

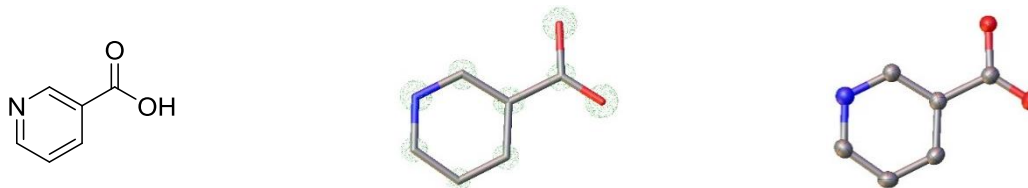
**Chem, Volume 6**

**Supplemental Information**

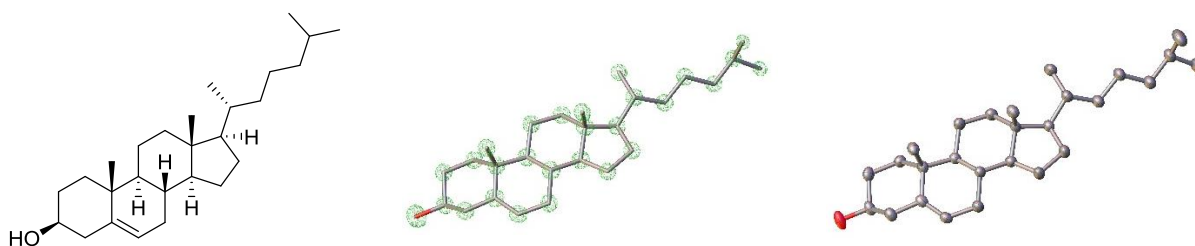
**Encapsulated Nanodroplet Crystallization  
of Organic-Soluble Small Molecules**

**Andrew R. Tyler, Ronnie Ragbirsingh, Charles J. McMonagle, Paul G. Waddell, Sarah E. Heaps, Jonathan W. Steed, Paul Thaw, Michael J. Hall, and Michael R. Probert**

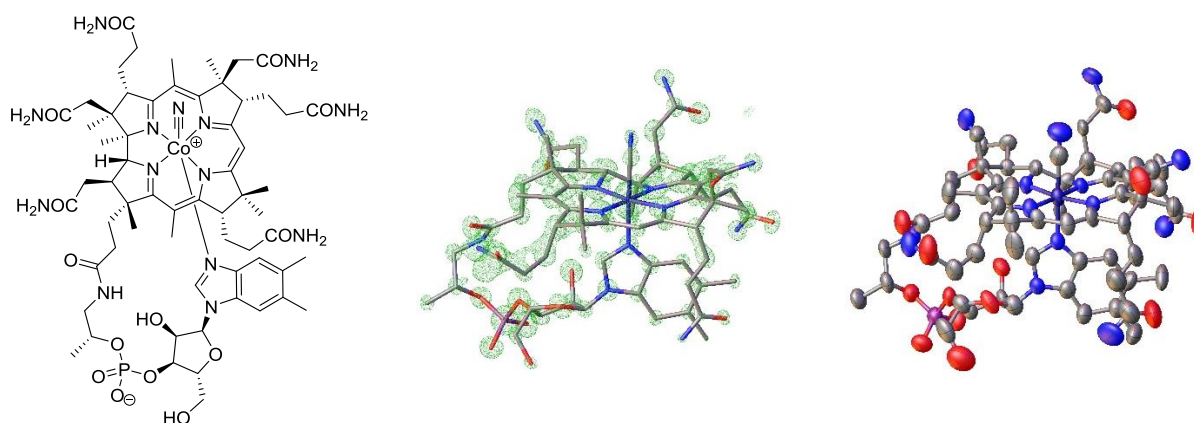
**Figure S1:** Additional compounds **S1-S4** for which single crystals were successfully grown via the ENaCt protocol and subsequently analysed by SCXRD. Molecular structures, electron density maps and refined crystallographic molecular models are shown. Each structure is provided with selected crystallographic information and ENaCt conditions for crystal growth.



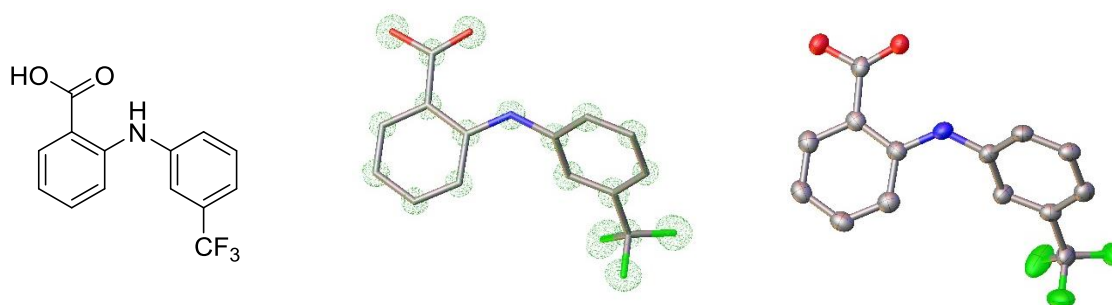
Nicotinic acid (**S1**):  $R_1 = 0.0594$ ,  $wR_2 = 0.1100$ , Residual  $e^-$  density ( $e^- \text{Å}^3$ ) min = -0.231 and max = 0.1171  
200 nL Mineral oil, 50 nL of 40 mg/mL of nicotinic acid in DMSO, 100 nL toluene.



Cholesterol (**S2**):  $R_1 = 0.0594$ ,  $wR_2 = 0.1118$ , Residual  $e^-$  density ( $e^- \text{Å}^3$ ) min = -0.162 and max = 0.199  
200 nL FC-40, 50 nL of 25 mg/mL of cholesterol in DMF, 25 nL *n*-butanol.

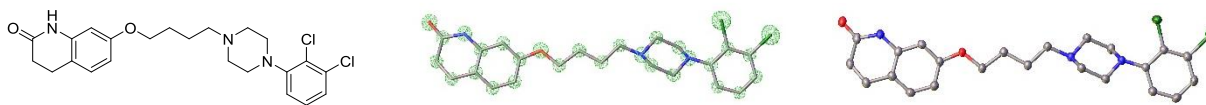


Vitamin B12 (**S3**):  $R_1 = 0.0902$ ,  $wR_2 = 0.1535$ , Residual  $e^-$  density ( $e^- \text{Å}^3$ ) min = -0.433 and max = 0.260,  
Flack = 0.071(12)  
200 nL Mineral oil, 50 nL of 50 mg/mL in DMSO, 50 nL ( $\pm$ )-2-methyl-2,4-pentanediol.

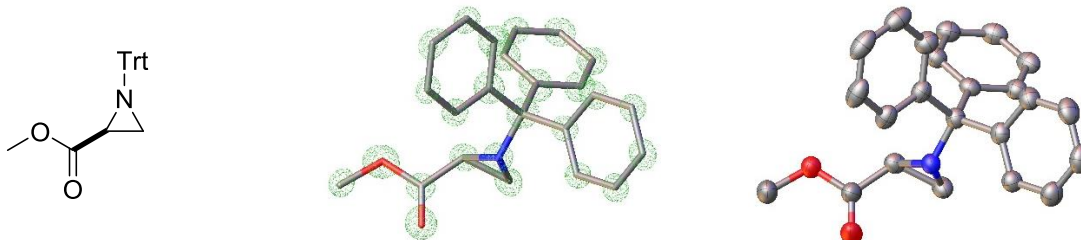


Flufenamic acid (**S4**):  $R_1 = 0.1093$ ,  $wR_2 = 0.1776$ , Residual  $e^-$  density ( $e^- \text{Å}^3$ ) min = -0.314 and max = 0.343  
200 nL PDMSO, 50 nL of 50 mg/mL flufenamic acid in DMF, 50 nL  $\text{H}_2\text{O}$ .

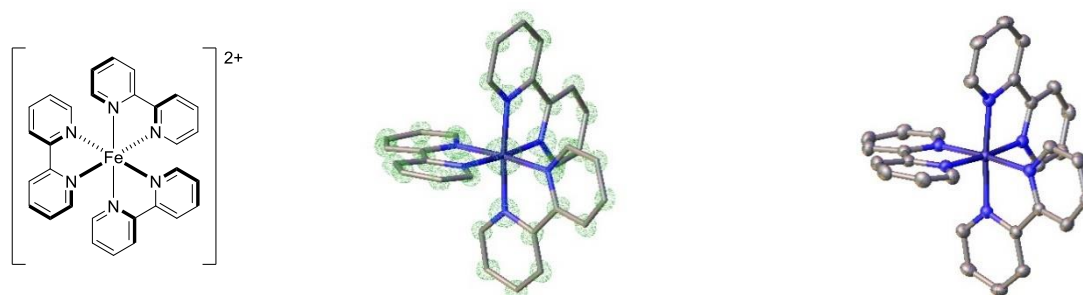
**Figure S2:** Additional compounds **S5-S7** for which single crystals were successfully grown via the ENaCt protocol and subsequently analysed by SCXRD. Molecular structures, electron density maps and refined crystallographic molecular models are shown. Each structure is provided with selected crystallographic information and ENaCt conditions for crystal growth.



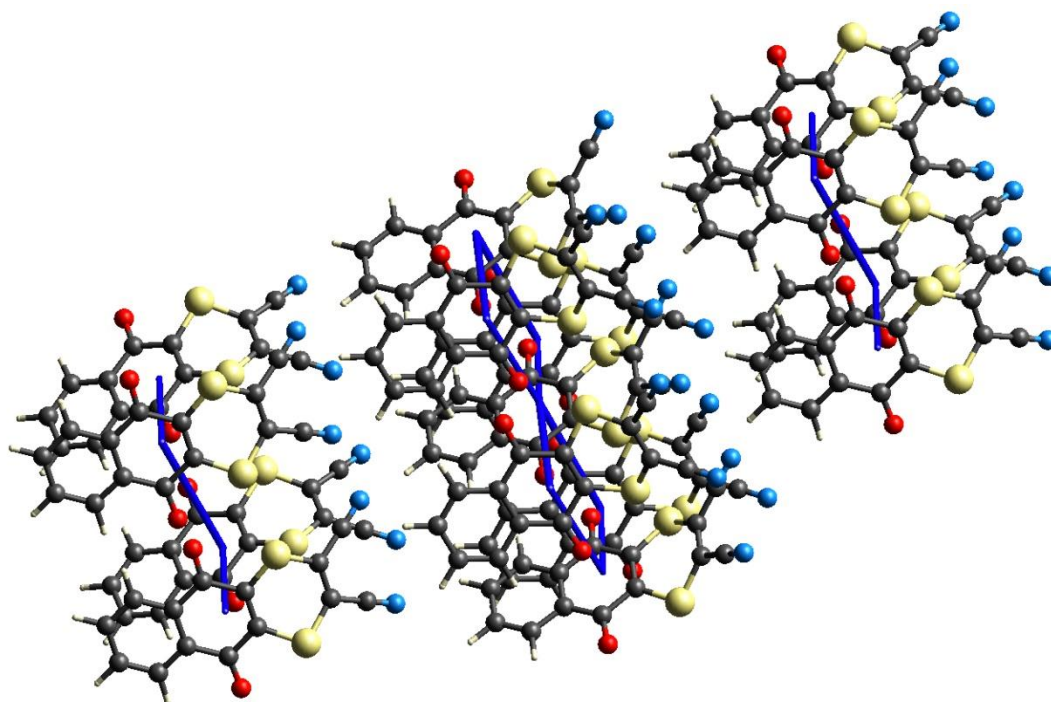
Aripiprazole (**S5**):  $R_1 = 0.0599$ ,  $wR_2 = 0.0947$ , Residual  $e^-$  density ( $e^- \text{Å}^{-3}$ ) min = -0.256 and max = 0.250  
200 nL Fomblin Y, 50 nL of 20 mg/mL of aripiprazole in DMSO.



Methyl (*S*)-1-tritylaziridine-2-carboxylate (**S6**):  $R_1 = 0.0461$ ,  $wR_2 = 0.0935$ , Residual  $e^-$  density ( $e^- \text{Å}^{-3}$ ) min = -0.129 and max = 0.130, Flack = 0.0(2)  
200 nL PDMSO, 50 nL of 40 mg/mL of methyl (*S*)-1-tritylaziridine-2-carboxylate in DMF, 100 nL H<sub>2</sub>O.



[Fe(bipy)<sub>3</sub>]<sup>2+</sup>·2PF<sub>6</sub><sup>-</sup> (**S7**):  $R_1 = 0.0331$ ,  $wR_2 = 0.0707$ , Residual  $e^-$  density ( $e^- \text{Å}^{-3}$ ) min = -0.288 and max = 0.299  
200 nL FC 40, 50 nL of 25 mg/mL of [Fe(bipy)<sub>3</sub>]<sup>2+</sup>·2PF<sub>6</sub><sup>-</sup> in DMSO (*Note*: counter ions omitted for clarity).



**Figure S3:** Total-energy-framework diagram for dithianon using cylinder size 50 for energies. Note energies less than  $15 \text{ kJmol}^{-1}$  have been omitted for clarity. Energies were calculated for the 150 K structure obtained by single crystal X-ray diffraction using the B3LYP/6-31G(d,p) functionals.

## Supplemental Experimental Procedures

### Materials.

Commercially available oils and chemical substrates were purchased from Sigma Aldrich, Fluorochem, Alfa Aesar or Molecular Dimensions.

5-Methyl-2-[(2-nitrophenyl)amino]-3-thiophenecarbonitrile (ROY) was provided by J.W. Steed, metal complex ( $[\text{Fe}(\text{bipy})_3]^{3+}$ ) was provided by M.R. Probert, methyl (S)-1-tritylaziridine-2-carboxylate was provided by A.R. Tyler, and methyl 4-(5,5-difluoro-5H-4 $\lambda^4$ ,5 $\lambda^4$ -dipyrrolo[1,2-c:2',1'-f][1,3,2]diazaborinin-10-yl)benzoate was provided by M.J. Hall.

Laminex™ glass 96 well plates with a 100 micron spacer and cover slips were purchased from Molecular Dimensions. Glass coated 96 well plates were obtained from Fisher Scientific.

### Sample preparation

Stock solutions of each substrate were freshly prepared for each crystallisation experiment. Samples were weighed into a screw top vial, dissolved in a suitable solvent at the desired concentration and then warmed where appropriate (40 °C, 30 min) to ensure that all solids were dissolved. Stock solutions were left to cool (1 hour, r.t.) and assessed *via* optical microscopy to ensure premature crystallisation had not occurred before being used in any ENaCt experiment.

### Crystallisation of substrates

Crystallisation experiments were completed using a TTP LabTech mosquito® liquid handling robot using LAMINEX™ glass 96 well plates with a 100 micron spacer and sealed with a glass cover slip. An appropriate volume (typically 200  $\mu\text{L}$ ) of each oil was first dispensed onto a LAMINEX™ plate (aspirate 1.0 mm/min, dispense 1.0 mm/min). After which 50  $\mu\text{L}$  of substrate solution was injected into each oil droplet (aspirate 20 mm/min, dispense 20 mm/min), followed by an additional injection of a secondary solvent if required (aspirate 20 mm/min, dispense 20 mm/min). Plates were then sealed with a glass cover slip, stored in the dark at room temperature and inspected for crystal growth at regular intervals. Visualisation of the experiment wells was carried out with a Nikon SMZ1000 microscope fitted with a cross polariser. Photographs were taken with a GXCAM-U3-5 5.1MP camera.

### Single crystal X-ray diffraction studies

Upon observation of suitable crystals, wells were opened, with use of a tungsten carbide scribe to remove a small portion of the glass cover slide, and the crystal manipulated using Mitegen Kapton microtools. Crystals were transferred to a glass slide and extracted under oil (Fomblin YR-1800) with a standard Mitegen Kapton loop, and mounted onto either an in-house diffractometer (Rigaku Oxford Diffraction Xcalibur, Atlas, Gemini ultra, or Bruker D8 Vantage, Photon 2, dual Incoatec  $\text{I}\mu\text{S}$  (Ag/Cu) both equipped with an Oxford Cryosystems Cryostream open-flow cooling device and maintained at 150 K) or stored in a liquid  $\text{N}_2$  storage dry shipper for investigation on beamline I19 of Diamond Light Source, *via* remote access.

### Ordinal logistic regression

Let the outcome of each experiment be encoded by an ordinal variable Y where:

$Y = \{1, \text{ if no solid (still solvated)}; 2, \text{ if phase separation}; 3, \text{ if solid, but not crystalline}; 4, \text{ if small crystal}; 5, \text{ if large crystal}.$

A proportional odds model for ordinal logistic regression was used to assess the relationship between the response variable Y and covariates, which comprised two quantitative variables (volume of solvent and volume of oil) and three categorical variables (solvent, type of oil and compound). Inference was performed in the Bayesian framework, carrying out the necessary numerical computation using rstan<sup>S1</sup>, the R interface to the Stan software<sup>S2</sup>. Use of FC40 and, in particular, PDMSO both appear to have a positive relationship with the response variable; the posterior probabilities that the corresponding regression coefficients are greater than zero, indicating positive association, are each equal to 1.00, to two decimal places.

## Supplemental References

S1. Stan Development Team (2016). RStan: the R interface to Stan. R package version 2.12.1.

S2. Carpenter, B., A. Gelman, M. D. Hoffman, D. Lee, B. Goodrich, M. Betancourt, M. A. Brubaker, J. Guo, P. Li and A. Riddell (2017). Stan: A probabilistic programming language. *Journal of Statistical Software* 76 (1), 1–32.