

Supplementary information submitted to Scientific Reports

## **Gene products and processes contributing to lanthanide homeostasis and methanol metabolism in *Methylobacterium extorquens* AM1**

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Table S1. Growth parameters for strains grown in methanol medium with and without La<sup>3+</sup>.

Strain	Growth Rate (h <sup>-1</sup> ) <sup>‡</sup>	
	MeOH	MeOH La <sup>3+</sup>
<b>Growth of control strains</b>		
Wild type	0.14 ± 0.01	0.16 ± 0.01
<i>mxoF</i>	NG	0.16 ± 0.01
<i>xoxF1</i>	30 h lag, 0.09 ± 0.01	6-9 h lag, 0.07 ± 0.00
<i>xoxF1 xoxF2</i>	NG*	6 h lag, 0.04 ± 0.01
<i>mxoF xoxF1 xoxF2</i>	NG	6 h lag, 0.04 ± 0.00
<b>Growth of strains defective in methanol metabolism independent of La<sup>3+</sup></b>		
<i>pqqBCDE</i>	NG	NG
<i>pqqF</i>	NG	NG
<i>cycK</i>	NG	NG
<i>ccmB</i>	NG	NG
<i>ccmC</i>	NG	NG
<i>META1_2359</i>	NG*	NG*
<i>META1_3908</i>	24 h lag, 0.05 ± 0.00	15 h lag, 0.07 ± 0.00
<i>META1p2024</i>	9 h lag, 0.09 ± 0.01	6 h lag, 0.10 ± 0.01
<i>META1_0863</i>	0.13 ± 0.00	0.14 ± 0.00

<sup>‡</sup>Data for a minimum of three biological replicates are reported.

\* indicates if a suppressor mutation allowed eventual growth of the strain.

NG indicates no growth. Methanol is abbreviated as MeOH.

Table S2. Strains and plasmids used in this study.

Strain or plasmid	Description	Reference
<i>M. extorquens</i>		
AM1	Rif <sup>R</sup> derivative (wild type)	1
CM194.1	$\Delta mxaF$	2
ES2813	$\Delta mxaF \Delta fae$	This study
ES2432	$\Delta xoxF1$	This study
ES2435	$\Delta xoxF2 \Delta xoxF1$	This study
ES2439	$\Delta mxaF \Delta xoxF2 \Delta xoxF1$	This study
ES2386	<i>pqqBCDE::Km</i>	This study
ES2360	<i>pqqF::Km</i>	This study
ES3941	<i>cycK::Km</i>	This study
ES3968	<i>ccmB::Km</i>	This study
ES3965	<i>ccmC::Km</i>	This study
ES2817	$\Delta xoxG$	This study
ES2821	$\Delta xoxJ$	This study
ES2785	$\Delta META1p0863$	This study
ES3947	<i>MexAM1_META1p1771::Km</i>	This study
ES3282	$\Delta orf6$	This study
ES2781	$\Delta orf7$	This study
ES3962	<i>MexAM1_META1p2359::Km</i>	This study
ES3959	<i>hss::Km</i>	This study
ES3257	<i>MexAM1_META1p3908::Km</i>	This study
ES2861	<i>MexAM1_META1_3909::Km</i>	This study
ES3950	<i>lutA::Km</i>	This study
ES4122	<i>lutB::Km</i>	This study
ES4444	<i>lutC::Km</i>	This study
ES4389	<i>lutD::Km</i>	This study
ES3953	<i>lutE::Km</i>	This study
ES4249	<i>lutF::Km</i>	This study
ES3956	<i>lutG::Km</i>	This study
ES2879	<i>lutH::Km</i>	This study
ES3983	$\Delta mxaF lutA::Km$	This study
ES4124	$\Delta mxaF lutB::Km$	This study
ES4447	$\Delta mxaF lutC::Km$	This study
ES4392	$\Delta mxaF lutD::Km$	This study
ES3986	$\Delta mxaF lutE::Km$	This study

Table S2. (continued)

Strain or plasmid	Description	Reference
<b><i>M. extorquens</i></b>		
ES4246	$\Delta mxaF$ <i>lutF::Km</i>	This study
ES3989	$\Delta mxaF$ <i>lutG::Km</i>	This study
ES2962	$\Delta mxaF$ <i>lutH::Km</i>	This study
ES2231	AM1 / pCM62	This study
ES4293	<i>lutA::Km</i> / pCM62	This study
ES4295	<i>lutA::Km</i> / pES667	This study
ES5029	<i>lutB::Km</i> / pCM62	This study
ES5030	<i>lutB::Km</i> / pES810	This study
ES4301	<i>lutE::Km</i> / pCM62	This study
ES4305	<i>lutE::Km</i> / pES666	This study
ES5031	<i>lutF::Km</i> / pCM62	This study
ES5032	<i>lutF::Km</i> / pES815	This study
ES4309	<i>lutG::Km</i> / pCM62	This study
ES4315	<i>lutG::Km</i> / pES668	This study
ES1819	AM1 / pES502	3
ES4192	$\Delta xoxF2$ $\Delta xoxF1$ / pES502	This study
ES4194	$\Delta xoxG$ / pES502	This study
ES4196	$\Delta xoxJ$ / pES502	This study
ES3743	AM1 / pHV25	This study
<b><i>E. coli</i></b>		
TOP10	Cloning strain (Sm <sup>R</sup> )	Invitrogen
S17-1	Helper strain (Tp <sup>R</sup> , Sm <sup>R</sup> )	4
<b>Plasmid</b>		
pRK2013	Conjugative helper plasmid (Km <sup>R</sup> )	5
pCM184	Allelic exchange suicide vector (Km <sup>R</sup> , Tc <sup>R</sup> , Ap <sup>R</sup> )	6
pHV2	Modified pCM184 expressing <i>sacB</i> (Km <sup>R</sup> , Tc <sup>R</sup> , Ap <sup>R</sup> )	This study
pCM157	Modified pCM62 expressing <i>cre</i> (Tc <sup>R</sup> )	6
pCM639	Modified pCM638 carrying IS <i>phoA</i> /hah-Tc (Tc <sup>R</sup> )	2
pAP5	Modified pCM62 carrying promoter-less <i>venus</i> (Tc <sup>R</sup> )	5
pCM62	Broad-host-range shuttle vector (Tc <sup>R</sup> )	7
pES502	pAP5 carrying <i>mxo</i> promoter region upstream of <i>venus</i> (Tc <sup>R</sup> )	5
pHV25	pAP5 carrying <i>lutH</i> promoter region upstream of <i>venus</i> (Tc <sup>R</sup> )	This study
pES667	pCM62 carrying <i>lutA</i> downstream of P <sub>lac</sub> promoter (Tc <sup>R</sup> )	This study
pES810	pCM62 carrying <i>lutB</i> downstream of P <sub>lac</sub> promoter (Tc <sup>R</sup> )	This study
pES666	pCM62 carrying <i>lutE</i> downstream of P <sub>lac</sub> promoter (Tc <sup>R</sup> )	This study

Table S2. (continued)

Strain or plasmid	Description	Reference
<b>Plasmid</b>		
pES815	pCM62 carrying <i>lutF</i> downstream of P <sub>lac</sub> promoter (Tc <sup>R</sup> )	This study
pES668	pCM62 carrying <i>lutG</i> downstream of P <sub>lac</sub> promoter (Tc <sup>R</sup> )	This study

## References

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Table S3. Primers used in this study.

Primer name	Sequence (5' to 3') <sup>‡</sup>	Plasmid	Gene
<b>Strain construction (pCM184 backbone)</b>			
xoxF_1740ULEcoRI	tagaattcGCCCCGATGGCAGGAATTAAGT	pES319-320	<i>xoxF1</i>
xoxF_1740URKpnI	atggtaccCTGCTTCGGCTCGTACTTCCACA		
xoxF_1740DLHpaI	tagttaacGCCTGTTCTACGTCCCCACCAAC		
xoxF_1740DRSacI	tagagctcCTGCACCGACGAGACCAAGAAGA		
exaF_1139ULEcoRI	gtgaattcCCTCCGACAGCCTACTGATGAAC	pES3-4	<i>exaF</i>
exaF_1139URKpnI	caggtacCGTCTTGGCGTCGTTGAGGATGTC		
exaF_1139DLHpaI	cagttaacGCCGTTCTGTGCCAACATCAACTG		
exaF_1139DRSacI	tagagctcGTTGATGCCAGCCGTCGTCCTTGG		
pqqBCDE_1748ULEcoRI	tagaattcTCAACATCCGCCAAGTCAGCTTC	pES369-458	<i>pqqBCDE</i>
pqqBCDE_1748URKpnI	atggtaccATATCGGGAGAGGCGTTCAGCAG		
pqqBCDE_1748DLHpaI	tatagttaacGACGAATGTGCAAAACCCGTTGA		
pqqBCDE_1748DRSacI	tatagagctcACGTCCGGTCGGATAGTCGTTGTC		
pqqF_2330ULEcoRI	tagaattcACCGTCCCGCCATCACCTATCGC	pES789-555	<i>pqqF</i>
pqqF_2330URKpnI	atggtaccGTGATTCAGGCCTCGCTGCGGTC		
pqqF_2330DLHpaI	tagttaacGGTGACCGGCTACCTGACCAAGG		
pqqF_2330DRSacI	tagagctcGTAGCGCAGGCTGGCGATCATCT		
lysR_0863ULEcoRI	atgaattcCAGGCGCTGTCGAAGCATTCAAG	pES431-432	<i>MexAM1_META1p0863</i>
lysR_0863URKpnI	atggtaccGGGTGTTCTCCAGCCTGAGCTTC		
lysR_0863DLHpaI	tagttaacAGCGAGATGCCGACGATGGAGAG		
lysR_0863DRSacI	tagagctcATGGACGTTGGAGAGGTGGATCT		
sacB_BegNcoI	tagcaccatggCGATCCTTTTTTAACCCATCACATA	pHV2	
sacB_EndNcoI	tataccatggGATATCGGCATTTTCTTTTGCG		

Table S3. (continued)

Primer name	Sequence (5' to 3') <sup>‡</sup>	Plasmid	Gene
<b>Strain construction (pHV2 backbone)</b>			
fae1_1767UBMunI	tata <b>caattg</b> CGCGGTTTCATCTCGCCTATCTCA	pES498-499	<i>faeI</i>
fae1_1767UENcoI	tata <b>ccatgg</b> CCGTGCTTGTGTTGACCAGACC		
fae1_1767DBAgeI	ctct <b>accggt</b> TTGTCACGGAGCAGCGTAAGTCG		
fae1_1767DESacI	tata <b>gagctc</b> GTGACGCTGTGGGTCGATGC		
cycK_1293UBAatII	tata <b>gagctc</b> CCAGCCTGAAGGAACGAGAAGAAGC	pES604-605	<i>cycK</i>
cycK_1293UENcoI	tata <b>ccatgg</b> CGGGCATCACAGCTTGCACCA		
cycK_1293DBBshTI	tata <b>accggt</b> CGGTCGGCGTCAGCCAGAT		
cycK_1293DESacI	tata <b>gagctc</b> GTGACGACGATGCGCCGGAC		
ccmB_2825UBAatII	tata <b>gagctc</b> CCATGTGGCATCACCGAATCAATGC	pES622-623	<i>ccmB</i>
ccmB_2825UENcoI	taat <b>ccatgg</b> TCAGGAAGAAGACGAGCGAGAGC		
ccmB_2825DBBshTI	tata <b>accggt</b> GCTGATGGTCCCGACTCTGATCTT		
ccmB_2825DESacI	tata <b>gagctc</b> AACCGATGTGAGGCCGATCTCC		
ccmC_2732UBAatII	tata <b>gagctc</b> AGAGACTGAGCGGGACCAAGC	pES620-621	<i>ccmC</i>
ccmC_2732UENcoI	tata <b>ccatgg</b> CCAGTAGGTCCCCCACATCGG		
ccmC_2732DBBshTI	tata <b>accggt</b> CTCGATGCTCTACCCGCTGCTC		
ccmC_2732DESacI	atat <b>gagctc</b> GGTCGCGTGCGGGATCTG		
ABC2_2359UBAatII	atat <b>gagctc</b> TGCGTATGCGATGGTTCAGACAA	pES618-619	<i>MexAM1_META1p2359</i>
ABC2_2359UENcoI	tata <b>ccatgg</b> AGCGGAAACACGATGAGTGGAATCG		
ABC2_2359DBBshTI	tata <b>accggt</b> CTGGACGGCATCTCCTTCACGG		
ABC2_2359DESacI	tata <b>gagctc</b> CGAGAGCATCCCGTTGATCCGAA		

Table S3. (continued)

Primer name	Sequence (5' to 3') <sup>‡</sup>	Plasmid	Gene
<b>Strain construction (pHV2 backbone)</b>			
25pep_3908UBBglII	ctag <b>agatct</b> GGTACACAGGTTGCGCCGGTTTTTC	pES467-468	<i>MexAM1_META1p3908</i>
25pep_3908UENcoI	tata <b>ccatgg</b> GCCTCGTCCTCGTCCTTCTTCTT		
25pep_3908DBAgeI	atata <b>accggt</b> CCTACAAGCCCAAGGCGATGATC		
25pep_3908DESacI	tata <b>gagctc</b> CGGTCAGGAAGACGACGAACAGG		
25mem_3909UBBglII	tata <b>agatct</b> GACCTACAAGCCCAAGGCGATGA	pES469-470	<i>MexAM1_META1p3909</i>
25mem_3909UENcoI	atata <b>ccatgg</b> CGGTCAGGAAGACGACGAACAGG		
25pep_3909DBAgeI	tata <b>accggt</b> CACGCCGATGGTGATCGCCTATG		
25pep_3909DESacI	tata <b>gagctc</b> CGCTGCCGTTCTCACTTCCCTTG		
hss_2024UBAatII	tata <b>gagctc</b> CGTCATGGGATTAAGTGCGAAACAGG	pES616-617	<i>hss</i>
hss_2024UENcoI	atata <b>ccatgg</b> CGGCTCTTGTCGTA CT CGAAATGGC		
hss_2024DBBshTI	tata <b>accggt</b> GACCTGCCACTACGCCTACCAC		
hss_2024DESacI	atata <b>gagctc</b> TGCGAAGGCCCGAACCGTAAC		
xoxG_1741UBBglII	tata <b>agatct</b> GCAACTTCATCGCCTGGGACAAC	pES459-460	<i>xoxG</i>
xoxG_1741UEKpnI	acgt <b>ggtagc</b> CGTCGTTGGTGATGTGGAAGGTC		
xoxG_1741DBAgeI	atata <b>accggt</b> GCGAGAAGGAAGACAAGCCCG		
xoxG_1741DESacI	tata <b>gagctc</b> AATCGAGCGGGTGG AATGAGAA		
xoxJ_1742UBBglII	atata <b>agatct</b> TCTTCTTGGTCTCGTCGGTGCAG	pES461-462	<i>xoxJ</i>
xoxJ_1742UEKpnI	tata <b>ggtagc</b> CGCAGCTTGACCTTCAGCTCGTC		
xoxJ_1742DBAgeI	tata <b>accggt</b> GCCACAACGAGAACGACTGGAAG		
xoxJ_1742DESacI	tata <b>gagctc</b> CTATCTCGGGCTCGGCAGCAATA		



Table S3. (continued)

Primer name	Sequence (5' to 3') <sup>‡</sup>	Plasmid	Gene
<b>Strain construction (pHV2 backbone)</b>			
xoxD_1771UBAatII	tatagacgctGCCATGCCGAGTTCGAGGTG	pES608-609	<i>MexAM1_META1p1771</i>
xoxD_1771UENcoI	attaccatggACCCTTGCCCTCGGTCTTCTC		
xoxD_1771DBBshTI	atataccggtGTGAAGGCCAAGGCCGAGAAG		
xoxD_1771DESacI	tatagagctcAGTCGGATTCGCGGTTGGCTAC		
orf6_1746UBMunI	atatacaattgGAGCAGCCGCAGATGAAGAACAC	pES493-494	<i>MexAM1_META1p1746</i>
orf6_1746UENcoI	atataccatggGTCGGTCCGATAGTCGTTGTCGT		
orf6_1746DBAgeI	atacaccggtCCTGCCCTATGACCGCTACGTT		
orf6_1746DESacI	atatagagctcCCGGCATTCCAACGAGATGTAGA		
orf7_1747UBMunI	atatacaattgTTACGGCTGGGCCTATGTGAACC	pES495-496	<i>MexAM1_META1p1747</i>
orf7_1747UENcoI	atataccatggGTGTTCTTCATCTGCGGCTGCTC		
orf7_1747DBAgeI	atataccggtCCAACCCGCGCTACATCAAGAAC		
orf7_1747DESacI	atatagagctcTGACGTTGAGGATGCCATTACC		
lutA_1778UBAatII	tatagacgctGGACGGGCTTTAACTCGGTTTCC	pES610-611	<i>lutA</i>
lutA_1778UENcoI	tataccatggGGCGAAGGTCTGCTTGGTGAAG		
lutA_1778DBBshTI	taataccggtGACTGGGGCGATTATGTCCCGT		
lutA_1778DESacI	tatagagctcCCGTCGCTCAAACAAGAAACAGATGA		
lutB_1779UBAatII	atatagagctcGCAAGTTCGGCCTCAAGATCACC	pES638-639	<i>lutB</i>
lutB_1779UENcoI	atataccatggGGCCTTCTCGTTGGTGACGTAGG		
lutB_1779DBAgeI	atataccggtCTCTACACGACCAACGGCACCTC		
lutB_1779DESacII	atatagagctcCCCCCTCACTTCACCGTAATCGT		

Table S3. (continued)

Primer name	Sequence (5' to 3') <sup>‡</sup>	Plasmid	Gene
<b>Strain construction (pHV2 backbone)</b>			
lutC_1780UBMunI	atat <b>caattg</b> GAGGTGATCGACAACGTGCTGGT	pES642-643	<i>lutC</i>
lutC_1780UENcoI	atat <b>ccatgg</b> GTTCTGCCGCTTCTTGGTGAGGT		
lutC_1780DBMluI	atata <b>acgct</b> CGGCACGATTACGGTGAAGTGAG		
lutC_1780DESacI	atat <b>gagctc</b> GCTGGCGTGGTAATGCAGGTTTT		
lutD_1781UBBgIII	atata <b>gatct</b> TGTCCGAGAAGGCGATGCAAG	pES644-645	<i>lutD</i>
lutD_1781UEMunI	atat <b>caattg</b> CAAGCACCACGAGGGCAAGC		
lutD_1781DBAgeI	atata <b>ccggt</b> GAGCAAGGCGAGGGCGAGTT		
lutD_1781DESacI	atg <b>gagctc</b> ATCGAGGTGCCGAGCGTGTT		
lutE_1782UBAatII	tata <b>gagctc</b> GTCCGAGAAGGCGATGCAAGAAC	pES612-613	<i>lutE</i>
lutE_1782UENcoI	tata <b>ccatgg</b> CGGCCCTCCTGATTGTTGTAGAGC		
lutE_1782DBBshTI	tata <b>accggt</b> CGACAAGAGAGCCGTGAAGTCCG		
lutE_1782DESacI	tata <b>gagctc</b> CAGAGCGGATATAGGGCAGAAGAGG		
lutF_1783UBAatII	tatc <b>gagctc</b> ACCCGGCTCTACAACAATCAGGAG	pES640-641	<i>lutF</i>
lutF_1783UENcoI	atat <b>ccatgg</b> TTCTGGTGGAAGAAACGCAGCAG		
lutF_1783DBAgeI	atata <b>ccggt</b> CGAATTGGTGCGTTTCGCTCTCT		
lutF_1783DESacI	atat <b>gagctc</b> TCGCAGACGTAGGAGACCGACTG		
lutG_1784UBAatII	tata <b>gagctc</b> GCTGGTCTGGCTGTTTCATCTTCG	pES614-615	<i>lutG</i>
lutG_1784UENcoI	atat <b>ccatgg</b> TAGCCCAGCGTCGTCACCTTG		
lutG_1784DBBshTI	tata <b>accggt</b> CGGACAAGGTGAGCGAGTTCAAGGA		
lutG_1784DESacI	tata <b>gagctc</b> GCAACATATTTCTGCCGTTCCGTCAA		

Table S3. (continued)

Primer name	Sequence (5' to 3') <sup>‡</sup>	Plasmid	Gene
<b>Strain construction (pHV2 backbone)</b>			
lutH_1785UBBgII	atata <b>gatct</b> TGCCGAATGATCCCGGTAAAGTC	pES465-466	<i>lutH</i>
lutH_1785UENcoI	tata <b>ccatgg</b> CGAACGAGGTGTTGAAACGAGCA		
lutH_1785DBAgeI	atata <b>ccgg</b> tCCCTGCTCGCTCACCAACTTCTT		
lutH_1785DESacI	atata <b>gagctc</b> TTCGGCCTCGCTTAGATTGCGTA		
<b>Transcriptional reporter fusion (pAP5 backbone)</b>			
lutH_1785BegAclI	tataa <b>acgtt</b> GCAGCCGTCGAAGCGATTTGC	pHV25	<i>lutH</i>
lutH_1785EndEcoRI	tata <b>gaattc</b> CCGAACGAGGTGTTGAAACGAGC		
<b>Complementation (pCM62 backbone)</b>			
lutA_1778BegKpnI	atata <b>ggtacc</b> CTTATGCTTCTCTCGTTGAGGCCG	pES667	<i>lutA</i>
lutA_1778EndSacI	atc <b>ggagctc</b> ATCGCCCGGATCGCTTCC		
lutB_1779BegKpnI	tattt <b>ggtacc</b> AAGCGATCCGGGCGATGC	pES810	<i>lutB</i>
lutB_1779EndSacI	tattt <b>gagctc</b> ACCTTTCGGGACGATGCCTTAGC		
lutE_1782BegKpnI	atata <b>ggtacc</b> GAGCAAGGCGAGGGCGAGTT	pES666	<i>lutE</i>
lutE_1782EndKpnI	atata <b>gagctc</b> GGCGGGATTCGAGGGAGACC		
lutF_1783BegKpnI	tattt <b>ggtacc</b> AAGGGTCGCATCGTCGCCCGTGG	pES815	<i>lutF</i>
lutF_1783EndSacI	tattt <b>gagctc</b> ACGAGCCGTAGCCCAGCGTCG		
lutG_1784BegKpnI	atata <b>ggtacc</b> GCCGCAATCGCCTACGATCC	pES668	<i>lutG</i>
lutG_1784EndSacI	atata <b>gagctc</b> CTTCGTCGCTTCCGACCGAAAG		

<sup>‡</sup>Restriction endonuclease sites are in bolded font. Capitalized letters represent bases that anneal to the target DNA sequence.