

## Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

### Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size ( $n$ ) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided  
*Only common tests should be described solely by name; describe more complex techniques in the Methods section.*
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g.  $F$ ,  $t$ ,  $r$ ) with confidence intervals, effect sizes, degrees of freedom and  $P$  value noted  
*Give  $P$  values as exact values whenever suitable.*
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's  $d$ , Pearson's  $r$ ), indicating how they were calculated

*Our web collection on [statistics for biologists](#) contains articles on many of the points above.*

### Software and code

Policy information about [availability of computer code](#)

Data collection	Sequence data was collected using the manufacturers software: Illumina was collected using HiSeq 2500 Control Software (HCS) v2.0.12 and Real Time Analysis (RTA) v1.18.61. PacBio sequence was collected using RSII Instrument Control Software v2.3.0 and SMRT Analysis v2.3.0
Data analysis	All software used was listed and described in the manuscript. The following programs were used: ALLPATHS (vLG); MST map (v1.0.0-2015); Meraculous assembler (2.2.2.5 release); HiRiseTM assembler; HipMer genome assembler (v0.9.6); ALLMAPS (v0.7.5); PASA (v2.3); EXONERATE (v2.4.0); RepeatMasker (v4.0.5); FGENESH+ (v2.0); FGENESH EST (v2.6); IQTREE v. 1.6.7; Progressive Cactus (v0.1); ZEN 2.3 Pro (Zeiss); Photoshop CS3 (Adobe); NOVOPlasty v.2.7.1; blasr v.5.3.2; samtools v.1.4.1; Canu v.1.7.1; IGV v.2.3.8; MAFFT v.7.215; trimAl v.1.2rev59; Maffilter (v1.3); MafStrander (v1.0); FigTree (v. 1.4.0); STRUCTURE HARVESTER (v. 0.9.94); DISTRUCT (v. 1.1); BEAST 2.4.7 software; polyCRACKER; TRACER v.1.6; sourmash (v2.0.0); Plotly Dash (v0.30.0); GET_HOMOLOGUES-EST v09052018; Rstat package v. 4.1.0; Picard tools (v2.18); GATK (v4.0); BBtools (v. 38.0); HTSeq (v. 0.9.1); DESeq2 v. 1.24.0; bedtools (v2.27.1); Biopython (v. 1.7.0); PAML (v. 4.9h)

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research [guidelines for submitting code & software](#) for further information.

### Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

Genome assemblies and annotations can be downloaded from Phytozome [<https://phytozome.jgi.doe.gov/>]. The direct link for the B. hybridum ABR113 reference

genome is [[https://phytozome-next.jgi.doe.gov/info/Bhybridum\\_v1\\_1](https://phytozome-next.jgi.doe.gov/info/Bhybridum_v1_1)], the direct link for the *B. stacei* ABR114 reference genome is [[https://phytozome-next.jgi.doe.gov/info/Bstacei\\_v1\\_1](https://phytozome-next.jgi.doe.gov/info/Bstacei_v1_1)] and the direct link for the *B. distachyon* Bd21 reference genome is [[https://phytozome-next.jgi.doe.gov/info/Bdistachyon\\_v3\\_1](https://phytozome-next.jgi.doe.gov/info/Bdistachyon_v3_1)]. The other genome assemblies and annotations created in this study (listed in supplementary data 1) can be downloaded from the *B. hybridum* genome page [[https://phytozome-next.jgi.doe.gov/info/Bhybridum\\_v1\\_1](https://phytozome-next.jgi.doe.gov/info/Bhybridum_v1_1)] through the download directory labelled “Additional genomes used in Gordon et al. Nat. Comm. 2020” direct link [<https://genome.jgi.doe.gov/portal/pages/dynamicOrganismDownload.jsf?organism=Bhybridum>]. Note that a free account is required to download data from Phytozome. The raw reads for the genomic sequences and RNA sequences are available from NCBI or ENA. The samples numbers and links are provided in Supplementary data 4 and 9. Seeds for the lines used in this study are available from the USDA National Plant Germplasm Service or by request from the authors. The source data underlying Figs 6,7 and 8 and Supplementary Figs 9,11 and 12 are provided as an Excel file named Source Data file.

## Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Life sciences       Behavioural & social sciences       Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://nature.com/documents/nr-reporting-summary-flat.pdf)

## Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	<input type="text" value="na"/>
Data exclusions	<input type="text" value="no data was excluded"/>
Replication	<input type="text" value="all replicates were included"/>
Randomization	<input type="text" value="na"/>
Blinding	<input type="text" value="na"/>

## Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

### Materials & experimental systems

n/a	<input type="checkbox"/> Involved in the study
<input checked="" type="checkbox"/>	<input type="checkbox"/> Antibodies
<input checked="" type="checkbox"/>	<input type="checkbox"/> Eukaryotic cell lines
<input checked="" type="checkbox"/>	<input type="checkbox"/> Palaeontology and archaeology
<input checked="" type="checkbox"/>	<input type="checkbox"/> Animals and other organisms
<input checked="" type="checkbox"/>	<input type="checkbox"/> Human research participants
<input checked="" type="checkbox"/>	<input type="checkbox"/> Clinical data
<input checked="" type="checkbox"/>	<input type="checkbox"/> Dual use research of concern

### Methods

n/a	<input type="checkbox"/> Involved in the study
<input checked="" type="checkbox"/>	<input type="checkbox"/> ChIP-seq
<input checked="" type="checkbox"/>	<input type="checkbox"/> Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/> MRI-based neuroimaging