## **Supplementary Information**

## Multifunctional N-P-doped carbon dots for regulation of apoptosis and autophagy in B16F10 melanoma cancer cells and *in vitro* imaging applications

Vivek K. Bajpai<sup>1</sup>, Imran Khan<sup>2,3</sup>, Shruti Shukla<sup>4</sup>, Sung-Min Kang<sup>5</sup>, Faisal Aziz<sup>3</sup>, Kumud Malika Tripathi<sup>6</sup>, Deepika Saini<sup>7</sup>, Hye-Jin Cho<sup>8</sup>, Nam Su Heo<sup>9</sup>, Sumit K. Sonkar<sup>7,\*</sup>, Lei Chen<sup>10,\*</sup>, Yun Suk Huh<sup>2,\*</sup>, Young Kyu Han<sup>1,\*</sup>

<sup>1</sup> Department of Energy and Materials Engineering, Dongguk University-Seoul, 30 Pildong-ro 1-gil, Seoul 04620, Republic of Korea

<sup>2</sup> Department of Biological Engineering, Biohybrid Systems Research Center (BSRC),

Inha University, 100 Inha-ro, Nam-gu, Incheon 22212, Republic of Korea

<sup>3</sup> The Hormel Institute, University of Minnesota, Austin, MN, 55912, USA

<sup>4</sup> Department of Food Science and Technology, National Institute of Food Technology Entrepreneurship and Management (NIFTEM), Sonipat, Haryana 131028, India

<sup>5</sup> Wallace H. Coulter Department of Biomedical Engineering, Georgia Institute of Technology and Emory School of Medicine, Atlanta, Georgia, 30332, USA

<sup>6</sup> Department of Chemistry, Indian Institute of Petroleum and Energy, Visakhapatnam 531035, Andhra Pradesh, India.

<sup>7</sup> Department of Chemistry, Malaviya National Institute of Technology, Jaipur, Jaipur 302017, India

<sup>8</sup> Reliability Assessment Center for Chemical Materials, Korea Research Institute of Chemical Technology (KRICT), 141 Gajeong-ro, Yuseong-gu, Daejeon 305-600, Republic of Korea

<sup>9</sup> Research Center for Materials Analysis, Korea Basic Science Institute (KBSI), Daejeon 34133, Republic of Korea
 <sup>10</sup> College of Food Science, Fujian Agriculture and Forestry University, Fuzhou, Fujian

<sup>10</sup> College of Food Science, Fujian Agriculture and Forestry University, Fuzhou, Fujian 350002, China

Running head: Anticancer potential of N-P-doped carbon nanodots

Corresponding authors: Sumit K. Sonkar (E-mail: <u>sksonkar.chy@mnit.ac.in);</u> Lei Chen (E-mail: <u>chenlei841114@hotmail.com);</u> Yun Suk Huh (E-mail: <u>yunsuk.huh@inha.ac.kr);</u> Young-Kyu Han (E-mail: <u>ykenergy@dongguk.edu</u>)

row min		16				10		ro	w max
		CT1	Т29	549	eLa	16F1			
	ld	Ĭ	Ţ	Â	Ť	à	ld		
		100.00	100.00	100.00	100.00	100.00	Cont	irol	
		99.88	99.12		98.21	98.51	1 µl/r	nL	
		99.64	98.14	98.64	97.23	95.61	2.5 µ	l/mL	
		91.66	93.13	92.44	95.64	92.14	5 μl/ι	nL	
		85.36	88.66	89.64	88.44	84.61	7.5 µ	l/mL	
		80.65	82.32	85.81	86.32	80.63	10 µl	/mL	
		75.66	78.63		85.12	75.65	12 µl	/mL	
		70.23	72.61	73.62	81.63	70.44	15 µl	/mL	
		68,55	58.66	58.63	78.96	65.98	20 µl	/mL	
			51.94	50.23	68.31	55.20	25 µl	/mL	
		48.10	44.68	45.17	58.12	50.02	30 µl	/mL	
		25.26	29.14	37.36	45.47	32.14	60 µl	/mL	
		15.21	22.34	28.23	25.16	12.54	90 µl	/mL	
			17.61	12.21	12.77	0.00	120 µ	ıl/mL	

**Figure S1** Heatmap showing the anticancer activities (% cell viability) of NPCDs (0-120  $\mu$ L/mL) on different cancer cells HCT116, HT29, A549, Hela, and B16F10 cells.



**Figure S2** Anticancer activity of NPCDs evaluated by MTT assay, effect of NPCDs (0- $120 \mu$ L/mL) on B16F10 melanoma cells.



**Figure S3** Cytotoxicity of NPCDs on BEAS-2B human lung epithelial cells and HACAT as human keratinocytes.



**Figure S4.** Autophagy attenuated by siRNA ATG5. The efficiency of siRNA mediated knockdown of ATG5 in B16F10 cells was examined by RT-PCR.

Sr. No.	Primary Antibody used	Dilution used	Secondary antibody used	Company
1	Bax (Rabbit)	1:1000	Anti-rabbit IgG,	Cell signaling
			HRP-linked	technology
2	Bcl2 (Rabbit)	1:1000	Anti-rabbit IgG,	Cell signaling
			HRP-linked	technology
3	Caspase 3 (Pabhit)	1.1000	Anti-rabbit IgG,	Cell signaling
	Caspase-5 (Rabbil)	1.1000	HRP-linked	technology
4	R actin (Pabhit)	1.1000	Anti-rabbit IgG,	Cell signaling
	p-actili (Rabbit)	1.1000	HRP-linked	technology
5	CDK 2 (Pabbit)	1:1000	Anti-rabbit IgG,	Cell signaling
			HRP-linked	technology
6	CDK (Rabbit)	1.1000	Anti-rabbit IgG,	Cell signaling
		1.1000	HRP-linked	technology
7	CDV 6 (Dabbit)	1.1000	Anti-rabbit IgG,	Cell signaling
		1.1000	HRP-linked	technology
8 pź	n21 (Dabbit)	1.1000	Anti-rabbit IgG,	Cell signaling
	μετ (παυριί)	1.1000	HRP-linked	technology
9	LC 2 (Dabbit)	1.1000	Anti-rabbit IgG,	Cell signaling
	LC-3 (Rabbit)	1.1000	HRP-linked	technology
10		1:1000	Anti-rabbit IgG,	Cell signaling
	poz (Rabbil)		HRP-linked	technology
11		1.1000	Anti-rabbit IgG,	Cell signaling
	ΑΙ G-ο (Κάρριι)	1:1000	HRP-linked	technology

**Table S1** Primary and secondary antibodies used in the study.

Secondary antibody dilution used 1:10,000 (Cell Signaling Technology).

Restore<sup>™</sup> PLUS Western Blot Stripping Buffer.

Primers	Sequence	Applications	
atg5-f	AGGCAACCTGACCAGAAACA	Real time PCR	
atg5-r	GAGGAAAGCAGAGGTGATGC	Real time PCR	
p62-f	TGCCCAGACTACGACTTGTG	Real time PCR	
p62-r	AGTGTCCGTGTTTCACCTTCC	Real time PCR	
lc3-f	AGCAGCATCCAACCAAAATC	Real time PCR	
lc3-r	CTGTGTCCGTTCACCAACAG	Real time PCR	
caspase3-f	TGAGCCATGGTGAAGAAGGA	Real time PCR	
caspase3-r	TCGGCCTCCACTGGTATTTT	Real time PCR	
βactin-f	TCCCTGTATGCCTCTGGTCGT	Real time PCR	
βactin-r	AAGCTGTAGCCTCTCTCGGTC	Real time PCR	

 Table S2 Primers used in this study.