Gasdermin-D-dependent IL-1 α release from microglia promotes protective immunity

during chronic Toxoplasma gondii infection

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Supplementary Figure 1 I Microglia and macrophages in the infected brain differ in IL-1 expression. ad, $CX_3CR_1^{Cre-ERT2}$ x ZsGreen^{fl/stop/fl} mice were left naïve or infected with 10 cysts of Me49 strain *T. gondii* parasites for 4 weeks. (n = 4 mice per group) **a**, Representative image of IL-1 α in naïve brain colocalizing with microglia, scale bar is 50 µm. **b-d**, Brains were harvested and analyzed by flow cytometry with intracellular cytokine staining. Data are presented as mean values +/- SEM. Numbers of IL-1 α^+ (**b**), IL-1 β^+ (**c**), and double positive (**d**) cells were quantified in both ZsGreen⁺ and ZsGreen⁻ populations in naïve and infected mice. Cells were pre-gated on singlets/live/ZsGreen. **e-g**, Brains from naïve or chronically infected mice were analyzed by flow cytometry. **e**, Representative plots of IL-1 α expression for naïve samples, previously gated on live/singlets. **f-g**, Representative plots of IL-1 α (**f**) and IL-1 β (**g**) expression for infected samples.



Supplementary Figure 2 I Brain IFN-*γ* **responses and peripheral immune responses are not impaired in IL-1R1 KO mice during chronic** *T. gondii* **infection.** WT and IL-1R1 KO mice were infected i.p. with 10 cysts of the Me49 strain of *T. gondii*. 6 weeks p.i. brains (**a-b**) and spleens (**c-h**) were harvested and processed for flow cytometry. Immune cell populations were enumerated. **a-b**, Brains were harvested and digested. Isolated cells were incubated at 37°C for 5 hours with a mix of PMA/ionomycin and brefeldin A. Intracellular cytokine staining was performed and analyzed by flow cytometry. Cells were pre-gated on singlets/Live/CD3+ and percent (**a**) and number (**b**) of IFN-*γ*⁺ CD4 and CD8+ T cells were determined. **c**, Total immune cells, pre-gated on singlets/live, p = 2.64x10⁻⁶ **d**, DCs, pre-gated on singlets/live/CD45+/Dump⁻(CD3/NK1.1/B220), p = 6.19x10⁻⁴ **e**, Monocytes/ macrophages, pre-gated on singlets/live/CD45+/CD11c⁻/CD11b⁺/CD45^{hi}, p = 0.001 **f**, CD8+ T cells, pre-gated on singlets/live/CD3⁺, p = 5.52x10⁻⁴ **h**, Tregs, pre-gated on singlets/live/CD3⁺, p = 9.63x10⁻⁴ **a-b**, A representative experiment is shown and statistics were performed using a two-tailed Student's T test. (n = 9 mice) Data are presented as mean values +/- SEM. **c-h**, Paired averages compiled from 3-6 experiments. Statistics were performed using a randomized block ANOVA. (n = 49 mice)



Supplementary Figure 3 I IFN- γ response and monocyte/macrophage response during acute infection are not impaired in IL-1R1 KO mice. WT and IL-1R1 KO mice were infected i.p. with 10 cysts of the Me49 strain of *T. gondii*. Mice were sacrificed 12 days p.i. **a**, Peritoneal lavage was performed and peritoneal exudate cells (PECs) were isolated and analyzed by flow cytometry. For Ly6G⁺ cells p = 0.03, for DCs p = 0.01. **b**, Spleen cells were isolated and analyzed by flow cytometry. **c**, Serum was harvested at the time of sacrifice and IFN- γ in the serum was analyzed by ELISA. A representative experiment is shown (n = 4 mice per group). Statistics were performed using a two-tailed Student's t-test between groups for each measure. Data are presented as mean values +/- SEM.



Supplementary Figure 4 I IL-1R1 is expressed by endothelial cells in the brain. a, Brains from chronically infected C57B6/J mice were harvested, fixed, and stained with antibodies against CD31 (red) and IL-1R1 (green). Representative of two independent experiments. **b-c**, Brains from uninfected C57B6/J mice were harvested and processed for flow cytometry analysis. Cells were previously gated on Singlets/Live/CD45⁻ and then were gated on CD31⁺ (**b**) and IL-1R1 (**c**) expression on the CD31⁺ population.



Supplementary Figure 5 | The brain endothelium is activated during chronic *T. gondii* infection. a-d, WT C57B6/J mice were either left naïve or infected i.p. with the Me49 strain of T. gondii. 4 weeks p.i. mice were sacrificed and brains were harvested for flow cytometry analysis. (n = 2 mice per group) a-b, Samples were pre-gated on singlets/live/Hoescht+/CD45-/CD31+ and then ICAM-1 expression was assessed. Representative plots from naïve (a) and infected (b) mice are shown. c-d, Samples were pregated as in a and then VCAM-1 expression was assessed. Representative plots from naïve (c) and infected (d) mice are shown. e, Histogram showing ICAM-1 expression on IL-1R1 positive and negative endothelial cells, the FMO is shown in filled gray f, Brains from chronically infected C57B6/J mice were harvested, fixed, and stained with antibodies against laminin (gray), IL-1R1 (red), and VCAM-1 (green). Scale bar = 50 μ m g-i, C57B6/J mice were infected i.p. with 10 cysts of the Me49 strain of *T. gondii.* 4 weeks p.i. mice were treated with either control IgG or 200 μ g each of α -LFA-1 and α -VLA-4 blocking antibodies on days 1 and 3 of treatment, and were sacrificed on day 5. Brains were harvested and processed for flow cytometry. (n = 9 mice) g, Cells were previously gated on singlets/live/CD11c /CD45+ and the numbers of CD11b+CD45^{hi} cells are shown. p = 0.004. Of the CD45^{hi} cells numbers of Ly6C^{hi} cells, p = 0.0001 (h) and iNOS⁺ cells, p = 0.005 (i) were enumerated. Statistics were performed using a two-tailed Student's T-test. Data are presented as mean values +/- SEM.



Supplementary Figure 6 I Example gating strategy for brain immune populations. Myeloid and T cell populations were identified using two separate panels. **a-b**, for all panels, samples were first gated on singlets and then cells which excluded the live/dead dye. **c-e**, to identify T cell populations, live cells were plotted to gate on either CD8+CD3+ (**c**) or CD4+CD3+ (**d**) cells. **e**, to identify Tregs, CD4+ T cells were gated on Foxp3+. **f-j**, to identify myeloid cell populations live cells were first gated on CD45+ (**f**). CD45+ cells were then gated on CD11c and MHCII (**g**), CD11c+MHCII hi cells were called DCs. CD11c- cells were then gated by CD45 and CD11b (**i**). CD11b+CD45hi cells were then gated on iNOS+ (**j**).