## **Supporting Information for**

# **Discovery of Selective, Substrate-Competitive and Passive Membrane Permeable Glycogen Synthase Kinase-3β Inhibitors: Synthesis, Biological Evaluation, and Molecular Modeling of New C-Glycosylflavones**

Zhibin Liang and Qing X. Li\*

Department of Molecular Biosciences and Bioengineering, University of Hawaii at Manoa, Honolulu, Hawaii 96822, United States

\*Corresponding author. Tel:  $+1$  (808) 965-2011; Fax:  $+1$  (808) 965-3542. E-mail address: qingl@hawaii.edu

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### **S1. Docking Method Validation by Re-docking and Cross-docking Experiments with Known GSK-3β Inhibitors**

We chose two X-ray crystallographic structures of GSK-3 $\beta$  (PDB codes 1PYX<sup>1</sup> and 1H8F<sup>2</sup>). The docking method was validated by re-docking and cross-docking experiments using both ATP-competitive and substrate-competitive inhibitors.

Re-docking the native ligand ANP to 1PYX resulted in a binding pose at the ATP site of GSK-3β with a RMSD of 0.33 Å as compared to its original crystal structure.<sup>1</sup> Under the same parameters, docking with the known ATP-competitive inhibitors, staurosporine and luteolin, led to the binding to the ATP site of GSK-3β, of which the binding poses and positions are similar to the reported data.<sup>1, 3</sup> The ligands HEPES, manzamine A and isoorientin are known GSK-3 $\beta$ inhibitors bound to the substrate site. Docking these ligands to 1PYX confirmed their binding to the substrate site, and the predicted poses are comparable to the reported data.<sup>2, 4, 5</sup> Cross-docking of the same ligands to the 1H8F demonstrated that ANP, staurosporine, and luteolin reside the ATP site of GSK-3β, whereas HEPES, manzamine A, and isoorientin are in the substrate site. Re-docking experiment using the native ligand HEPES of 1H8F resulted in a binding pose at the substrate site of GSK-3 $\beta$  with a RMSD of 0.62 Å as compared to its original crystal structure.<sup>2</sup>



**Predicted docking poses of ANP (A), staurosporine (B), luteolin (C), HEPES (D), manzamine A (E), and isoorientin (F) in GSK-3β (PDB code 1PYX). Ligands in magenta color are original structures in X-ray crystallographic complex, ligands in green color are docking structures.**



**Predicted docking poses of ANP (A), staurosporine (B), luteolin (C), HEPES (D), manzamine A (E), and isoorientin (F) into GSK-3β (PDB code 1H8F). Ligands in magenta color are original structures in X-ray crystallographic complex, ligands in green color are docking structures.**



## **S2. Inhibitions and Docking Scores of** *C***-Glycosylflavones and Reference Inhibitors to GSK3β**

*<sup>a</sup>* IC50 values were the mean of quadruplicate of each of two independent experiments. *<sup>b</sup>* Docking scores were binding affinities calculated by AutoDock Vina. <sup>*c*</sup> IC<sub>50</sub> value from ref 6.



**Linear plots of pIC50 (-LogIC50) versus calculated binding affinity (PDB codes 1PYX and 1H8F) for GSK-3β inhibitors**

#### **S3. Predicted Docking Poses of Flavones with GSK-3β (PDB Code 1PYX)**

Docking poses of flavones **1** (A), **9** (B), **17** (C), **21** (D), **3** (E), and **4** (F) with GSK-3β (PDB code 1PYX). Surface representation of the ATP site of GSK-3β is shown in orange, the substrate site of GSK-3β is shown in green. Within the substrate site, surface features are shown with acidic residues in red, basic residues in blue, and hydrophobic residues in green.  $Mg^{2+}$  ions are shown as bright green spheres. The dotted lines represent interactions with  $Mg^{2+}$  ions and hydrogen bonds with side chain residues of GSK-3β.





**S4. Summary of Molecular Interaction Distances of Compound 30 with GSK3β (PDB Code 1PYX) in Molecular Docking**

#### **S5. GSK-3α Homology Modeling Method**

The SWISS-MODEL<sup>7</sup> template library (SMTL version 2017-09-21, PDB release 2017-09-15) was searched with BLAST and HHBlits for evolutionary related structures matching the target sequence GSK-3α (UniProt Code P49840). For each identified template, the template's quality has been predicted from features of the target-template alignment. The templates with the highest quality have then been selected for model building. Overall 4470 templates were found to match the target sequence. The top templates are:



Template	<b>Sequence</b> identity	Oligo- state	Found by	Method	<b>Resolution</b>	<b>Sequence</b> similarity	Range	Coverage	<b>Description</b>
$1$ pyx. $1.A$	82.97	homodimer	<b>BLAST</b>	X-rav	$2.40 \text{ Å}$	0.55	$98 -$ 449	0.77	$GSK-3\beta$
<b>Built with</b>	Oligo-State	Ligands	<b>GMOE</b>	<b>OMEAN</b>					
ProMod3 Version 1.0.2.	monomer	$Mg^{2+}$ ion	0.72	$-0.90$					

The GSK-3α homology model was built based on the GSK-3β template PDB code 1PYX:

#### Model Quality Estimation:

The global and per-residue model quality has been assessed using the QMEAN scoring function. For improved performance, weights of the individual QMEAN terms have been trained specifically for SWISS-MODEL.



## **S6. Protein Sequence Alignment Between Human GSK-3α (UniProt Code P49840) and Human GSK-3β (PDB Code 1PYX)**

Identical residues are unpainted, nonidentical residues are yellow highlighted, unmatched residues are green highlighted; key residues comprised of ATP site are colored in red, key residues comprised of substrate site are colored in blue. It is obvious that the kinase catalytic domain comprised of both ATP and substrate pocket sequences (D121-H208 and C241-L270 in GSK-3α; D58-H145 and C178-L207 in GSK-3β) are conserved as high as 93% identical. Only nine amino acid residues are nonidentical in these regions between two isoforms.



#### **S7. HPLC Analysis for Purity Determination of Compounds 5-31**

The purity of compounds **5**-**31** was determined by analytical reverse phase Agilent HPLC with a diode array detector (Waters XSELECT CSH Fluoro-Phenyl column,  $150 \times 4.6$  mm,  $3.5$ μm, isocratic elution with 35% aqueous acetonitrile containing 0.1% formic acid for over 20 min, detection at 210 nm, flow rate of 0.8 mL/min). All tested compounds were over 95% purity.











#### **S8. In Vitro Kinase Selectivity Screening for Compound 30**

The Kinase Selectivity Profiling System includes kinase and substrate pairs. Percentage kinase activities upon treatment of compound **30** at 5 μM in a panel of human protein kinases were measured by the ADP-Glo Kinase Assay Kit according to the manufacturer's protocol. The kinase inhibitor staurosporine was used at 1 μM as a reference control. Each data point was collected in quadruplicate of two independent experiments.





#### **S9. LC-MS Method for Determination of the PAMPA Permeability**

At the end of the incubation period, 10 μL sample from each donor and acceptor well was quantified by LC-ESI-QTOF-MS analysis under a positive mode and used a 7-point calibration with appropriate dilution of the samples. A sum of pseudo-molecular ions  $[M+H]^+$ ,  $[M+Na]^+$  and  $[M+K]^+$  for each target analyte was extracted from total ion chromatogram (TOC) for quantification. Samples were run in quadruplicate.

Permeability of the compounds (*P*e) were calculated using the following formula:

Permeability (cm/s):  $P_e = \{-\ln[1 - C_A(t)/C_{eq}]\}/[A^*(1/V_D+1/V_A)^*t]$ 

where A = filter area (0.3 cm<sup>2</sup>), V<sub>D</sub> = donor well volume (0.3 mL), V<sub>A</sub> = acceptor well volume  $(0.2 \text{ mL})$ , t = incubation time (5 hours = 18000 seconds), C<sub>A</sub>(t) = compound concentration in acceptor well at time t,  $C_D(t)$  = compound concentration in donor well at time t,

$$
C_{eq} = [C_D(t)^*V_D + C_A(t)^*V_A]/(V_D + V_A).
$$

Mass retention ( $R\%$ ) is calculated using the following formula:

 $R\% = \{1 - [C_D(t)*V_D+C_A(t)*V_A]/(C0*V_D)\} * 100$ 

where C0 is the initial compound concentration in donor well.

## **S10. Structures of the Reference Compounds**









NH

**Atenolol**



**Staurosporine**

**TDZD-8 Theophylline**

**Desipramine**

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