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Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see <u>Authors & Referees</u> and the <u>Editorial Policy Checklist</u>.

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St	at	ıstı	ICS

For	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
	A description of all covariates tested
	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
\boxtimes	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
\boxtimes	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
	\boxtimes Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated
	Our web collection on statistics for biologists contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection PClamp 10.6 (Clampex), GrassLab, ANYmaze 4.94h beta, NIS-Elements 5.11, LAS-X 3.4.1

Data analysis Clampfit 11.0.3, Grass

Clampfit 11.0.3, GrassLab, Mini Analysis 6.0.7, ImageJ 1.52a (NIH), NIS-Element Viewer 4.5, GraphPad PRISM 6 and 8, SigmaPlot 10 and 12, MatLab R2012a

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data $% \left(1\right) =\left(1\right) \left(1\right) \left($
- A description of any restrictions on data availability

The authors declare that all relevant data in this study are included within the article and supplementary information. The source data underlying Figures 1e-h, 2a-h, 3c, f, g, j, k, 4a, b, d, e, g, h, j, k,5a-e, 6f-m, and 7d, h-l and Supplementary Figures 1a, c-f, 3b, c, 4c, e, f, 5c-f, 6b-d, 7a, b, 8a, b, 9a, b, 10a-e, 11a, b, and 13b, c are provided as a Source Data file. Source Data are provided with the paper. All requests for raw data and materials are promptly reviewed by the Hospital for Sick Children to verify whether the request is subject to any intellectual property or confidentiality obligations. Any materials that can be shared will be released via a Material Transfer Agreement.

Field-sne	ecific reporting	
	ne below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.	
Life sciences	Behavioural & social sciences	
	the document with all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>	
Life scier	nces study design	
All studies must dis	sclose on these points even when the disclosure is negative.	
Sample size	The number of experiments were based on our previous publications on EEG recordings (Snead et al., J Neurosci. 2000, 20:6218-24), slice recordings, immunostaining and behavioral tests (Zhou et al., Nat Neurosci. 2018, 21:50-62).	
Data exclusions	No data exclusion except for those animals that died or disabled before the completion of the experiment.	
Replication	Immunostaining and bright field sample images are representative of at least three individual experiments. Replication numbers in all other experiments are indicated in figure legends.	
Randomization	All allocation was random. The experimenters did not know the genotype or treatment groups during the experiment.	
Blinding	Investigators were blinded to the sample allocation, experimentation and data analysis	
We require informati	g for specific materials, systems and methods on from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, ted is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.	
Materials & ex	perimental systems Methods	
n/a Involved in th	ne study n/a Involved in the study	
Antibodies	ChIP-seq	
Eukaryotic	cell lines	
Palaeonto	logy MRI-based neuroimaging	
Animals ar	nd other organisms	

Antibodies

Antibodies used

Clinical data

Human research participants

Commercial antibodies were used in this study: anti-GFP (Aves Labs, Cat #GFP1010), anti-Neuroligin 2 (Synaptic System, Cat#129 203), anti-Parvalbumin (Sigma Aldrich, Cat#P3088, Clone PARV-19), anti-GFAP (Cell Signaling Technology, Cat#3670, Clone GA5, Lot Number 6).

The following secondary antibodies were used: donkey anti-chicken IgG, Alexa Fluor 488 (Jackson ImmunoResearch Laboratories, Cat#703-545-155), donkey anti-rabbit IgG, Alexa Fluor 568 (Invitroge, Cat#A10042), goat anti-mouse IgG, Alexa Fluor 488 (Invitrogen, Cat# A32723), and goat anti-mouse IgG, Alexa Fluor 546 (Invitroge, Cat# A-11030).

Validation

Validation information was based on the manufacturer's websites:

anti-GFP (Aves Labs, Cat #GFP1010) (https://www.aveslabs.com/collections/epitope-tag-6xhis-beta-gal-actin-and-gfp-antibodies/products/green-fluorescent-protein-gfp-antibody): The antibody was analyzed by western blot and immunohistochemistry using transgenic mice expressing the GFP gene.

anti-Neuroligin 2 (Synaptic System, Cat#129 203) (https://www.sysy.com/products/neuroligin2/facts-129203.php): The antibody reacts with human, rat, mouse, monkey, ape, cow. Specific for neuroligin 2, no cross reactivity to neuroligins 1, 3, 4. (K.O. verified), can be used for western blot, ELISA and immunostaining analysis.

anti-Parvalbumin (Sigma Aldrich, Cat#P3088, Clone PARV-19) (https://www.sigmaaldrich.com/catalog/product/sigma/p3088? lang=en®ion=CA): The antibody reacts with a variety of species and used for immunohistochemistry in brain tissues. anti-GFAP (Cell Signaling Technology, Cat#3670, Clone GA5, Lot Number: 6) (https://www.cellsignal.com/products/primary-antibodies/gfap-ga5-mouse-mab/3670?site-search-type=Products): The antibody detects endogenous levels of total GFAP protein from human, mouse and rat. Can be used for western blot, immunohistochemistry and immunofluorescence.

Animals and other organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research

Laboratory animals

Neuroloigin (1,2,3, in the 129SvxC57BL/6 genetic background) KO mice, FMR1 KO mice and FVB mice were purchased from the Jackson Laboratory. PAK3 transgenic mice were made in house. The age of the mice ranged from 6-18 weeks with similar

number of males and females. This information is indicated in the Methods of the paper.

Wild animals No wild animals were used in the study.

Field-collected samples No field collected samples were used in the study.

Ethics oversight All animal protocols used in this study were approved by the Lab Animal Use Committee of the Hospital for Sick Children,

Toronto, Canada

Note that full information on the approval of the study protocol must also be provided in the manuscript.