

Figure S1. EspG₅mtub complements secretion of a *M. marinum* Δ espG₅. Secretion analysis of *M. marinum* WT, Δ espG₅ and Δ espG₅ complemented with WT espG₅mmar or espG₅mtub. Proteins were visualized by SDS-PAGE and immunoblotting using antibodies against EsxN and PE_PGRS proteins (ESX-5 substrates), GroEL2 (lysis and whole cell loading control) and EspG₅.

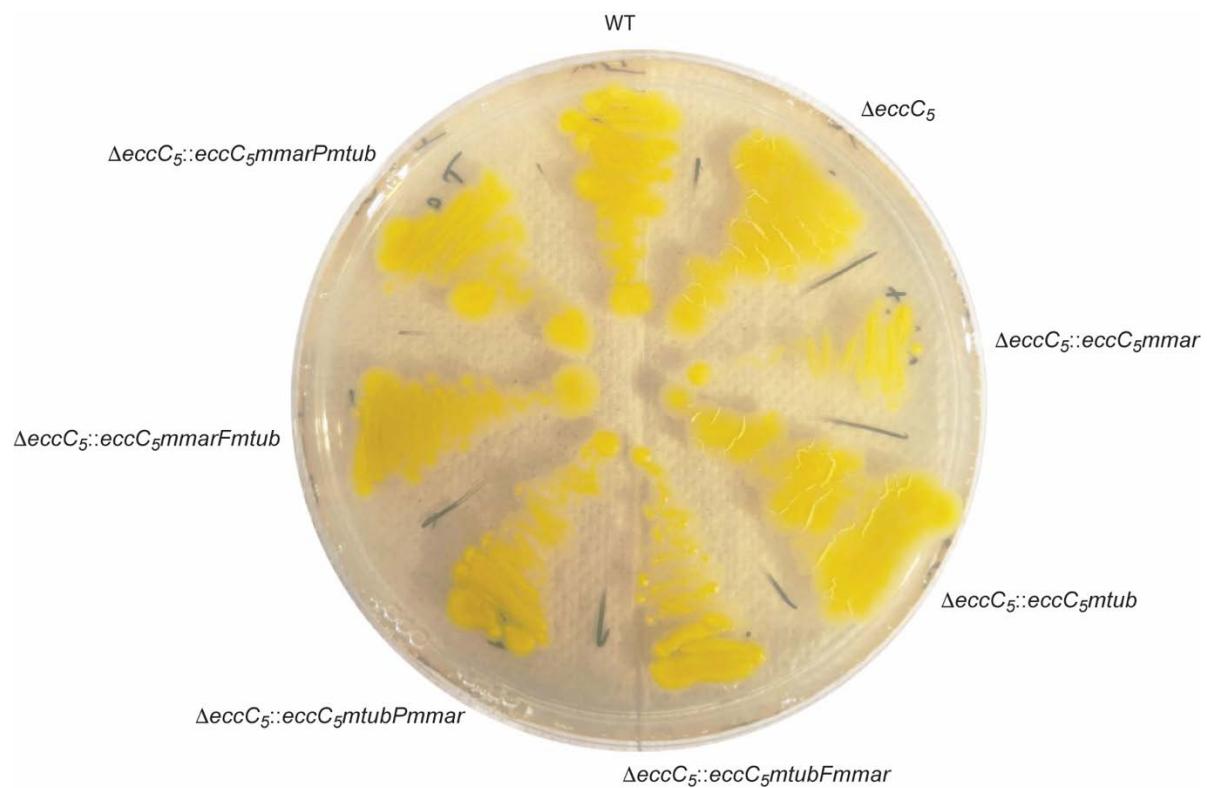


Figure S2. Linker 2 of EccC_{5mtub} affects the macroscopic appearance of *M. marinum*. Colony morphology of *M. marinum* WT, $\Delta eccC_5$ and $\Delta eccC_5$ complemented with constructs depicted in Fig. 2B. Strains were streaked on 7H10 agar media supplemented with 10% Middlebrook OADC (BD Biosciences).

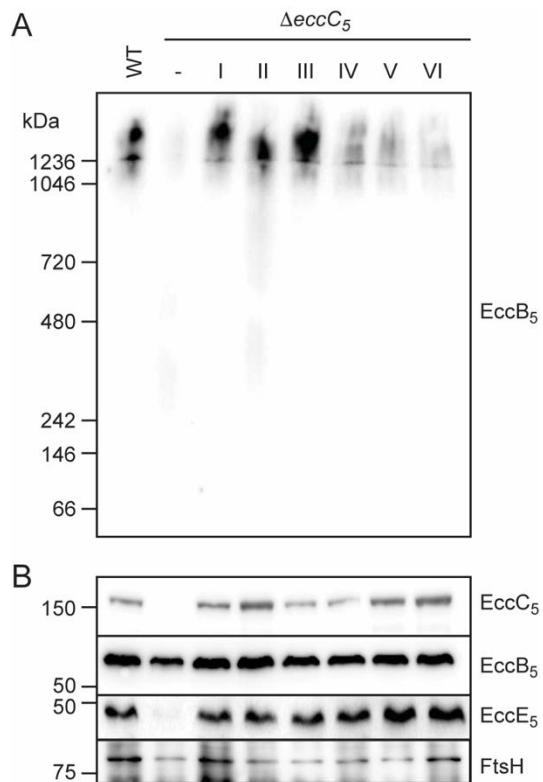


Figure S3. ESX-5 membrane complex analysis. (A) BN-PAGE and immunoblot analysis of DDM solubilized cell envelope fractions of *M. marinum* WT, $\Delta eccC_5$ and $\Delta eccC_5$ complemented with constructs depicted as in Fig. 2B. (B) SDS-PAGE and immunoblot analysis of the same set of samples as in A. Proteins were visualized using antibodies against EccB₅, EccC₅, EccE₅ (membrane proteins forming the ESX-5 membrane complex) and FtsH (membrane protein loading control).

A

B

EsxM_Mtub	MASRFMTDPHAMRDAGRFEVHAAQTVDEEARRMWASAQNISGAGWSGMAEATSLDTMTXM	60
EsxM_Mmar	VTARFMTDPHAMRDAGRFEVHAAQTVDEEARKMWASSQNIAGAGWSGMSASATSLDTMGQM	60
EsxB_TC	MAPQSAVDRAAMAQAAQDIEQSANAIR-GMQNQLASAKDQLRSHWEGDASMAFEAVFNRF	59
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EsxM_Mtub	NQAFRNIVNMHLHGVRDGLVRDANNYEQQEQASQQILSS-----	98
EsxM_Mmar	NTAFRNIVNMHLHSVRDGLVRDANNYEQQEQASQQVLRG-----	98
EsxM_Mtub	NQAFRNIVNMHLHGVRDGLVRDANNYEQQEQASQQILSS-----	98
EsxB_TC	NEDFSRVLKALDGMHESLVTRITYEARREAAQSVNRVQALLNG	104
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Figure S4. Divergence of Linker 2 sequences. (A) Alignment of the linker 2 region from *M. tuberculosis* H37Rv EccC₅, *M. marinum* M EccC₅ and *T. curvata* EccC. The C-terminal part of NBD1 and the N-terminal part of NBD2 are highlighted in blue. The sequences that were swapped between constructs EccC_{5mmar}P_{mtub} and EccC_{5mtub}P_{mmar} (partial linker 2 swap) are depicted in bold. The sequences that were swapped between constructs EccC_{5mmar}F_{mtub} and EccC_{5mtub}F_{mmar} (full linker 2 swap) are underlined. The missing structural feature of the linker 2 in the *T. curvata* EccC crystal structure (Rosenberg et al., 2015) is stroked through. (B) Sequence alignment between EsxB of *T. curvata* and EsxM from *M. marinum* and *M. tuberculosis*. Alignments were produced using Clustal Omega.

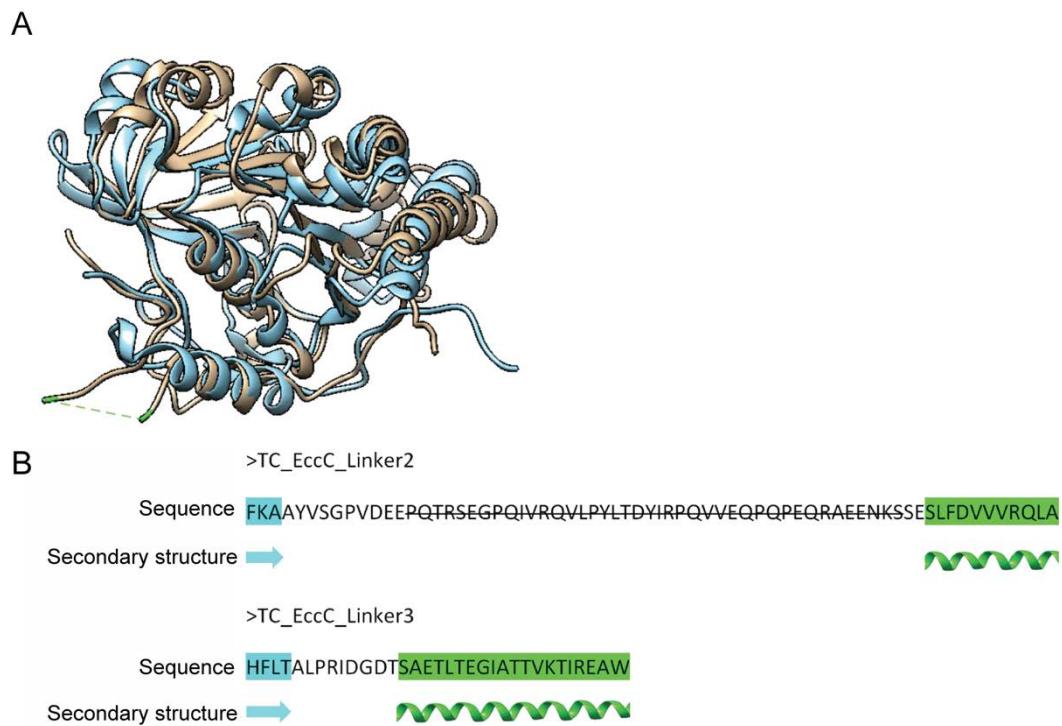


Figure S5. Comparison of linker 2 and linker 3. (A) Superimposition of NBD1 (light brown) and NBD2 (light blue) from *T. curvata* (Rosenberg et al., 2015), emphasizing the missing structural feature in the linker 2 region, in green. (B) linker 2 and linker 3 loop sequences with the adjacent upstream β -sheet (blue) and downstream α -helix (green). The missing structural feature is stroked through in the linker 2 sequence.

	Walker B	
EccCab1_M_tub	TLFVVVDEFAELLQSHPDFIGLFDRICRVGRSLRVHLLATQSLQTGGVRIDKLEPNLTY	652
EccCab1_M_marinum	TLFVVVDEFAELLQSHPDFIGLFDRICRVGRSLRVHLLATQSLQTGGVRIDKLEPNLTY	652
EccC2_M_tub	VLLVIVDEYLELFAHKWIDLIIHIGQEGRGANVFFMLGGQRDLSS--LQKVKSNIAF	687
EccC3_M_marinum	TLFVVADEFTLMALADHPPEYAEFLDYVARKGRSFRIHILFASQTLDVGK--IKDIDKNTSY	654
EccC3_M_tub	TLFVVADEFTLMALADHPPEYAEFLDYVARKGRSFRIHILFASQTLDVGK--IKDIDKNTAY	649
EccC4_M_marinum	TLFTIIVDEFAELLSQHPDFAEVFVAIGRLGRSLGMHLLLASRLEEGR--LRGLEAHLSY	584
EccC4_M_tub	TLFIVVDEFSELLSQQHPEFVDVFLAIGRVGRSLGMHLLLASRLEGR--LRGLETHLSY	594
EccC5_M_marinum	MLVVVIDEFYEWFRIMPTAVDVLDISIGRQGRAYWIHLMMASQTIESR--AEKLMENMGY	666
EccC5_M_tub	MLVVVIDEFYEWFRIMPTAVDVLDISIGRQGRAYWIHLMMASQTIESR--AEKLMENMGY	665
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EccCab1_M_tub	RIALRTTSSHESKAVIGTPEAQYITNK--ESVGFL-RVMEDPVKFSTFYISGPYMPPA	709
EccCab1_M_marinum	RIALRTTSSHESKAVIGTPEAQYITNK--ESVGFL-RVMEDPIKFSTLYISGPYVPAA	709
EccC2_M_tub	RIALRAESGDDSRVEIGSDAAYHLP SK--ENGFALL-KVGRDLEPFRCFYLSAPFVVPK	744
EccC3_M_marinum	RIGLKVASPVSRSRQIIGVEDAYHIEAGKEHKGVLFLVPAPGAAPIRFRSTYVDGIYEPPQ	714
EccC3_M_tub	RIGLKVASPVSRSRQIIGVEDAYHIESGKEHKGVLFLVPAPGAAPIRFRSTYVDGIYEPPQ	709
EccC4_M_marinum	RVCLKTLSAIESRTALGTLDFELPNT---PGSGLL-SSPTAELTRFETAFVSGPVPAGP	640
EccC4_M_tub	RMCLKTWSASESRNVNLGTQDAYQLPNT---PGAGLL-QTGTGELIRFQTAFVSGPLRRAS	650
EccC5_M_marinum	RLVLKARTAGA-AQAAGVPNAVNLPQ---AGLGYF-RKSLEDIIRFQAEFLWRDYFQPG	721
EccC5_M_tub	RLVLKARTAGA-AQAAGVPNAVNLPQ---AGLGYF-RKSLEDIIRFQAEFLWRDYFQPG	720
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EccCab1_M_tub	AGVETNGEA-GGPGQQTT-----RQAARIHRFTAAPVLEAP-----	745
EccCab1_M_marinum	T-AETNGDG-SGPSTQFA-----KRALQIREFTAAPVLEAL-----	744
EccC2_M_tub	KKEVART----IDMTLTQPRLYDWQYQPLDAADEA--LATAAAADAEPDEF-----	790
EccC3_M_tub	QTAKTRVVHN-----IPEPKLFTATAVEPDPTV--ISEV	746
EccC4_M_tub	TAKAVVQS-----VPEPKLFTAAAVEPDPTV--IADT	741
EccC5_M_tub	PDADSV-AL-----PAAVRPFTSEATGDVRT-----GG	667
EccC5_M_tub	PSAVHP-VA-----PPSVRPFTTHAAAPVTA-----GP	677
EccC5_M_tub	ITVDGEEAPVLVHSIDYIRPQLFTNSFTPLETEVGGPEIDKVVVAHANGEVVEEVA---	777
EccC5_M_tub	VSIDGEEAPALVHSIDYIRPQLFTNSFTPLETEVSGGPDIEPVVAQPNGEVESDDIEGGE	780
	. :	
	Start EccC _{b1}	
EccCab1_M_tub	TPMTAEPEVRTLREVVLQDQLGTA-ESRAYKM WL PPLTNPVPLNELIARDR-----R-	795
EccCab1_M_marinum	TPMTAEPEVRTLREVVLQDQLSTV-ESRAYKM WL PPLVDPPTLDELVARDR-----R-	794
EccC2_M_tub	LYYDDGFKKKKIVDVLRESLYNVPHRSPPR WL APLEPDEPVDRLVAAYRGKPWHVDYQ	850
EccC3_M_marinum	EEEHPVGPPrKLATI GE QLARY-GPQAPR WL PPLDEPIPLSTLLATAG-----VA	797
EccC3_M_tub	DEQEPADPPRKLATI GE QLARY-GPQAPR WL PPLDEPIPLSTLLATAG-----VA	792
EccC4_M_marinum	ADGADTATGPTVLTQVLDRIAGH-GPRAHQV WL PPLDRAPALASLLSDGA-----AE	718
EccC4_M_tub	VGGTAEVPTPTVHLAVLDRLVGH-GPAAHQV WL PPLDEPPMLGALLRDAE-----PA	728
EccC5_M_marinum	EAEEEGIRVKVGTVIIDQLRRI-NFEPYRL WQ PPLTQPVAIDDVLNRFLGHWPQKEYGS	836
EccC5_M_tub	DEDEEGVRTPKVGTVIIDQLRKI-KFEPYRL WQ PPLTQPVAIDDVLNRFLGRPWKEYGS	839
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	Walker A	
EccCab1_M_tub	-QPLRFALGIMDEPRRLQDVGVVDVSGAGG GNIGIGG GAPQT KS TLLQTVMMSAATHSP	854
EccCab1_M_marinum	-QPLRFALGIMDEPRRLQDVGVVDVSGAGG GNIGIGG GAPQT KS TLLQTVMMSAATHSP	853
EccC2_M_tub	NPGLMFPGVMDIPEESQQVVHAVA--LRSNIIIVGAKQRGKTTLMAIMCSAATMYTP	908
EccC3_M_marinum	QRQWRWPLGEIDKPFEMRMRDPLVFDATSSAGNVLVIGHGPKS KS TKALQSFMLSAASLHSP	857
EccC3_M_tub	PRQWRWPLGEIDKPFEMRMRDPLVFDARSSAGNVMVIGHGPKS KS TKALQTFILSAASLHSP	852
EccC4_M_marinum	HAELAVPIGVVDRPFDQSRTPLTIDLTAAAGNVAVVGAQPSG KS TTLSTLITALAATHDP	778
EccC4_M_tub	QAELEVPIGVIDRPFEQSRVPLTIDLSGAACNVAVVGAQPSG KS TKALRTLIMALAATHDA	788
EccC5_M_marinum	ARNLVFPIGVIDRPFKHDPQPPWTVDTSGP <i>G</i> S <i>N</i> V <i>L</i> I <i>L</i> G <i>A</i> GG <i>S</i> CK TTALQTLISSAALTHTP	896
EccC5_M_tub	ACNLVFP <i>I</i> GI <i>D</i> RP <i>Y</i> KHDQPPWTDTSGPG <i>A</i> N <i>V</i> <i>L</i> <i>I</i> <i>L</i> G <i>A</i> GG <i>S</i> CK TTALQTLICSAALTHTP	899
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Figure S7. Alignment of all EccC homologs of *M. marinum* M and *M. tuberculosis* H37Rv. Part of the linker 2 region, which shows no sequence homology and overlaps largely with the missing structural feature of the *T. curvata* EccC and the “Partial linker 2 swap” constructs, is highlighted in grey. the C-terminal part of NBD1 and the N-terminal part of NBD2 are highlighted in blue. Walker A and B motifs are highlighted in green. The first residue of EccC_{b1} is depicted in red. The alignment was produced using Clustal Omega.

Table S1. List of plasmids used in this study.

Nr.	Plasmid	Features	Source
1.	pMV- <i>espG_{5mtub}</i>	<i>hsp60</i> promotor, L5 <i>attP</i> & <i>aph</i> , <i>oriE</i> , <i>hyg^R</i>	This study
2.	pMV- <i>espG_{5mmar}</i>	<i>hsp60</i> promotor, L5 <i>attP</i> & <i>aph</i> , <i>oriE</i> , <i>hyg^R</i>	Phan et al., 2017
3.	pMV- <i>eccc_{5mtub}</i>	<i>hsp60</i> promotor, L5 <i>attP</i> & <i>aph</i> , <i>oriE</i> , <i>kan^R</i>	This study
4.	pMV- <i>eccc_{5mmar}</i>	<i>hsp60</i> promotor, L5 <i>attP</i> & <i>aph</i> , <i>oriE</i> , <i>kan^R</i>	Ates et al., 2015
5.	pMV- <i>eccc_{5mtub}F_{mmar}</i>	<i>hsp60</i> promotor, L5 <i>attP</i> & <i>aph</i> , <i>oriE</i> , <i>kan^R</i>	This study
6.	pMV- <i>eccc_{5mtub}P_{mmar}</i>	<i>hsp60</i> promotor, L5 <i>attP</i> & <i>aph</i> , <i>oriE</i> , <i>kan^R</i>	This study
7.	pMV- <i>eccc_{5mmar}F_{mtub}</i>	<i>hsp60</i> promotor, L5 <i>attP</i> & <i>aph</i> , <i>oriE</i> , <i>kan^R</i>	This study
8.	pMV- <i>eccc_{5mmar}P_{mtub}</i>	<i>hsp60</i> promotor, L5 <i>attP</i> & <i>aph</i> , <i>oriE</i> , <i>kan^R</i>	This study

Table S2. List of primers used in this study.

Nr.	Name	Sequence (5' - 3')
1.	EsxMtb_XmnI_F	GACTGAAAGAATTCCAT ATGGCCTCACGTTTAT GAC
2.	EspG5tub_HindIII_R	GACTAACGTTCTACACTCTGCTGTGTTTC
3.	EccC5tub_XmnI_F	GACTGAAAGAATTCCAT ATGAAGCGT GGTTTGCCC
4.	EccC5tub_HindIII_R	GACTAACGTTCTACCGACGCACCTCGG
5.	C5tub_MunI.fw	CCCGCAATTGTTCACCAACTCATTAC
6.	C5tub_interswap.fw	GAAGAAGGCATCCGAGT GCCGAAGGTT GGGAC GGT
7.	C5_swap_1_FW 69C	ATATGGCCGGCAGGGCCGT GCCTATTG
8.	C5_swap_1_RV 67C	GTCGATCGCGACGGGCTG
9.	C5_swap_2_FW 62C	CCCGTCGCGATCGACGACTTG
10.	C5_swap_2_RV 65C	CTGATCACCGCGGCCATG
11.	C5 mar Tb full domain 1 FW	ATCCGGAGGAAAGAATT CCTATTGCCGCAAGGCCGACG
12.	C5 mar Tb full domain 1 RV	GTCGATGCTGACGCCGGGCTGGAAAGTAGTCC
13.	C5 mar Tb full domain 2 FW	GGCGTCAGCATCGACGGCGA
14.	C5 mar Tb full domain 2 RV	ACC CGTTGACCAGGTCGTCGATGGCGAC
15.	C5 mar Tb full domain 3 FW	GACCTGGTCAACC GGTTCTCG
16.	C5 mar Tb full domain 3 RV	CGACATCGATAAGCTTTACTTCGACCTCGGTC
17.	C5 mar partial TB interdom 1 RV	GAACGAGTTGGTAAACAACTGC GGGCGAAT G
18.	C5 mar partial TB interdom 2 FW	CATT CGCCCGCAGTTGTTACCAACTCGTTC
19.	C5 mar partial TB interdom 2 RV	GTACCCACCTGGGGTGCGCACCCCCCTC
20.	C5 mar partial TB interdom 3 FW	GCACCCCCAAGGTGGGTACCGTGATC