

Figure S1. Muscles and motor neurons differentiate normally in *mis12* mutants, related to

Figure 1

(A-C) Motor neurons were labeled and counted by using *OK6-GAL4* to drive *UAS-mCD8::GFP* in either *mis12*^{f03756}/+ (control, **A**) or *mis12*^{f03756}/*mis12*^{f03756} (**B**) embryos. The number of neurons in the three indicated segments were counted (**C**). It was not always possible to discern all 8 motor neurons in each segment and therefore some embryos were scored at fewer than the predicted 24 motor neurons; the counts were nonetheless equivalent

between the genotypes. (D,E) Muscles were stained with rhodamine-phalloidin (green). In each hemisegment of 21-h AEL embryos of *mis12*^{f03756} homozygotes, the normal pattern of muscle fibers was present. Muscles are numbered according to the nomenclature of Crossley (1978) to facilitate the comparison. (F, G) Rhodamine-phalloidin stained muscles 6 and 7 in two adjacent segments of control and *mis12*^{f03756} homozygous embryos. Nuclei were stained with Hoechst dye (blue) and innervating axons with anti-HRP (Magenta). (H) Quantification of nuclei in muscle 6; unpaired t test with Welch's correction. Scale bar 20µm.

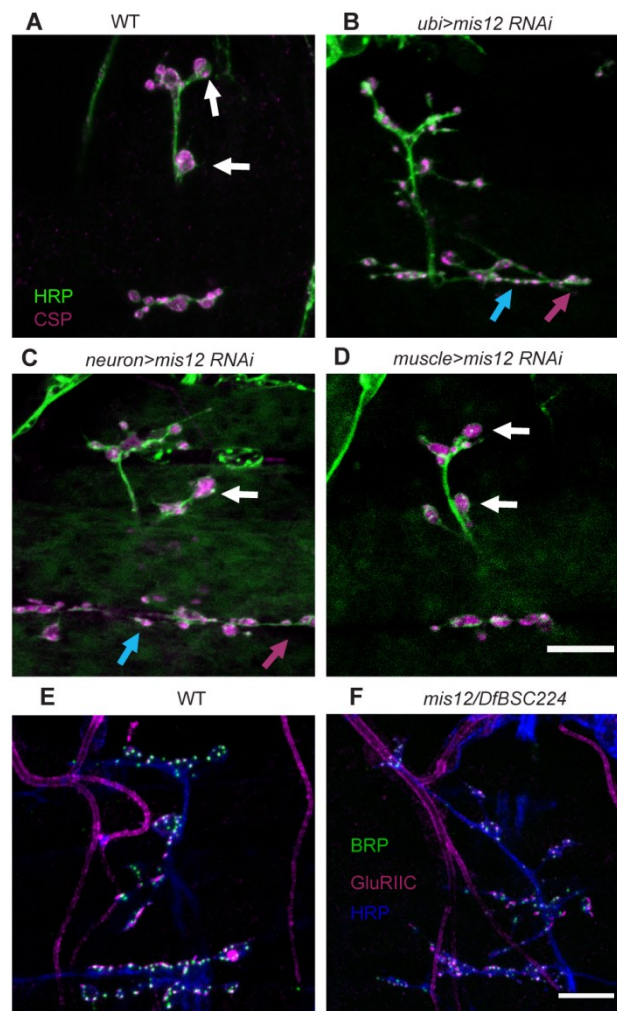


Figure S2. Knockdown of *mis12* ubiquitously or selectively in neurons causes NMJ growth defects and further characterization of *mis12* synapses, related to Figure 1

(A-D) Embryonic neuromuscular junctions on muscles 6, 7, 12, and 13 stained with anti-HRP

and anti-CSP. RNAi against *mis12* expressed either ubiquitously (B) or in neurons (C), but not in muscle (D), gave rise to overly long neurites (magenta arrows) with many small clusters of vesicles (blue arrows) in place of the normal large boutons (white arrows) as quantified in Figure 1. (E, F) Immunolocalization of the presynaptic active zone marker Brp (green) and postsynaptic receptor GluRIIC (magenta), co-stained with anti-HRP (blue) as a marker of neuronal membranes. Neuromuscular junctions on muscles 6, 7, 12, and 13 are shown for wild type (A) and *mis12*^{f03756}/*DfBSC224* embryos (B). Scale bar, 10 μ m.

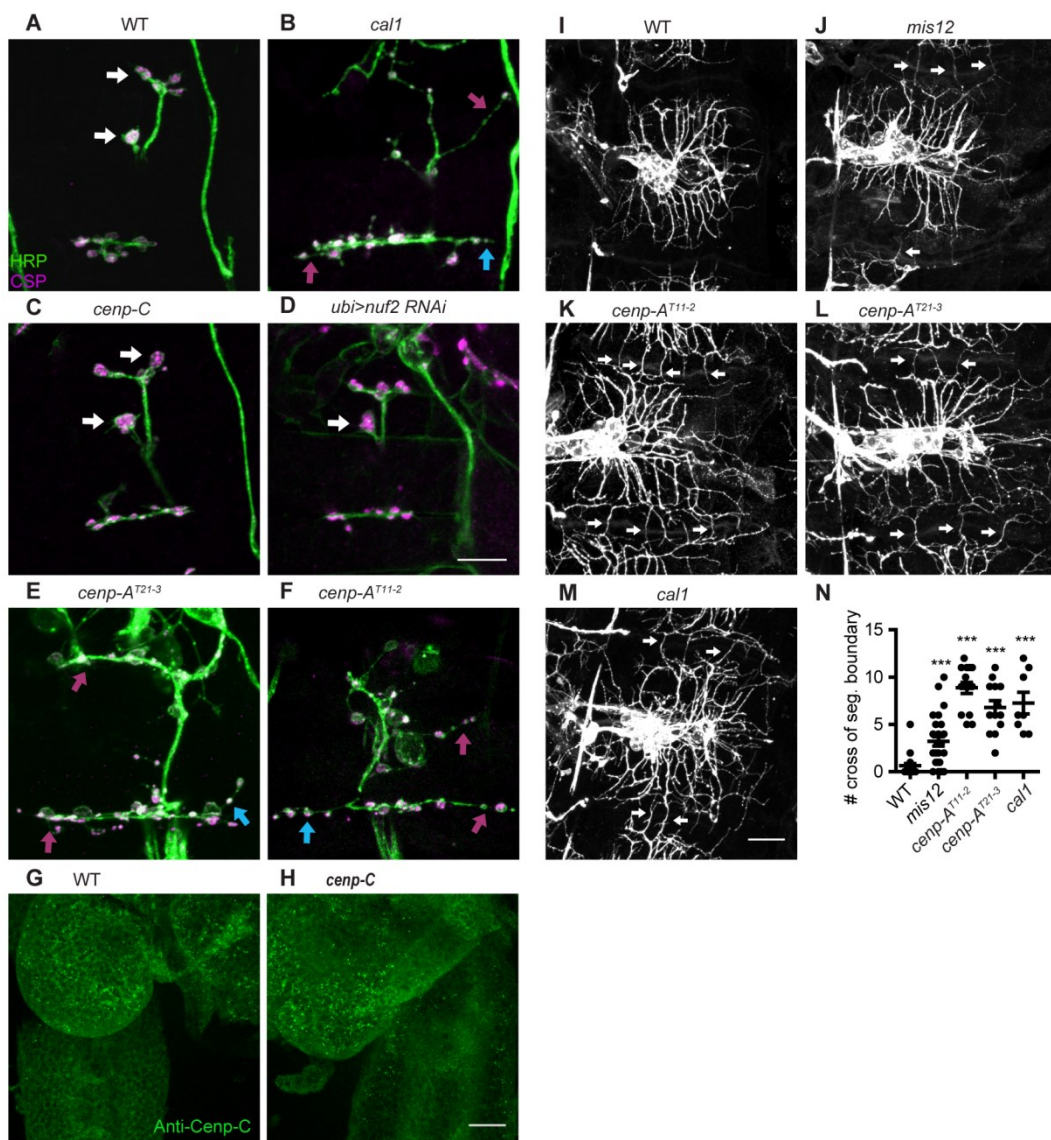


Figure S3. Kinetochores at the NMJ and in sensory neuron dendrites, related to

Figure 2

(A-F) anti-HRP and CSP staining of junctions on muscles 6,7,12, and 13, as in Figure 2, for Wild-type **(A)**, *cal1*^{c03646} **(B)**, *cenp-C*^{pr141} **(C)**, and ubiquitous RNAi knockdown of *nuf2* **(D)**, and *Cenp-A* (*cid*^{T21-3} and *cid*^{T11-2}) **(E,F)**. White arrows mark normal boutons. *Cenp-A* and *cal1* mutant embryos had overly long neurites (magenta arrows) and many small boutons (blue arrows), but these phenotypes were not present in *Cenp-C* mutants or the *nuf2* knockdown, genotypes that also retained the large boutons that characterize wildtype synapses. Quantification appears in Figure 2. **(G,H)** Cenp-C staining of embryonic brain indicates that extensive maternally contributed Cenp-C protein perdures into late stages of embryonic development (21 h AEL) in *cenp-C*^{pr141} and likely offers an explanation for the lack of a synaptic phenotype in that genotype. **(i-M)** anti-HRP staining of dorsal sensory neuron clusters for Wild-type **(I)**, *mis12*^{f03756} **(J)**, *Cenp-A* (*cid*¹¹⁻²) **(K)**, *cenp-A* (*cid*^{T21-3}) **(L)** and *cal1*^{c03646} **(M)**. White arrows indicate dendrites that cross the segment boundary. **(N)** Quantification of the number of dendrites that crossed the segment boundary in the indicated genotypes. Each genotype was compared with WT using one-way ANOVA with Dunnett's multiple comparisons test. ****p*<0.001. Error bars, mean ± s.e.m. Scale bar, 10 μm (A-F and I-M) and 20 um (G, H).

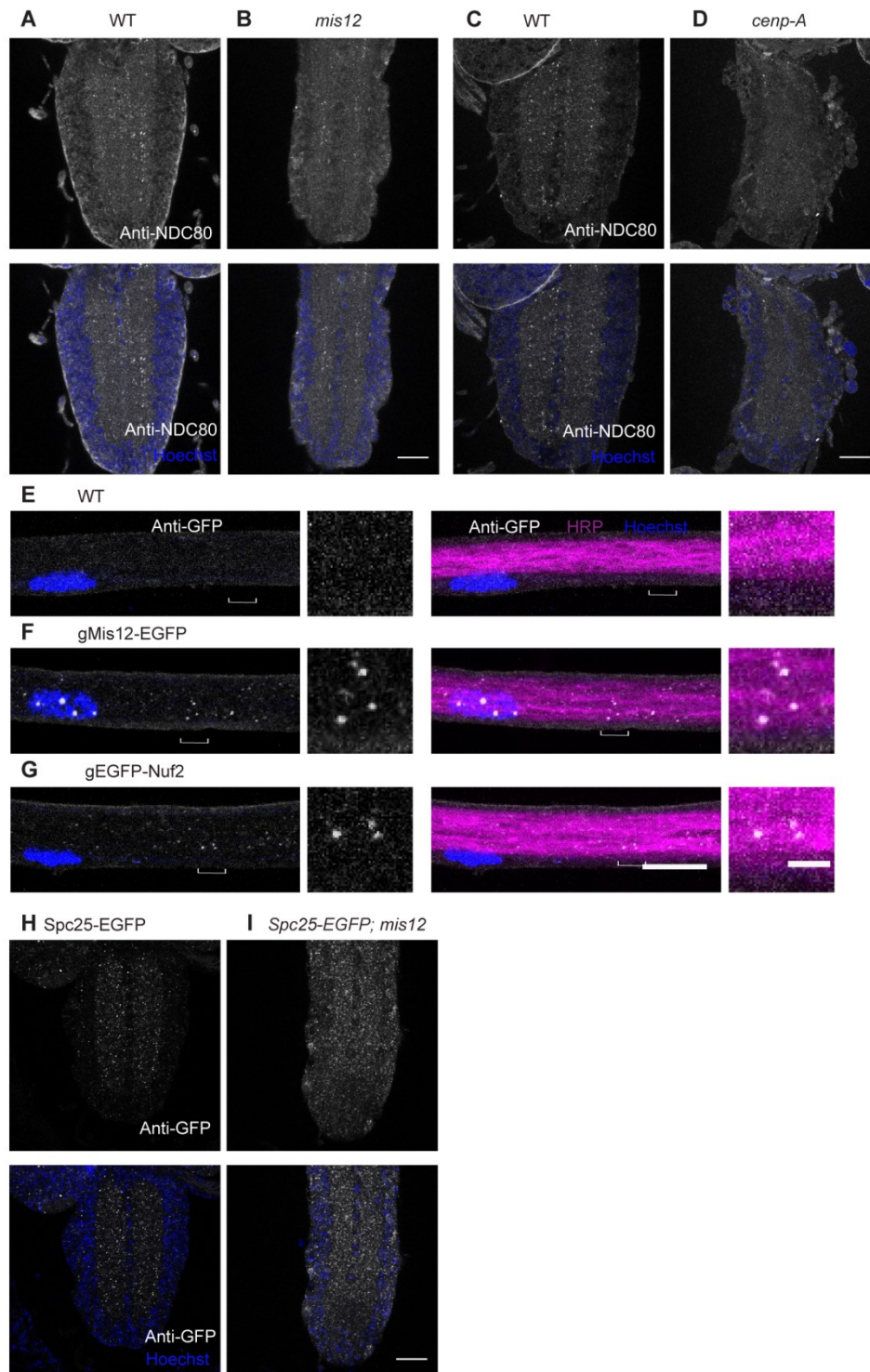


Figure S4. Kinetochores protein localization in VNC and axons, related to Figure 3

(A-D) Ventral nerve cords stained with anti-Ndc80 (white) and Hoechst dye (blue). In wildtype embryos Ndc80 puncta are seen in the synaptic neuropil. This signal is reduced in

embryos mutant for *mis12* **(B)** or *cenp-A/cid* **(D)**. Scale bar, 20 μm . **(E-G)** In axons of third instar larvae, Mis12-EGFP **(F)** and EGFP-Nuf2 **(G)** formed sparse puncta (white) in axonal cytoplasm. The Mis12 signal was not as bright as those present, and presumably centromeric, in nearby Hoechst-stained (blue) glial nuclei. Axons are labelled with anti-HRP (magenta). Right panels show enlargements of the axonal regions marked by brackets. Scale bar, 10 μm , 2 μm in right-hand panels. **(H,I)** EGFP tagged Spc25 was expressed under control of its endogenous promoter. In the neuropil of the embryonic ventral nerve cord Spc25-EGFP formed bright puncta of GFP immunoreactivity (white) in a region devoid of nuclei (Hoechst dye, blue). In a *mis12* mutant background, the GFP-tagged spc25 was more abundant and broadly distributed, including the retention of spc25 in the cell bodies. Scale bar, 20 μm .