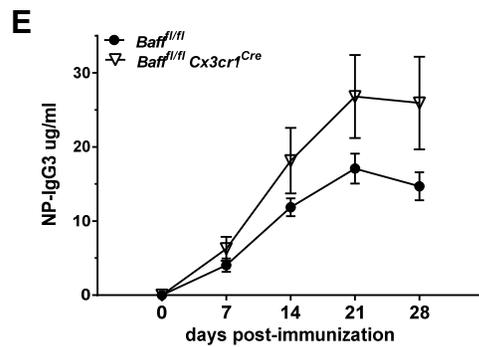
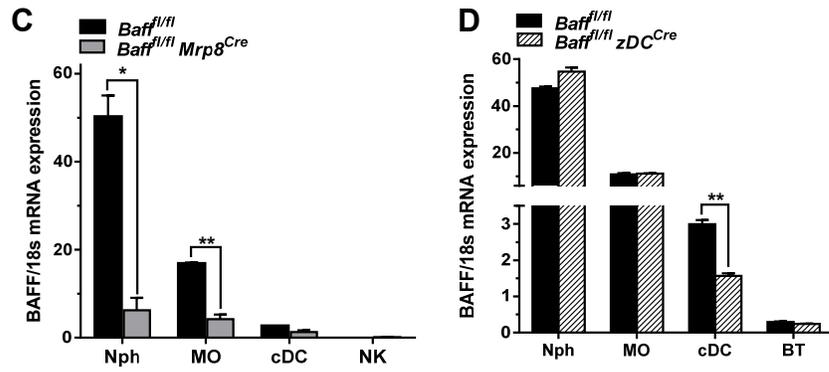
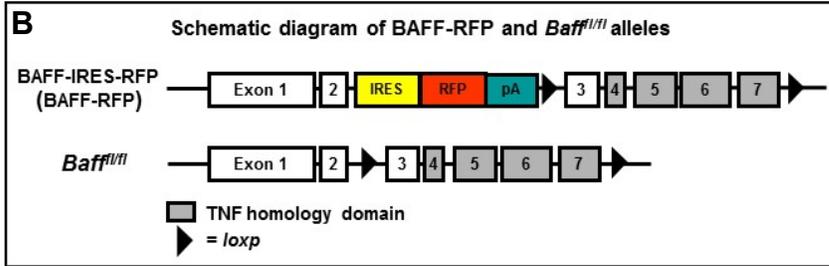
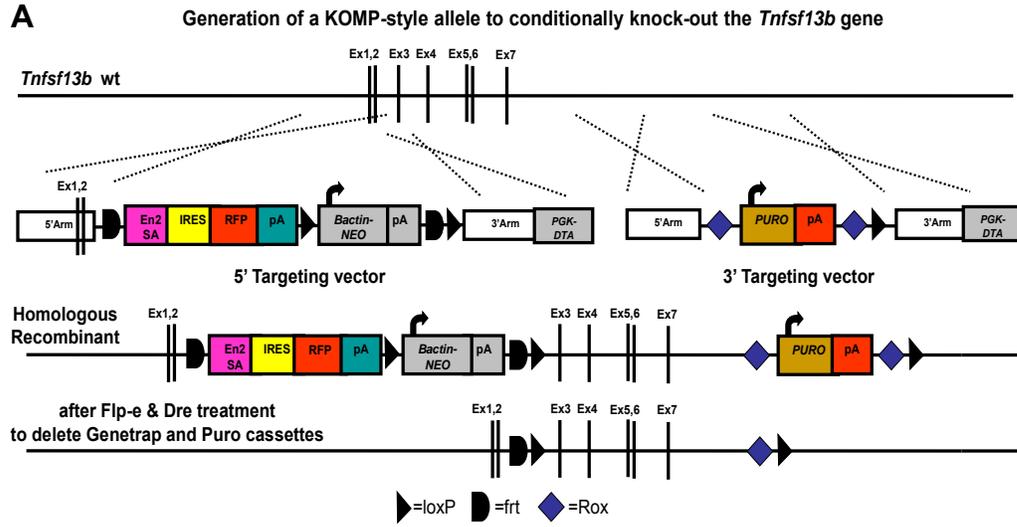


Supplemental Fig. S1



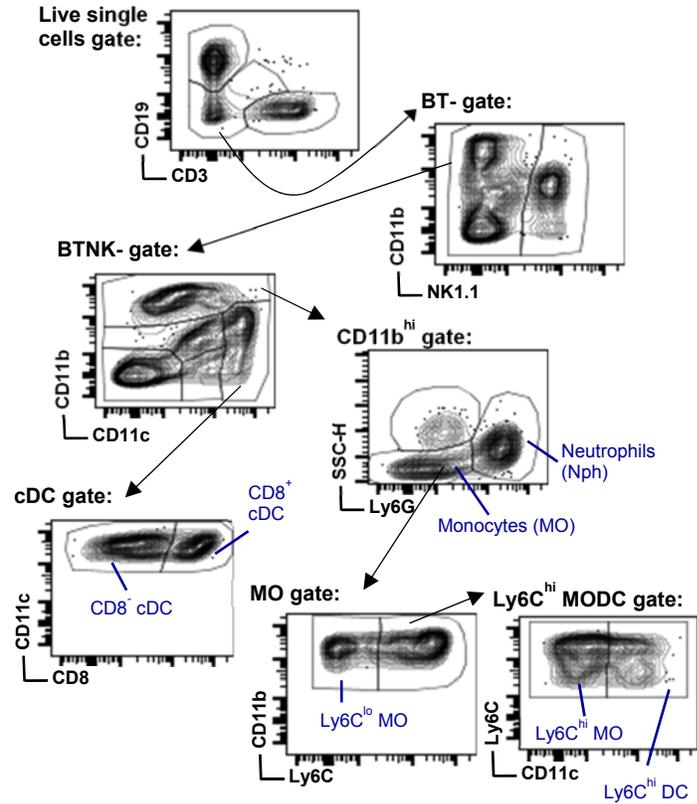
Supplemental Figure S1. *Tnfsf13b* (BAFF) reporter mice expressing RFP, *Baff^{fl/fl}* mice, and verification of conditional KO mice. A. Schematic of the two-construct approach used to generate a KOMP-style allele to conditionally knock-out the *Tnfsf13b/Baff* gene. The *Tnfsf13b-5'* construct included a Genetrap Cassette expressing the TagRFPT (red fluorescence protein (RFP), under the control of an internal ribosome entry site (IRES) and flanked by *frt* sites for selective deletion, a loxP site and arms of homology of ~5kb each. The *Tnfsf13b-5'* construct was inserted between exons 2 and 3 of *Tnfsf13b* gene. Neomycin (*Neo*) and Diphtheria Toxin A (DTA) elements for positive and negative selection are indicated. The *Tnfsf13b-3'* construct was inserted after the last exon 7 in the intergenic region, and included the other loxP site, arms of homology (~5kb each), a Puromycin (*Puro*) and a DTA element. The *Tnfsf13b-3'* construct was electroporated first into ES cells, and a positive clone electroporated with the *Tnfsf13b-5'* construct. Homologous recombination of the two constructs generated the heterozygous BAFF-IRES-RFP (BAFF-RFP) reporter mice where *Tnfsf13b* is functionally knocked-out and the reporter expresses the IRES-RFP under *Tnfsf13b* promoter and the loxP sites between exon 3 and 7. The germline BAFF-RFP^{+/-} mice were bred with B6N(B6J)-Tg(CAG-Flpo) mice to excise the Genetrap Cassette and *Neo*, and then with B6;129-Tg(CAG-dre) mice to remove *Puro*. The Flpo and Dre recombinases were bred out to generate *Baff^{fl/fl}* mice expressing wild type *Baff* with floxed exons 3-7. **B.** Schematic diagram of BAFF-IRES-RFP (BAFF-RFP) and *Baff^{fl/fl}* alleles. **C, D,** Splenocytes harvested from *Baff^{fl/fl}* (**C, D**), *Baff^{fl/fl} Mrp8^{Cre}* mice (**C**), or *Baff^{fl/fl} zDC^{Cre}* mice (**D**), enriched with CD11b⁺ CD11c⁺ magnetic bead positive selection, sorted on a FACS Aria cell sorter and assessed for *Baff* mRNA by qPCR. **C, D,** Data show means ± SEM of relative *Baff* mRNA from one experiment pooling cells from six mice/group (**C**), or from one representative of two experiments pooling cells from six-seven mice/group (**D**). Selective depletion of BAFF in Nphs from *Baff^{fl/fl} Mrp8^{Cre}* mice was confirmed with sorted BM cells. **C, D,** Sorted cells from *Mrp8^{Cre}* mice and *zDC^{Cre}* mice showed similar *Baff* mRNA levels as *Baff^{fl/fl}* mice (not shown). **C, D,** Statistics were determined with two-tailed unpaired Student's *t*-test, * *p*<0.05, ** *p*<0.01. **E,** *Baff^{fl/fl}* mice crossed with *Cx3cr1^{Cre}* mice (1) to generate *Baff^{fl/fl} Cx3cr1^{Cre}* mice. *Baff* mRNA in sorted splenic MOs from *Baff^{fl/fl} Cx3cr1^{Cre}* mice was lower than in control MOs, confirming that BAFF was

knocked down just in the appropriate subset. *Baff^{fl/fl}* mice and *Baff^{fl/fl} Cx3cr1^{Cre}* mice were immunized i.p. with 20 µg of NP-Ficoll, bled at the indicated time points, NP-specific IgG3 was analyzed by ELISA. Data show means ± SEM of NP-IgG3 and are from one experiment; *Baff^{fl/fl}*, N=8; *Baff^{fl/fl} Cx3cr1^{Cre}*, N=6. Statistics were performed with 2-way ANOVA with Tukey's multiple comparison test.

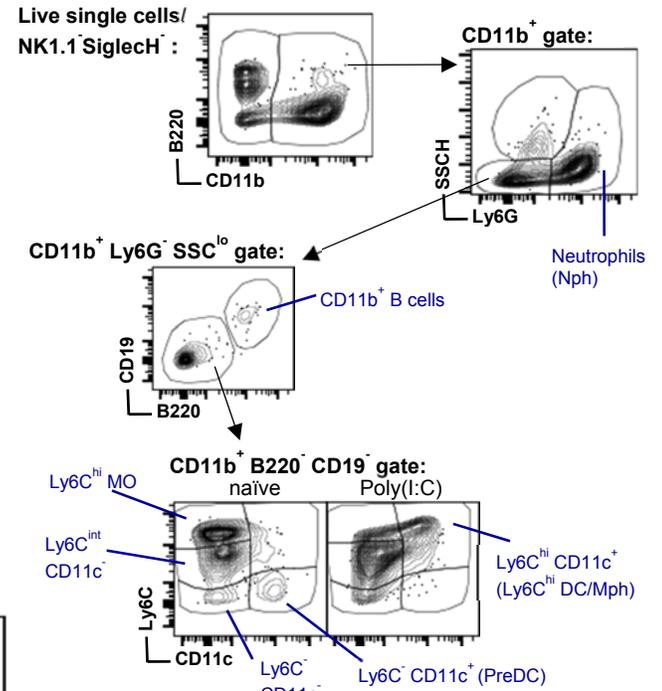
1. Yona, S., K.-W. Kim, Y. Wolf, A. Mildner, D. Varol, M. Breker, D. Strauss-Ayali, S. Viukov, M. Guilliams, A. Misharin, D. A. Hume, H. Perlman, B. Malissen, E. Zelzer, and S. Jung. 2013. Fate Mapping Reveals Origins and Dynamics of Monocytes and Tissue Macrophages under Homeostasis. *Immunity* 38: 79–91.

Supplemental Fig. S2

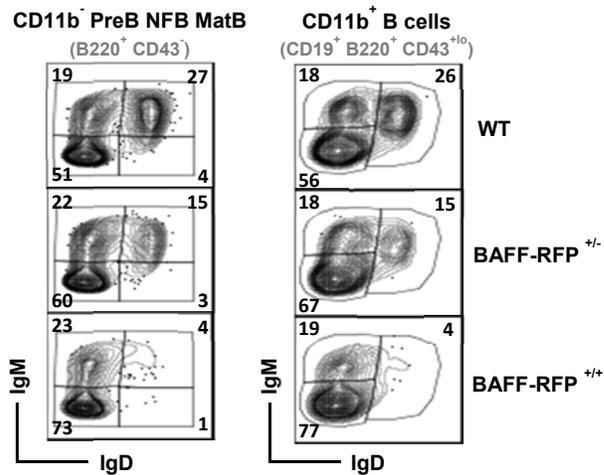
A Gating strategy for myeloid cell populations in Spleen



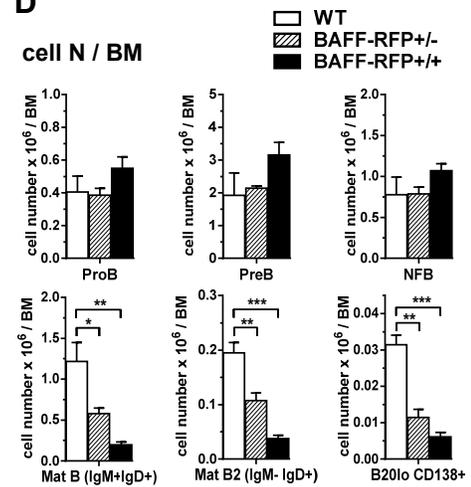
B Gating strategy for myeloid cell populations in BM



C BM

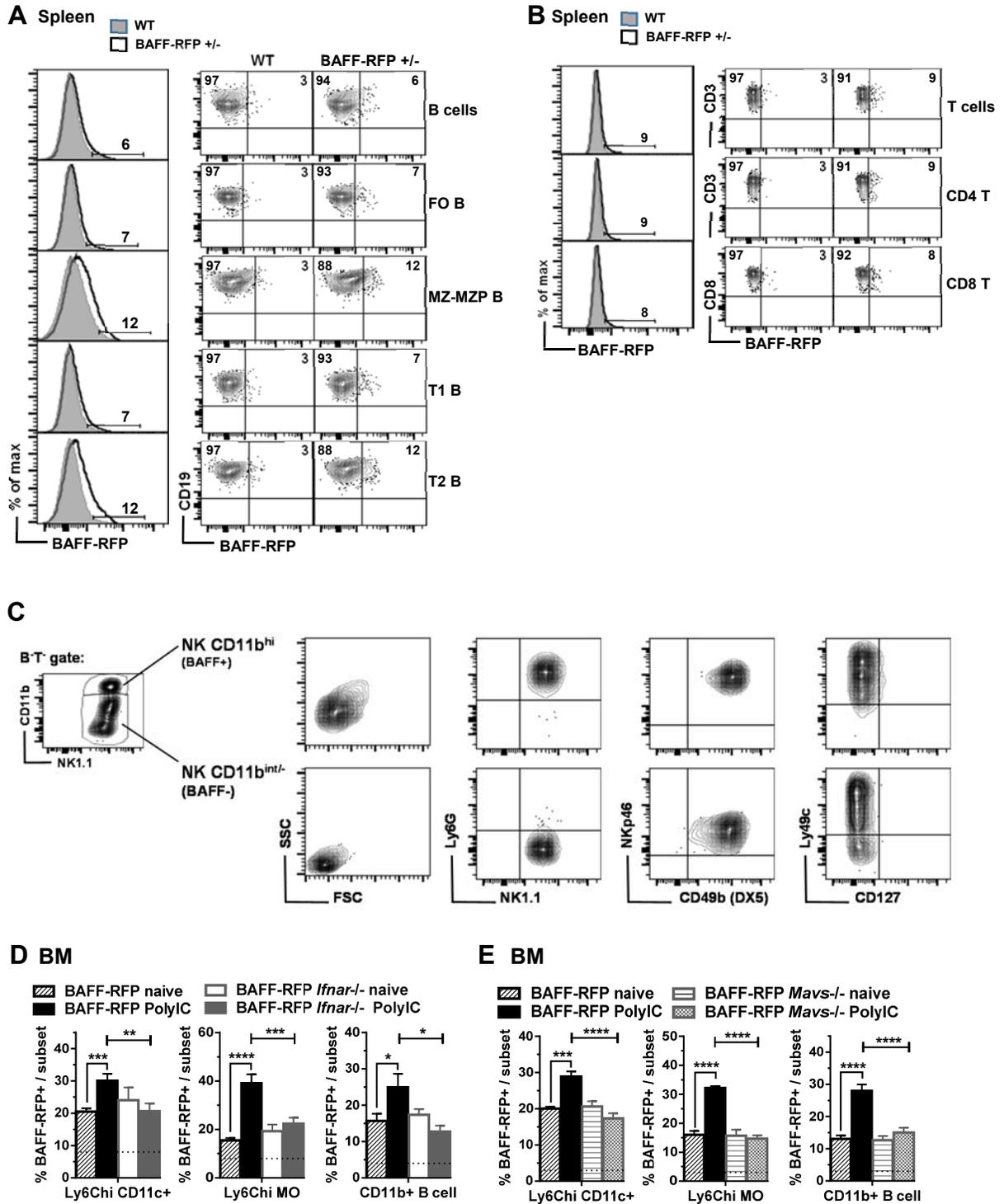


D



Supplemental Figure S2. Analyses of myeloid cells and B cells in spleens and BMs from WT, BAFF-RFP^{+/-} and BAFF-RFP^{+/+} mice. Gating strategy to identify myeloid cell populations in WT splenic (A) and BM cells (B). Debris, doublets and nonviable cells were excluded. A, splenic B cells (CD19⁺CD3⁻) and T cells (CD19⁻CD3⁺) were defined first; in non-B cells and non-T cells (CD19⁻CD3⁻ gate) populations were defined as follows: **NK cells**, NK1.1⁺CD11b^{+/-}; neutrophils (**Nphs**), CD11b^{hi}Ly6G^{hi}Ly6C^{int}SSC^{int}NK1.1⁻; Ly6C^{hi} monocytes (**Ly6C^{hi} MOs**), CD11b^{hi}Ly6C^{hi}CD11c⁻SSC⁻Ly6G⁻NK1.1⁻; Ly6C^{hi} dendritic cells (**Ly6C^{hi} DCs**), CD11b^{hi}Ly6C^{hi}CD11c^{hi}SSC⁻Ly6G⁻NK1.1⁻; **Ly6C^{lo} MOs**, CD11b^{int}CD11c⁻Ly6C^{lo}SSC⁻Ly6G⁻NK1.1⁻; plasmacytoid DCs (pDCs), SiglecH⁺CD11b⁻CD11c^{lo}B220⁺Ly6G⁻NK1.1⁻; **CD8⁺ cDCs**, CD11c^{hi}CD8⁺B220⁻Ly6G⁻NK1.1⁻; **CD8⁻ cDCs**, CD11c^{hi}CD8⁻B220⁻Ly6G⁻NK1.1⁻. B, For the CD11b⁺B220⁻CD19⁻ gate, BM cells are shown from mice treated or not for 24 h with Poly(I:C). BM NK cells (NK1.1⁺SiglecH⁻) and pDCs (NK1.1⁻SiglecH⁺) were defined and then gated out. In the NK1.1⁻SiglecH⁻ gate CD11b⁺ gated myeloid populations were defined as follows: neutrophils (**Nphs**), CD11b⁺Ly6G^{hi}Ly6C^{int}SSC^{int}; **CD11b⁺ B cells**, B220⁺CD19⁺CD11b⁺Ly6G⁻SSC⁻. In the **CD11b⁺B220⁻CD19⁻Ly6G⁻SSC⁻** gate we identified 5 populations of myeloid cells and precursors defined as follows (See also **Table S1**): **CD11b⁺ myeloid precursors**, Ly6C⁻CD11c⁻CD115⁻CX3CR1⁻CCR2⁻MHCII⁻; **Nph precursors**, Ly6C^{int}CD11c⁻CD115⁻CX3CR1⁻CCR2⁻MHCII⁻; **Ly6C^{hi} MOs**, CD11c⁻CD115⁺CX3CR1⁺CCR2^{hi}MHCII⁻; **preDCs**, CD11c⁺Ly6C⁻CD115⁺CX3CR1^{hi}CCR2⁺MHCII⁺. After treatment with Poly(I:C) a BM Ly6C^{hi} DC/Mph precursor (CD11c⁺CD115^{hi}CX3CR1^{hi}CCR2^{hi}MHCII⁺) was substantially upregulated. In the NK1.1⁻SiglecH⁻CD11b⁻ gate, B cell precursors were defined as described in Methods. C, D, B cell populations from naïve WT mice, BAFF-RFP^{+/-} mice and BAFF-RFP^{+/+} mice. C, representative plots of B220⁺CD43⁻ (PreB NFB MatB) cells include preB cells, NF B cells, and mature B cells (*left panels*), and CD11b⁺ B cells (*right panels*). D graphs show means ± SEM of cell numbers. Data are from one of two independent experiments performed with three mice per group. Statistics were performed by one-way ANOVA corrected with Holm-Sidak method for multiple comparisons; * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

Supplemental Fig. S3



Supplemental Figure S3. BAFF-RFP in B cell and T cell subsets, BAFF⁺ CD11b^{hi} NK cell phenotypes and requirement for MAVS for BAFF up-regulation in BM myeloid and B cell subsets by Poly(I:C). **A** and **B**, BAFF-RFP expression detected by flow cytometry in splenic B cell subsets (**A**) and T cell subsets (**B**) from naïve BAFF-RFP^{+/-} mice. RFP background control in WT (C57BL/6) mice is also shown. **A** and **B**, BAFF-RFP levels are shown as histograms on the left, BAFF-RFP^{+/-} mice (*black empty histograms*), WT mice (*grey filled histograms*); and on the right as dot plots. Data show one representative of more than three experiments. For gating strategy of B cell subsets and T cell subsets see Methods. **C**. Phenotype of NK CD11b^{int/-} cells and NK CD11b^{hi} cells upregulated in spleen after Poly(I:C) i.p. injection. NK cell subset phenotypes: CD11b^{int/-} NK cells, NK1.1⁺Ly6G⁻CD49b⁺NKp46^{lo}Ly-49C^{+/-}CD127⁻; CD11b^{hi} NK cells, NK1.1⁺Ly6G^{hi}CD49b⁺NKp46^{hi}Ly-49C^{hi}CD127⁻. **D** and **E**, BAFF-RFP mice and BAFF-RFP *Ifnar*^{-/-} mice (**D**) and BAFF-RFP *Mavs*^{-/-} mice (**E**) were left untreated or treated i.p. with 200µg/mouse Poly(I:C) for 24 hrs. **D** and **E**, bar graphs show means ± SEM of % of BAFF-RFP⁺ BM cells and summarize data from two independent experiments (N=6-7). Dotted lines show RFP⁺ background in WT and *Ifnar*^{-/-} (**D**) or *Mavs*^{-/-} (**E**) mice that were included as controls in each experiment. Statistics were performed by one-way ANOVA corrected with Holm-Sidak method for multiple comparisons; * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$.

Supplemental Table S1. Phenotype of myeloid precursors in BM

Cell Phenotype	Cell Name	Cell marker					
		CD115	CX3CR1	CCR2	Ly6G	MHCII	BAFF-RFP
Ly6C ⁻ CD11c ⁻	CD11b ⁺ myeloid precursor	-	-	-	-	-	++
Ly6C ^{int} CD11c ⁻	Nph prec	-	-	-	+	-	++
Ly6C ^{int} CD11c ⁻	Nph	-	-	-	+++	-	+++
Ly6C ^{hi} CD11c ⁻	Ly6C ^{hi} MO	++	++	++++	-	-	+/-
Ly6C ^{hi} CD11c ⁺	Ly6C ^{hi} DC/Mph	+++	+++	+++	+	+	++
Ly6C ⁻ CD11c ⁺	preDC	++	+++	+	-	+	+

+ and - indicates MFI levels of cell markers