Enzyme	Amino acid length	Gene	Chromosomal localization	Substrate
ST8Sia I	356	ST8SIA1	12p12.1-p11.2	GM3
ST8Sia II	375	ST8SIA2	15q26	N-glycan on NCAM
ST8Sia III	380	ST8SIA3	18q21	NeuAcα2,3Galβ1,4GlcNAc-
ST8Sia IV	359	ST8SIA4	5q21	N-glycan on NCAM
ST8Sia V	376	ST8SIA5	18q12.1-q12.3	GM1b, GT1b, GD1a, GD3
ST8Sia VI	398	ST8SIA6	10p13	NeuAcα2,3(6)Gal-
ST6GalNAc I	600	ST6GALNAC1	17q25	GalNAcα1,O-Ser/Thr
ST6GalNAc II	374	ST6GALNAC2	17q25	Galβ1,3GalNAcα1,O-Ser/Thr
ST6GalNAc III	305	ST6GALNAC3	1p31.1-p31.2	NeuAcα2,3Galβ1,3GalNAc-(Lipid)*
ST6GalNAc IV	302	ST6GALNAC4	9q34	NeuAcα2,3Galβ1,3GalNAc-(Protein)*
ST6GalNAc V	336	ST6GALNAC5	1p31.1	GM1b
ST6GalNAc VI	299	ST6GALNAC6	9q34	GM1b, GT1b, GD1a
ST6Gal I	406	ST6GAL1	3q27-q28	Galβ1,4GlcNAc-
ST6Gal II	529	ST6GAL2	2q11.2-q12.1	Galβ1,4GlcNAc-
ST3Gal I	340	ST3GAL1	8q24.2	Galβ1,3GalNAc-(Protein)*
ST3Gal II	350	ST3GAL2	16q21–22.3	Galβ1,3GalNAc-(Lipid)*
ST3Gal III	375	ST3GAL3	1pter-p32.3	Galβ1,3(4)GlcNAc-
ST3Gal IV	329	ST3GAL4	11q23-q24	Galβ1,4(3)GlcNAc-
ST3Gal V	362	ST3GAL5	2p24.3-p24.1	Lac-Cer
ST3Gal VI	331	ST3GAL6	3p21.1-q13.2	Galβ1,4GlcNAc-

Table S1. Human sialyltransferases (edited from Takashima et al., 2002 [8]).

* preferential but not specific substrate. Lac-Cer (lactosylceramide), GM (monosialoganglioside), GD (disialoganglioside), GT (trisialogangliosid).



Figure S1. Gating strategy for flow cytometry. (**a**) Forward scatter (FSC) area against width was used for doublet discrimination whereas FSC versus sideward scatter (SSC) was used for removal of debris. 10,000 events were recorded in the P1 gate and analyzed in the FL1 channel. A representative histogram for Sambucus Nigra Lectin (SNA) is shown as an example. (**b**) Representative histograms from the staining with SNA, Maackia Amurensis Lectin II (MAL II) or GD3 are shown in black. The red histogram represents the streptavidin (for SNA and MAL II) or secondary antibody control (for GD3).



Figure S2. Sialyltransferase expression in NK cells. RNA was isolated from the human NK cell lines NK-92, NKL, KHYG-1 and primary NK cell. cDNA was synthesized and PCR reactions were performed with primer pairs for all 20 human sialyltransferases. (a) Representative agarose gels are shown from 3 independent experiments performed with cDNA derived from the NK cell lines. (b) Gels from 3 independent experiments performed with cDNA derived from 3 healthy blood donors are shown. For product sizes see Table 1.



Figure S3. Expression of *IFN-* γ in NK-92 cells after activation with IL-2. NK-92 cells were incubated without IL-2 for 24 h. Afterwards, cells were either left untreated (control) or treated with 1000 U/ml IL-2 for 4 h. cDNA was synthesized and quantitative real-time PCR reactions were performed. Data were normalized to Beta-2 microglobulin (*B2M*) expression. Graphs show average mean ± SD of 4 independent experiments.