

Supplemental Information

Essential role of *Salmonella* Enteritidis DNA adenine methylase in modulating inflammasome activation

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Xilong Kang, Chuang Meng, Xinan Jiao, Zhiming Pan

Figure S1

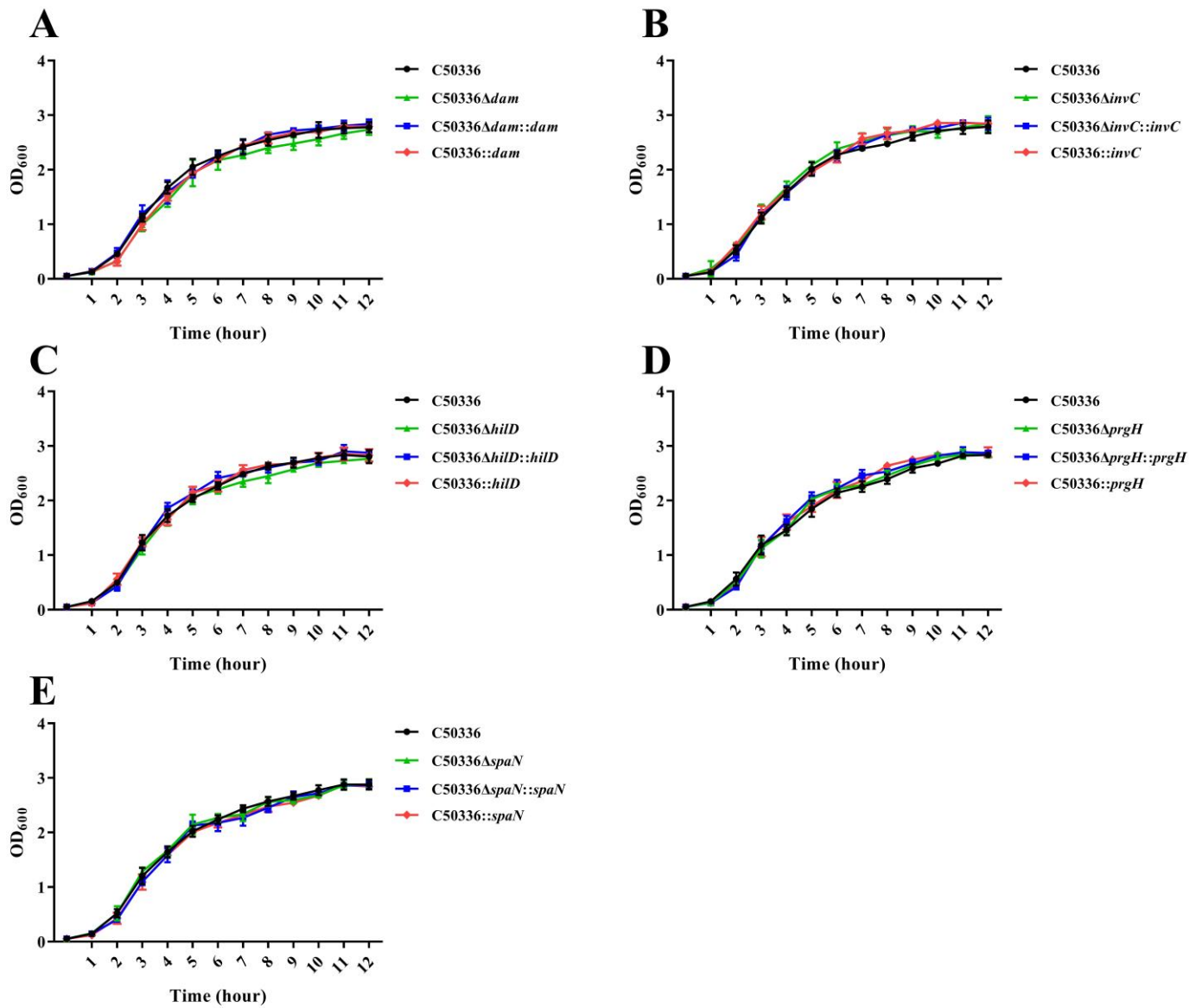


Figure S2

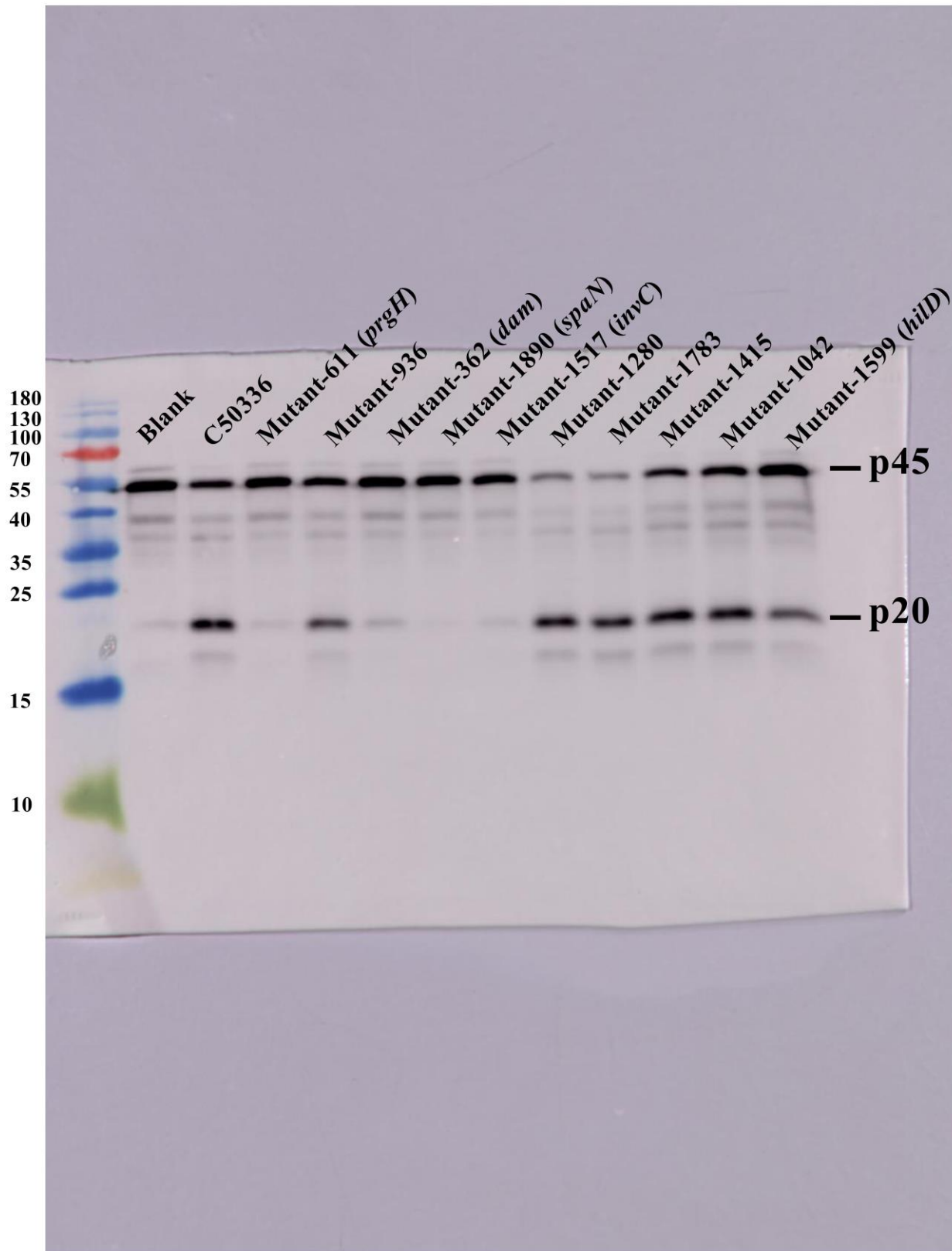


Figure S2

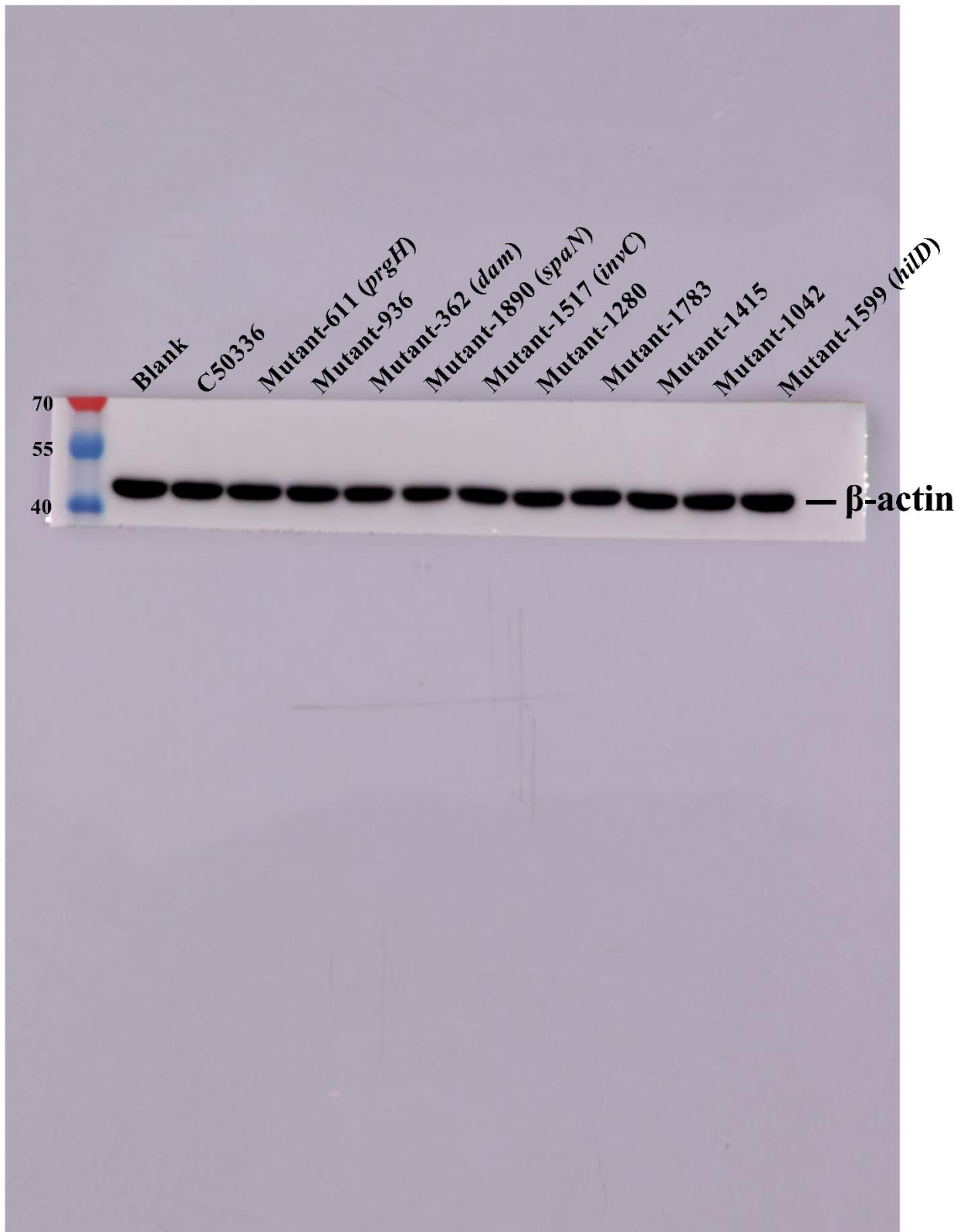


Figure S3

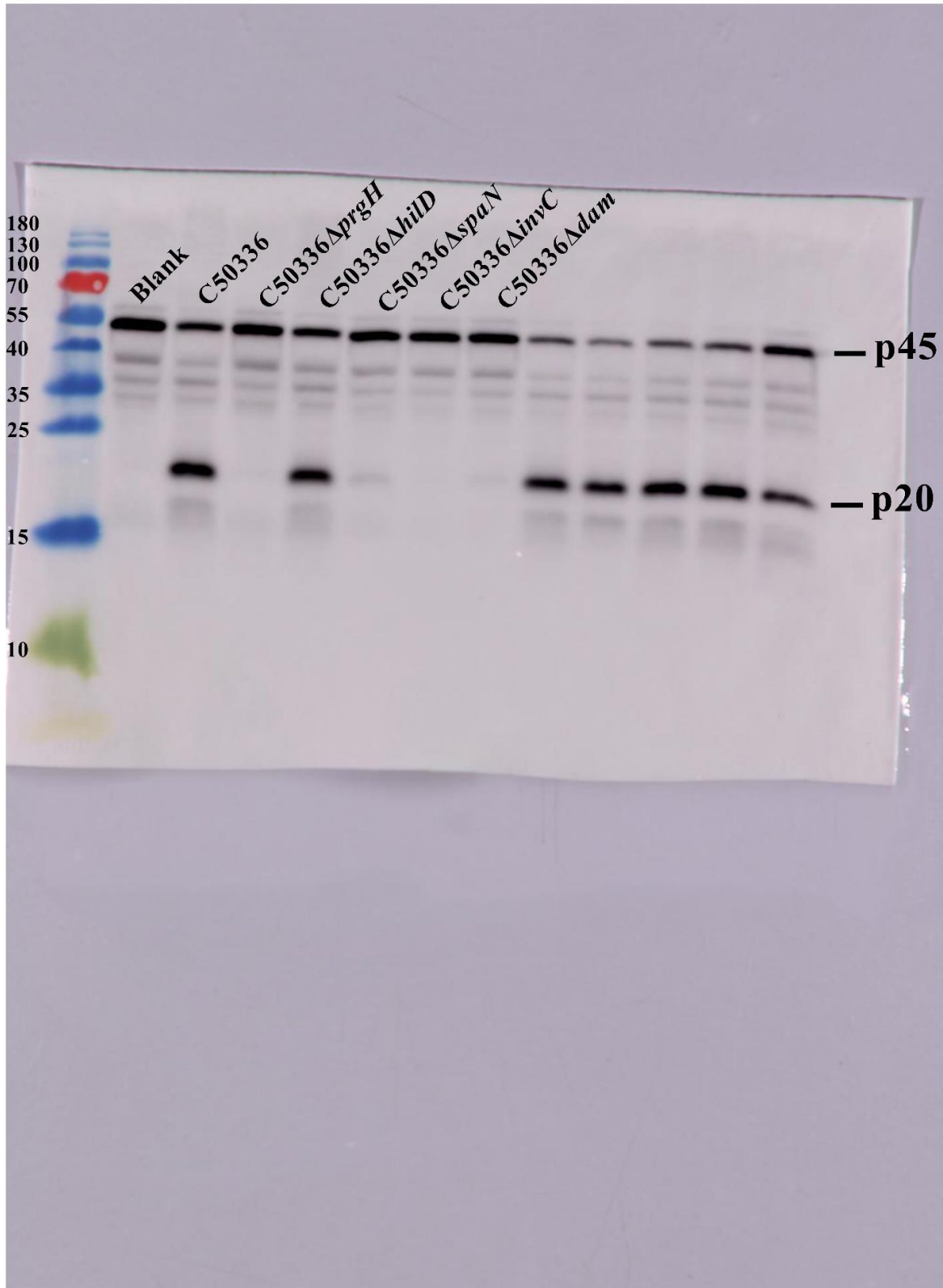


Figure S3

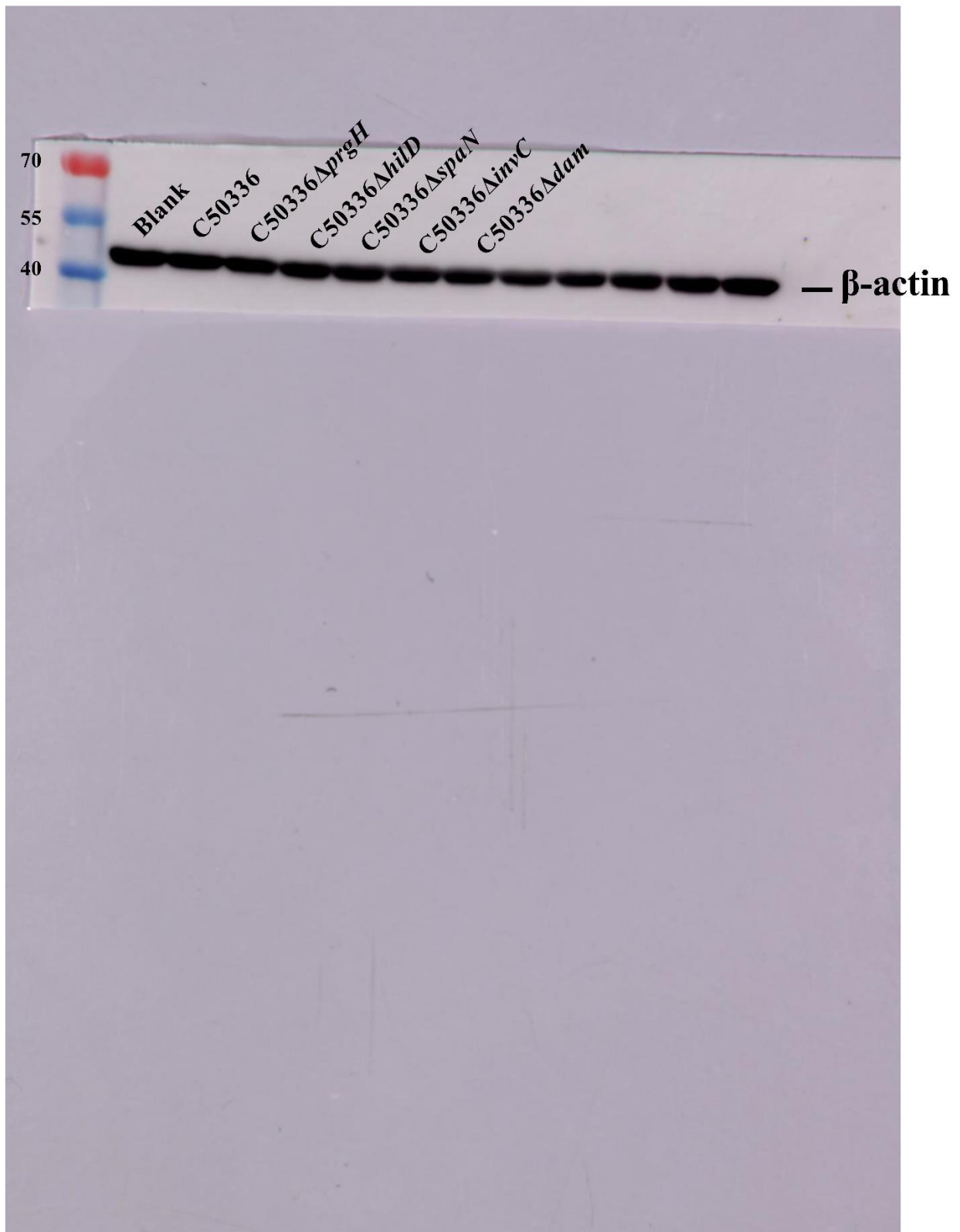


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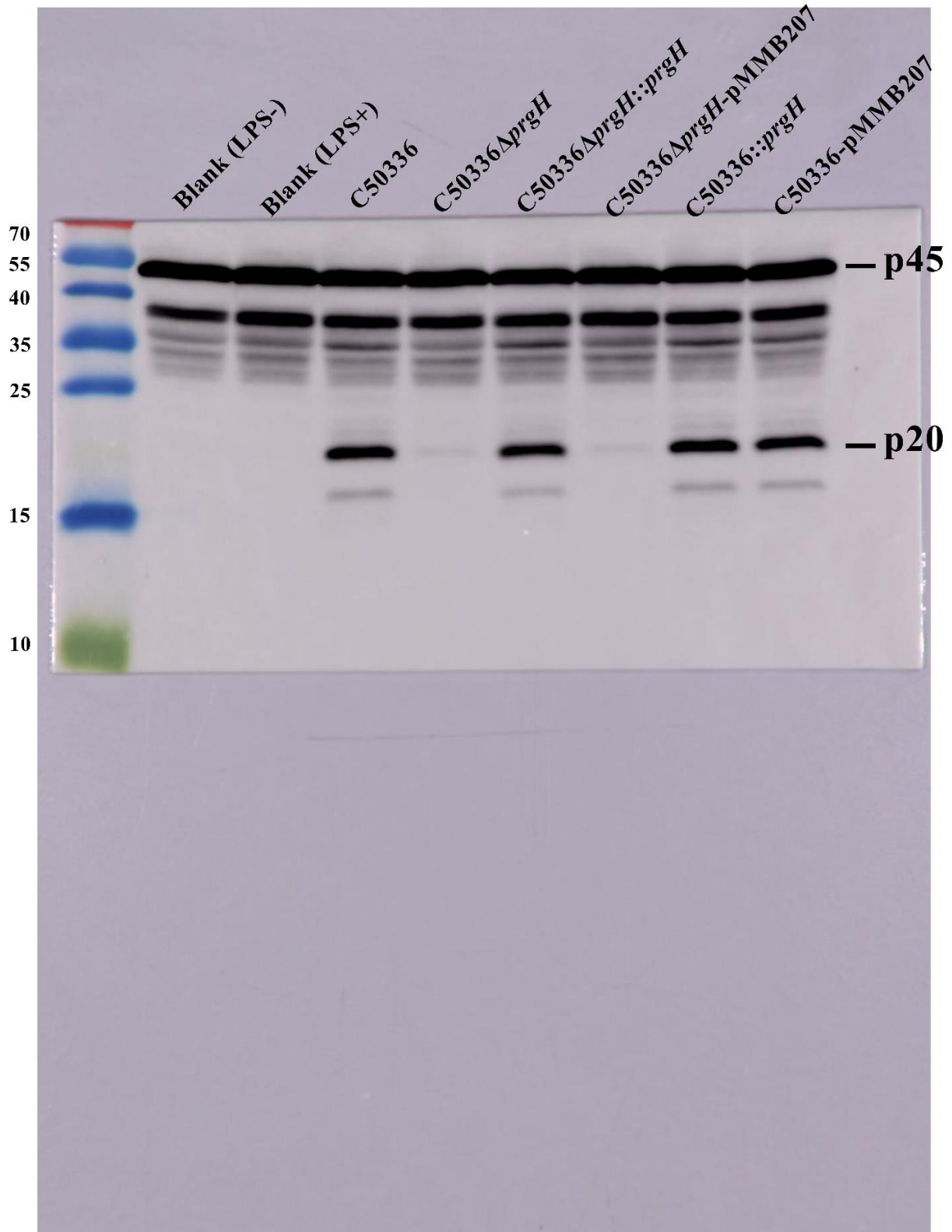


Figure S4



Figure S4

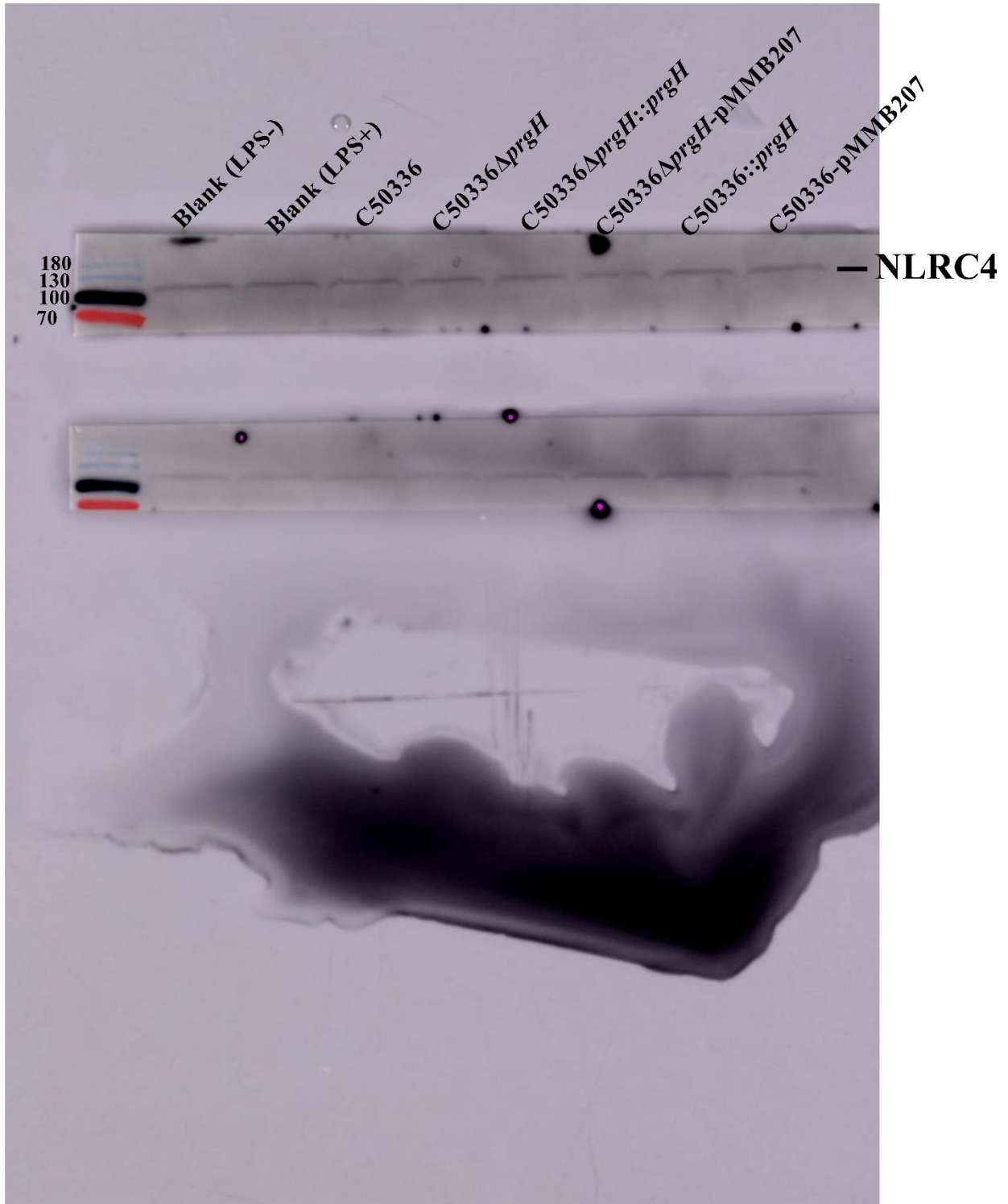


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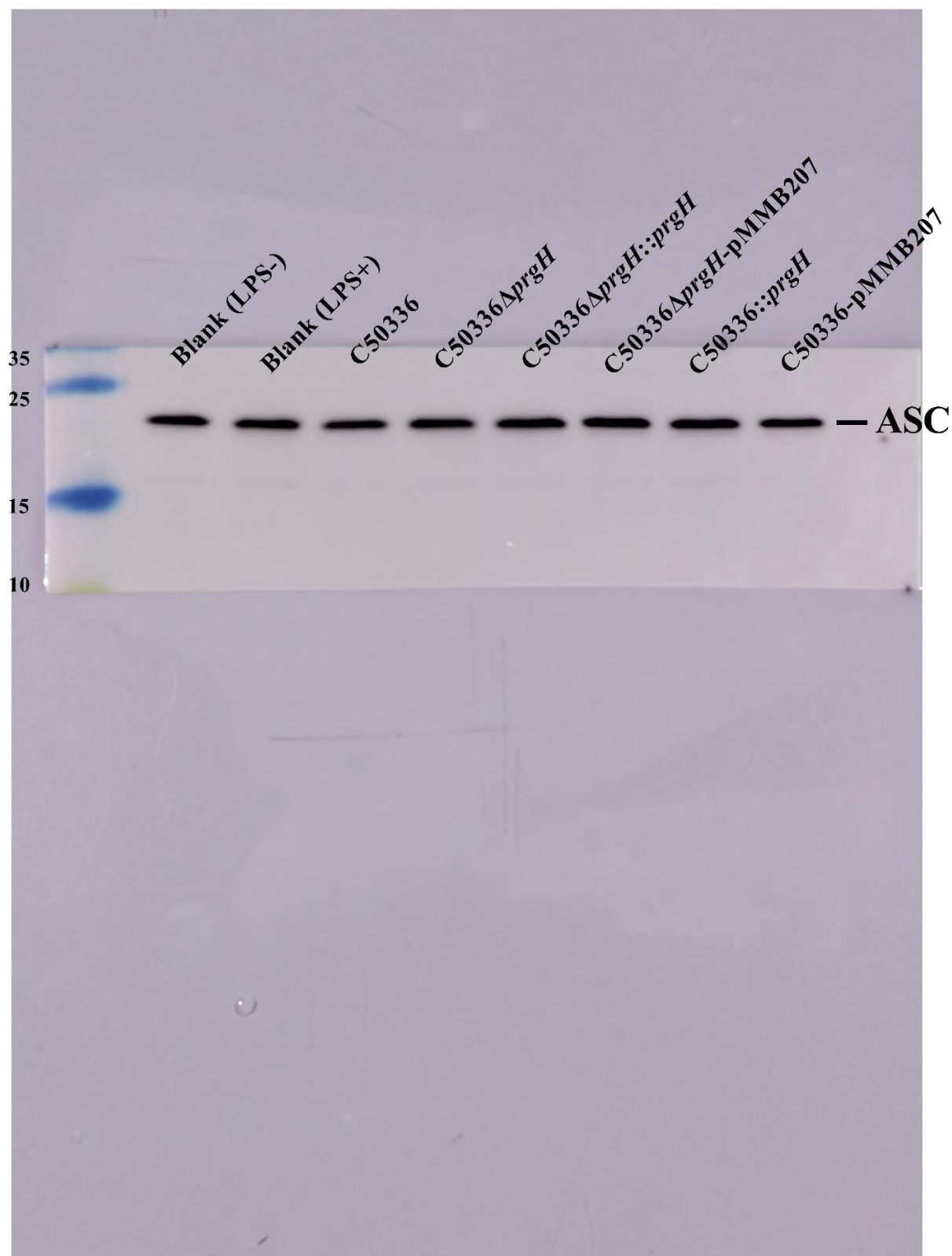


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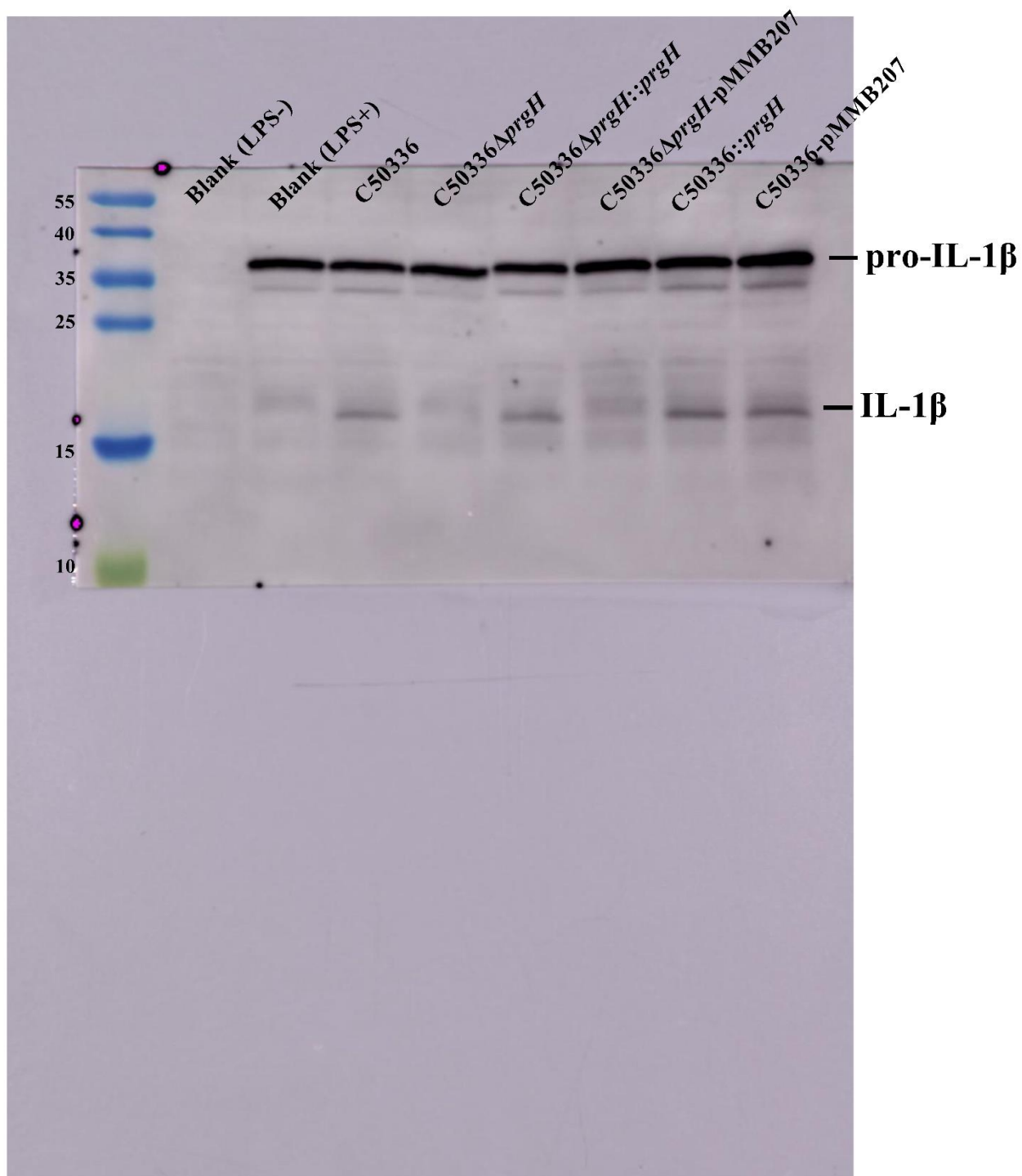


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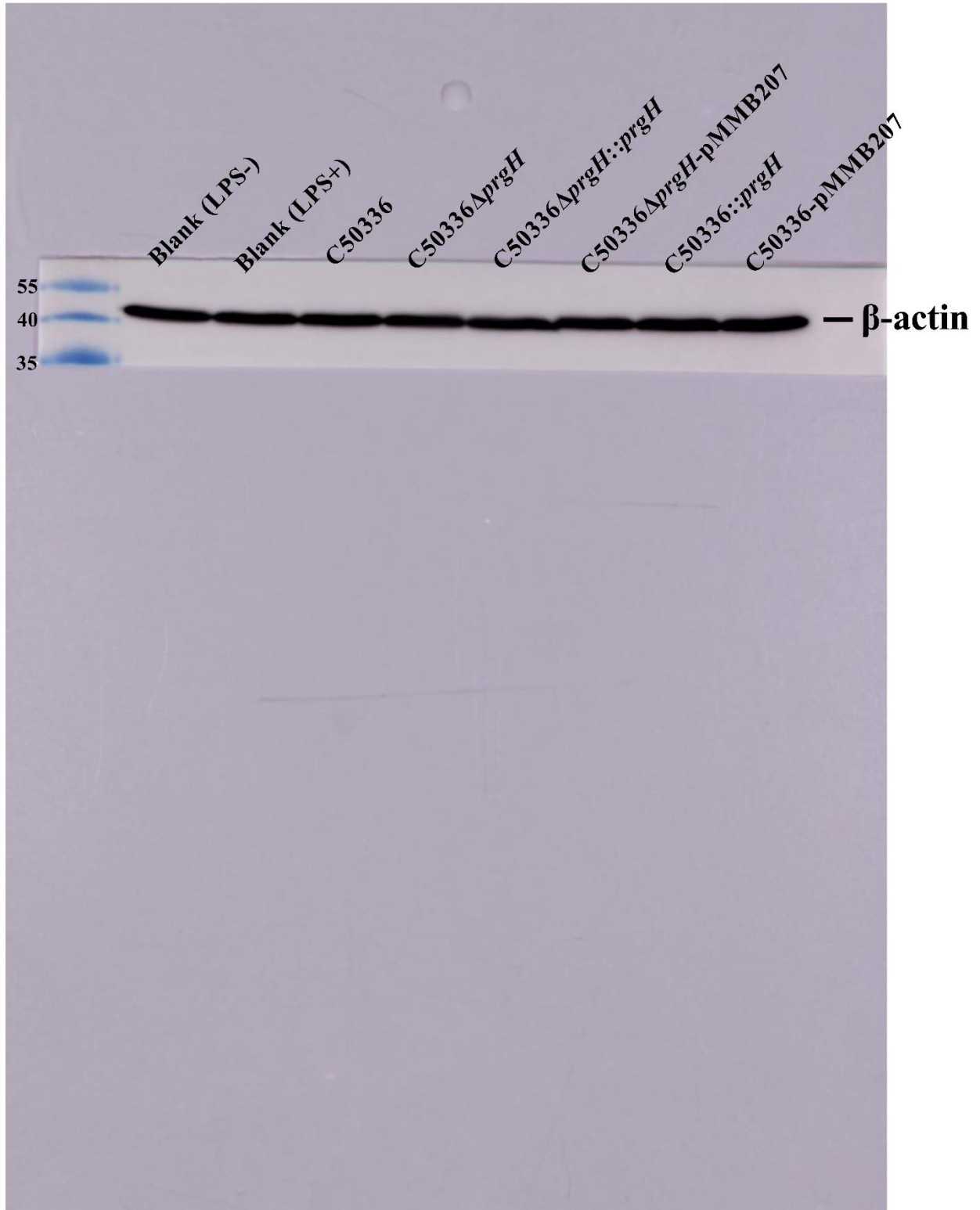


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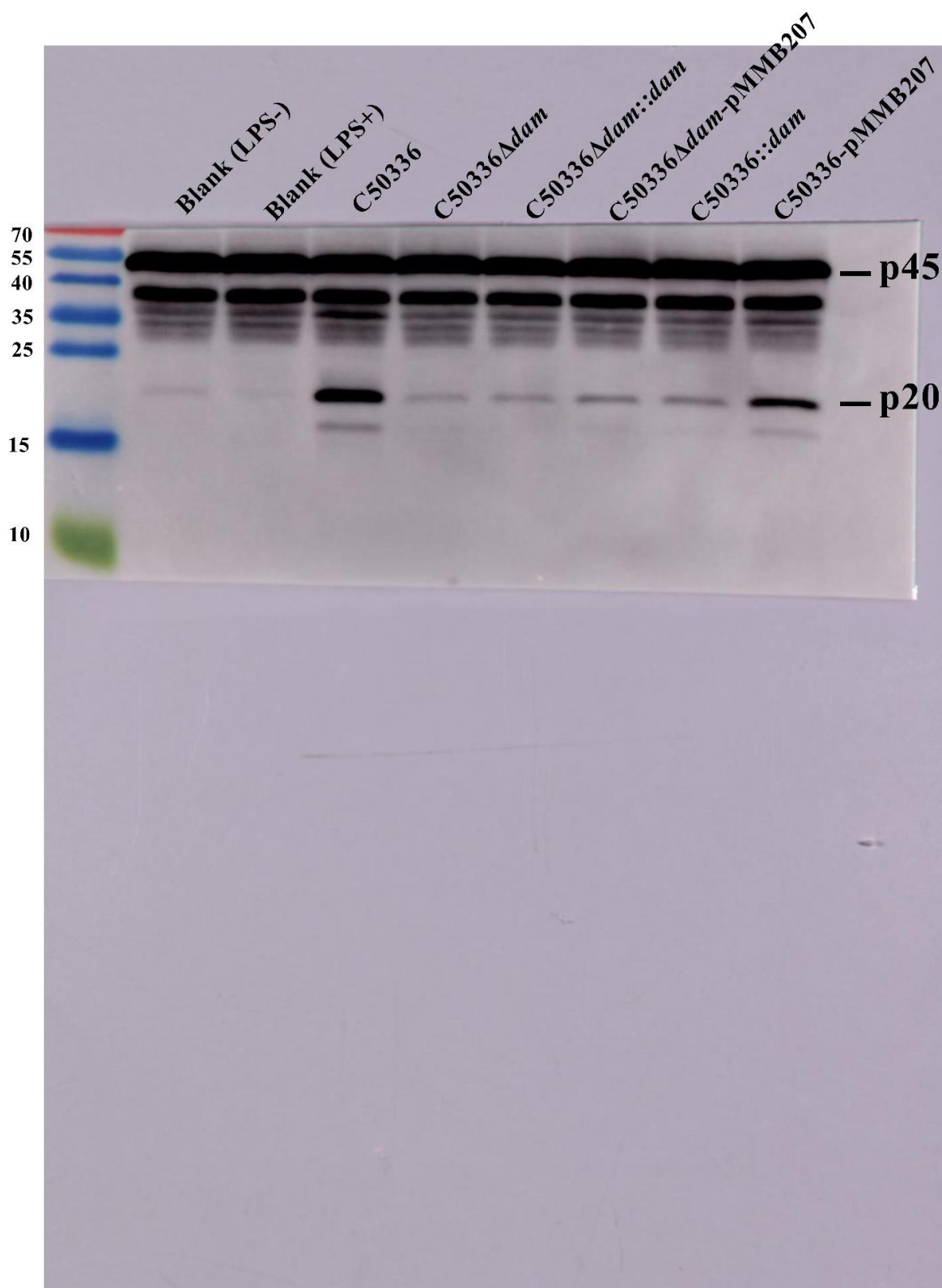


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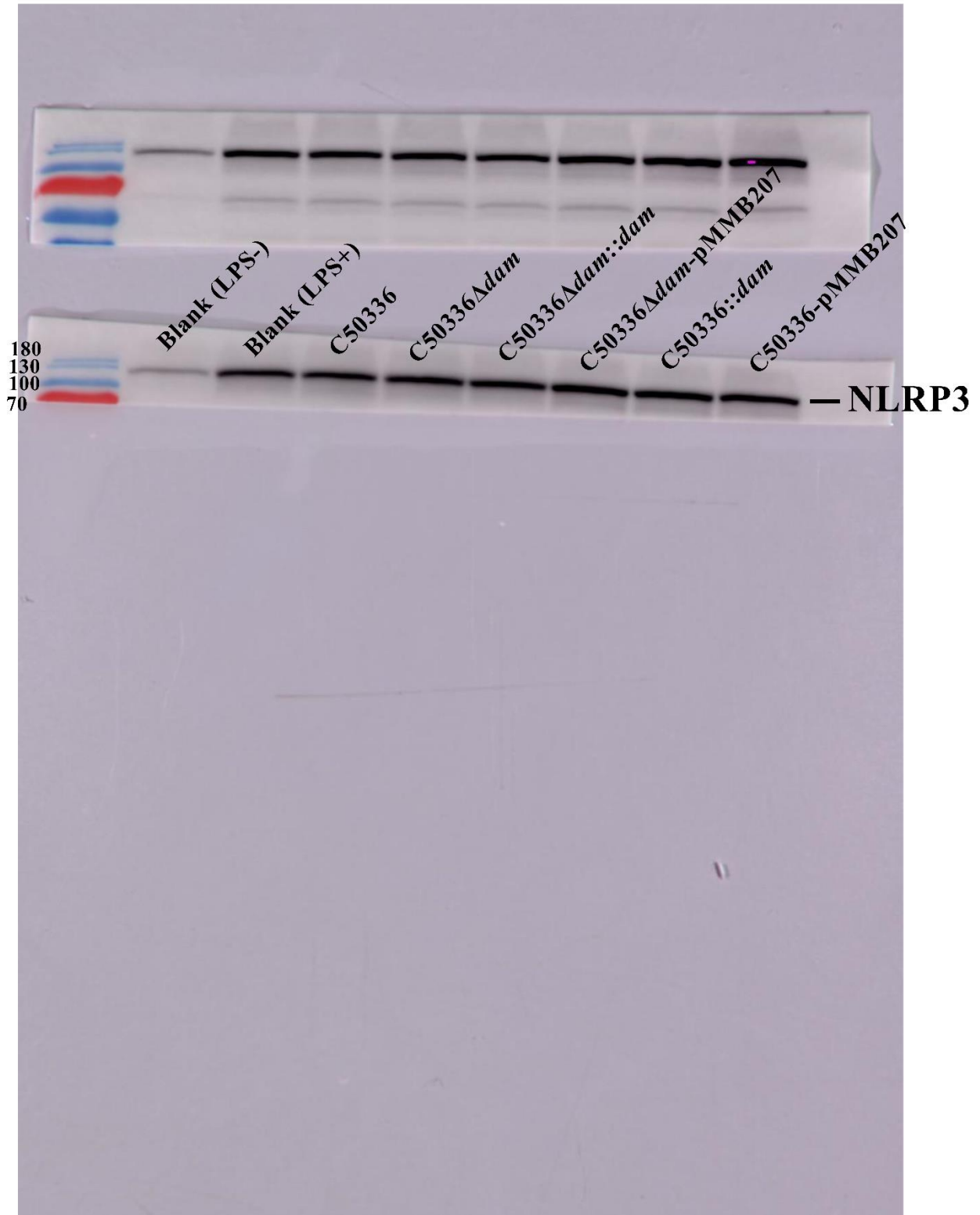


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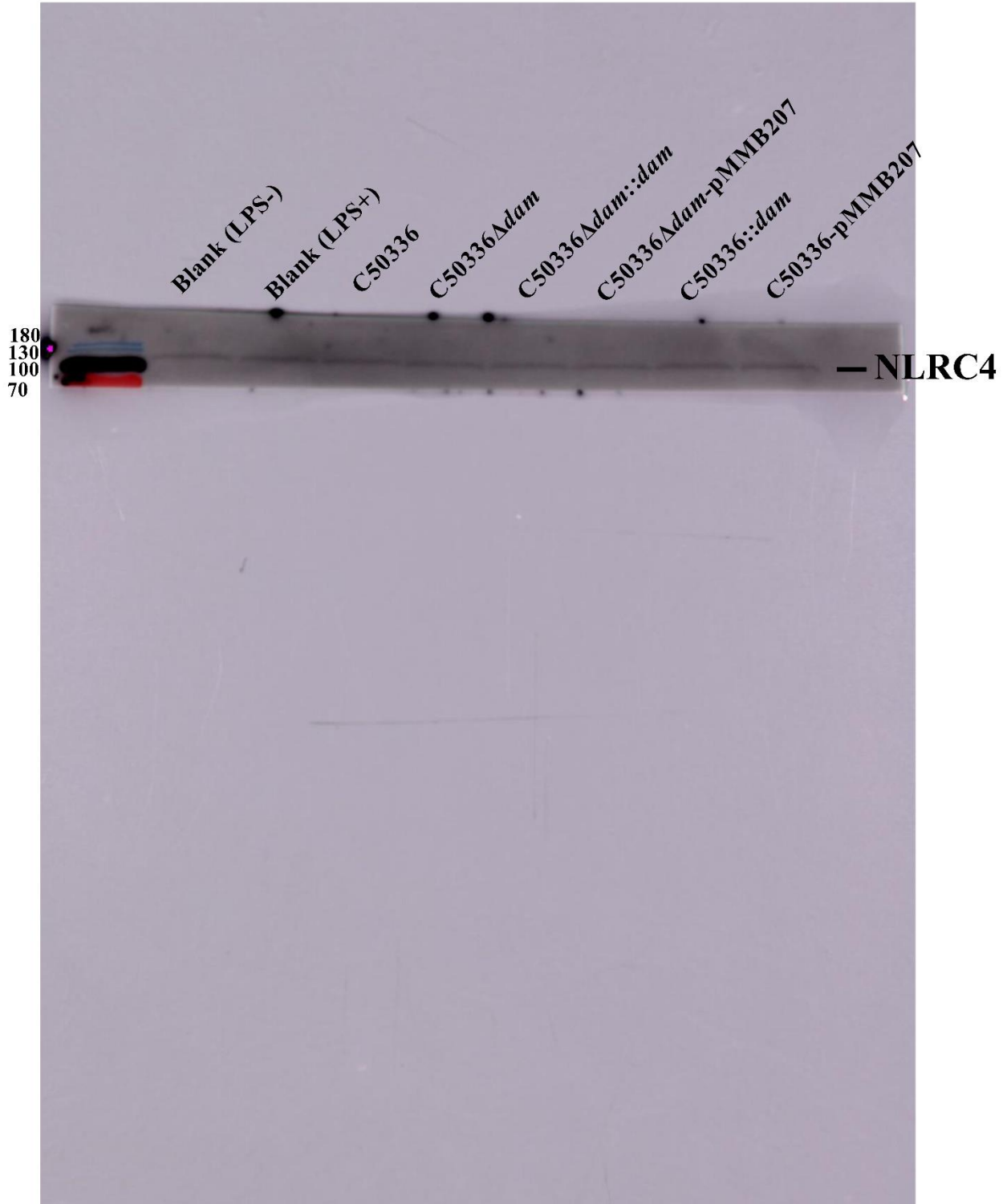


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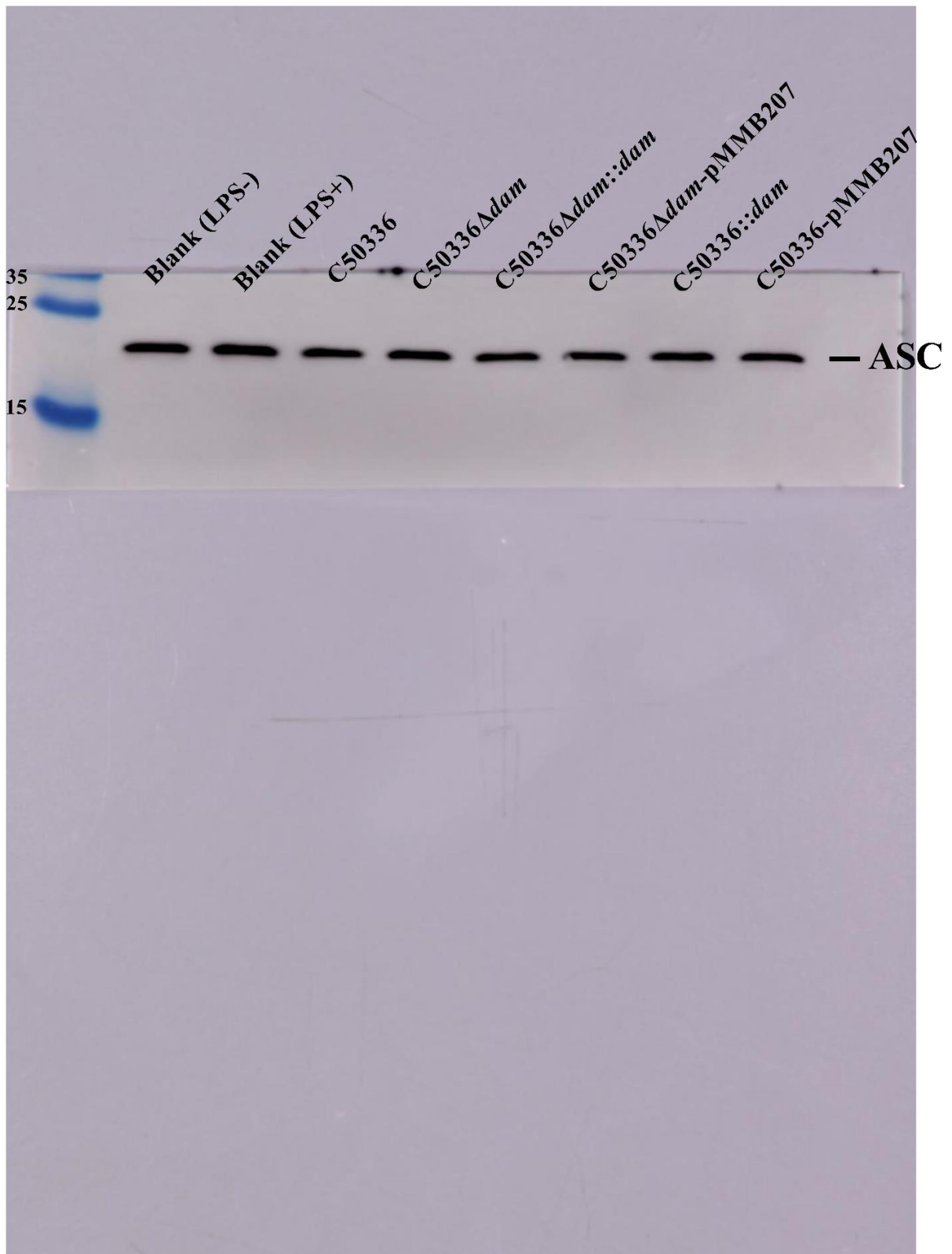


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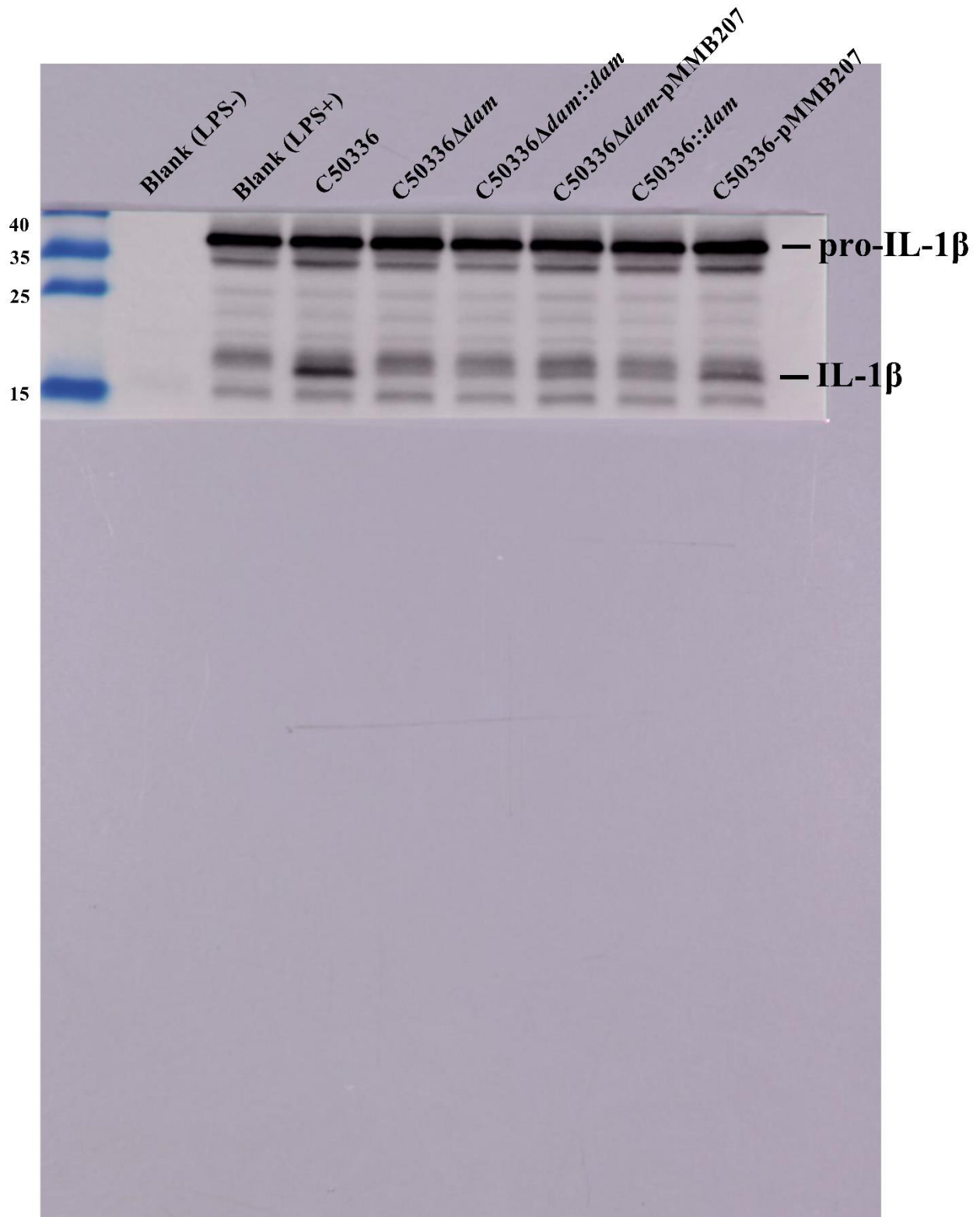


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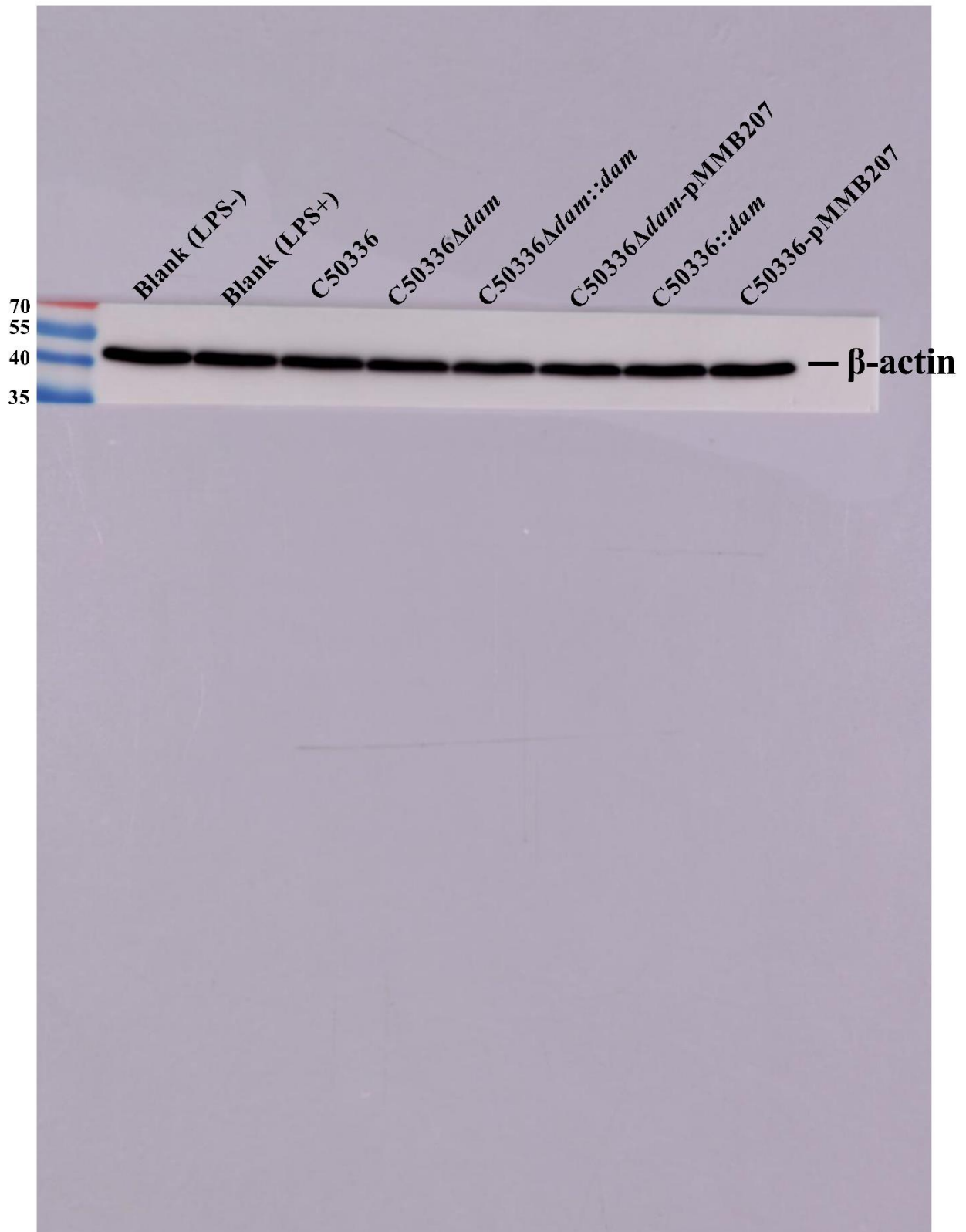


Figure S6

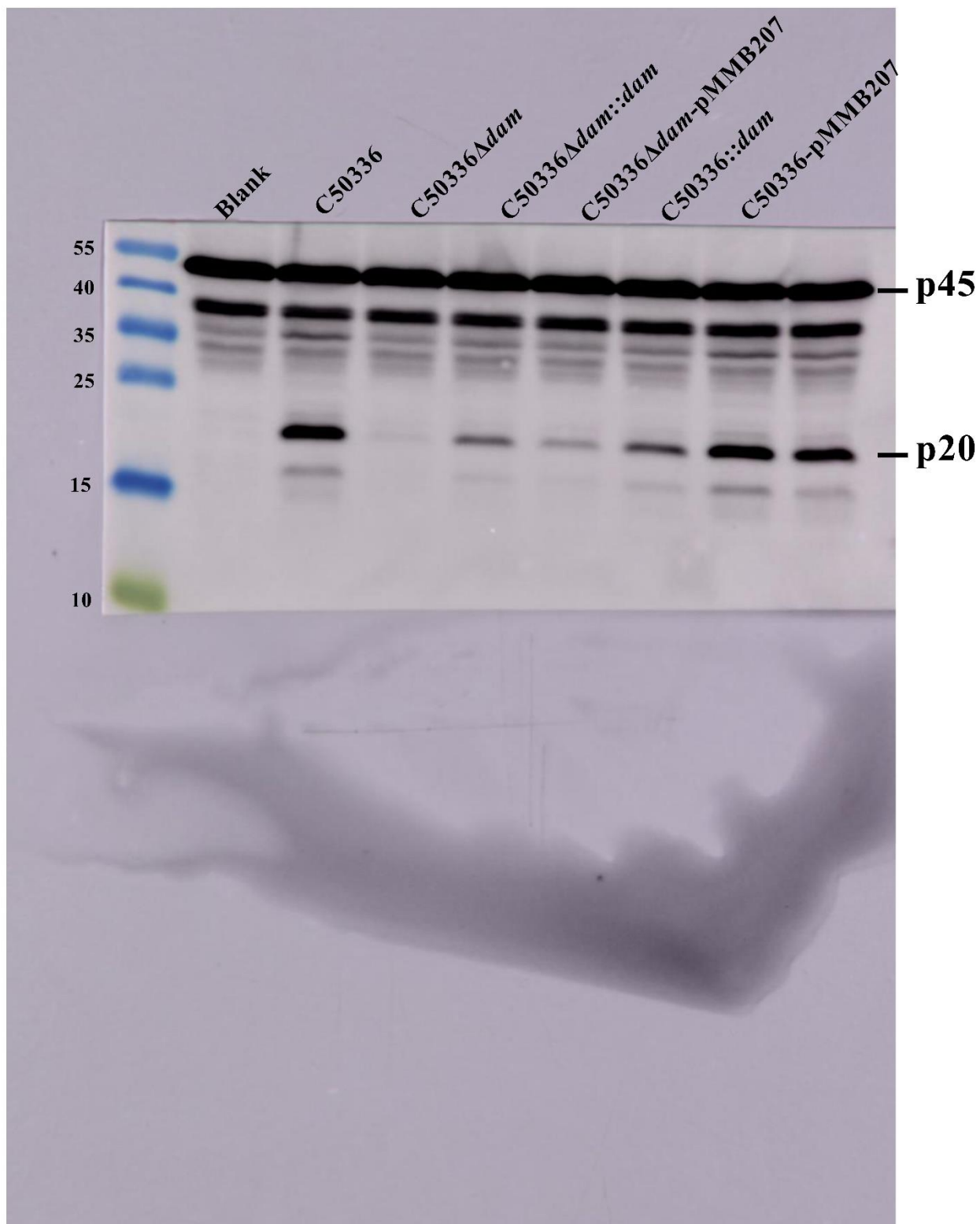


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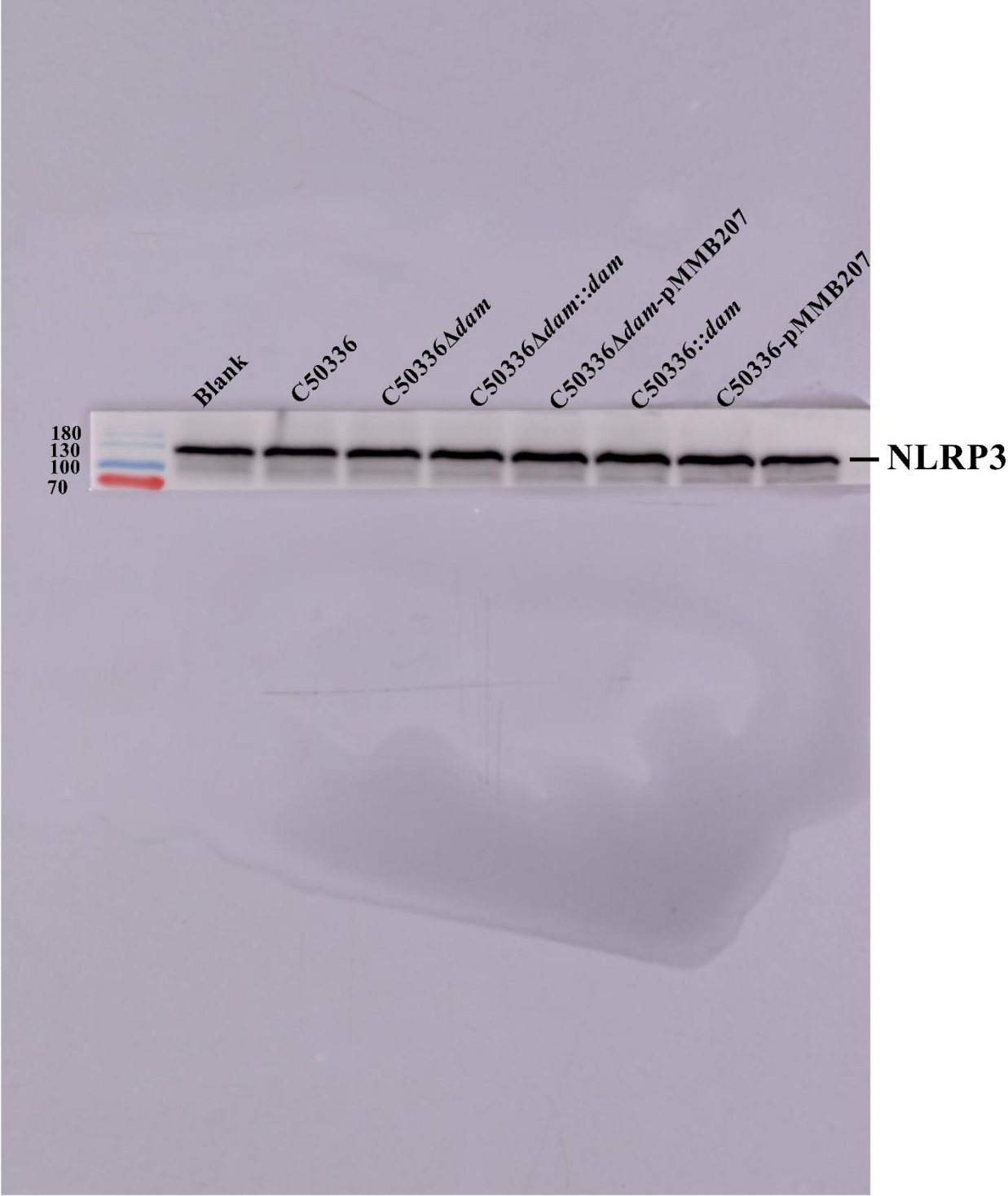


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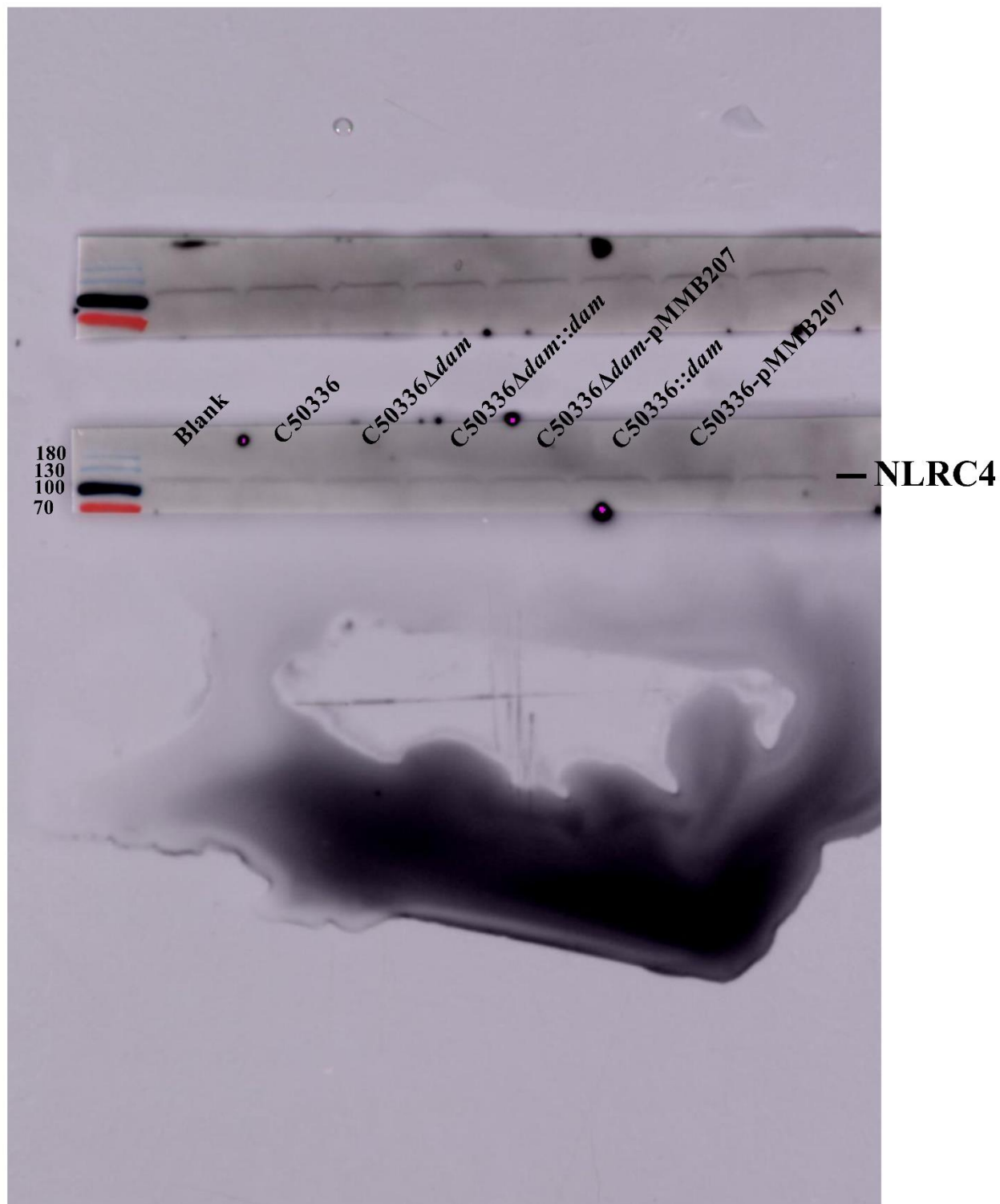


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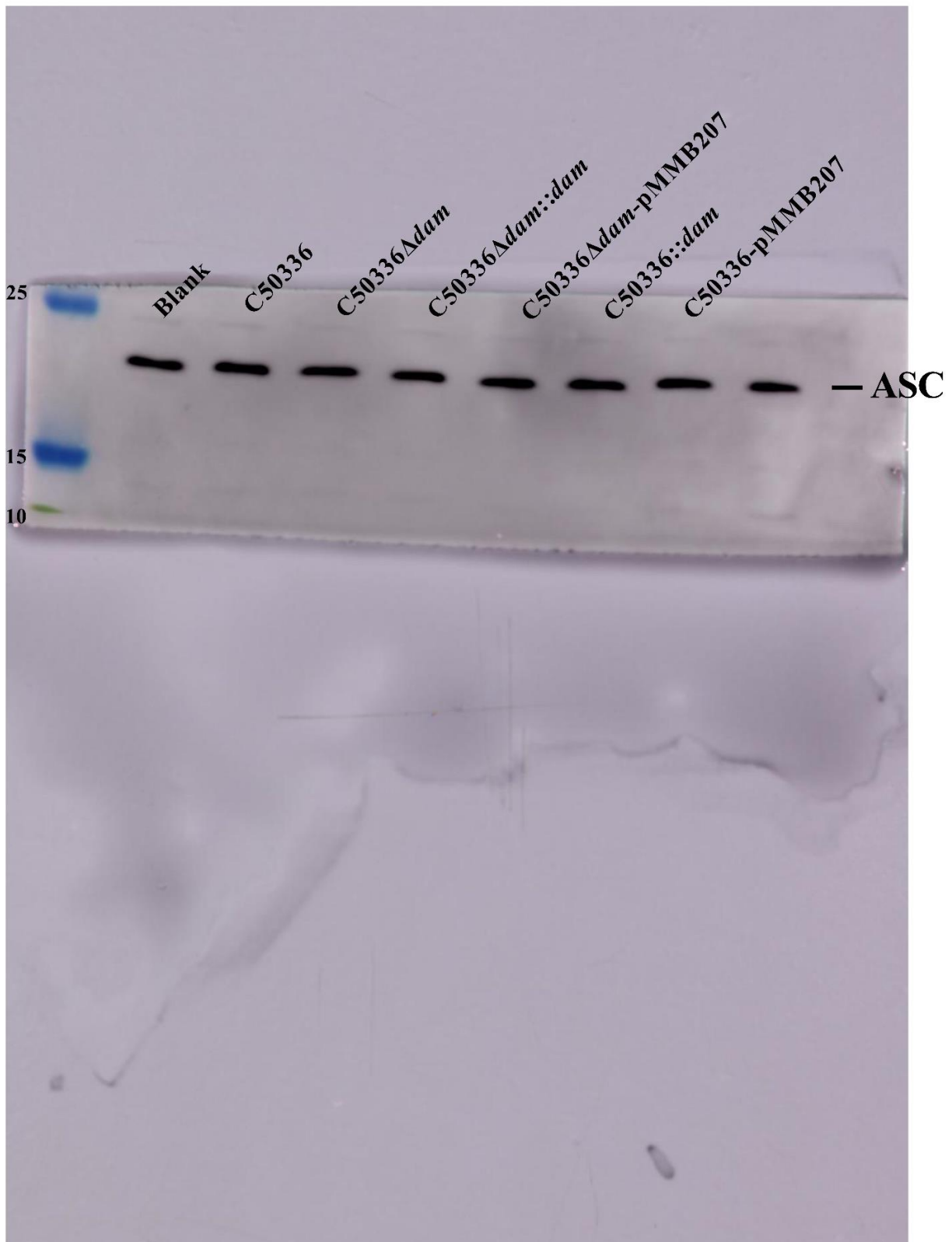


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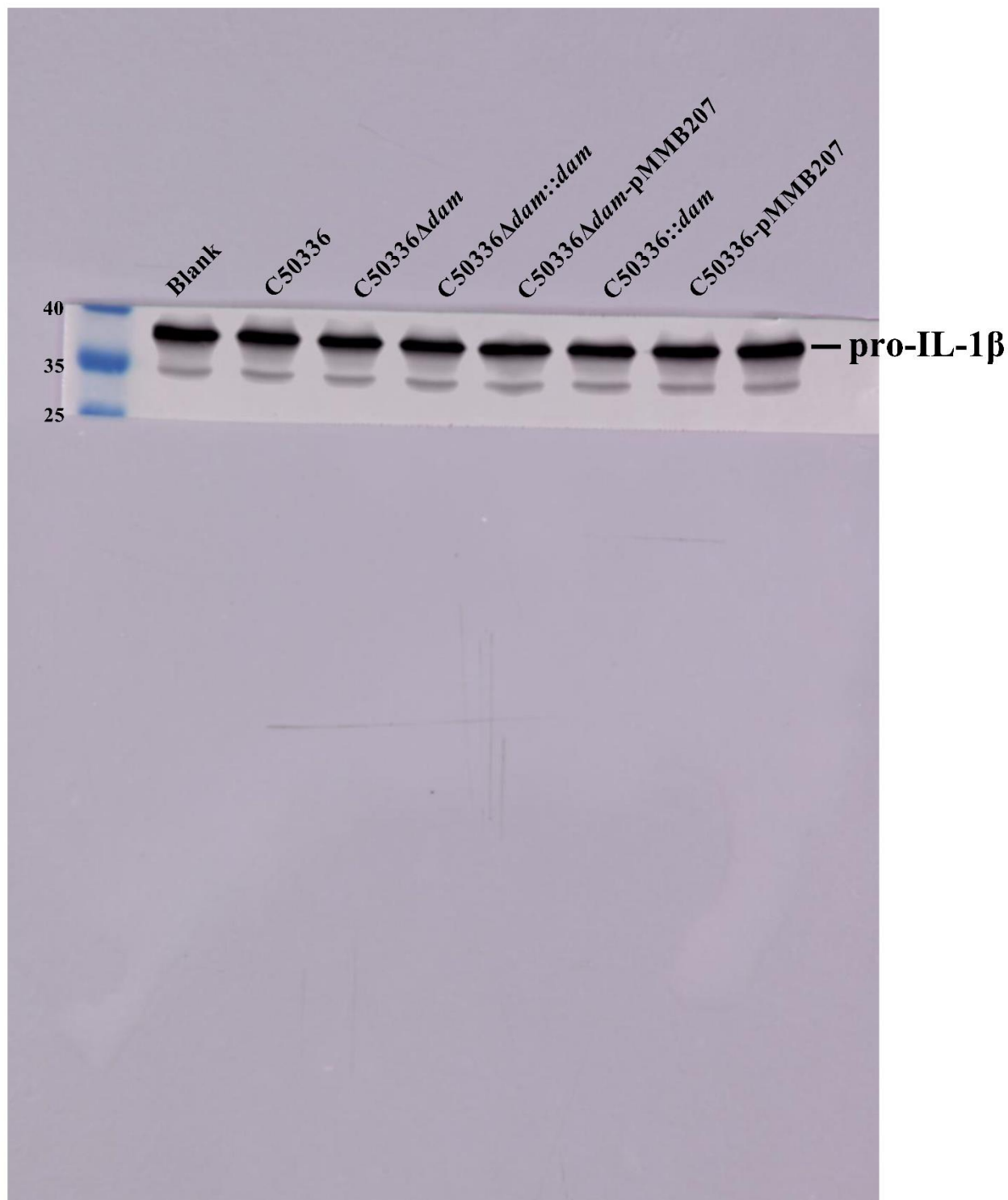


Figure S6

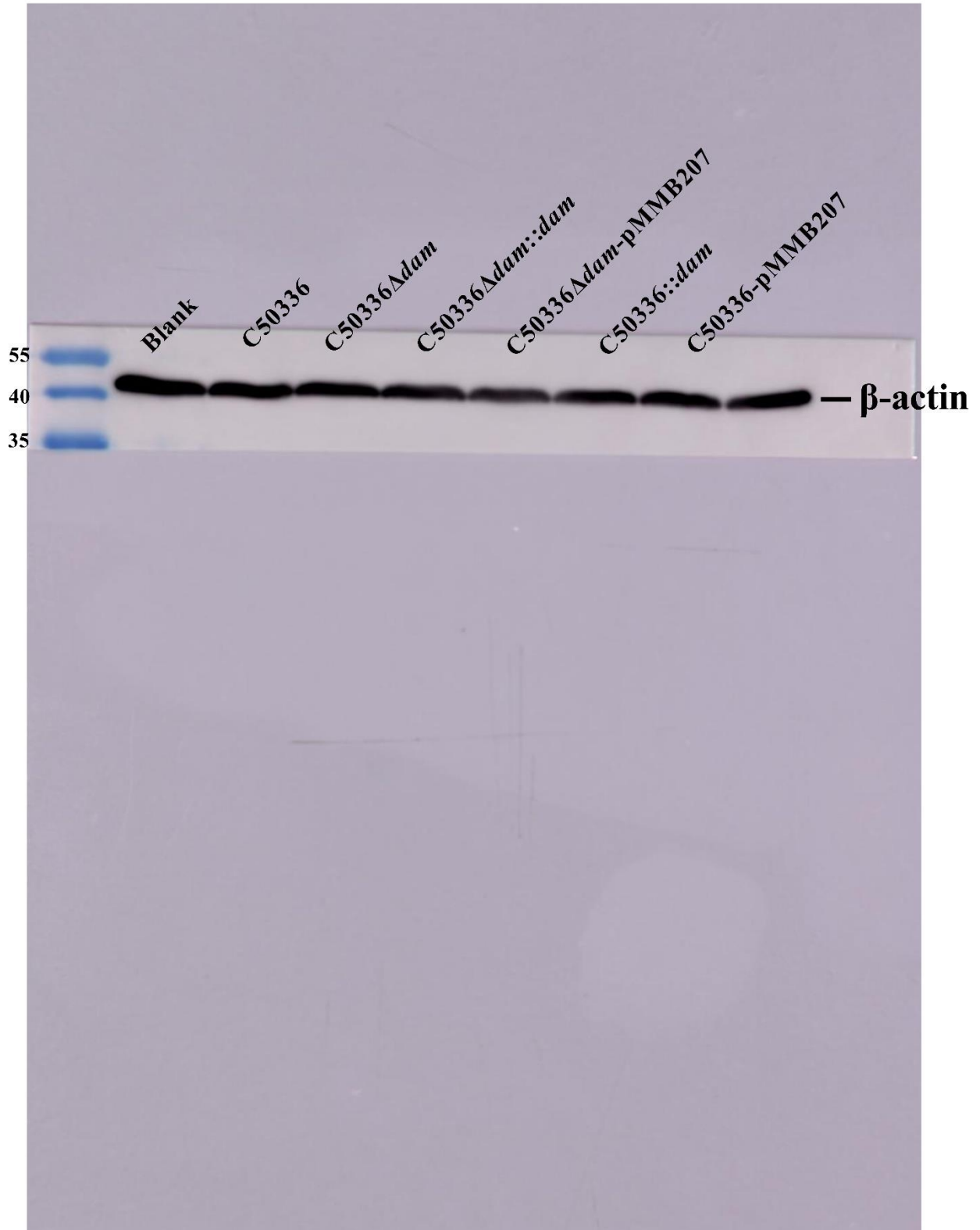


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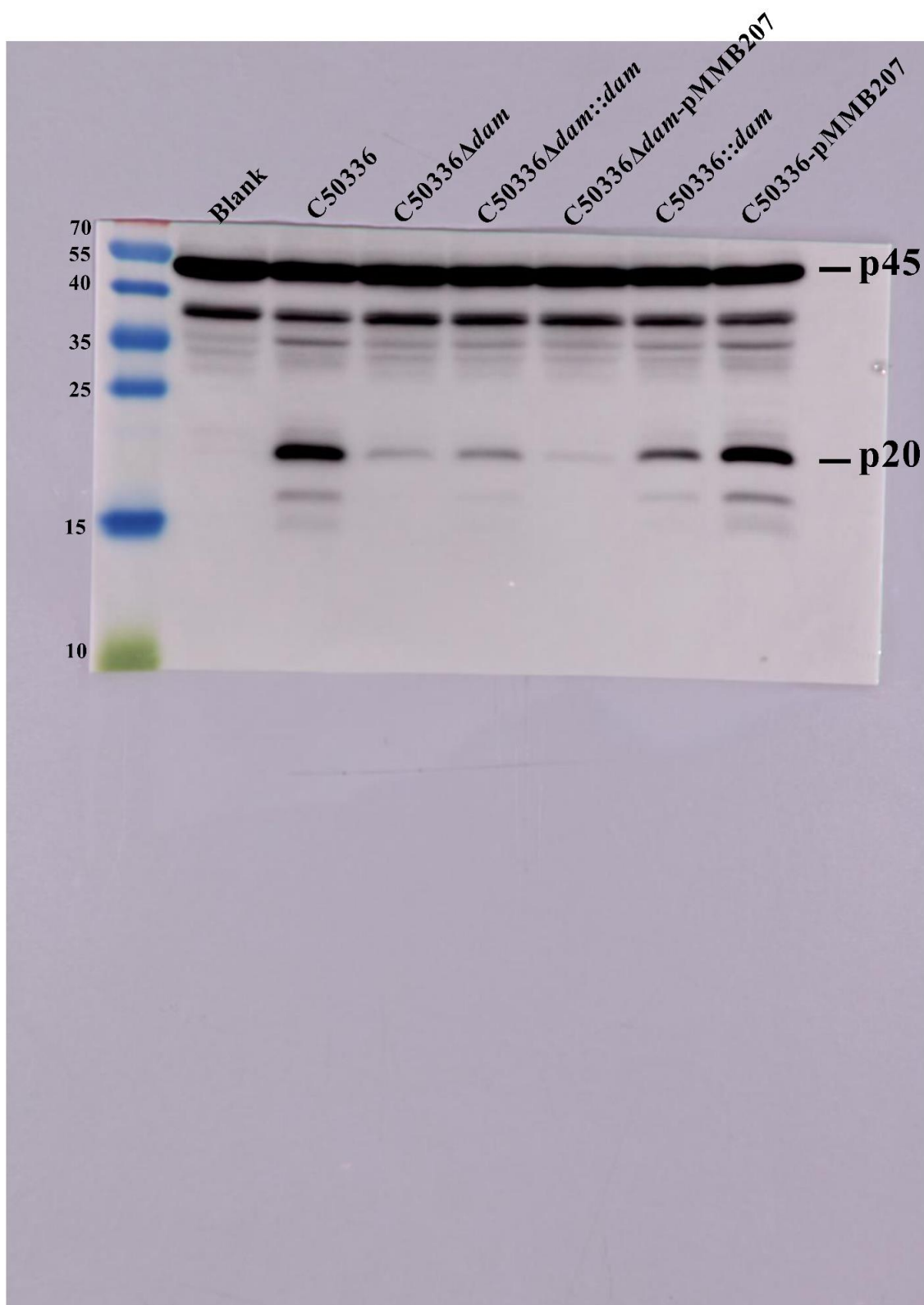


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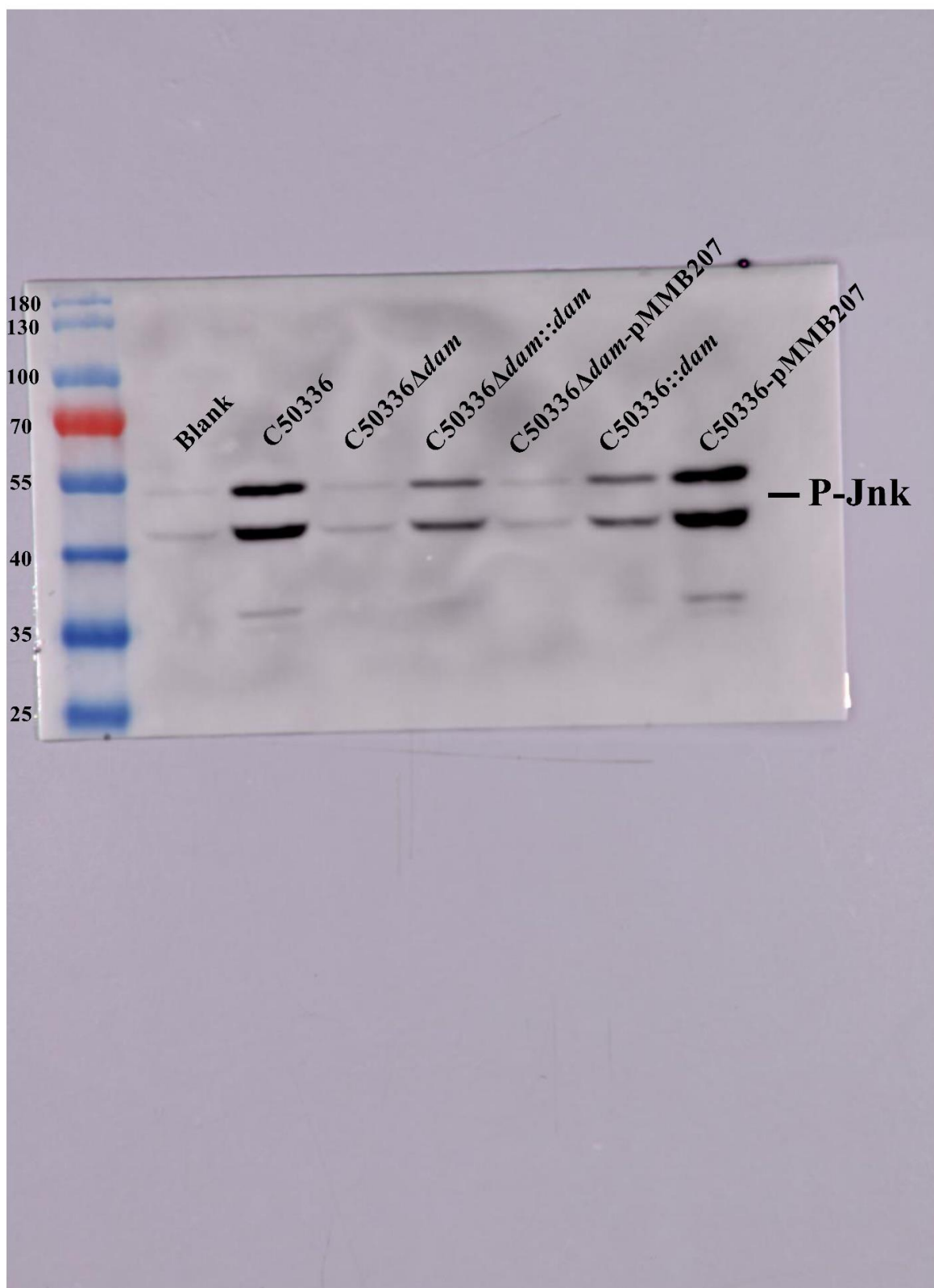


Figure S7

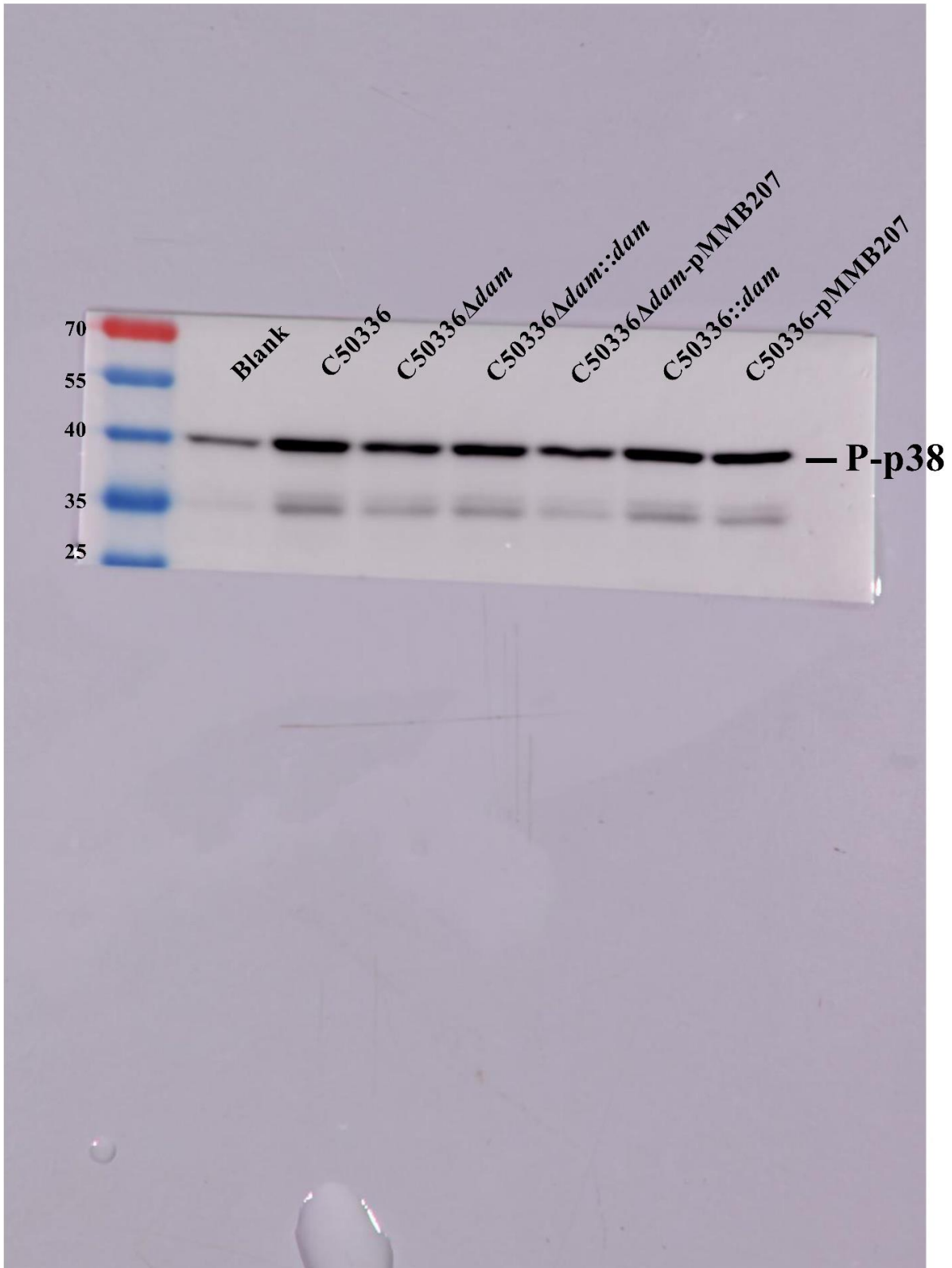


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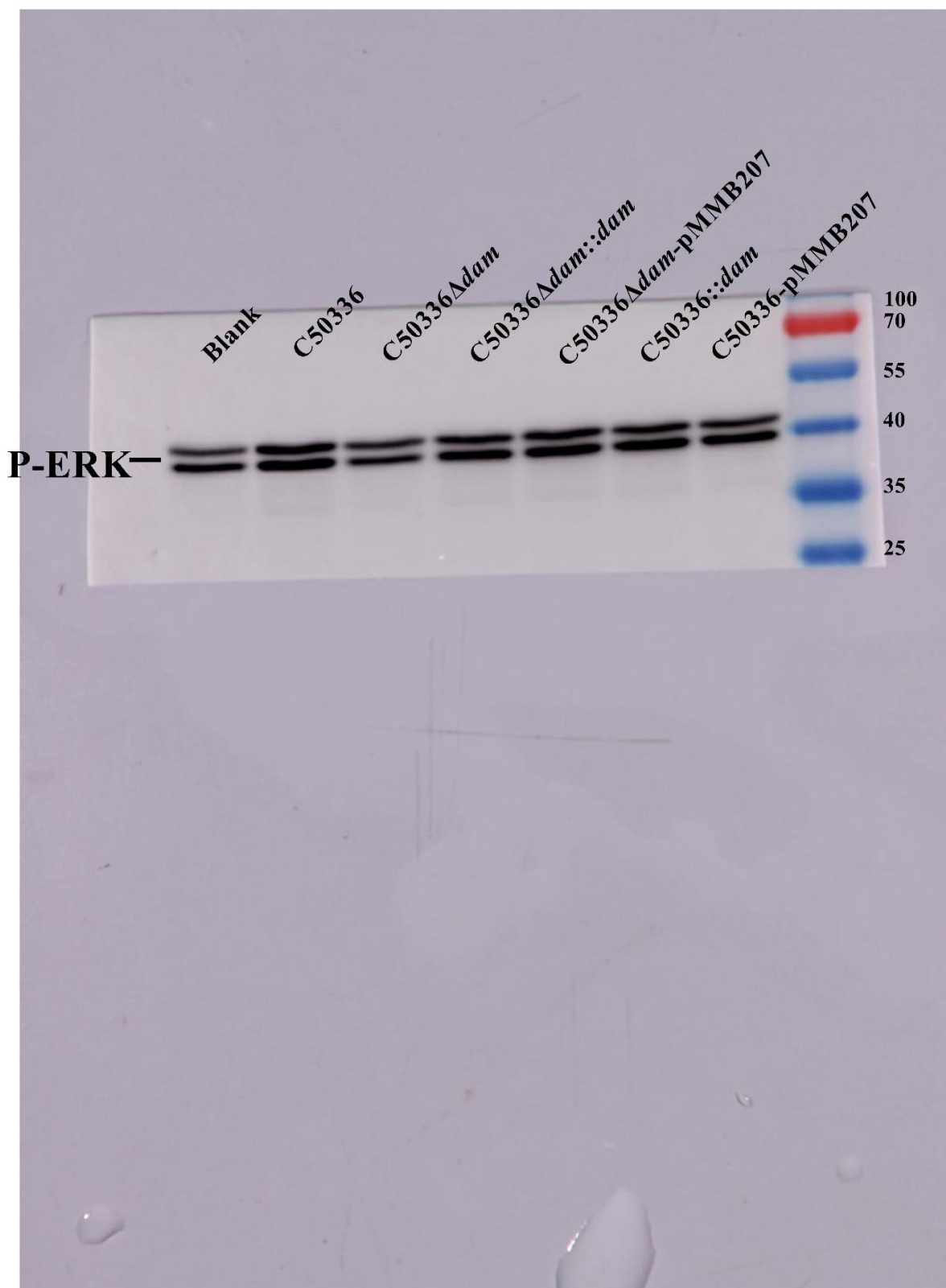


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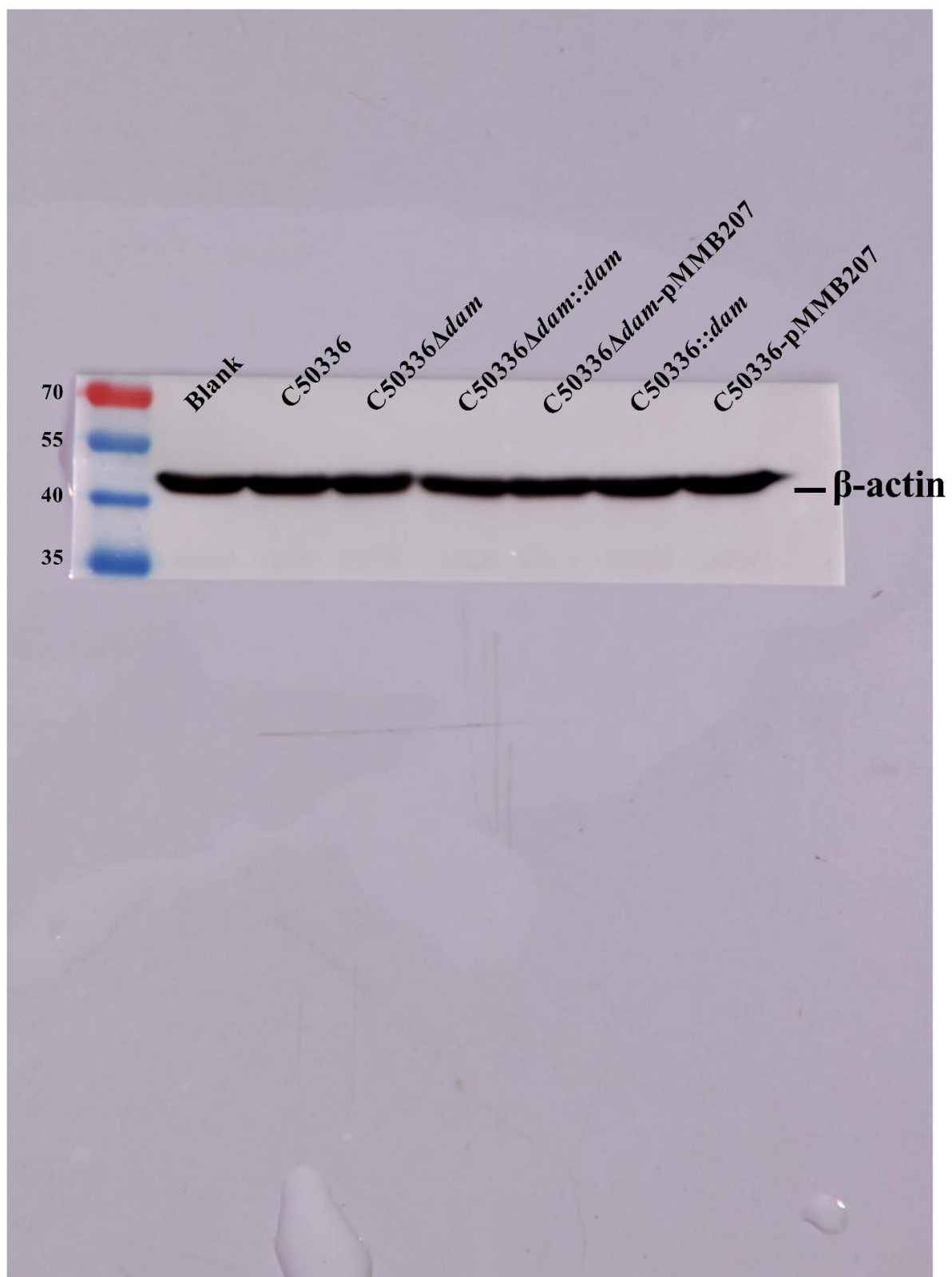


Figure S8

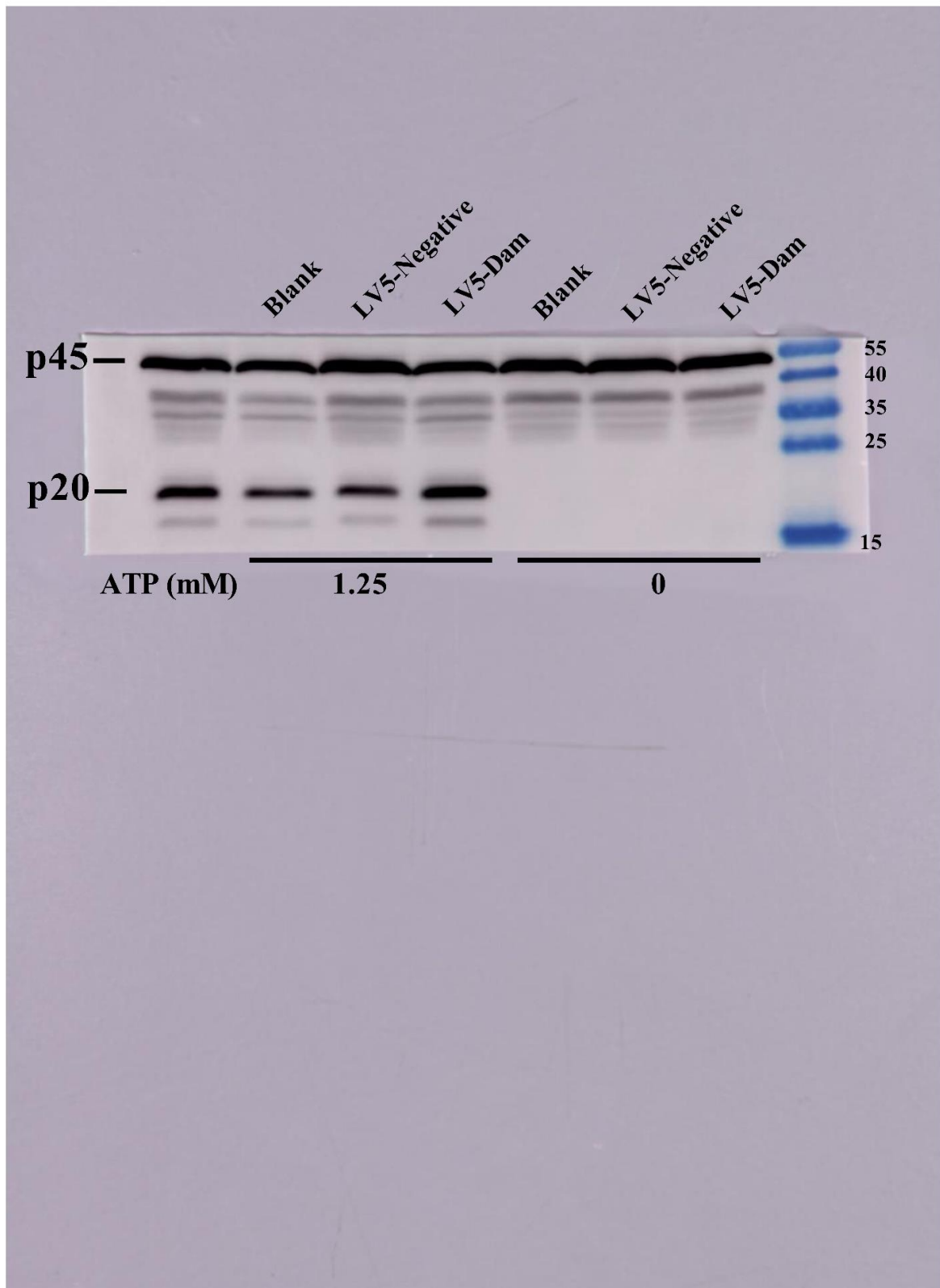


Figure S8

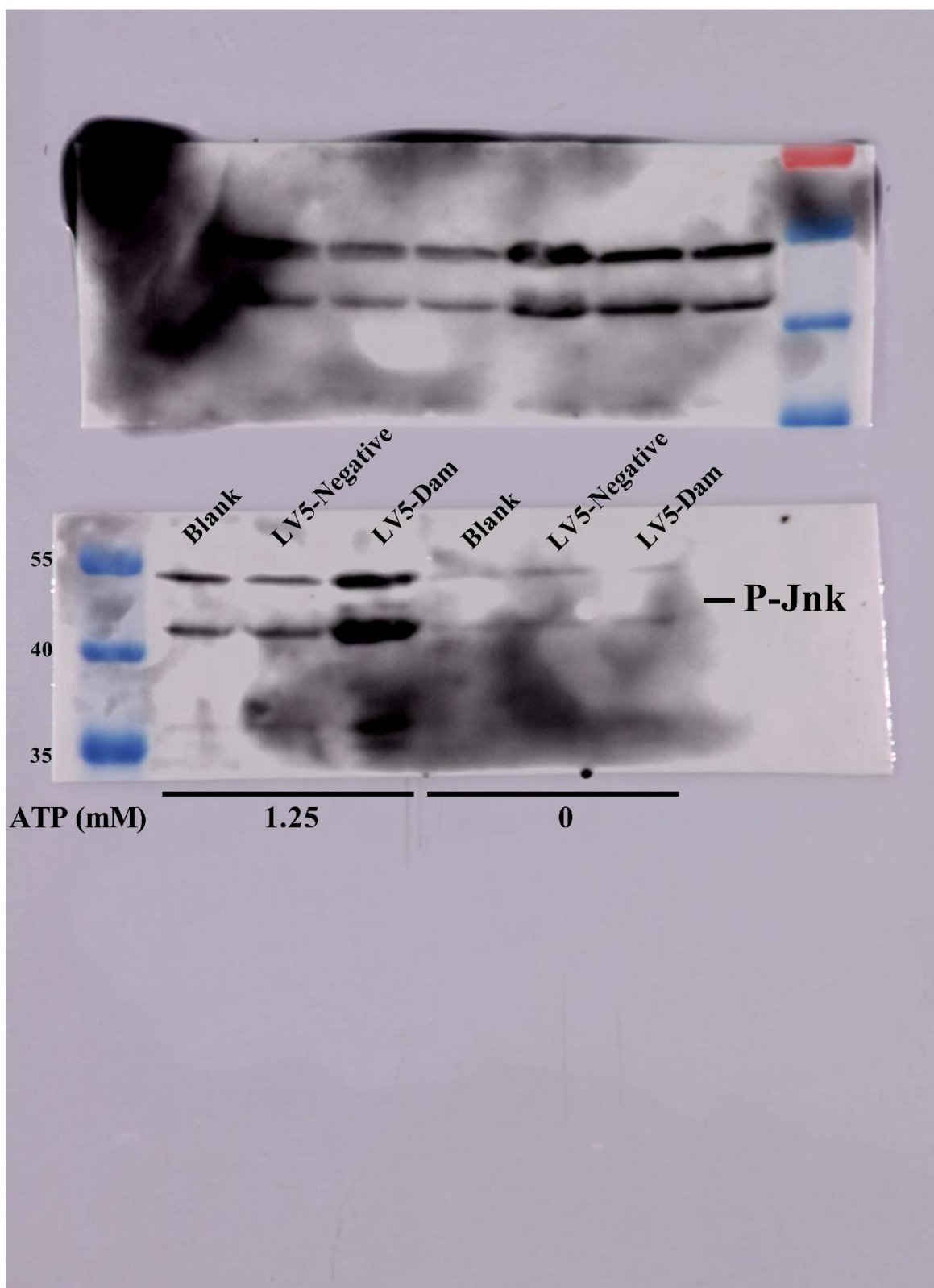


Figure S8

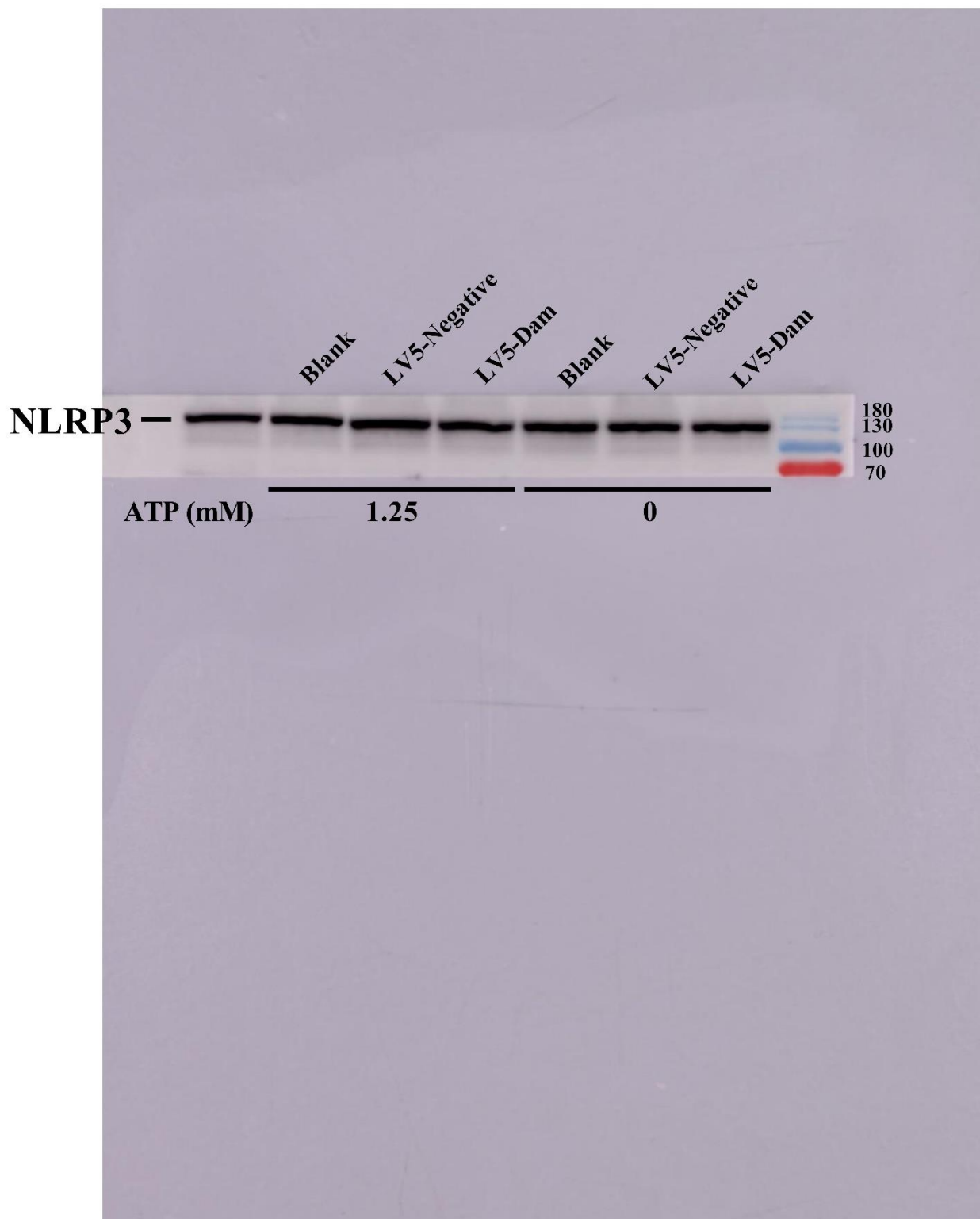


Figure S8

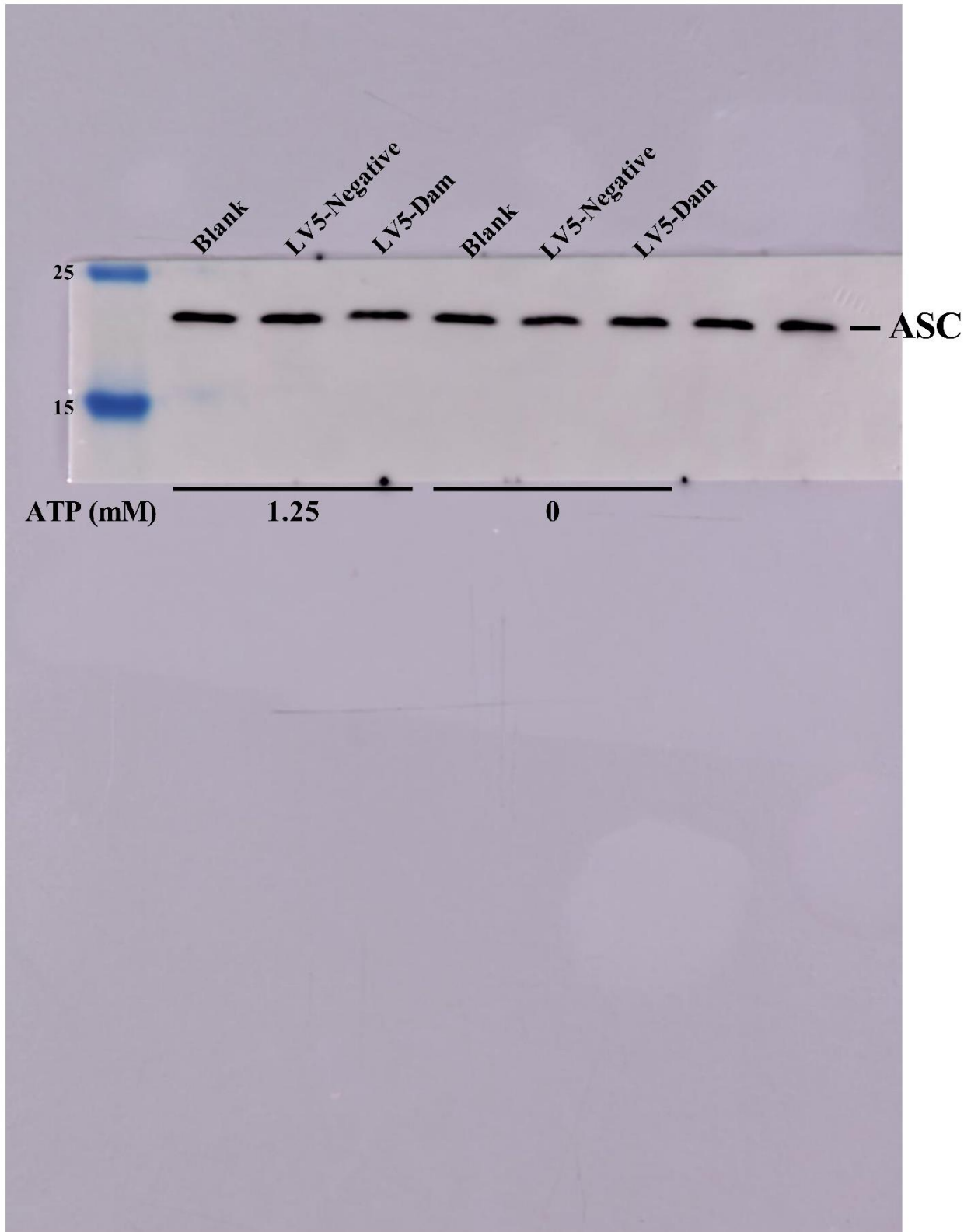


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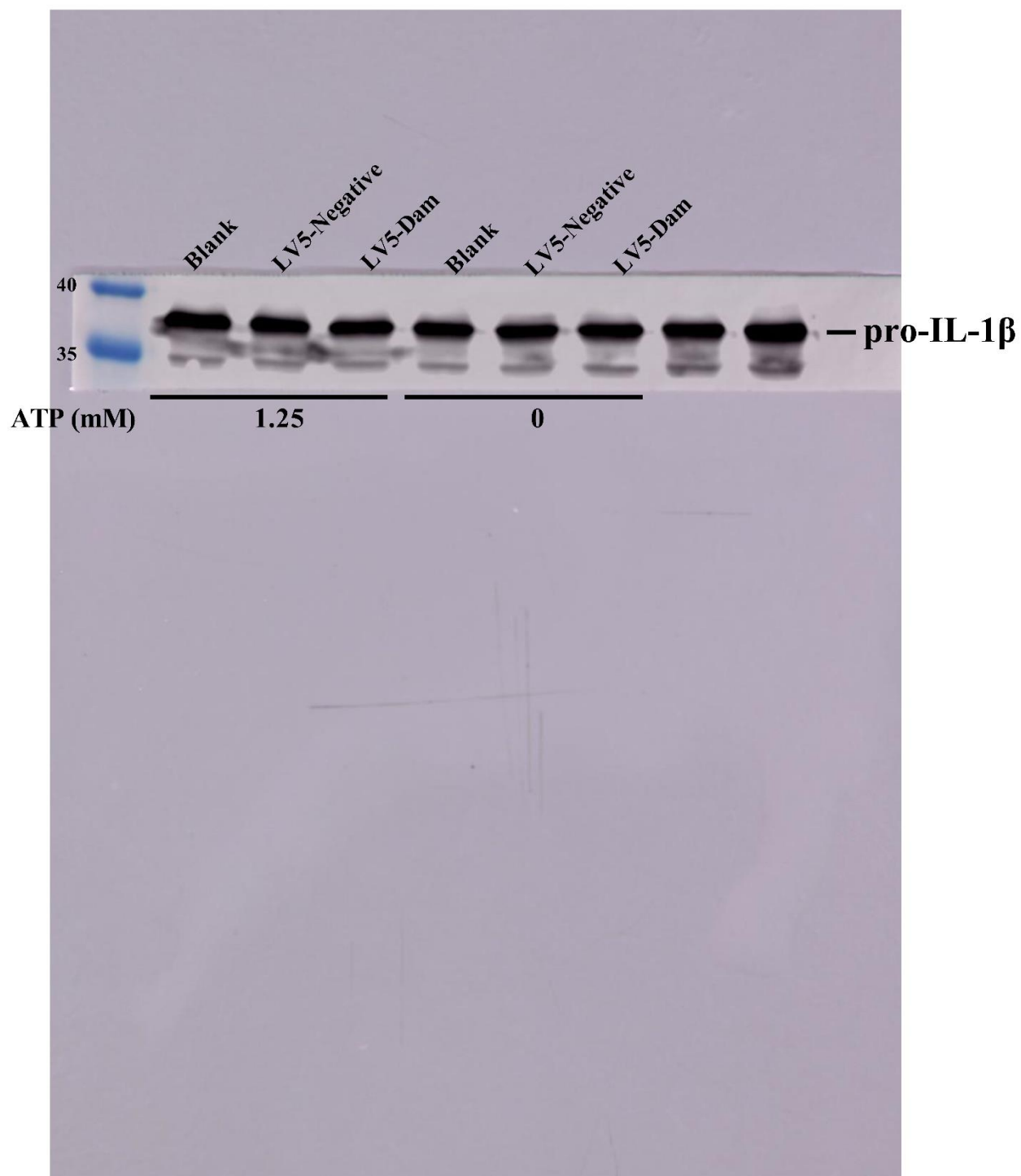
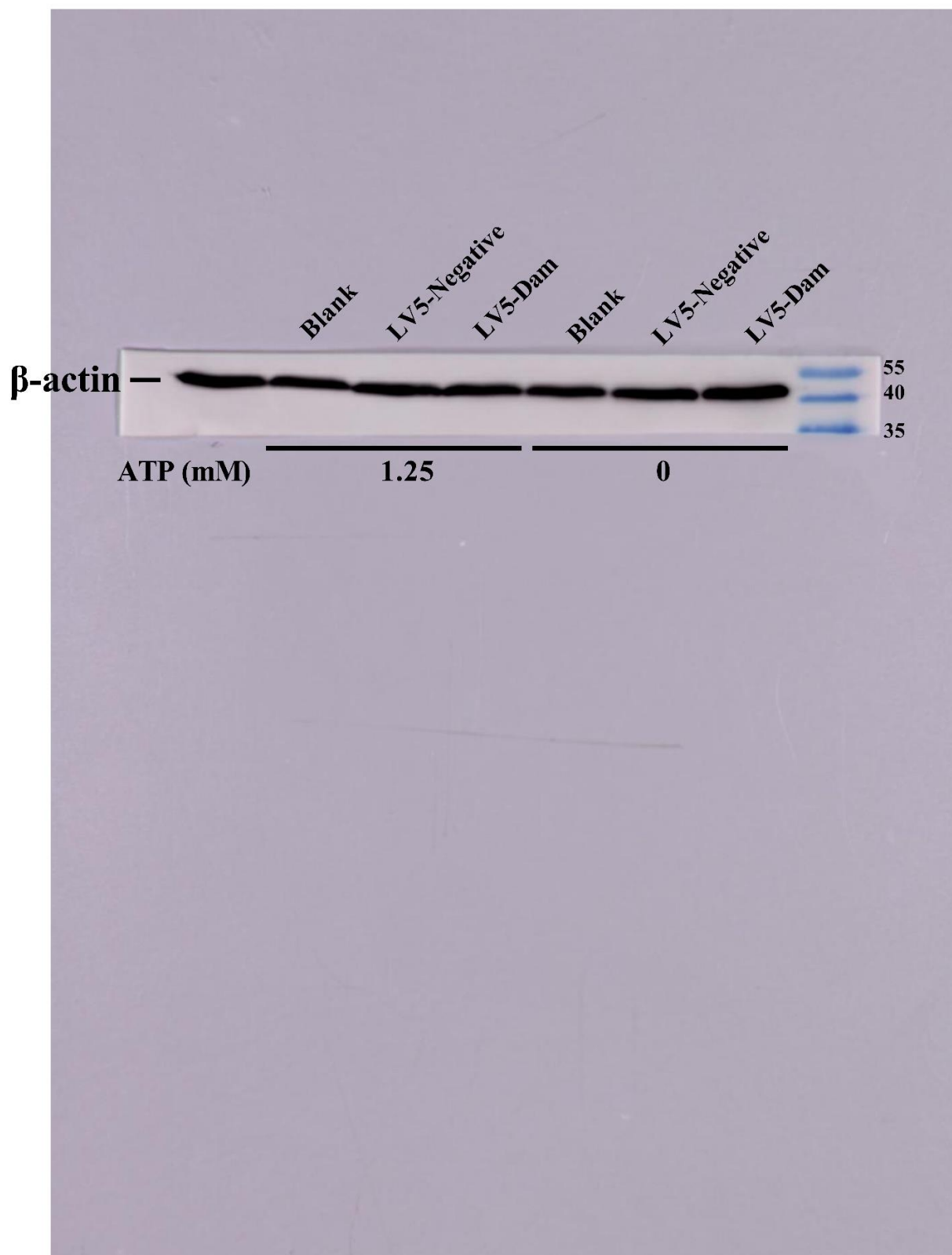


Figure S8



Supplemental figure legends

Fig. S1 Growth curves of *dam* (a), *invC* (b), *hilD* (c), *prgH* (d), and *spaN* (e) gene deletion mutants, complementation, and overexpression strains. Bacteria were grown in liquid LB medium at 37 °C for 12 h with agitation, and the OD₆₀₀ values of triplicate cultures in LB medium were determined in 1-h intervals.

Fig. S2 Two rounds of screening to identify the genes involved in regulating inflammasome activation *in vitro*. J774A.1 cells were pre-treated with LPS (1 µg/mL, 5 h) and then infected with candidate transposon mutants at an MOI of 20 for 4 h, uninfected cells was used as a negative control (Blank). The activation of caspase-1 (p20) was examined via western blot. β-actin was blotted as a loading control. Molecular mass markers in kDa are indicated on the left.

Fig. S3 Deletion mutants of *dam*, *invC*, *prgH*, and *spaN* failed to induce inflammasome activation. J774A.1 cells were pre-treated with LPS (1 µg/mL, 5 h) and then infected with WT strain C50336 and *dam*, *invC*, *hilD*, *prgH*, and *spaN* gene deletion mutants at an MOI of 20 for 4 h, uninfected cells was used as a negative control (Blank). The activation of caspase-1 (p20) was examined via western blot. β-actin was blotted as a loading control. Molecular mass markers in kDa are indicated on the left.

Fig. S4 Deletion of *prgH* did not influence the synthesis of inflammasome components. C57BL/6 BMDMs were pre-treated with LPS (1 µg/mL) for 5 h (untreated and uninfected BMDMs was used as a negative control, Blank LPS-), and then infected with C50336, C50336Δ*prgH*, C50336Δ*prgH*::*prgH*, C50336Δ*prgH*-pMMB207, C50336::*prgH*, or C50336-pMMB207 at an MOI of 20 for 4 h, uninfected BMDMs

was used as another negative control (Blank LPS+). Bacteria bearing pMMB207 plasmids were cultured with IPTG (0.5 mM). The expression of caspase-1, NLRP3, NLRC4, ASC, and pro-IL-1 β were analyzed by immunoblotting. β -actin was blotted as a loading control. Molecular mass markers in kDa are indicated on the left.

Fig. S5 Overexpression of Dam inhibited inflammasome activation. C57BL/6 BMDMs were pre-treated with LPS (1 μ g/mL) for 5 h (untreated and uninfected BMDMs was used as a negative control, Blank LPS-), and then infected with C50336, C50336 Δ dam, C50336 Δ dam::*dam*, C50336 Δ dam-pMMB207, C50336::*dam*, or C50336-pMMB207 at an MOI of 20 for 4 h, uninfected BMDMs was used as another negative control (Blank LPS+). Bacteria bearing pMMB207 plasmids were cultured with IPTG (0.5 mM). The expression of caspase-1, NLRP3, NLRC4, ASC, and pro-IL-1 β were analyzed by immunoblotting. β -actin was blotted as a loading control. Molecular mass markers in kDa are indicated on the left.

Fig. S6 The ability of the *dam* complementation strain and overexpression strain cultured without IPTG to activate the inflammasome was improved. C57BL/6 BMDMs were pre-treated with LPS (1 μ g/mL) for 5 h (untreated and uninfected BMDMs was used as a negative control, Blank LPS-), and then infected with C50336, C50336 Δ dam, C50336 Δ dam::*dam*, C50336 Δ dam-pMMB207, C50336::*dam*, or C50336-pMMB207 at an MOI of 20 for 4 h uninfected BMDMs was used as another negative control (Blank LPS+). Bacteria bearing pMMB207 plasmids were cultured without IPTG. The expression of caspase-1, NLRP3, NLRC4, ASC, and pro-IL-1 β were analyzed by immunoblotting. β -actin was blotted as a loading control. Molecular mass markers in kDa are indicated on the left.

Fig. S7 The deletion of Dam blocked the Jnk pathway in infected J774A.1 cells. J774A.1 cells were pre-treated with LPS (1 $\mu\text{g}/\text{mL}$, 5 h) and then infected with C50336, C50336 Δdam , C50336 $\Delta dam::dam$, C50336 Δdam -pMMB207, C50336::dam, or C50336-pMMB207 at an MOI of 20 for 4 h, uninfected cells was used as a negative control (Blank). Bacteria bearing pMMB207 plasmids were cultured without IPTG. The activation of caspase-1, phosphorylated Jnk (P-Jnk), phosphorylated p38 (P-p38), and phosphorylated ERK1/2 (P-ERK1/2) were analyzed by immunoblotting. β -actin was blotted as a loading control. Molecular mass markers in kDa are indicated on the left.

Fig. S8 Dam independently promoted NLRP3 inflammasome activation. J774A.1 cells were transduced with LV5-Dam or LV5-negative lentivirus, untreated cells was used as a negative control (Blank). Cells were then pre-treated with LPS (1 $\mu\text{g}/\text{mL}$, 5 h) and stimulated with or without ATP (1.25 mM) for 1 h. The expression of caspase-1, NLRP3, ASC, pro-IL-1 β , and P-Jnk were analyzed by immunoblotting. β -actin was blotted as a loading control. Molecular mass markers in kDa are indicated on the left.