

Neuron, Volume 107

Supplemental Information

**MCH Neurons Regulate Permeability
of the Median Eminence Barrier**

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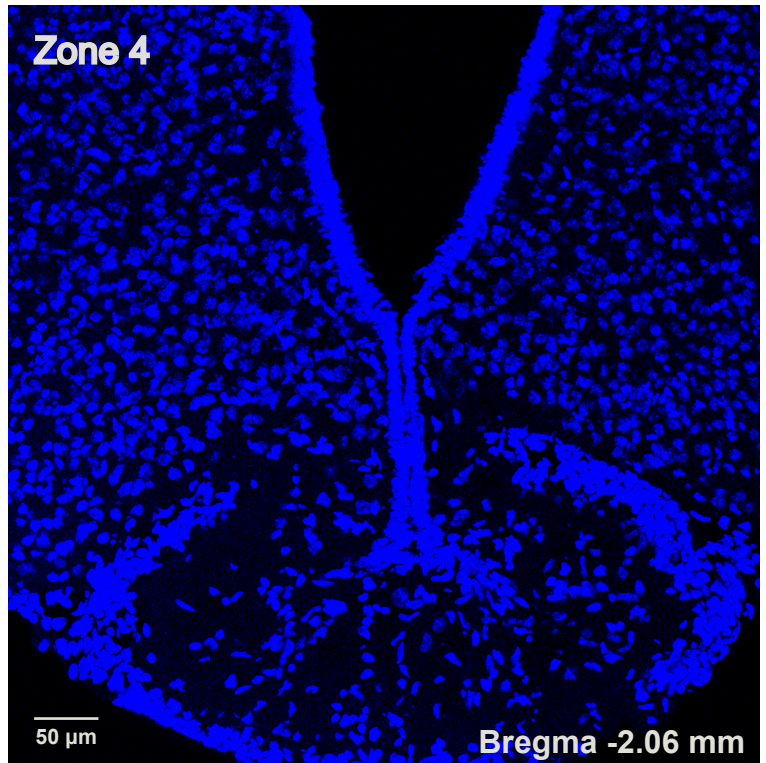
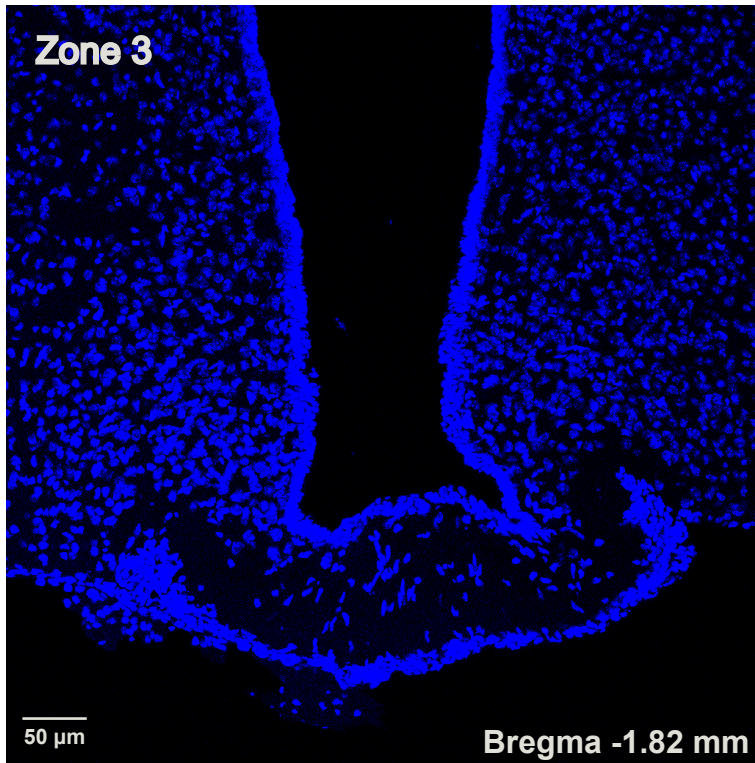
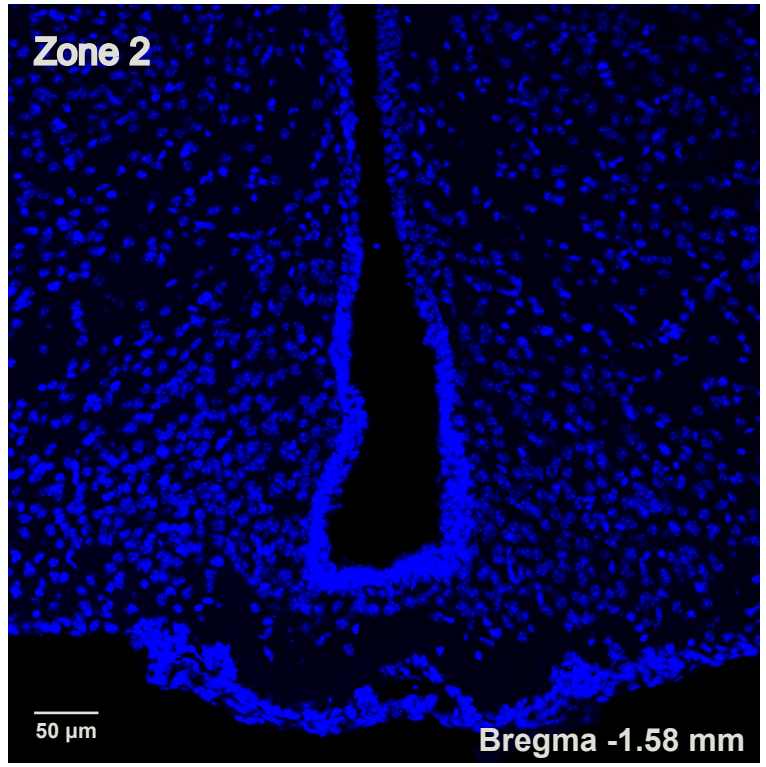
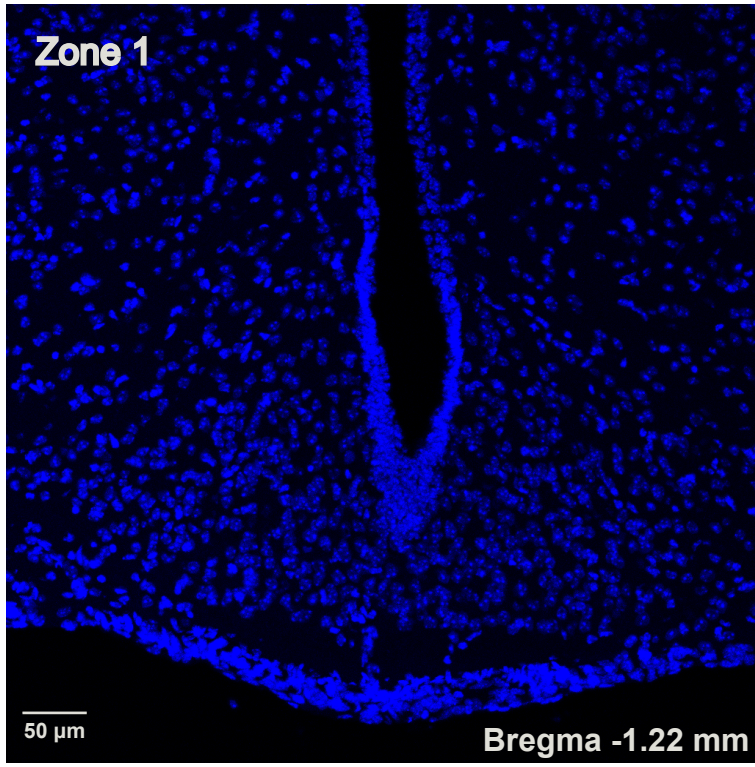


Figure S1. Related to Figure 1. Classification of four zones in the ARC. Representative anatomical location for each zone of the ARC from rostral to caudal. Scale bar: 50 μ m.

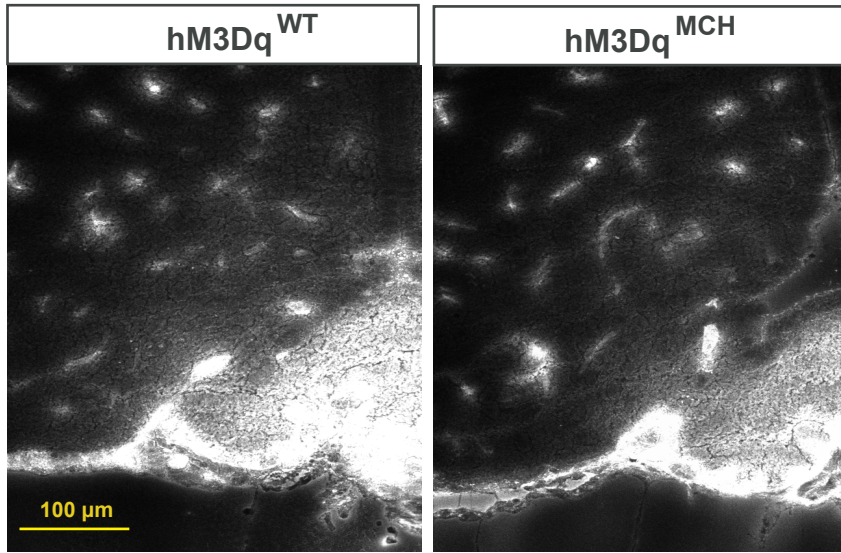
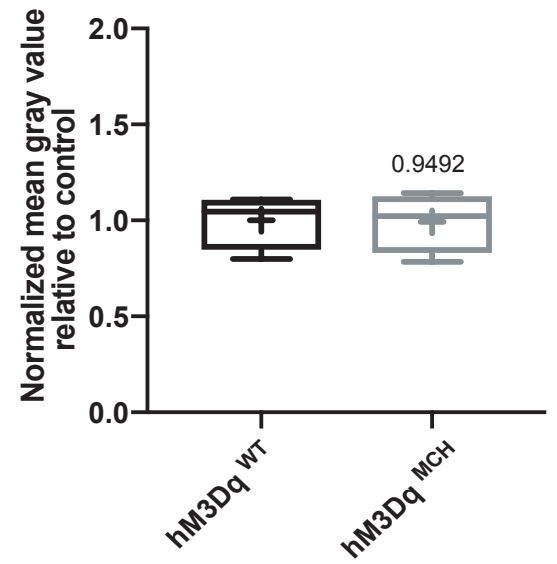
A**B**

Figure S2. Related to Figure 2. Evans blue diffusion showed no detectable difference in vehicle-treated mice. (A) Representative images of Evans blue dye diffusion into the ARC region of fasted and 4 hrs vehicle-injected control and hM3Dq^{MCH} mice. (B) Quantification of Evans blue dye diffusion in the ARC region in vehicle-treated control (n=4) and hM3Dq^{MCH} (n=4) mice. Statistics: unpaired Student's T-test.

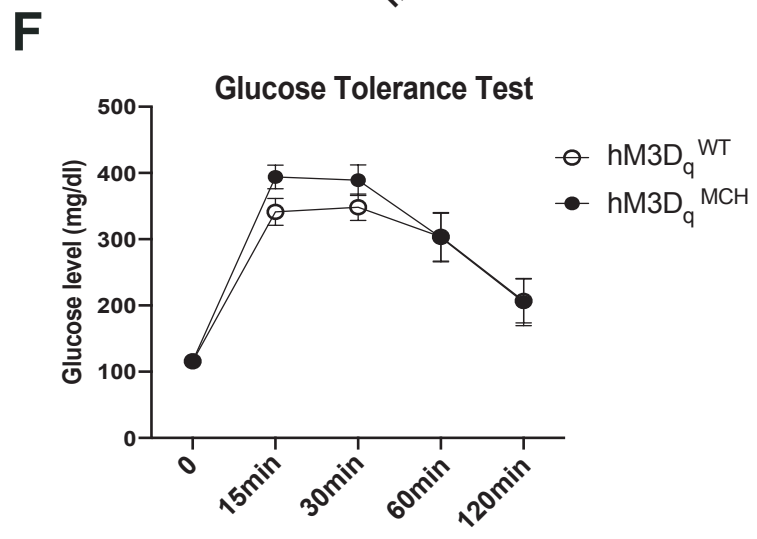
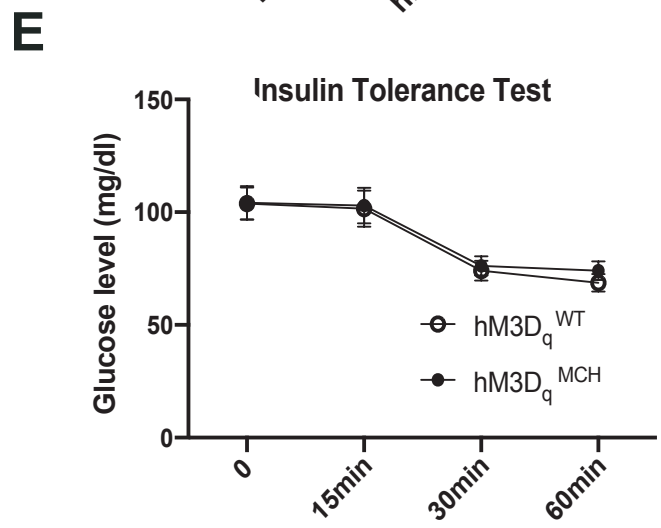
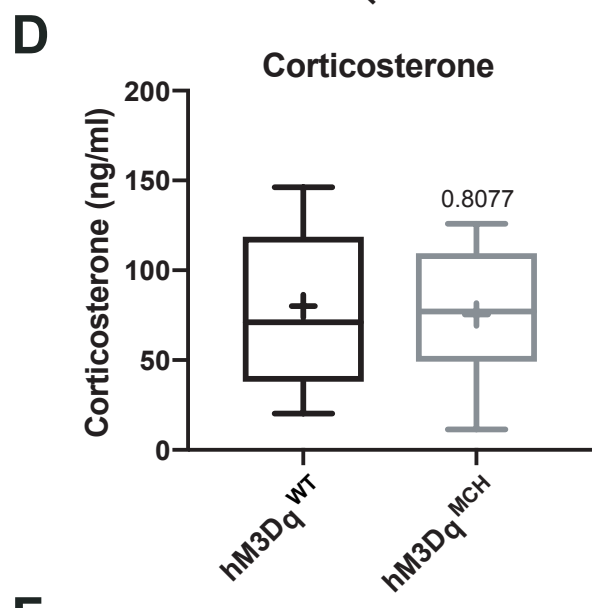
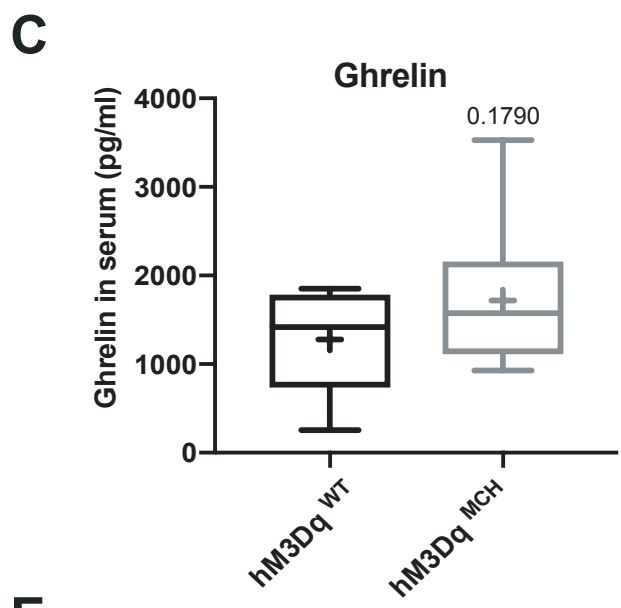
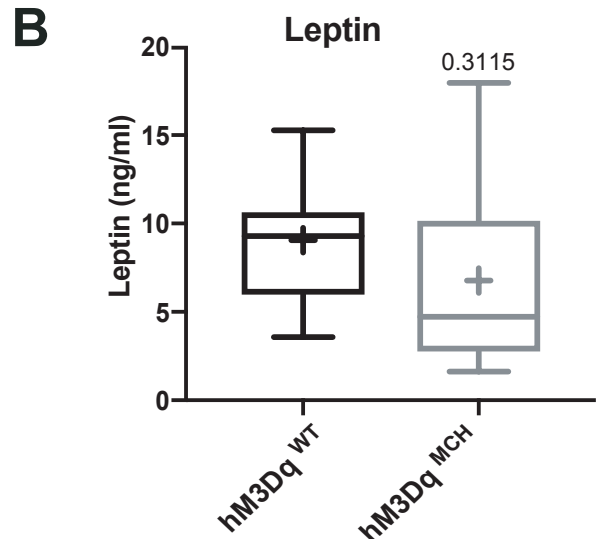
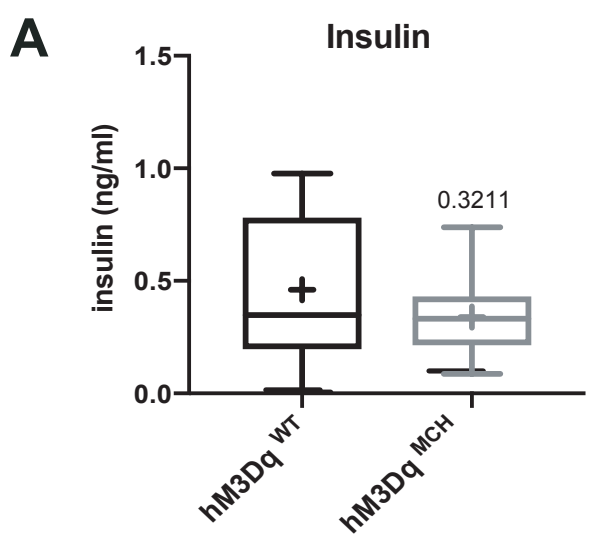


Figure S3. Related to Figure 2 and 3. Systemic metabolic parameters in CNO-treated control and hM3Dq^{MCH} mice. (A)-(D) Serum Insulin, Leptin, Ghrelin and Corticosterone levels of fasted and 4 hrs CNO-treated control (n=8-10) and hM3Dq^{MCH} mice (n=10-13). (E) and (F) Insulin tolerance test and Glucose tolerance test in fasted and 4 hrs CNO-treated control (n=10) and hM3Dq^{MCH} mice (n=13). Statistics: unpaired student T-test, for A-D; Two way ANOVA Sidack Post hoc test, for E and F.

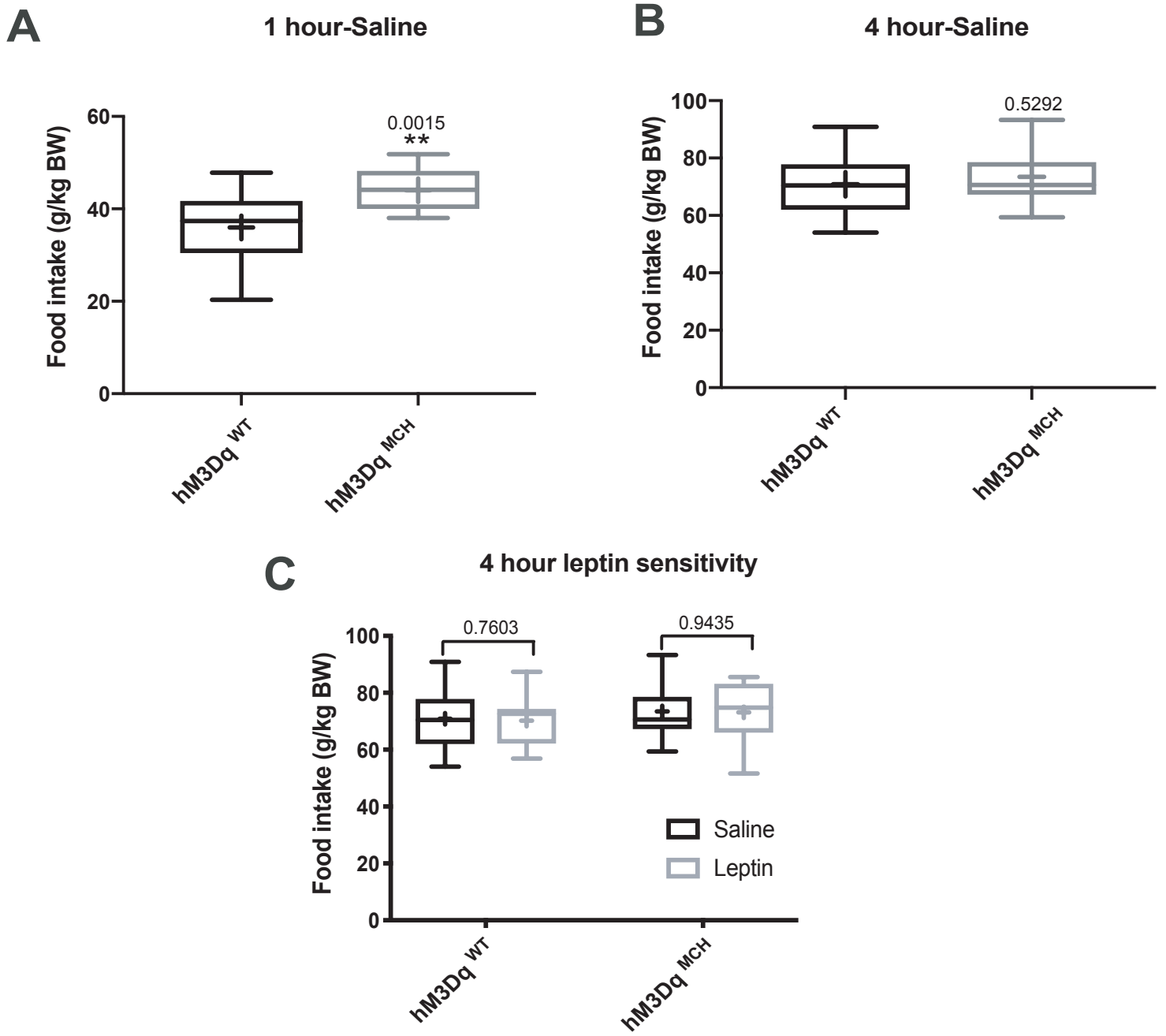


Figure S4. Related to Figure 4. Feeding regulation in hM3Dq^{MCH} mice . (A) and (B), 1 hr and 4 hrs food intake in fasted and 4 hrs CNO-treated control (n=13) and hM3Dq^{MCH} mice (n=13) with saline (leptin vehicle) injected at half hour before refeeding. (C), Leptin sensitivity at 4 hrs refeeding in fasted and 4 hrs CNO-treated control and hM3Dq^{MCH} mice. Statistics: unpaired Student's T-test, **p < 0.01, except paired Student's T-test in leptin sensitivity.

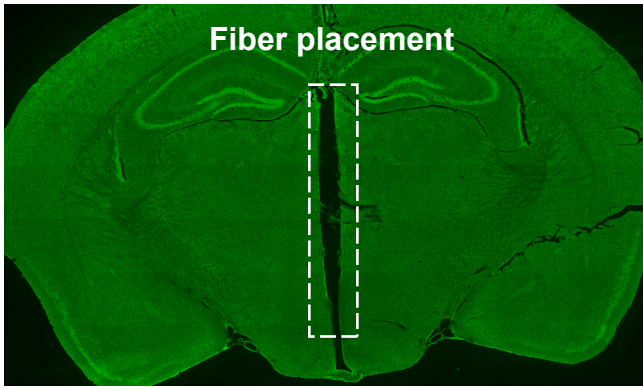
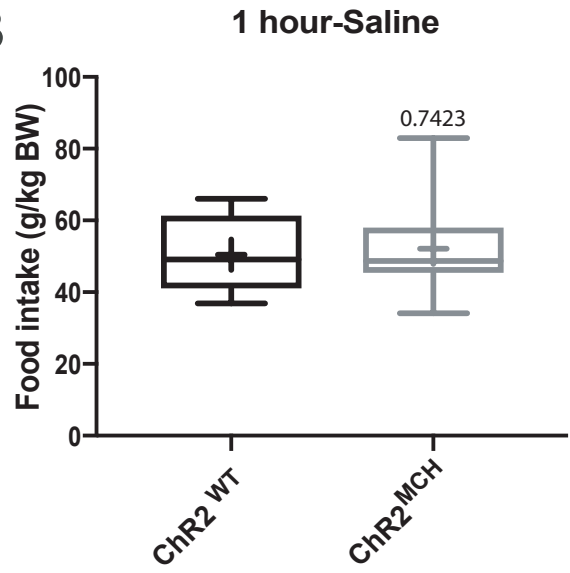
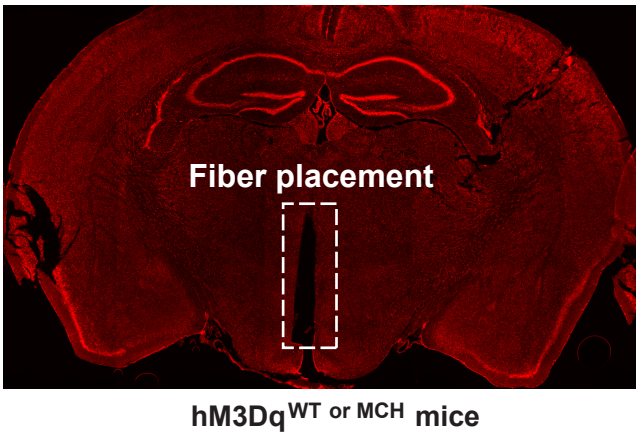
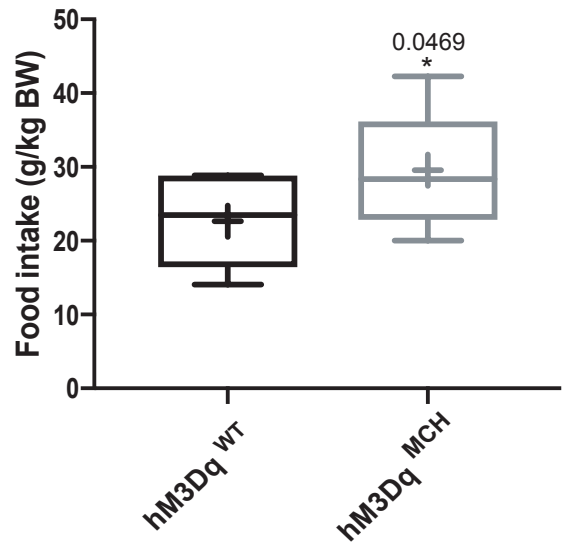
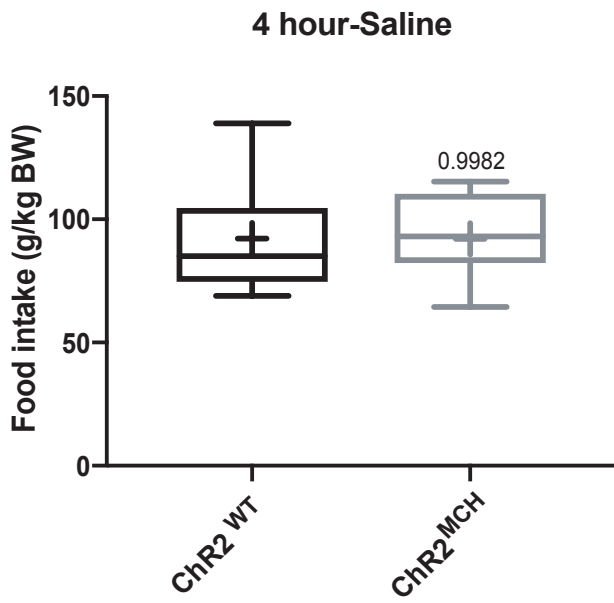
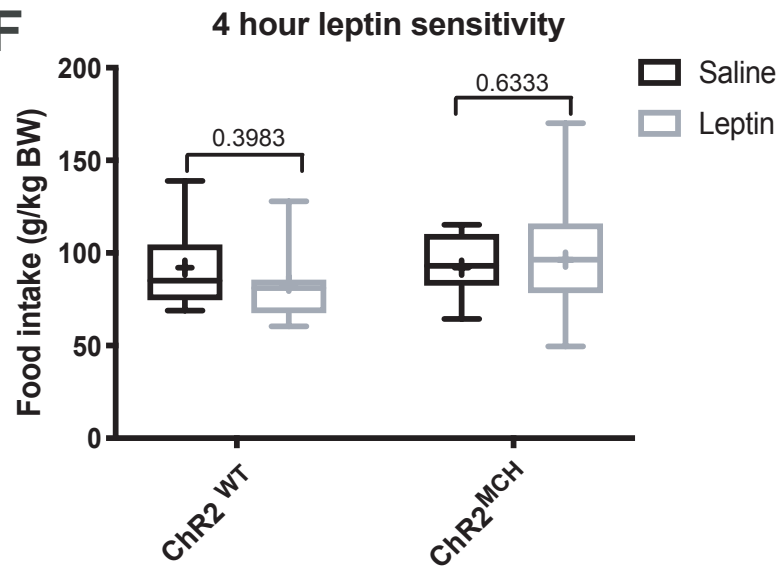
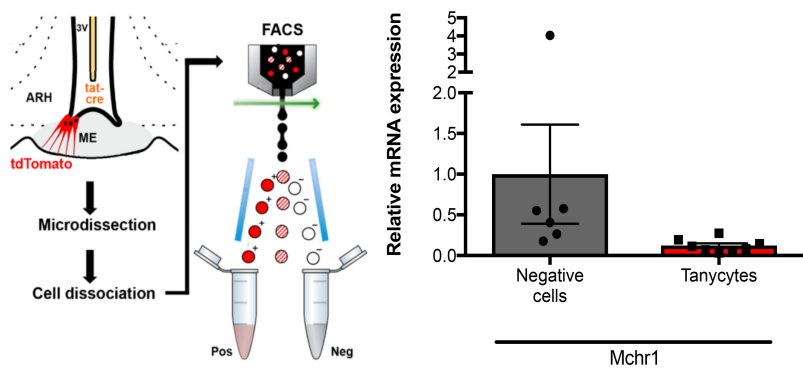
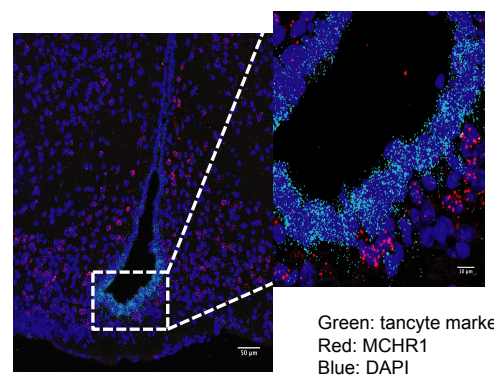
A**B****C****D****E****F**

Figure S5. Related to Figure 5. Optogenetic MCH-ME projection stimulation does not change refeeding upon the saline injection. (A) Placement of optical fiber implantation into third ventricle in optogenetic experiments. Green: neuronal tracer stained section from perfused mice with optical fiber implanted. Dashed white square illustrates the fiber trace. (B) 1 hr refeeding in fasted and 3.5 hrs light illuminated control (n=12) and ChR2^{MCH} mice (n=11) injected with saline (leptin vehicle) half hour before refeeding. (C) Placement of optical fiber into control and hM3Dq^{MCH} mice. Red: neuronal tracer stained section from perfused mice with optical fiber implanted. (D) 1 hr food intake in fasted and 4 hrs CNO-treated and fiber implanted control (n=6) and hM3Dq^{MCH} mice (n=8). (E) and (F) 4 hr refeeding in fasted and 3.5 hrs light illuminated control (n=12) and ChR2^{MCH} mice (n=11) injected with saline (leptin vehicle) half hour before refeeding and 4 hrs leptin sensitivity in those mice. Statistics: unpaired Student's T-test, *p < 0.05, except paired Student's T-test in leptin sensitivity.

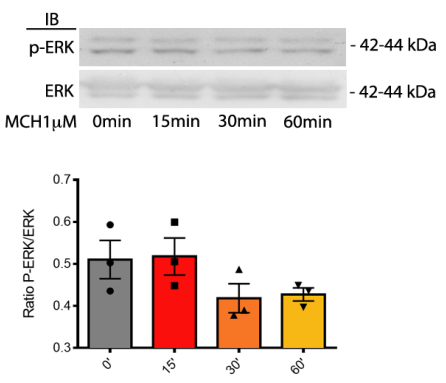
A qPCR quantification of Mchr1 expression from sorted tancytes



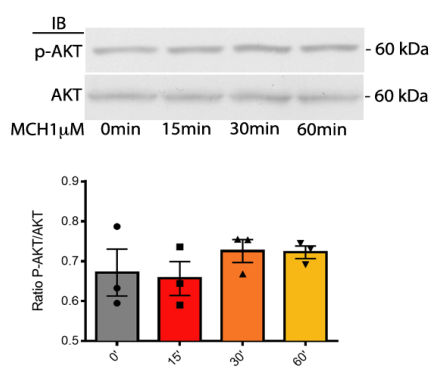
B MCHR1 mRNA *in situ*



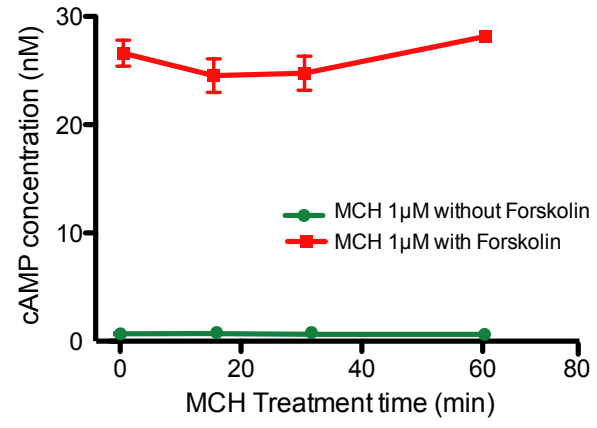
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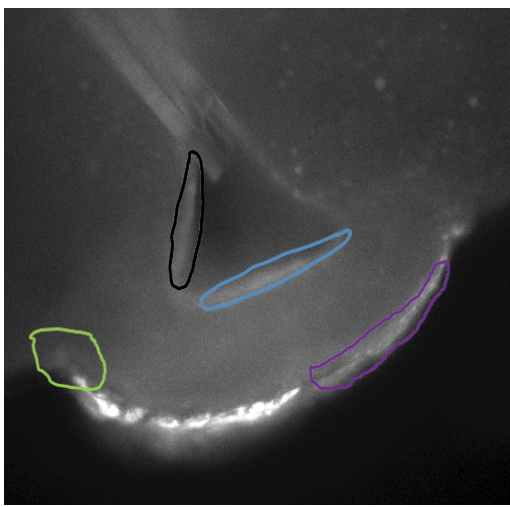
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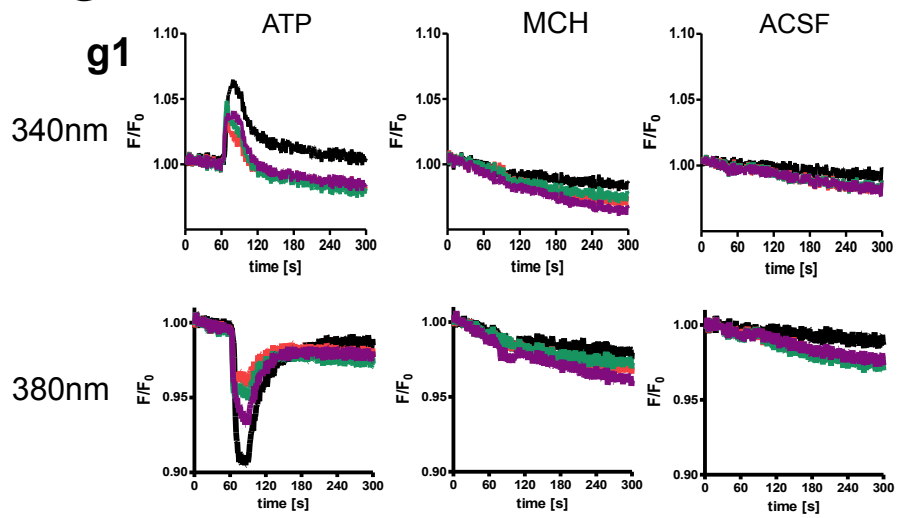
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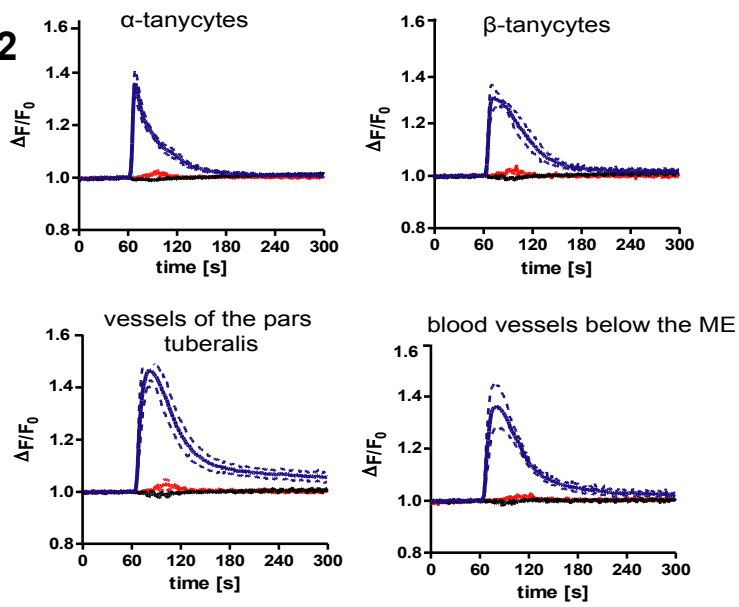
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G



g2



H

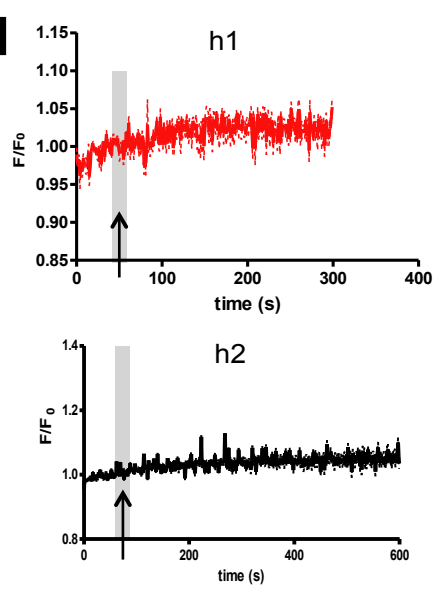
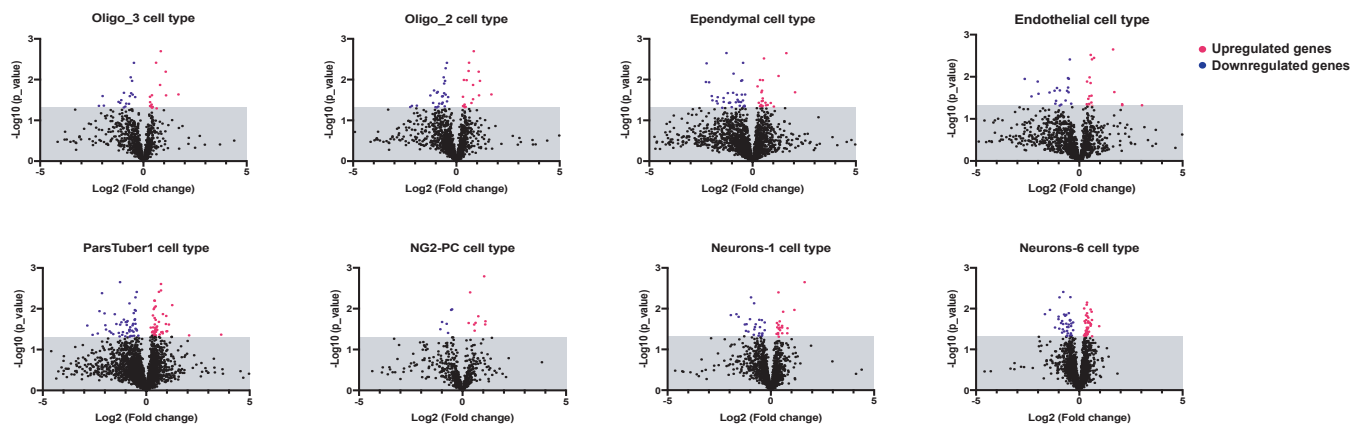


Figure S6. Related to Figure 6 and 7. MCH does not alter tanycyte signaling.

(A) Illustration of FACS sorted purified tanycytes from the ME (left) and qPCR quantification of Mchr1 expression in purified tdTomato marked tanycytes and non-tdTomato cells of the ME (right). (B) mRNA expression of Mchr1 mRNA by *in situ* hybridization on ME sections. Red, Mchr1; Green, tanycyte marker, Ppp1r1b; Blue, DAPI. (C) Western blotting of ERK and pERK expression after 0, 15 min, 30 min and 60 min of 1 μ M MCH incubation (upper panel) and ratio of pERK/ERK expression (lower panel). (D) Western blotting of AKT and pAKT protein expression after 0, 15 min, 30 min and 60 min of 1 μ M MCH incubation (upper panel) and ratio of pAKT/AKT comparison (lower panel). (E) cAMP concentration changes after 1 μ M MCH 0, 15 min, 30 min and 60 min of incubation with and without forskolin. (F) Illustration of the regions of interest for calcium imaging in ME slices. (G) Representative time course of fluorescence changes in a living brain slice loaded with Fura2 emitted with 340 nm (g1 upper panel) and 380 nm (g1 lower panel) after direct stimulation with ATP (1 mM, 2 μ l over 30 s; a and b), MCH (1 mM, 2 μ l over 30 s; c and d) and ACSF (2 μ l over 30 s; e and f). Black, alpha tanycytes; red, beta tanycytes; green, blood vessels in the pars tuberalis; purple, blood vessels beneath the median eminence. g2, Average time curves of $\Delta F/F_0$ in α -tanycytes, β -tanycytes, vessels of the pars tuberalis and blood vessels beneath the median eminence of 3 brain slices loaded with Fura2 after direct stimulation with ATP (1 mM, 2 μ l over 30 s; blue), MCH (1 mM, 2 μ l over 30 s; red) and ACSF (2 μ l over 30 s; black). (H) Average of $\Delta F/F_0$ in ME-tanycytes of 3 brain slices loaded with Fura2 after direct stimulation with Glutamate (100 μ M, 100 μ l over 30 s; red) (h1) and baclofen (1 mM, 100 μ l over 30 s; black) (h2).

A**B**

Upregulated genes in endothelial cells

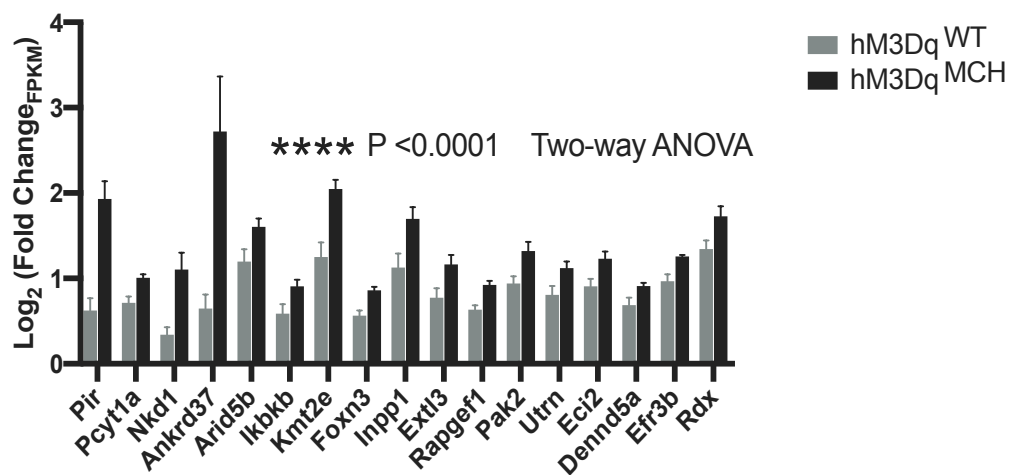
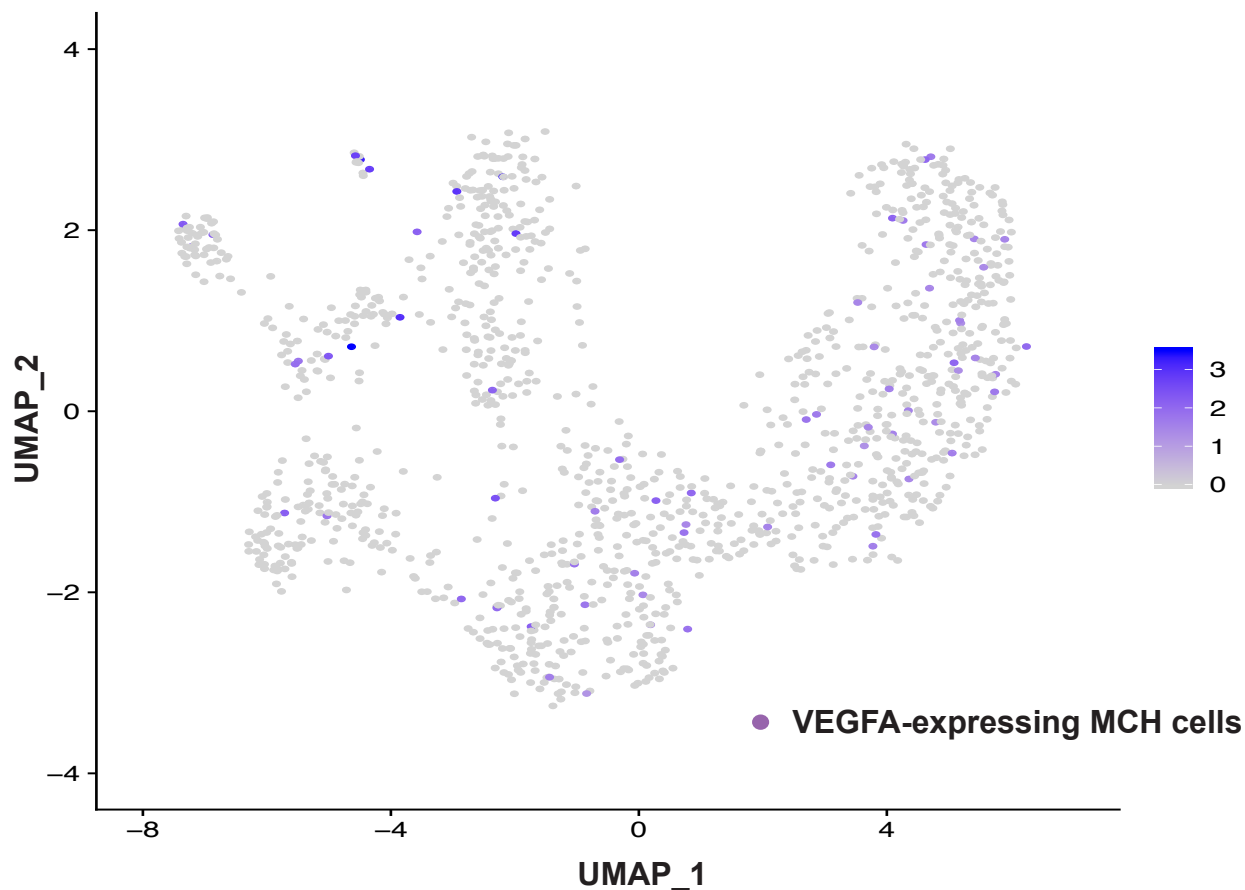
**C**

Figure S7. Related to Figure 6. PhosphoRibotrap-based analysis of all cell types regulated by MCH neuron activation. (A) Neuronal, glia and endothelial cell types in the ARC were regulated in 1 hr CNO treated hM3Dq^{MCH} mice. (B) Enriched gene expression of endothelial cell cluster in control and hM3Dq^{MCH} mice. (C) VEGFA expressing cells are distributed cross all the clusters of MCH neurons. Purple dots represent VEGFA positive cells.