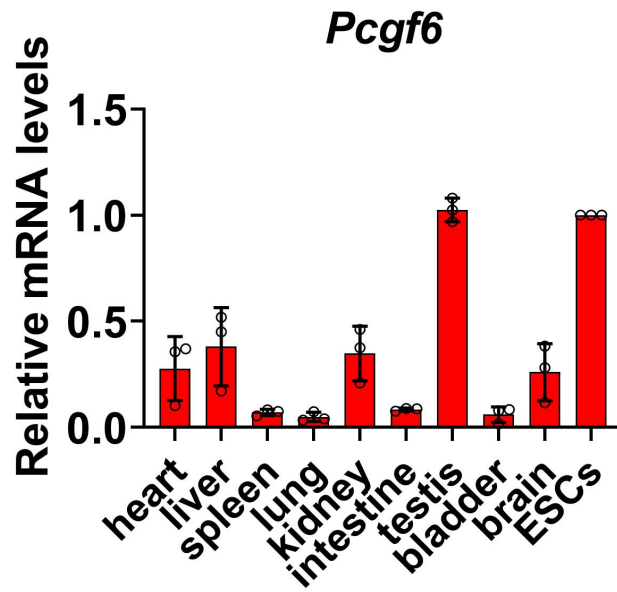


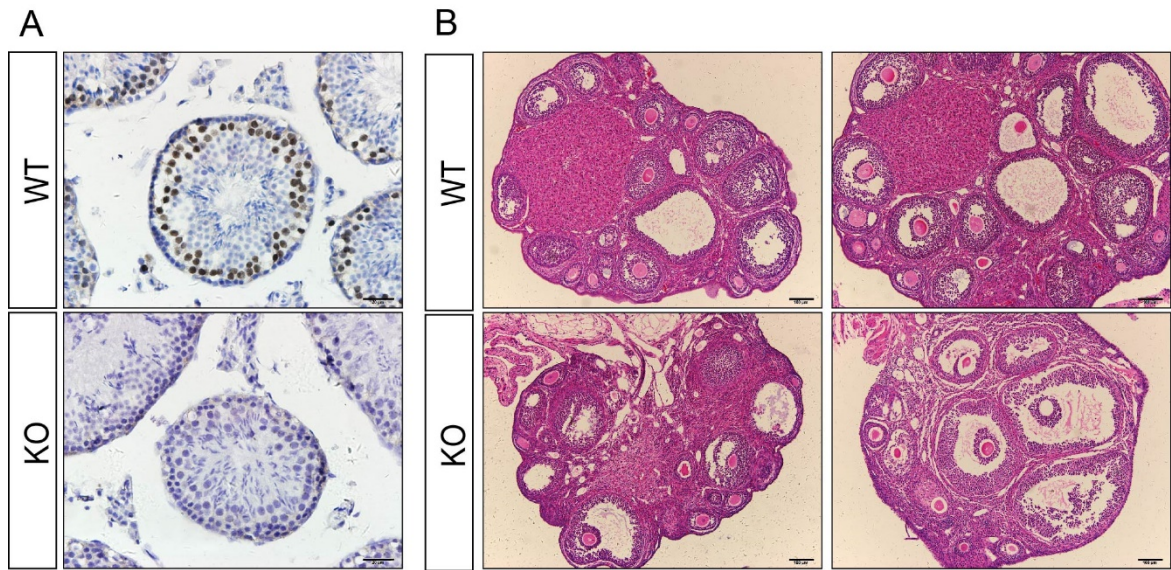
## Supporting Information

### **The polycomb group protein PCGF6 mediates germline gene silencing by recruiting histone-modifying proteins to target gene promoters**

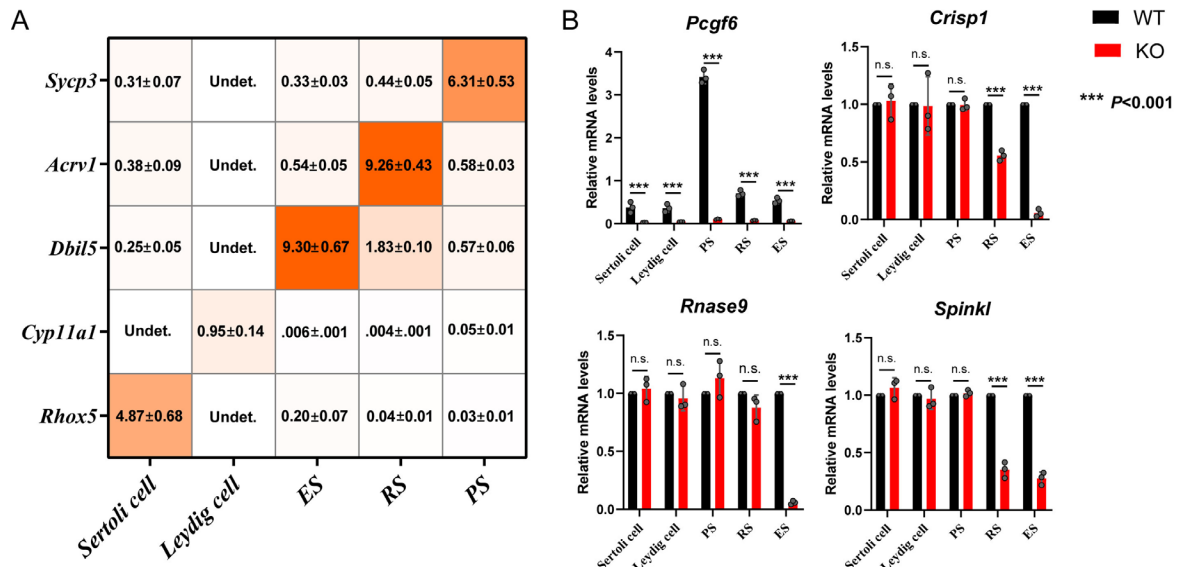
Mengjie Liu<sup>1</sup>, Yaru Zhu<sup>1</sup>, Fei Xing<sup>2</sup>, Shuang Liu<sup>1</sup>, Yin Xia<sup>3</sup>, Qing Jiang<sup>4</sup>, Jinzhong Qin<sup>1\*</sup>



**Figure S1. Transcriptional levels of *Pcgf6* in various tissues of wild-type mice.** *Actin* was used as a control for normalization. The *Pcgf6* mRNA levels of wild-type ESCs were considered as 1. Data are from 3 biological replicates each with 3 technical replicates. Error bars are SD.

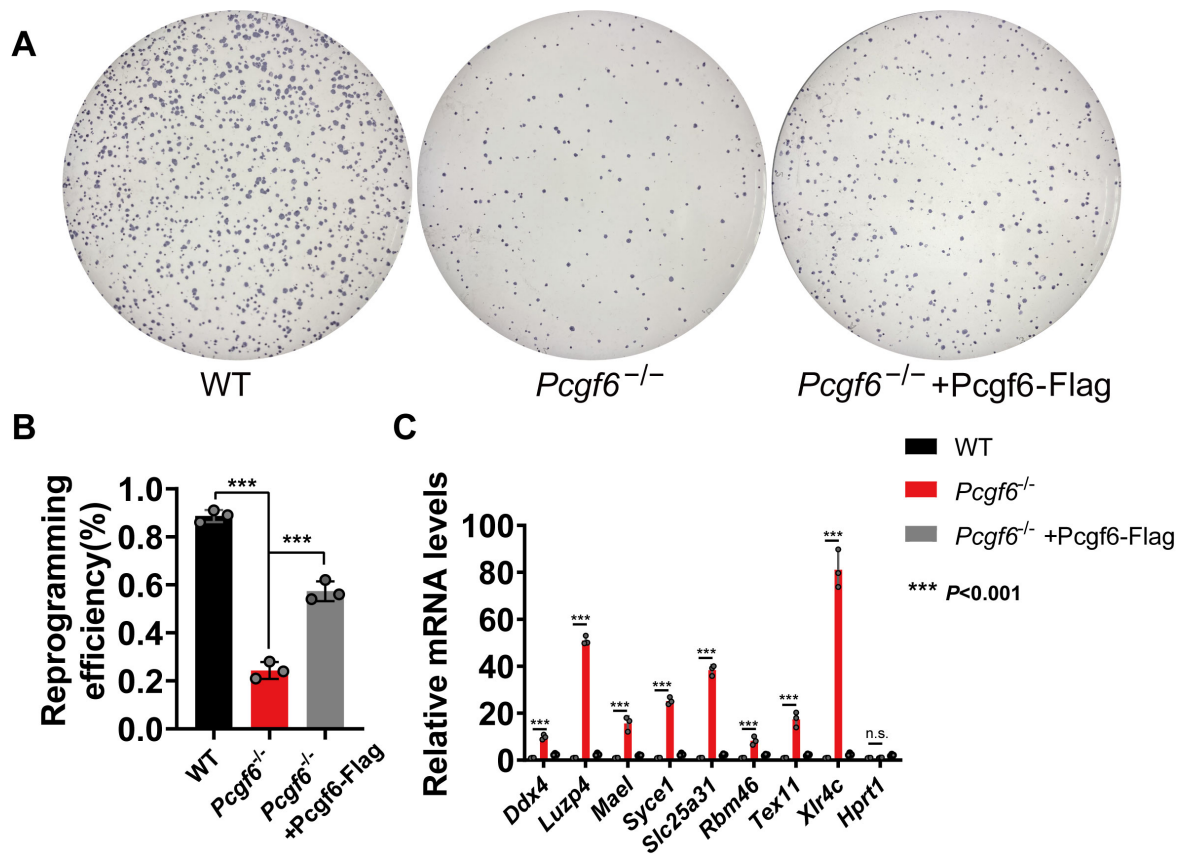


**Figure S2. Staining of tissue sections.** A. Immunohistochemistry staining of representative sections of the testes with the anti-Pcgf6 antibody. Scale bars represent 20  $\mu\text{m}$ . B. HE-staining of representative sections of the ovaries of the WT and *Pcgf6*<sup>-/-</sup> mice. Scale bars represent 100  $\mu\text{m}$ .

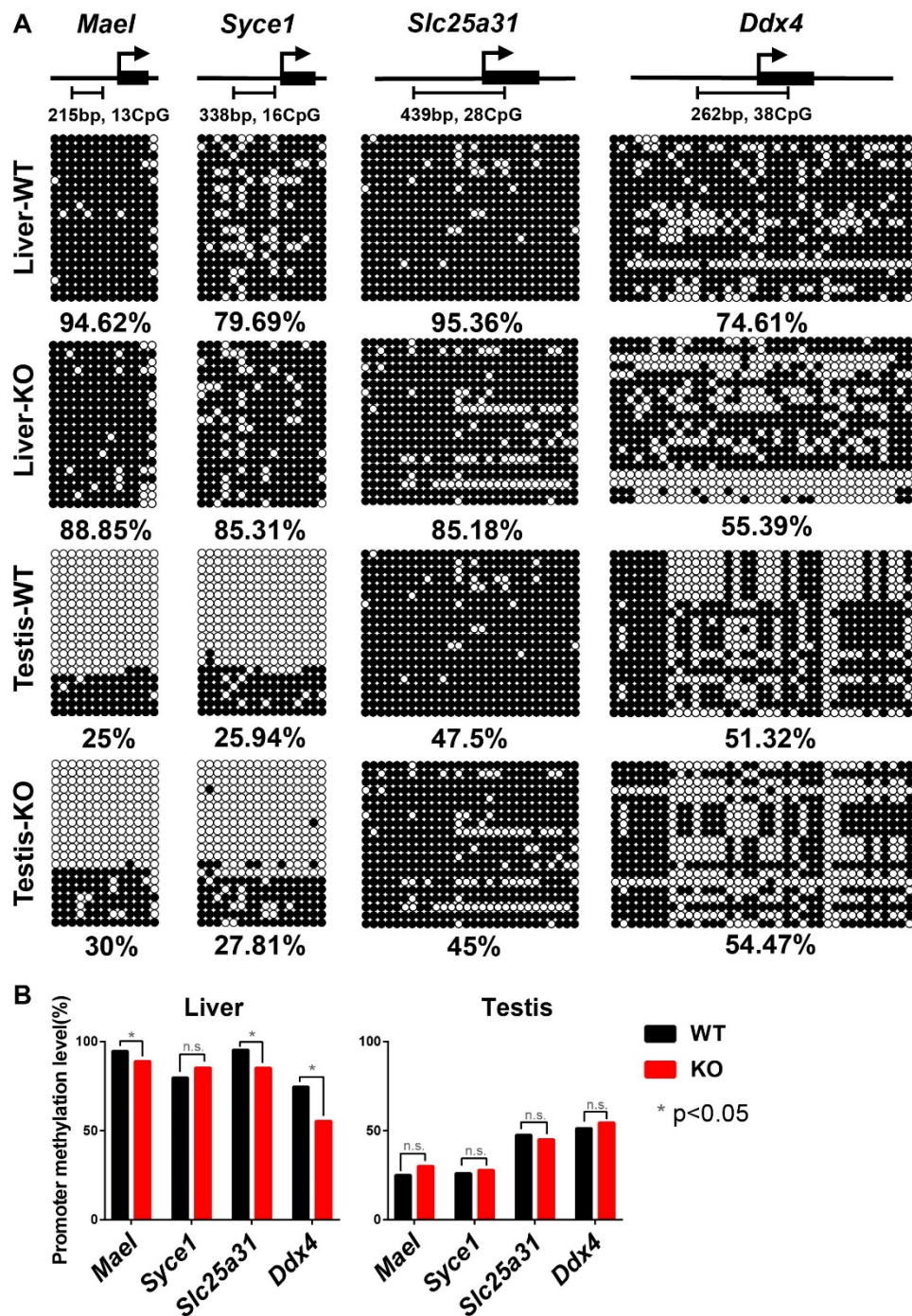


**Figure S3. Transcriptional levels of *Pcgf6* targeted genes in various testicular cell types.**

A. Expression levels of markers genes in purified cell populations. Values represent qRT-PCR analyses of RNA from purified mouse testicular cell populations with Sertoli, Leydig, and germ cell-specific markers, normalized to *Rps2* RNA. The values shown are representative of means  $\pm$  SD of six individuals, relative to those for total testes RNA. Primer sequences are available in Supplementary Table S1. PS, pachytene spermatocyte; RS, round spermatid; ES, elongating spermatid; Undet., undetected. B. The relative mRNA levels of indicated genes in purified testicular cell populations of wildtype and *Pcgf6*<sup>-/-</sup> mice. Error bars indicate  $\pm$  SD of 3 independent biological samples. n.s. means non-significant (two-tailed Student's t-test).

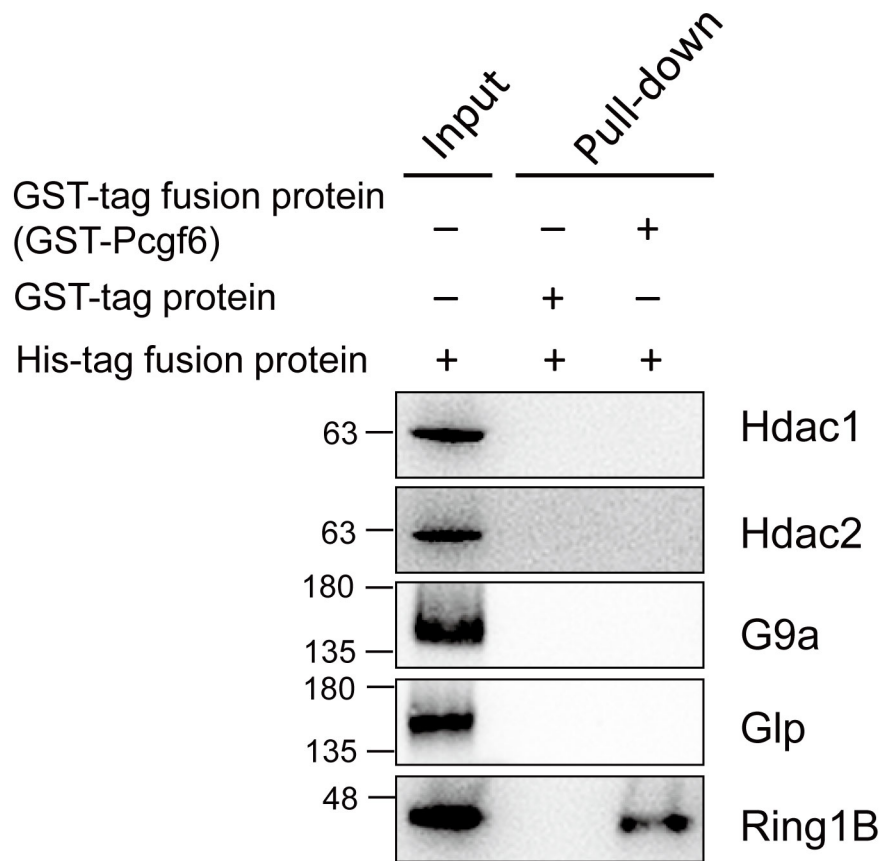


**Figure S4. Deficiency of *Pcgf6* decreases iPS colony numbers.** A. Effects of *Pcgf6* knockout on somatic cell reprogramming. iPS cell colonies were visualized in 60 mm dishes by AP staining at day 21. B. Quantification (average and SD) of iPS cell colonies from three independent experiments at day 21. C. Relative expression of the indicated germline-related genes in MEFs of the indicated genotypes determined by qRT-PCR. *Actin* was used as a control for normalization. Error bars indicate  $\pm$ SD of 3 independent experiments.

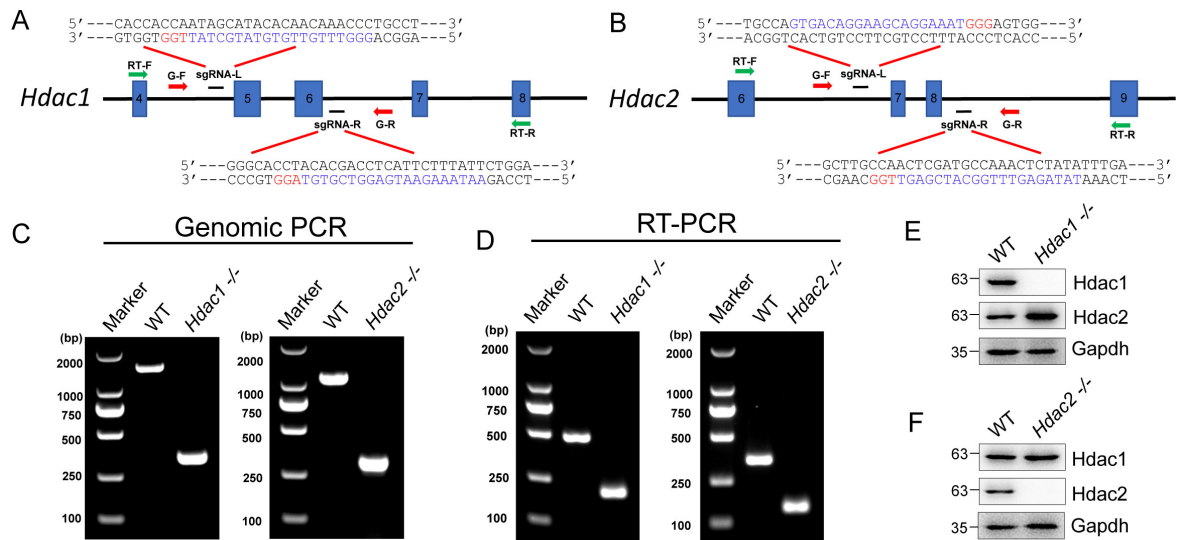


**Figure S5. DNA methylation levels of the germline genes are modestly decreased in the liver after *Pcgf6* depletion.**

A. Methylation status of CGI promoters of four up-regulated germline genes in the liver of *Pcgf6*<sup>-/-</sup> mice. Testis was used as a control. Cartoons demonstrate locations of CGIs relative to TSSs (arrows). CpG nucleotides are shown by black (methylated) or white (unmethylated) circles. Methylation extent is given as a percentage of methylated CpG. B. Bar graph ( $\chi^2$  test) of the CpG methylation levels.

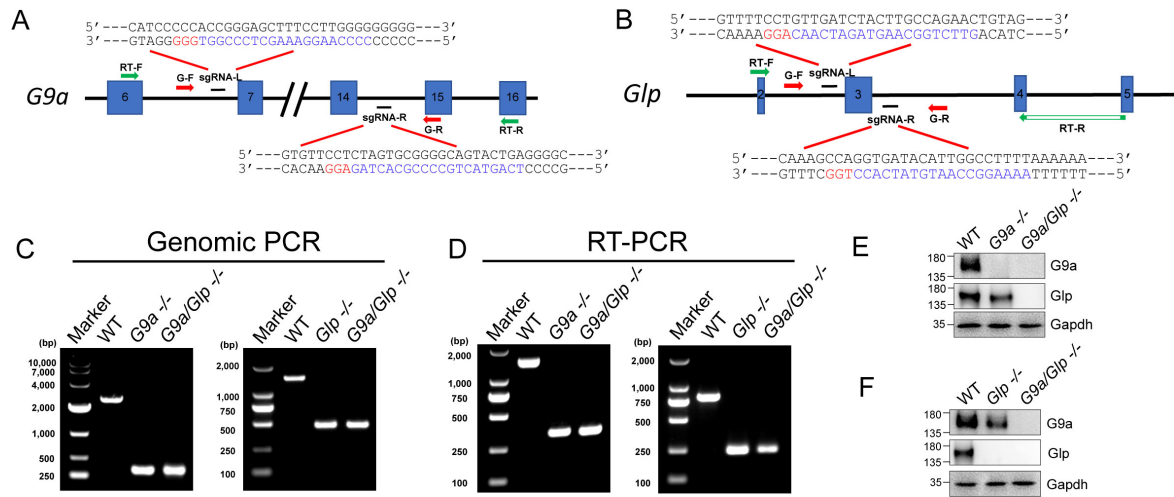


**Figure S6. Pcgf6 does not directly interact with Hdac1/2 and G9a/Glp.** His-tag fusion proteins were probed for interaction with GST-Pcgf6 in pull-down assays. GST tag alone is used as a negative control. The specific interaction of those proteins with GST-Pcgf6 was detected by Western blot with specific antibodies for Hdac1, Hdac2, G9a, Glp or Ring1B respectively. 10% of the fusion protein used in each assay was loaded as input. Assays were done independently for each protein. The results are representative of two independent experiments.

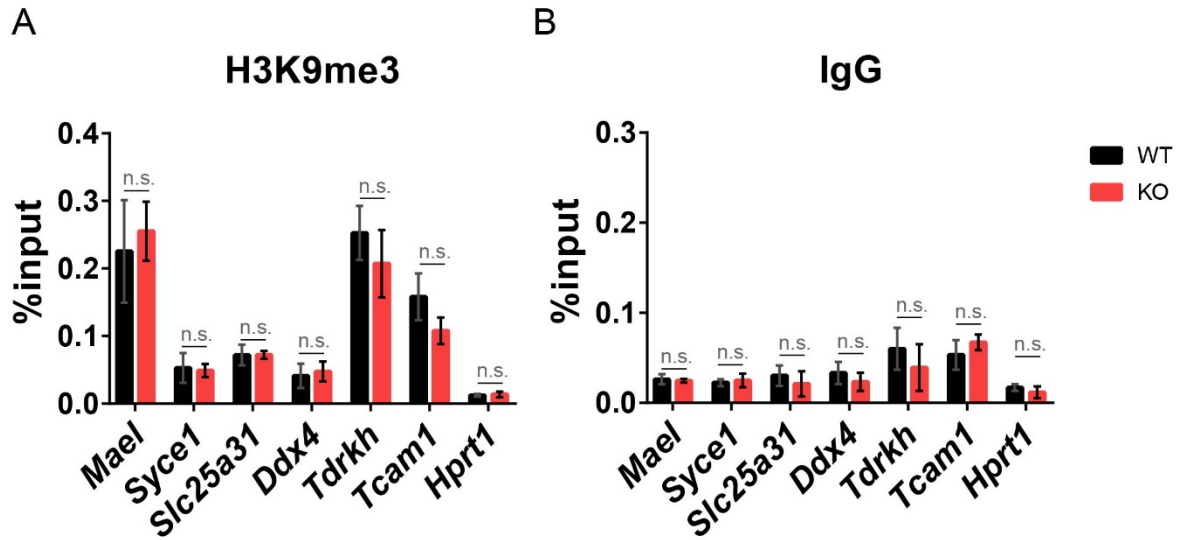


**Figure S7. Generation of *Hdac1* or *Hdac2* knockout ESCs.** A, B. The schematic of *Hdac1* and *Hdac2* gene targeting strategies. The exons 5-6 of *Hdac1* or the exons 7-8 of *Hdac2* were deleted by a pair of sgRNAs, respectively. The location of genomic PCR primers was shown by red arrows, and RT-PCR primers were represented by green arrows. C. Genomic PCR for analysis of *Hdac1/2* gene status. D. RT-PCR analysis for residual *Hdac1/2* mRNA. E, F. The absence of the proteins Hdac1 or Hdac2 in knockout ESCs analyzed by Western blot. Gapdh served as a loading control.

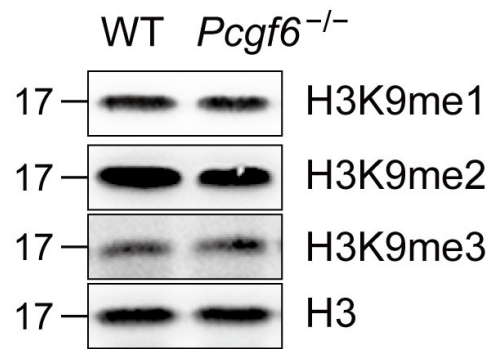




**Figure S8. Generation of *G9a* and *Glp* knockout ESCs.** A, B. The schematic of *G9a* and *Glp* gene targeting strategies. The exons 7-14 of *G9a* or the exons 3 of *Glp* were deleted by a pair of sgRNAs, respectively. The locations of genomic PCR primers were showed by red arrows, and RT-PCR primers were represented by green arrows. C. Genomic PCR for analysis of *G9a* and *Glp* gene status. D. RT-PCR analysis for residual *G9a* and *Glp* mRNA. E, F. The absence of the proteins G9a or Glp in knock-out ESCs analyzed by Western blot. Gapdh was used as a loading control.



**Figure S9. ChIP of H3K9me3 followed by qPCR analysis in ESCs of indicated genotypes.** Purified rabbit IgG was used as a negative control. Error bars indicate  $\pm$  SD of 3 biological repeats. n.s. means non-significant (two-tailed Student's t-test).



**Figure S10. Western blot showing the methylation levels of H3K9 in WT and *Pcgf6*<sup>-/-</sup> ESCs.**

**Table S1. List of Primers Used in this Study.**

Using	Target	Forward	Reverse
Genomic PCR	<i>Pcgf6</i>	TGGGGAGGGAAGTTAAGAGG	R1: TGAGCCCAAGAGTGTGGTA R2: CTTGAACTCACGGCAATCCT
	<i>Hdac1</i>	CTTATGGTGCACATGCAGACG	GATCAGGAGGCCGTGACAAT
	<i>Hdac2</i>	AGCAGAAGAAGGCCTAGTCG	TCAGACAACAAGGTACAGGTC
	<i>G9a</i>	GCTTTTCGTGGAGTGTGGTT	AGTTGGGGTCCAGGTTGTC
	<i>Glp</i>	ATGCCAGAAGAAAGCATTGG	ACTTCACTCAATGCCAGGA
RT-PCR	<i>Hdac1</i>	TGAGGACTGTCCGGTGTTTG	CACTGCACTAGGCTGGAACA
	<i>Hdac2</i>	CAACAGATCGCGTGATGACC	CGGATTGTGTAGCCTCCTCC
	<i>G9a</i>	GAACTCTGGTAGCCTGTCCG	GTTACCACGGCCTCCATTA
	<i>Glp</i>	AGTTCTGGCCAAGCAAGAGA	ACAAAAGGCAAGCAAGCTGT
qRT-PCR	<i>Pcgf6</i>	GCCAGTCCCTTCAAGCAAAG	GCCCGATAGTTGCTTCTCCT
	<i>Ddx4</i>	TGTCAGACGCTCAACAGGAT	ACTGGATTGGGAGCTTGTGA
	<i>Luzp4</i>	CAGACGTGACAGTAGGGGAA	GGCCATGAGATCTGCCAATG
	<i>mael</i>	TCTAAGACCTGGGTGCGAAG	ACTGTTGCTGGCCTCATAGT
	<i>Rbm46</i>	ATTGCAAATGGATCCAGAG	TCCCAGAAGAGAGGGTAGCA
	<i>Syce1</i>	CTGAGGTTGGATTTTGAAGA	TCCTTGCTGCTGTCCAAAAC
	<i>Sle25a31</i>	CGATCCGGTGTCTTTCTCGA	ACATTTGCCAAATTGCCACG
	<i>Tex11</i>	CCTGAGGCTGACTTGACCAT	GGGGAGCCTCATCAACATATC
	<i>Hprt1</i>	TCAGTCAACGGGGACATAAA	GGGGCTGTACTGCTTAACCAG
	<i>Xlr4c</i>	CCACTTCTTGAAAGTCCAGCA	TCAGAGAGTTTTCCAGCCTGT
	<i>Xlr4b</i>	GAAACAATCTGGCCCTTGAA	CTTCGCTCATGCTGGACTTT
	<i><math>\beta</math>-actin</i>	AGCCATGTACGTAGCCATCC	CTCTCAGCTGTGGTGGTGAA
	<i>Mageb4</i>	TACTGCTGGCTTCCCTCAA	TCTCCCTTAGATGCTGGCTG
	<i>Zscan4c</i>	ACTCAGGAGCTGCAAAGTCT	ACCATGACAGGAGGCTTCAA
	<i>Tdrkh</i>	AATCATAGGGAGAGGCGGTG	ACTGATTGGCTGCTTTCGTG
<i>Taf7l</i>	GCATGGCATTACTCCACCAC	GGCATGGACCCTTGACTTTC	
<i>Hprt1</i>	TCAGTCAACGGGGACATAAA	GGGGCTGTACTGCTTAACCAG	

	<i>Acrv1</i>	TCAGCAACTTTCAAGCGAGTAT	CTCCTGAAGAGTGCTCACCTG
	<i>Cyp11a1</i>	CCAGTGTCCCCATGCTCAAC	TGCATGGTCCTTCCAGGTCT
	<i>Dbil5</i>	CCCAGGGCGACTGTAACATC	GCAATGTAGATCCTCATGGCAT
	<i>Rhox5</i>	CACCAGGACCAAAGTGGCC	GGTATGGAAGCTGAGGGTT
	<i>Rps2</i>	CTGACTCCCGACCTCTGGAAA	GAGCCTGGGTCTCTGAACA
	<i>Sycp3</i>	AGCCAGTAACCAGAAAATTGAGC	CCACTGCTGCAACACATTTCATA
	<i>Rnase9</i>	CCGAAGCTTCAGGGGAACTA	ACAGCGCCCCTTATAGTGAA
	<i>Crisp1</i>	GCTGTTCTTCTTGGCTGCTG	CTGCCAGATGGAGAAACCAT
	<i>Spink1</i>	TCTGACTTTGGTGCTTCTTCC	CTATCATCTGCACAAACTGGGT
Bisulfite sequencing	<i>Ddx4</i>	GGTTATGGAGTTAAGAGGTTT	ACCAAAACCAAACCTAAAA
	<i>Mael</i>	GGAATGTGAAATATTTAGGAA	TATAAAAATAAAAACAACCTAA AACTAAAT
	<i>Syce1</i>	TTGGTAATTGGGAATTGGG	AACTTCCAAACTATTAACA CACCTA
	<i>Sle25a31</i>	AGGAGTTAATAAGTTTAGGGTGAT	AACACCTACAAACCACACCAAC
CHIP-qPCR	<i>Ddx4</i>	CCTTGGAGAGAGAAACGGGA	GCGGGGACAACAAATAGCAT
	<i>Mael</i>	ATCACAGTGCTCTCCCCAC	CGCCCTTAGTAACCGAGCT
	<i>Syce1</i>	GCAGTGTCTAACAGCCTGGA	CCCTTCAGCCACTACCTCTG
	<i>Sle25a31</i>	ACACGTGTTATGGTCACATGC	GCCTTCTTTGAAGACTGCTTCT
	<i>Tdrkh</i>	GTGAGCGTTGATTGGTGAGG	TTCTCCCACACCCCAAACCTT
	<i>Tcam1</i>	TGTTACGCTTCTCCTCTCA	GAGTCAAAGCGGTCGTCTTG
	<i>Hprt1</i>	GAATCCTCTGGGAGACGACA	CGGAAAGCAGTGAGGTAAGC

**Table S2. List of antibodies used in this study.**

Description	Supplier	Product ID#	Applications
Anti-Pcgf6	Abcam	ab200038	WB, IP, CHIP
Anti-Ring1B	Proteintech	16031-1-AP	WB, CHIP
Anti-Rybp	Santa Cruz Biotechnology	sc-374235	WB
Anti-L3mbtl2	Santa Cruz Biotechnology	sc-365134	WB
Anti-Max	Santa Cruz Biotechnology	sc-197	WB
Anti-Hdac1	CST	5356	WB,CHIP
Anti-Hdac2	CST	5113	WB,CHIP
Anti-Hp1 $\gamma$	CST	2619	WB
Anti-E2f6	Santa Cruz Biotechnology	sc-390022	WB
Anti-G9a	RnD Systems	PP-A8620A-00	WB,CHIP
Anti-Glp	RnD Systems	PP-B0422-00	WB, CHIP
Anti-Cbx8	CST	14696	WB
Anti-Ezh2	CST	5246	WB
Anti-H3k9me1	Millipore	17-680	WB,CHIP
Anti-H3k9me2	Millipore	17-648	WB,CHIP
Anti-H3k9me3	Millipore	17-625	WB,CHIP
Anti-H3k27ac	Abcam	Ab4729	WB
Anti-H2AK119ub1	CST	8240	ChIP
Anti-H3ac	Millipore	06-599	WB,CHIP
Anti-H4ac	Millipore	06-866	CHIP
Anti-H3	Millipore	07-690	WB
Anti-Gapdh	Santa Cruz Biotechnology	sc-47724	WB

goat anti-mouse IgG-HRP	Santa Cruz Biotechnology	sc-2005	WB
goat anti-rabbit IgG-HRP	Santa Cruz Biotechnology	sc-2004	WB

**Table S3. The misregulated genes in the testis of *Pcgf6*<sup>-/-</sup> mice. Separate file (.xlsx)**

**Table S4. The misregulated genes in the liver of *Pcgf6*<sup>-/-</sup> mice. Separate file (.xlsx)**

**Table S5. The misregulated genes in the brain of *Pcgf6*<sup>-/-</sup> mice. Separate file (.xlsx)**