Supplementary Information

Highly parallel profiling of Cas9 variant specificity

Jonathan L. Schmid-Burgk^{123,43}, Linyi Gao^{123,43}, David Li^{123,4,5}, Zachary Gardner^{123,4}, Jonathan Strecker^{123,4}, Blake Lash^{123,4}, Feng Zhang^{123,45,6,7,*}

 ^a Broad Institute of MIT and Harvard Cambridge, MA 02142, USA
^a McGovern Institute for Brain Research
^a Department of Brain and Cognitive Sciences
^a Department of Biological Engineering
Massachusetts Institute of Technology, Cambridge, MA 02139, USA
^a These authors contributed equally
^e Howard Hughes Medical Institute, Cambridge, MA 02139, USA
^a Lead Contact

*Correspondence should be addressed to F.Z. (zhang@broadinstitute.org)



Supplemental Figure 1 I Extended validation and application of TTISS, Related to Figure 1 (A) TTISS results for multiplexing of 1, 3, 10, 30, and 60 gRNAs. The number of reads for each detected genomic locus is plotted. On-target sites are marked in red. (B) Quantitative TTISS results from three cell lines using 59 guides. (C) Detection of donor integration sites using prime editing targeting three genomic loci in HEK 293T cells. Spacer and extension sequences are provided in Supplemental Table 3. (D) Distribution of off-target sites per gRNA across 59 gRNAs detected by TTISS using WT SpCas9. (E) Comparison of GuideScan-predicted specificity scores to TTISS measured on-target fractions for 59 guides. (F) Comparison of Elevation specificity scores to TTISS measured on-target fractions for 47 guides which could be scored by the CRISPR ML online interface.



Supplemental Figure 2 I On-target and off-target activity of selected SpCas9 variants, Related to Figures 1 and 2. All indel frequencies were quantified by targeted deep sequencing. (A) Normalized indel frequencies for 59 target sites for WT, LZ3 Cas9, and seven previously reported SpCas9 specificity-enhancing variants. Each dot represents a different guide (mean of n = 2 replicates). The teal lines show the median activity for each Cas9 variant. Target sites were selected from the GeCKO library (Shalem et al. Science 2014), each targeting a different gene, without prior knowledge of activity. (B) Activity of SpCas9 variants at additional on-target and off-target sites. Guides g5-g11 were selected based on prior knowledge of low activity for eSpCas9(1.1) and SpCas9-HF1. (C) Crystal structure of SpCas9 (PDB ID: 5F9R) showing the position of the four mutations in LZ3. (D) Activity of double mutants of selected specificity-enhancing single mutants. (E) Epistasis plots of the variants in (D) for guides g1 and g2, where epistasis was calculated as fAB/(fA x fB), where fAB is the normalized indel frequency of the double mutant, and fA and fB are the normalized indel frequencies of the corresponding single mutants.





Supplemental Figure 3 I Extended assessment of +1 indel frequencies using TTISS, Related to Figure 3. (A) +1 insertion frequencies measured by TTISS or predicted by FORECasT, inDelphi, or Lindel are correlated to +1 frequencies measured by targeted indel sequencing for WT SpCas9 across 58 gRNAs. (B) TTISS-predicted +1 frequencies for SpCas9 variants calculated for 58 gRNAs plotted against TTISS-predicted +1 frequencies for WT SpCas9 across 59 guides, grouped by the nucleotide identity at the -4 position relative to the PAM. (D) Plot of +1 frequencies for LZ3 against +1 frequencies for WT SpCas9 as measured by targeted sequencing for 59 gRNAs. (E) Insertion and deletion length distributions of Cas9 variants across 59 guides from targeted sequencing. Indel length frequencies relative to total indels are shown on logarithmic scale.