	0	9.40	34.13
hsa-miR-147b-3p	GTGTGCGGAAATGCTTCTGCT	0	7.02	59.21	62.69
hsa-miR-150-3p_R-1	CTGGTACAGGCCTGGGGGACA	0	14.04	15.04	9.75
hsa-miR-155-3p	CTCCTACATATTAGCATTAACA	0	0	4.70	4.18
hsa-miR-186-3p_L-2R+1	CCAAAGGTGAATTTTTTGGGA	0	0	0	9.06
hsa-miR-188-5p_R+1	CATCCCTTGCATGGTGGAGGGT	0	0	6.58	0
hsa-miR-190a-3p_L+1	ACTATATATCAAACATATTCCT	0	0	45.12	34.13
hsa-miR-192-3p_R+1	CTGCCAATTCATAGGTCACAGT	0	0	0	4.18
hsa-miR-194-3p	CCAGTGGGGCTGCTGTATCTG	0	0	0	6.97
hsa-miR-196b-3p	TCGACAGCACGACACTGCCTTC	0	0	7.52	0
hsa-miR-205-3p_L-1R+1	ATTTTCAGTGGAGTGAAGTTCA	0	12.87	12.22	0
hsa-miR-219b-5p	AGATGTCCAGCCACAATTCTCG	0	14.04	0	0
hsa-mir-222-p5	AAGGTGTAGGTACCCTCAAT	0	0	7.52	16.02
hsa-miR-301a-5p	GCTCTGACTTTATTGCACTACT	0	0	0	16.02
hsa-miR-301b-3p	CAGTGAATGATATTGTCAAAGC	0	0	4.70	0
hsa-miR-338-3p_R+1	TCCAGCATCAGTGATTTTGTGA	0	0	11.28	79.41
hsa-miR-371b-5p	ACTCAAAAGATGGCGGCACTTT	0	0	0	6.27
hsa-miR-377-3p_R-2	ATCACACAAAGGCAACTTTT	0	0	0	4.88
hsa-miR-378b_R+3_1ss14GT	ACTGGACTTGGAGTCAGAAAGA	0	0	4.70	0
hsa-miR-379-5p	TGGTAGACTATGGAACGTAGG	0	0	0	3.48
hsa-miR-379-3p_1ss22TA	TATGTAACATGGTCCACTAACA	0	7.02	0	0
hsa-miR-382-3p_R+1	AATCATTCACGGACAACACTTT	0	0	0	4.18
hsa-miR-409-3p_L+1	CGAATGTGTCTCGGTGAACCCCT	0	0	0	6.27
hsa-miR-410-3p	AATATAACACAGATGGCCTGT	0	0	0	4.88
hsa-miR-422a_1ss10AG	ACTGGACTTGGGGTCAGAAGGC	0	0	0	3.48
hsa-miR-449c-5p_L-1R+1	AGGCAGTGTATTGCTAGCGGCTGTT	0	0	0	13.24
hsa-miR-450a-1-3p	ATTGGGAACATTTTGCATGTAT	0	0	4.70	4.88
hsa-miR-489-3p_R-1	GTGACATCACATATACGGCAG	0	0	12.22	0
hsa-miR-499a-3p_L+1R-4_1ss5AG	GAACGTCACAGCAAGTCTG	0	7.02	4.70	0
hsa-miR-512-5p_L-1R-1	ACTCAGCCTTGAGGGCACTTT	0	0	9.40	0
hsa-miR-514a-3p	ATTGACACTTCTGTGAGTAGA	0	0	9.87	4.88
hsa-miR-515-3p	GAGTGCCTTCTTTTGGAGCGTT	0	0	0	3.48
hsa-miR-518d-5p_R-1	CTCTAGAGGGAAGCACTTTCT	0	3.01	3.09	0
hsa-miR-520c-3p	AAAGTGCTTCCTTTTAGAGGGT	0	0	15.15	4.18
hsa-miR-518e-3p_R-1	AAAGCGCTTCCTTCAGAGT	0	11.70	24.44	3.48
hsa-miR-518f-5p_R-1	CTCTAGAGGGAAGCACTTTCT	0	3.01	3.09	0
hsa-miR-520b-3p_R+1	AAAGTGCTTCCTTTTAGAGGGT	0	0	15.15	4.18
hsa-miR-519e-5p_R-3	TTCTCCAAAAGGGAGCACT	0	0	4.70	0

hsa-miR-520a-5p_R-2	CTCCAGAGGGAAGTACTTT	0	10.53	26.32	6.97
hsa-miR-520a-3p_R-2	AAAGTGCTTCCTTTGGACT	0	16.38	15.98	22.29
hsa-miR-520d-3p_R+1	AAAGTGCTTCTTTGGTGGGTT	0	0	6.58	0
hsa-miR-539-3p_L-2R+2	CATACAAGGACAATTTCTTTTT	0	15.21	0	0
hsa-miR-545-5p	TCAGTAAATGTTTATTAGATGA	0	0	0	4.18
hsa-miR-548v	AGCTACAGTTACTTTTGCACCA	0	0	10.34	14.63
hsa-mir-548n-p3	CAATTACTTTTGCACCAACC	0	0.88	0	0
hsa-mir-548p-p5_1ss3TG	AAGTAATTGCAGTTTTTGTC	0	0	0	4.18
hsa-mir-548f-1-p5	TTAGGTTGGTGCAAAAGTAAT	0	0.34	0.47	0
hsa-miR-548b-5p_R-1	AAAAGTAATTGTGGTTTTGGC	0	0	0.67	4.88
hsa-miR-548i_R-2	AAAAGTAATTGCGGATTTTG	0	0	0	1.74
hsa-miR-548ah-5p	AAAAGTGATTGCAGTGTTTG	0	0	4.70	0
hsa-miR-548e-5p_L-1R-1	AAAAGCAATCGCGTTTTTG	0	8.19	0	9.06
hsa-mir-548f-4-p5	TTAGGTTGGTGCAAAAGTAAT	0	0.34	0.47	0

150 **Table S4:** Previous reported microRNAs dysregulated in breast cancer were listed and
151 compared with our miRNA basal abundance data. The inferential expression changes of
152 miRNAs in early breast cancer (BC) are analyzed according to reports and our miRNA
153 sequencing data.

miRNA name	miRNA sequence	normalized reads				Reported Expression in Breast cancer (BC)		Inferential Expression in early BC
		cfRNA4	cfRNA3	cellRNA3	cellRNA4	Results	Opposite Results	
hsa-let-7e-3p_R-2	CTATACGGCCTCCTAGCTTT	90	109	91	89	upregulated	downregulated	upregulated
hsa-let-7a-5p	TGAGGTAGTAGGTTGTATAGTT	69874	57909	83431	129086	upregulated	downregulated	downregulated
hsa-let-7c-5p	TGAGGTAGTAGGTTGTATGGTT	2273	2555	2570	2421	downregulated		downregulated
hsa-let-7c-3p_R-1	CTGTACAACCTTCTAGCTTTC	7	0	6	0			
hsa-let-7b-5p	TGAGGTAGTAGGTTGTGTGGTT	82701	116853	81208	48587	downregulated		downregulated
hsa-let-7b-3p_R-1_1ss21CT	CTATACAACCTACTGCCTTCT	3748	2485	1170	602			
hsa-miR-10b-5p_R-1	TACCCTGTAGAACCGAATTTGT	183	123	70	88	upregulated		
hsa-miR-16-5p	TAGCAGCACGTAAATATTGGCG	7284	7890	12245	13131			
hsa-miR-16-1-3p	CCAGTATTAAGTGTCTGCTGA	0	0	23	69	upregulated		upregulated
hsa-miR-16-2-3p_L+1R-1	ACCAATATTACTGTGCTGCTTT	221	392	564	694			upregulated
hsa-miR-18a-5p	TAAGGTGCATCTAGTGCAGATAG	82	91	794	714	upregulated	downregulated	upregulated
hsa-miR-18a-3p_R-2	ACTGCCCTAAGTGCTCCTTCT	140	171	49	44			upregulated
hsa-miR-19a-3p	TGTGCAAATCTATGCAAACTGA	196	291	1394	1409	downregulated		
hsa-miR-20a-5p	TAAAGTGCTTATAGTGCAGGTAG	4964	3870	31801	31944	downregulated		downregulated
hsa-miR-20a-3p	ACTGCATTATGAGCACTAAAG	17	25	155	114			
hsa-miR-20b-5p	CAAAGTGCTCATAGTGCAGGTAG	0	0	36	54	upregulated		upregulated
hsa-miR-21-5p	TAGCTTATCAGACTGATGTTGA	167598	225677	1216907	773412	upregulated		
hsa-miR-21-3p_R+1	CAACACCAGTCGATGGGCTGTC	123	188	2935	2382			upregulated
hsa-miR-25-5p_R+2	AGGCGGAGACTTGGCAATTGCT	35	42	28	43	upregulated		upregulated
hsa-miR-25-3p	CATTGCCTTGTCTCGGCTGTA	24937	34050	24624	23471			
hsa-miR-29c-5p_R-1	TGACCGATTCTCCTGGTGTT	672	542	1325	1091	upregulated		upregulated

hsa-miR-29c-3p_R-1	TAGCACCATTTGAAATCGGTT	2999	5933	15753	7487			
hsa-miR-30a-5p	TGTAAACATCCTCGACTGGAAG	143946	97103	178925	183651	downregulated		downregulated
hsa-miR-30a-3p_1ss22CT	CTTTCAGTCGGATGTTTGCACT	3262	3163	3579	5699			
hsa-miR-30b-5p	TGTAAACATCCTACACTCAGCT	383303	264659	372314	346935	downregulated		downregulated
hsa-miR-34a-5p_R+1	TGGCAGTGTCTTAGCTGGTTGTT	95	113	1263	1481		upregulated	
hsa-miR-34a-3p_R+1	CAATCAGCAAGTATACTGCCTT	0	0	0	8	downregulated		upregulated
hsa-miR-34c-5p	AGGCAGTGTAGTTAGCTGATTGC	0	0	6	42			
hsa-miR-34c-3p	AATCACTAACCCACACGGCCAGG	0	6	0	0			
hsa-miR-92a-1-5p	AGGTTGGGATCGGTTGCAATGCT	25	66	25	35			upregulated
hsa-miR-92a-3p	TATTGCACTGTCCCGCCTGT	71133	52695	36108	24183	downregulated	upregulated	downregulated
hsa-miR-92a-2-5p	GGGTGGGATTGTTGCATTAC	7	0	0	0			upregulated
hsa-miR-103a-2-5p	AGCTTCTTACAGTGCTGCCTTG	36	7	53	43			upregulated
hsa-miR-103a-3p	AGCAGCATTGTACAGGGCTATGA	38638	22562	35255	76850		upregulated	
hsa-miR-122-5p_R-1	TGGAGTGTGACAATGGTGT	0	156	0	29	upregulated		upregulated
hsa-miR-125b-5p_R-1	TCCCTGAGACCCTAACTTGTG	29145	29131	18317	8758			
hsa-miR-125b-2-3p_L-2	ACAAGTCAGGCTCTTGGGAC	937	560	444	336	upregulated		upregulated
hsa-miR-125b-1-3p	ACGGGTTAGGCTCTTGGGAGCT	10	25	10	10			upregulated
hsa-miR-126-5p_R+1	CATTATTACTTTTGGTACGCGA	0	0	7	23			
hsa-miR-126-3p	TCGTACCGTGAGTAATAATGCG	48	76	70	193	downregulated	upregulated	upregulated
hsa-miR-133a-3p_L-1R+1	TTGGTCCCTTCAACCAGCTGT	8	4	7	2	downregulated	upregulated	upregulated
hsa-miR-139-5p	TCTACAGTGCACGTGTCTCCAGT	0	9	22	10	downregulated		
hsa-miR-141-3p_R-1	TAACACTGTCTGGTAAAGATG	45058	66296	119600	65598	downregulated		downregulated
hsa-miR-143-5p_1ss22TA	GGTGCAGTGTCATCTCTGGA	39	50	207	151			
hsa-miR-143-3p_R+1	TGAGATGAAGCACTGTAGCTCT	2268	4251	12322	18655	downregulated		downregulated
hsa-miR-145-5p	GTCCAGTTTCCCAGGAATCCCT	138	168	112	367			upregulated
hsa-miR-145-3p	GGATTCTGGAAATACTGTCT	8	22	58	103		upregulated	upregulated
hsa-miR-148a-3p	TCAGTGCACACTACAGAACTTGT	386987	588485	918411	876058	downregulated		downregulated
hsa-miR-148b-5p_L+1R-1	GAAGTCTGTATACACTCAGG	870	649	598	1076			upregulated
hsa-miR-148b-3p_R-1	TCAGTGCATCACAGAACTTGT	20149	21809	40891	61755		downregulated	downregulated
hsa-miR-155-5p_R-1	TTAATGCTAATCGTGATAGGGGT	39	81	1697	1390		upregulated	upregulated
hsa-miR-155-3p	CTCCTACATATTAGCATTAAACA	0	0	5	4			upregulated
hsa-miR-181a-5p_R-1	AACATTCAACGCTGTCGGTGAG	24468	26168	17747	14616	downregulated	upregulated	downregulated
hsa-miR-182-5p_R-2	TTGGCAATGGTAGAACTCACA	29344	16727	23348	40420		upregulated	
hsa-miR-182-3p_L+2R-1	GGTGGTTCTAGACTTGCCAACT	62	28	76	79			upregulated
hsa-miR-191-5p_R-1	CAACGGAATCCCAAAGCAGCT	221594	117554	51911	87488		upregulated	
hsa-miR-191-3p_R-3	GCTGCGCTTGGATTTCGTC	30	14	7	0			upregulated
hsa-miR-192-5p	CTGACCTATGAATTGACAGCC	671	503	1714	2494		upregulated	upregulated
hsa-miR-192-3p_R+1	CTGCCAATCCATAGGTCACAGT	0	0	0	4			upregulated
hsa-miR-195-5p_R+1	TAGCAGCACAGAAATATTGGCA	1000	774	3561	1953		upregulated	upregulated
hsa-miR-195-3p_R-2	CCAATATTGGCTGTGCTGCT	50	42	43	40			upregulated
hsa-miR-199a-5p_R-1	CCCAGTGTTACAGACTACCTGTT	22	23	20	19		upregulated	upregulated
hsa-miR-199a-3p_R-1	ACAGTAGTCTGCACATTGGTT	642	716	717	1711			upregulated
hsa-miR-200c-5p	CGTCTTACCAGCAGTGTGG	18	6	65	25		upregulated	upregulated
hsa-miR-200c-3p	TAATACTGCCGGTAATGATGGA	764438	473643	254207	253470	upregulated	downregulated	downregulated

hsa-miR-200b-5p	CATCTTACTGGGCAGCATTGGA	5836	6310	2957	2449			upregulated
hsa-miR-200b-3p	TAATACTGCCTGGTAATGATGA	354432	251845	266377	258231		upregulated	downregulated
hsa-miR-200a-5p	CATCTTACCGGACAGTGCTGGA	2598	1846	1627	2221			upregulated
hsa-miR-200a-3p_R+1	TAACACTGTCTGGTAACGATGTT	345502	350091	385973	393857		upregulated	downregulated
hsa-miR-202		0	0	0	0		upregulated	upregulated
hsa-miR-203a-3p	GTGAAATGTTTAGGACCACTAG	797	2044	3199	1505		upregulated	upregulated
hsa-miR-205-5p	TCCTTCAATCCACCGGAGTCTG	15626	21105	19760	8789			downregulated
hsa-miR-205-3p_L-1R+1	ATTTCACTGGAGTGAAGTTCA	0	13	12	0		downregulated	
hsa-miR-210-5p_R-1	AGCCCCTGCCACCGCACACT	58	132	34	20		upregulated	upregulated
hsa-miR-210-3p	CTGTGCGTGTGACAGCGGCTGA	515	1797	4409	1792			upregulated
hsa-miR-215-5p_R+1	ATGACCTATGAATTGACAGACA	0	0	18	26		upregulated	upregulated
hsa-miR-222-p5	AAGGTGTAGGTACCCTCAAT	0	0	8	16		upregulated	upregulated
hsa-miR-222-3p_R+2	AGCTACATCTGGCTACTGGGTCT	679	783	2627	2580			upregulated
hsa-miR-299		0	0	0	0		upregulated	upregulated
hsa-miR-320d_R-1	AAAAGCTGGGTTGAGAGG	186	139	62	18		upregulated	upregulated
hsa-miR-320b_R-2	AAAAGCTGGGTTGAGAGGGC	1206	817	276	252			upregulated
hsa-miR-324-5p_R-1	CGCATCCCTAGGGCATTGGT	274	370	614	783		upregulated	upregulated
hsa-miR-324-3p_L-3R+1	ACTGCCCCAGGTGCTGTGGT	313	881	288	182			upregulated
hsa-miR-335-5p_R-2	TCAAGAGCAATAACGAAAAAT	27462	21489	35648	31513		downregulated	downregulated
hsa-miR-342-5p_R+1	AGGGGTGCTATCTGTGATTGAG	228	204	114	54		downregulated	
hsa-miR-342-3p	TCTCACACAGAAATCGCACCCGT	6948	7060	4489	5121			downregulated
hsa-miR-365b-5p_L+1R-2	GAGGGACTTTCAGGGGCAGCT	33	274	5	0			
hsa-miR-365b-3p	TAATGCCCTAAAAATCCTTAT	12813	10465	6151	8229		downregulated	downregulated
hsa-miR-365a-5p_R-3	AGGGACTTTTGGGGGCAGAT	36	118	0	14			
hsa-miR-373		0	0	0	0		upregulated	upregulated
hsa-miR-375-5p	GCGACGAGCCCTCGCACAAACC	19	12	0	0		upregulated	downregulated
hsa-miR-375-3p	TTTGTTCGTTCGGCTCGCGTGA	170762	305772	53533	42830			downregulated
hsa-miR-376c		0	0	0	0		upregulated	upregulated
hsa-miR-382-5p	GAAGTTGTTTCGGTGGATTTCG	17	0	0	10		upregulated	upregulated
hsa-miR-382-3p_R+1	AATCAATTCACGGACAACACTTT	0	0	0	4			upregulated
hsa-miR-409-5p_R-1	AGGTTACCCGAGCAACTTTGCA	14	0	0	23		upregulated	upregulated
hsa-miR-409-3p_L+1	CGAATGTGCTCGGTGAACCCCT	0	0	0	6			upregulated
hsa-miR-425-5p	AATGACACGATCACTCCCGTTGA	7590	6681	2323	3182		upregulated	
hsa-miR-425-3p_L+1R-1	CATCGGGAATGTCGTGTCGCC	1157	1007	629	752			
hsa-miR-451a_R-1	AAACCGTTACCATTACTGAGT	4331	6033	3030	806		upregulated	
hsa-miR-497-5p	CAGCAGCACACTGTGGTTTGT	170	598	1645	879		downregulated	
hsa-miR-523-3p_L-1R-2	AACGCGCTTCCCTATAGAGG	11	19	8	0		upregulated	upregulated
hsa-miR-526b-5p_R-1	CTCTTGAGGGAAGCACTTTCTG	7	25	32	14		upregulated	upregulated
hsa-miR-625-5p_R-1	AGGGGGAAAAGTTCTATAGTC	7	0	15	12		upregulated	upregulated
hsa-miR-625-3p	GACTATAGAACTTCCCCCTCA	162	58	46	28			upregulated
hsa-miR-628-5p_R-1	ATGCTGACATATTTACTAGAG	26	48	233	223		upregulated	upregulated
hsa-miR-628-3p_R+1	TCTAGTAAGAGTGGCAGTCGAA	137	171	106	112			upregulated
hsa-miR-652-3p_R+1	AATGGCGCCACTAGGGTTGTGA	6220	4833	2953	3025		downregulated	
hsa-miR-744-5p	TGCGGGGCTAGGGCTAACAGCA	820	462	480	1050		downregulated	

hsa-miR-744-3p_R-1_1ss21CT	CTGTTGCCACTAACCTCAACT	15	14	6	0		
hsa-miR-801		0	0	0	0	upregulated	upregulated
hsa-miR-1287-5p_R-2	TGCTGGATCAGTGGTTCGAG	203	455	418	260	downregulated	
hsa-miR-1287-3p	CTCTAGCCACAGATGCAGTGAT	0	16	5	0		

154 References

- 155 1. Martin M: **Cutadapt removes adapter sequences from high-throughput sequencing reads.** *2011*
156 2011, **17**(1):3.
- 157 2. Chen H, Smith AD, Chen T: **WALT: fast and accurate read mapping for bisulfite sequencing.**
158 *Bioinformatics (Oxford, England)* 2016, **32**(22):3507-3509.
- 159 3. Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R: **The**
160 **Sequence Alignment/Map format and SAMtools.** *Bioinformatics (Oxford, England)* 2009,
161 **25**(16):2078-2079.
- 162 4. Song Q, Decato B, Hong EE, Zhou M, Fang F, Qu J, Garvin T, Kessler M, Zhou J, Smith AD: **A**
163 **reference methylome database and analysis pipeline to facilitate integrative and comparative**
164 **epigenomics.** *PloS one* 2013, **8**(12):e81148.
- 165 5. Akalin A, Kormaksson M, Li S, Garrett-Bakelman FE, Figueroa ME, Melnick A, Mason CE:
166 **methylKit: a comprehensive R package for the analysis of genome-wide DNA methylation**
167 **profiles.** *Genome biology* 2012, **13**(10):R87.
- 168 6. Chiu RW, Akolekar R, Zheng YW, Leung TY, Sun H, Chan KC, Lun FM, Go AT, Lau ET, To WW
169 *et al*: **Non-invasive prenatal assessment of trisomy 21 by multiplexed maternal plasma DNA**
170 **sequencing: large scale validity study.** *BMJ (Clinical research ed)* 2011, **342**:c7401.
- 171 7. Yoon S, Xuan Z, Makarov V, Ye K, Sebat J: **Sensitive and accurate detection of copy number**
172 **variants using read depth of coverage.** *Genome research* 2009, **19**(9):1586-1592.
- 173 8. Ginestet C: **ggplot2: Elegant Graphics for Data Analysis by H. Wickham.** *Journal of the Royal*
174 *Statistical Society* 2011, **174**(1):245-246.
- 175 9. Nepomnyashchaya YN, Artemov AV, Roumiantsev SA, Roumyantsev AG, Zhavoronkov A:
176 **Non-invasive prenatal diagnostics of aneuploidy using next-generation DNA sequencing**
177 **technologies, and clinical considerations.** *Clinical chemistry and laboratory medicine* 2013,
178 **51**(6):1141-1154.