

## Figures

**Fig S1** TraDIS results at *K. pneumoniae* capsule loci

**Fig S2** Further validation of TraDIS hits

**Fig S3** Flow cytometry-based analysis of C3b-positive cell populations

**Fig S4** Flow cytometry-based analysis of C5b-9-positive cell populations, and cytoplasmic marker release from B5055  $\Delta rfaH$

## Tables

*Included in this document*

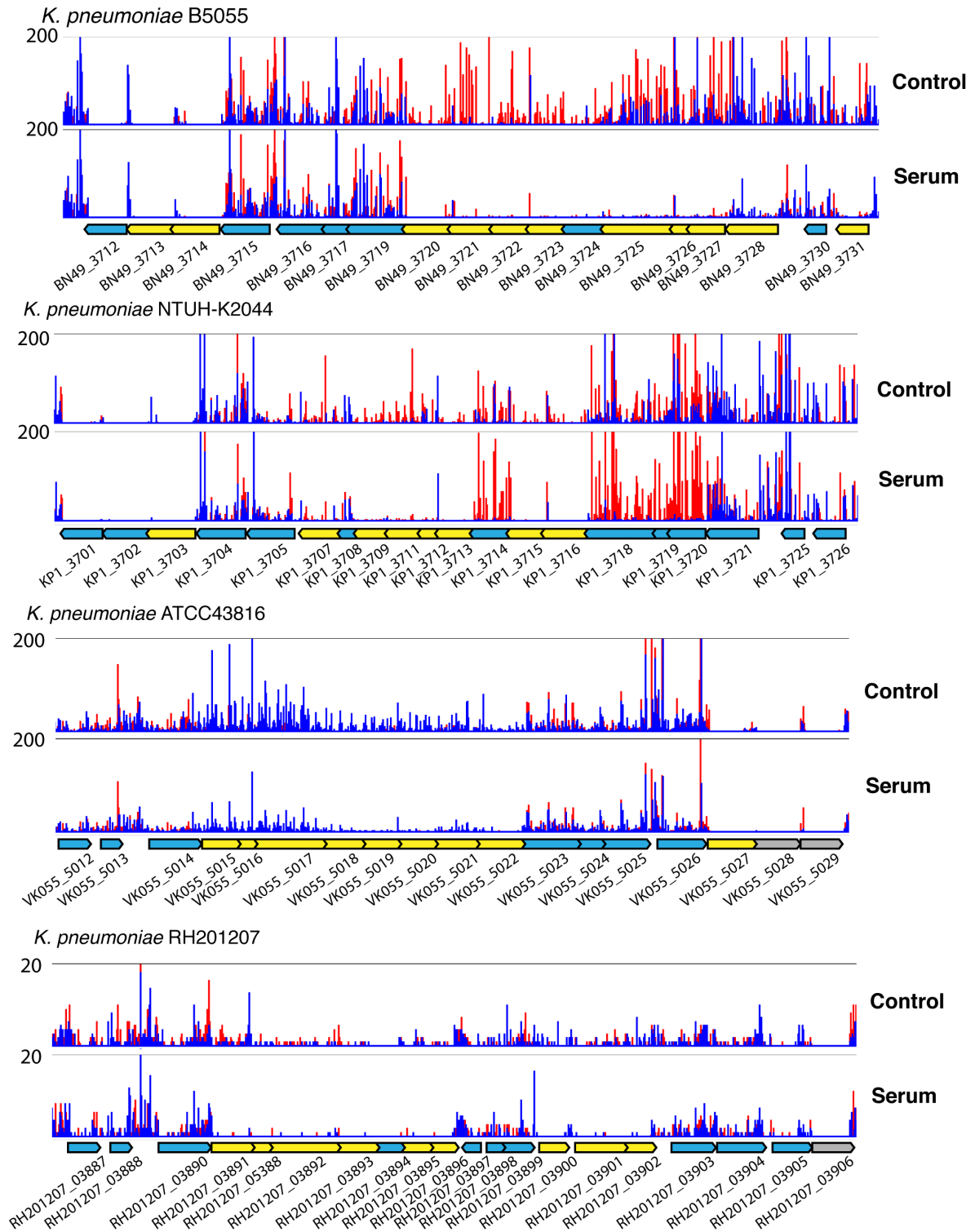
**Table S1** Strains, plasmids and primers used in this study

*Separate documents*

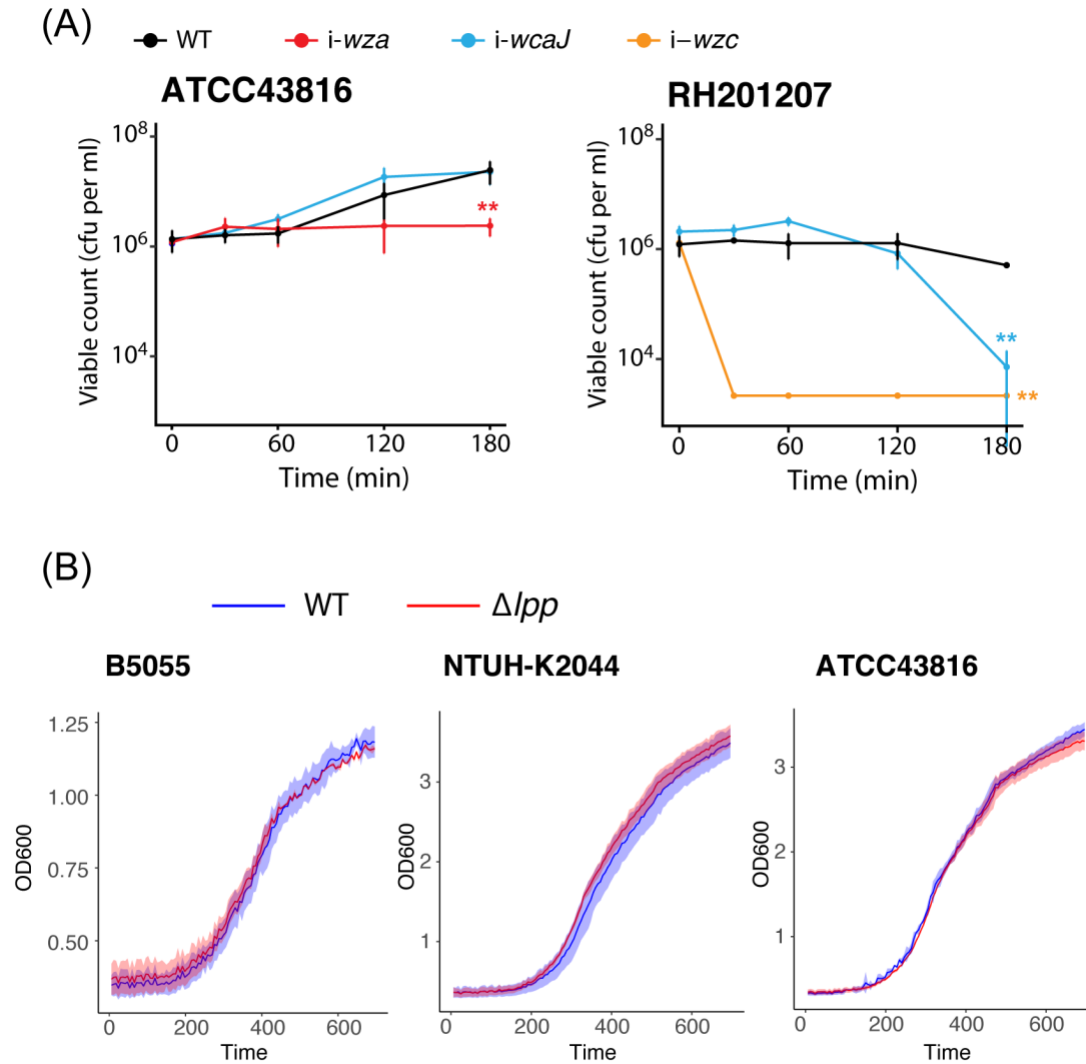
**Table S2** Sequencing data summary

**Table S3** TraDIS full results for four *K. pneumoniae* strains

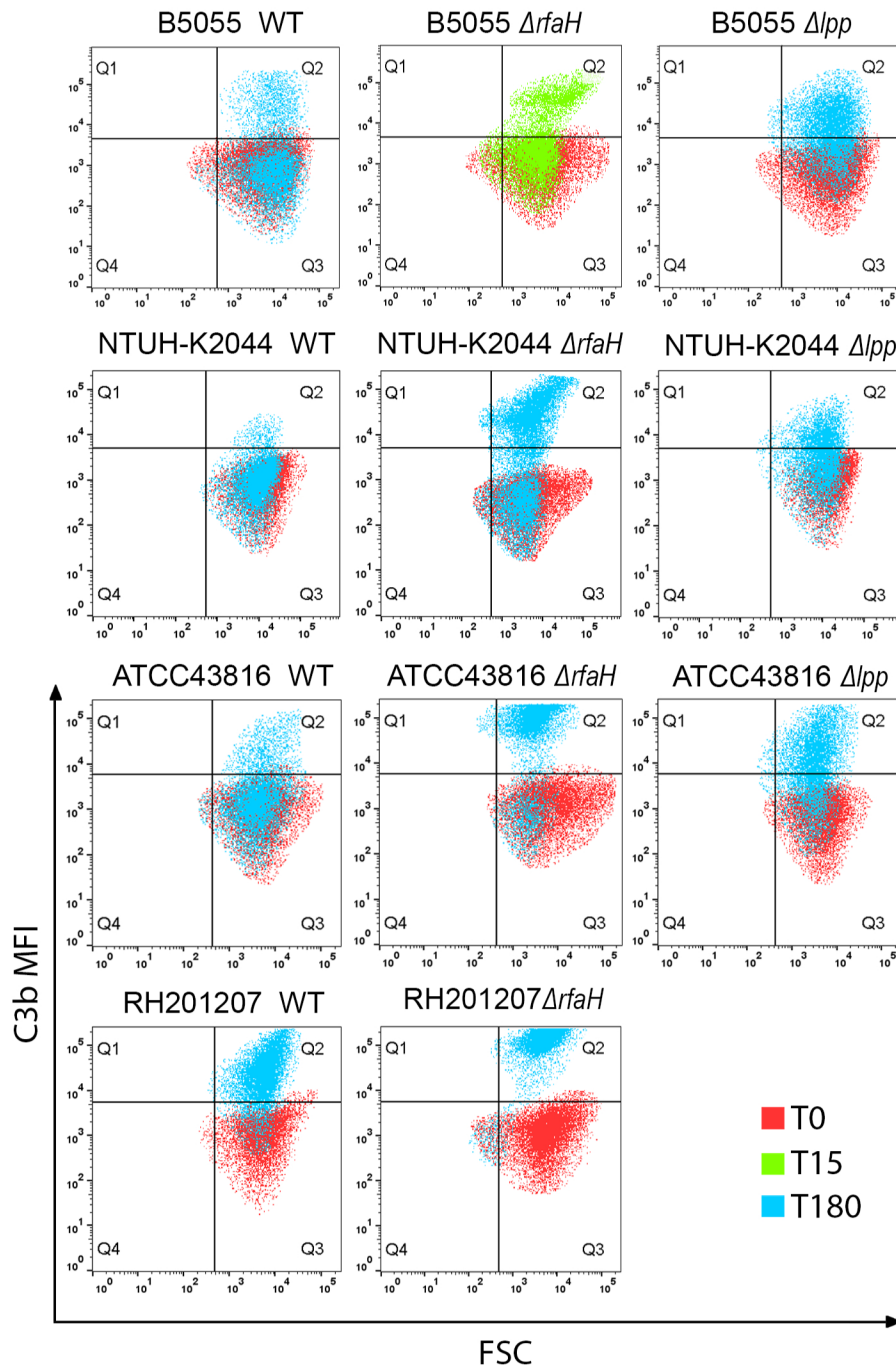
**Table S4** Genes identified as complement resistance determinants by TraDIS



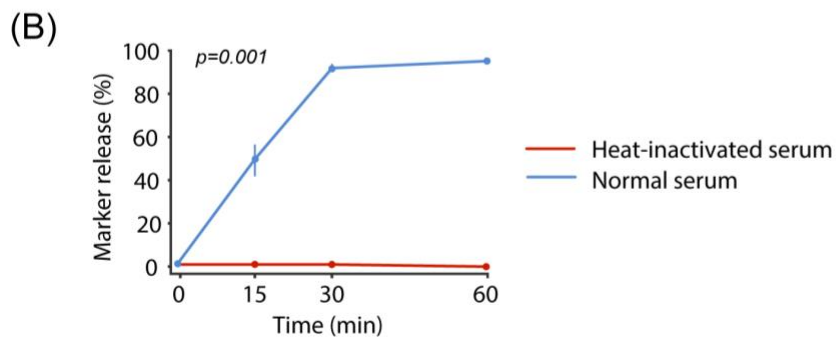
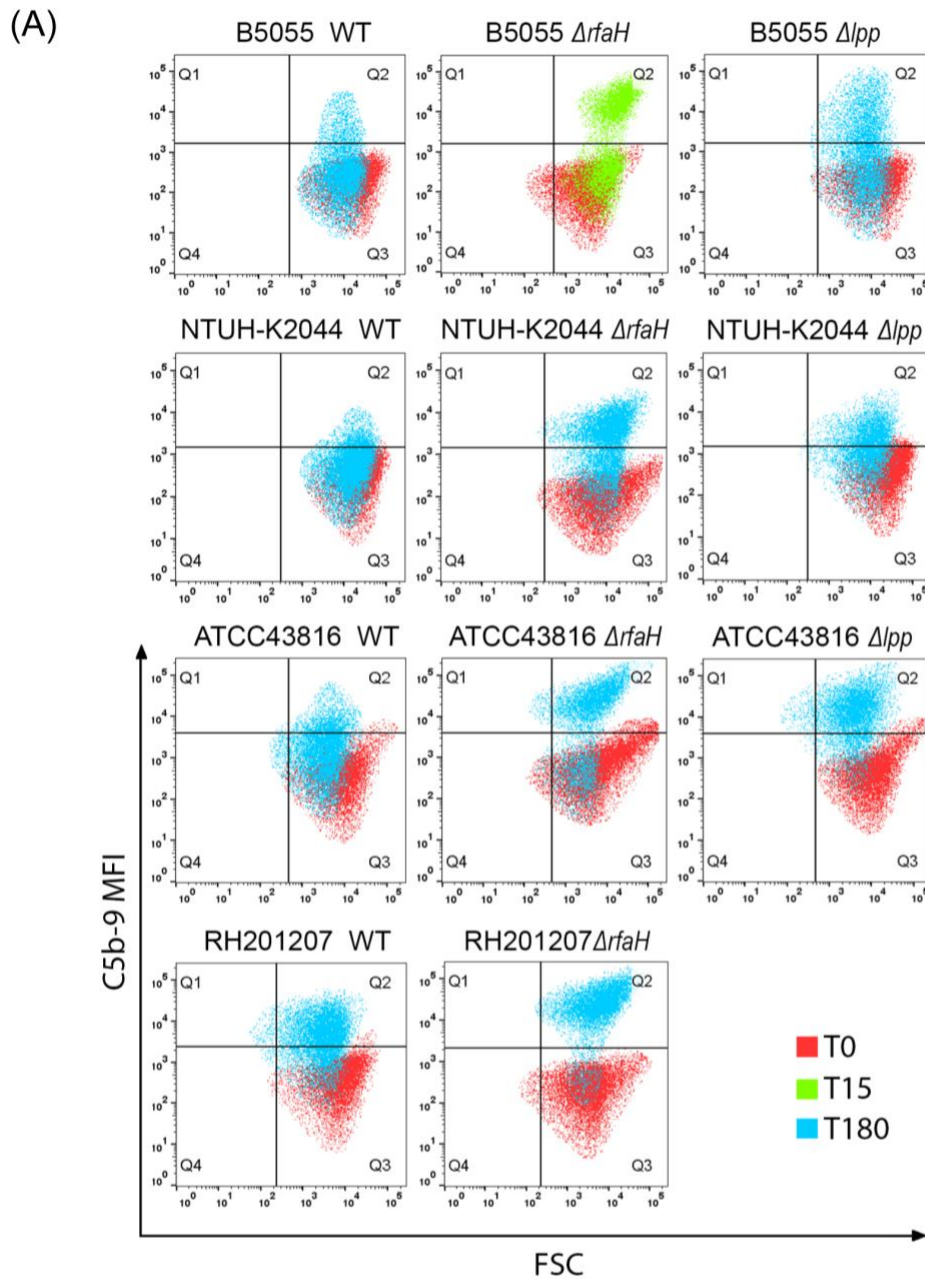
**Fig S1 TraDIS results at *K. pneumoniae* capsule loci** Insertion plots for TraDIS sequencing at the capsule locus of each strain. Read numbers correspond to the height of the vertical bars, which are coloured blue for plus strand and red for minus strand, respectively. Genes are coloured yellow for hits, grey for essential, and blue for not hits.



**Fig S2 Further validation of TraDIS hits** (A) Serum killing assays with additional capsule locus mutants of ATCC43816 ( $n = 5$ ) and RH201207 ( $n = 3$ ). Exponential phase cells were incubated with 66% human serum and survival monitored over 3 hours. Statistical significance as determined by two-factor repeated measures ANOVA was  $p < 0.0001$  for both ATCC43816 and RH201207 (note that these data are derived from the same set of experiments as that shown in Fig 4, and were analysed together). Strain-wise differences relative to wild type at  $t=180$  were determined by one-way ANOVA followed by Dunnett's test.  $**p < 0.01$  (B) Growth of *K. pneumoniae*  $\Delta lpp$  mutants ( $n = 2, 7$  technical replicates each). Strains were grown with shaking at  $37^\circ\text{C}$  in a 96-well plate ( $200\mu\text{l/well}$ ) in a Pherastar plate reader, and OD600 measured every 6 minutes. Growth of WT and  $\Delta lpp$  strains was compared by two-factor repeated measures ANOVA and was not found to be significant.



**Fig S3 Flow cytometry-based analysis of C3b-positive cell populations** Flow cytometry overlay plots illustrate the increase in C3b levels on the bacterial cell surface after 15 min (T15) or 180 min (T180) incubation in human pooled serum at 37°C, compared to T0. C3b positive populations are shown in Q2 quadrants. Data for the four *K. pneumoniae* wildtypes and  $\Delta rfaH$  and  $\Delta lpp$  mutants.



**Fig S4** (A) Flow cytometry-based analysis of C5b-9-positive cell populations. Flow cytometry overlay plots illustrate the increase in C5b-9 formation on the bacterial cell surface after 15 min (T15) or 180 min (T180) incubation in human pooled serum at 37°C, compared to T0. C5b-9 positive populations are shown in Q2 quadrants. Data for the four *K. pneumoniae* wildtypes and  $\Delta rfaH$  and  $\Delta lpp$  mutants. (B) Release of cytoplasmic GFP from B5055  $\Delta rfaH$  carrying the pFLS21 fluorescence marker

vector following incubation in normal or heat-inactivated human serum (n = 2). Results were compared by two-way repeated measures ANOVA.

**Table S1** Strains, plasmids and primers used in this study

<b>Bacterial strains</b>		
<b>Strain</b>	<b>Description/genotype</b>	<b>Source</b>
<i>Klebsiella pneumoniae</i> ATCC 43816	Hypervirulent, ST493, K-type 2, commonly used in mouse studies	American Type Culture Collection; <sup>1</sup>
<i>Klebsiella pneumoniae</i> NTUH-K2044	Hypervirulent, liver abscess isolate, ST23, K-type 1 reference strain	Jin-Town Wang, National Taiwan University Hospital; <sup>2</sup>
<i>Klebsiella pneumoniae</i> B5055	Mouse-virulent sequence type 66, K-type 2 reference strain	<sup>3</sup>
<i>Klebsiella pneumoniae</i> RH201207	Colistin-resistant UK gut isolate, ST258, K-type 106	Public Health England; <sup>4</sup>
<i>Escherichia coli</i> $\beta$ 2163	F <sup>-</sup> RP4-2-Tc::Mu $\Delta$ dapA::( <i>erm-pir</i> )	<sup>5</sup>
<i>K. pneumoniae</i> ATCC43816	$\Delta$ <i>lpp</i>	This study
<i>K. pneumoniae</i> ATCC43816	$\Delta$ <i>rfaH</i>	<sup>6</sup>
<i>K. pneumoniae</i> ATCC43816	Tn- <i>wza</i>	This study
<i>K. pneumoniae</i> ATCC43816	Tn- <i>wcaJ</i>	This study
<i>K. pneumoniae</i> B5055	$\Delta$ <i>lpp</i>	This study
<i>K. pneumoniae</i> B5055	$\Delta$ <i>rfaH</i>	This study
<i>K. pneumoniae</i> NTUH-K2044	$\Delta$ <i>lpp</i>	This study
<i>K. pneumoniae</i> NTUH-K2044	$\Delta$ <i>rfaH</i>	<sup>6</sup>
<i>K. pneumoniae</i> RH201207	$\Delta$ <i>rfaH</i>	This study
<i>K. pneumoniae</i> RH201207	Tn5- <i>wza</i>	This study
<i>K. pneumoniae</i> RH201207	Tn5- <i>wzc</i>	This study
<b>Plasmids</b>		
<b>Name</b>	<b>Description</b>	<b>Source</b>
pDS1028	Transposon delivery vector, Tn5-Cm <sup>R</sup> <i>ori6K</i> , pRL27-derived	<sup>7</sup>
pKNG101-Tc	Allelic exchange vector, Tc <sup>R</sup>	<sup>5</sup>
pBAD33	Overexpression vector, Ara-inducible, Cm <sup>R</sup>	<sup>8</sup>
pFLS16	pKNG101-Tc derivative, ATCC43816 <i>rfaH</i> flanking region	<sup>6</sup>
pFLS21	pDiGc derivative containing Tc <sup>R</sup> cassette from	
pFLS31	pKNG101-Tc derivative, ATCC43816/B5055 <i>lpp</i> flanking region	This study
pFLS33	pKNG101-Tc derivative, NTUH <i>lpp</i> flanking region, constructed with FS342-345	This study
pFLS34	pKNG101-Tc derivative, RH201207 <i>rfaH</i> flanking region, constructed with FS136-FS139	This study
pFLS37	pBAD33 derivative, ATCC43816 <i>lpp</i> gene, no promoter	This study
pFLS38	pBAD33 derivative, ATCC43816 <i>lpp</i> gene $\Delta$ K78, no promoter	This study
pMJD82	pACYC184 derivative, NTUH-K2044 <i>rfaH</i> gene with native promoter	This study
<b>Primers</b>		
<b>Name</b>	<b>Sequence 5' – 3'</b>	<b>Description</b>
FS57	gaccacacgtcgactagtgcnnnnnnnnnnnagag	Random-prime PCR
FS58	gaccacacgtcgactagtgcnnnnnnnnnnnagcc	Random-prime PCR
FS59	gaccacacgtcgactagtgcnnnnnnnnnnngatac	Random-prime PCR
FS60	gaccacacgtcgactagtgc	Random-prime PCR
FS108	aatgatacggcgaccaccgagatctacaccaggaaacttaacggctgacatgg	pDS1028 Tn 1
FS109	agcctctcaaagcaattttgagtgaca	pDS1028 Tn 2
FS136	tgtgactagtagcgccgagcgcctct	RfaH KO Fwd
FS137	tgacattaacggcgtaataactcattccactc	RfaH KO Rev overlap
FS138	tgtgactagtagcggtagcagctgga	RfaH KO Rev
FS139	gtggaatgagtcattacgccgtaaatgtcaa	RfaH KO Fwd overlap
FS342	gctactagtcgctgtgctgcaatac	Lpp KO Fwd
FS343	tagagggtattaatagattctgtaataaaaatgg	Lpp KO overlap 1
FS344	ttattaccagaactctattaataccctctagat	Lpp KO overlap 2
FS345	gctactagtcggccggaccgaacaa	Lpp KO Rev
FS335	ctctggtaccctcggggcgctttt	Lpp into pBAD33 Fwd, KpnI
FS336	ctctaagcttttacttacggtaagagtgagc	Lpp into pBAD33 Rev, HindIII
FS337	ctctaagcttttaccgtaagagtgagcctg	Lpp $\Delta$ C-terminal Lysine into pBAD33 Rev, HindIII
FS346	cctgatcagggttgagatgtgtataagacag	RH201207 Tn primer 1
FS347	attgtagcgccgccctatacctt	RH201207 Tn primer 2
MJD112	ccggatccggtaacggcgcgatgctgtaattttgtaaattag	RfaH Fwd
MJD113	ccgtcgacattagattttctggaagtcggtatttttaac	RfaH Rev

1. Broberg, C.A., Wu, W., Cavalcoli, J.D., Miller, V.L., Bachman, M.A. 2014. Complete Genome Sequence of *Klebsiella pneumoniae* Strain ATCC 43816 KPPR1, a Rifampin-Resistant Mutant Commonly Used in Animal, Genetic, and Molecular Biology Studies. *Genome Announcements*. 2 (5), e00924-14-e00924-14
2. Wu, K.M. et al. 2009. Genome sequencing and comparative analysis of *Klebsiella pneumoniae* NTUH-K2044, a strain causing liver abscess and meningitis. *Journal of Bacteriology*. 191 (14), 4492–4501
3. Nassif, X., Sansonetti, P.J. 1986. Correlation of the virulence of *Klebsiella pneumoniae* K1 and K2 with the presence of a plasmid encoding aerobactin. *Infection and immunity*. 54 (3), 603–608
4. Jana, B. et al. 2017. The secondary resistome of multidrug-resistant *Klebsiella pneumoniae*. *Scientific Reports*. 7, 42483
5. Demarre, G., Guérout, A.M., Matsumoto-Mashimo, C., Rowe-Magnus, D.A., Marlière, P., Mazel, D. 2005. A new family of mobilizable suicide plasmids based on broad host range R388 plasmid (IncW) and RP4 plasmid (IncP $\alpha$ ) conjugative machineries and their cognate *Escherichia coli* host strains. *Research in Microbiology*. 156 (2), 245–255
6. Dorman, M.J., Feltwell, T., Goulding, D.A., Parkhill, J., Short, F.L. 2018. The Capsule Regulatory Network of *Klebsiella pneumoniae* Defined by density-TraDISort. *mBio*. 9 (6), doi: 10.1128/mBio.01863-18
7. Monson, R. et al. 2015. A Plasmid-Transposon Hybrid Mutagenesis System Effective in a Broad Range of Enterobacteria. *Frontiers in Microbiology*. 6, 1442
8. Guzman, L.M., Belin, D., Carson, M.J., Beckwith, J. 1995. Tight regulation, modulation, and high-level expression by vectors containing the arabinose PBAD promoter. *Journal of bacteriology*. 177, 4121–4130