

SUPPLEMENTRY MATERIALS

Heterotypic immunity against vaccinia virus in an HLA-B*07:02 transgenic mousepox infection model

Running title: Ectromelia virus-derived CD8⁺ T cell epitopes

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TABLE S1: Properties of peptides chosen to generate binary encoded pB7.2 tetramers

	ORF/self ^a	VACV ^b	ECTV ^c	Source ^d	pB7.2 monomer exchange, % ^e
1	A24R ₁₀₀₂₋₁₀₁₀	KPYASKVFF	•	MS	70.83053
2	A34R ₈₂₋₉₀	LPRPDTRHL	•	MS	81.74804
3	A3L ₁₉₂₋₂₀₀	SPSNHHILL	•	MS	74.94446
4	A47L ₂₂₇₋₂₃₆	KPVSDLYTSM	•	MS	56.64893
5	A4L ₁₂₆₋₁₃₅	APASSLLPAL	NSH	MS	76.23163
6	B17L ₁₈₁₋₁₉₀	APYPGNVLVY	•	MS	82.74924
7	B18R ₃₀₅₋₃₁₃	RPADSITYL	HPS DSITYL	MS	94.84093
8	B18R ₃₀₅₋₃₁₃ ^f	RPLDSITYL	HPS DSITYL	MS	83.9946
9	B8R ₇₀₋₇₈	FPKNDFVSF	FPN NDFVSF	MS	94.85919
10	C10L ₄₁₋₄₉	LPMEDNSDI	•	MS	46.47672
11	D11L ₅₀₆₋₅₁₄	MPTVDEDLF	•	MS	58.24184
12	D5R ₃₇₅₋₃₈₃	LPKEYSSEL	•	MS	74.42226
13	D9R ₂₆₋₃₅	IPRSKDTTHVF	•	MS	84.6781
14	E1L ₁₀₋₁₈	LPNITLKII	•	MS	25.29541
15	E2L ₂₁₆₋₂₂₄	RPRDAIRFL	•	MS	71.43947
16	E9L ₁₇₅₋₁₈₃	FPSVFINPI	•	MS	57.55645
17	E9L ₁₇₅₋₁₈₃ ^f	FPSVFINPV	FPSVFINP I	MS	73.84057
18	E9L ₅₂₆₋₅₃₄	FPYEGGKVF	•	MS	111.9869

19	G5R ₃₄₁₋₃₄₉	LPCQLMYAL	•	MS	70.8617
20	G7L ₁₇₅₋₁₈₃	LPMIIGEPI	•	MS	70.068
21	H1L ₆₅₋₇₃	LPNSNINII	•	MS	59.41963
22	H5R ₈₉₋₉₇	SPSPGVGDI	NSH	MS	54.39731
23	I1L ₅₃₋₆₂	IPVDLVKSSF	•	MS	46.01204
24	I4L ₄₉₈₋₅₀₇	RPIGIGVQGL	•	MS	70.52276
25	I6L ₂₃₇₋₂₄₅	FPTNTLTSI	•	MS	57.23258
26	I6L ₂₇₂₋₂₈₀	IPKKIVSLL	•	MS	56.66139
27	I7L ₃₄₂₋₃₅₀	TPPKSFKSL	•	MS	57.33455
28	J3R ₈₋₁₆	KPFMYFEEI	•	MS	42.75962
29	K1L ₁₅₁₋₁₅₉	IPSTFDLAI	•	MS	48.3324
30	K6L ₁₇₋₂₅	KPITYPKAL	•	MS	78.04875
31	L4R ₃₇₋₄₅	FPRSMLSIF	•	MS	40.13252
32	N2L ₁₀₄₋₁₁₃	RPNQHHTIDL	RPN K HHTIDL	MS	86.15705
33	I4L ₆₇₀₋₆₇₈	LPEDIKRVY	•	MS	46.96676
34	D13L ₁₆₀₋₁₆₈	TPFDVEDTF	•	MS	52.06957
35	TA25R ₉₅₄₋₉₆₂	FPSSNETSI	•	MS	49.48632
36	D1R ₈₀₈₋₈₁₇	RPSTRNFFEL	•	MS	70.72724
37	B15R ₉₁₋₁₀₁	IPDEQKTIIGL	•	Predicted	76.69625
38	O1L ₅₄₉₋₅₅₇	IPITDSLSF	•	Public	64.66695
39	O1L ₅₄₉₋₅₅₇ ^f	IPITESLSF	IPIT D SLSF	Public	58.66162

40	B22R ₇₂₋₈₀	TVADVRRHCL	SID ^D VRRQ ^C CL	Public	79.57025
41	C1L ₁₀₂₋₁₁₁	KPKPAVRFAI	EY ^N RAVRFAE	Public	75.30461
42	D1R ₆₈₆₋₆₉₄	HPRHYATVM	•	Public	71.18591
43	F4L ₆₋₁₄	APNPNRFVI	•	Public	98.25802
44	J6R ₃₀₃₋₃₁₁	MPAYIRNTL	•	Public	86.58602
45	O1L ₃₃₅₋₃₄₄	RPMSLRSTII	RP ^I SLRSTII	Public	74.84205
46	J2R ₁₁₆₋₁₂₄	KPFNNILNL	•	Public	56.809954
47	TTF2 ₃₃₃₋₃₄₁	APAAPGLSL	na	Self	76.23394
48	HNRPM ₄₉₉₋₅₀₇	APIDRVGQTL	na	Self	72.42752
49	SAP145 ₁₈₋₂₇	APIQGNREEL	na	Self	69.33693
50	HK2 ₉₀₄₋₉₁₂	APKCDVSFL	na	Self	59.89381
51	CUL-4A ₅₋₁₄	APRKGSFSAL	na	Self	95.3412
52	ABCC12 ₁₃₄₇₋₁₃₅₃	APSAFGMLL	na	Self	67.52423
53	NLRP2 ₂₁₃₋₂₂₀	GPSGLGKTAI	na	Self	60.51939
54	TRIM27 ₄₆₈₋₄₇₆	GPVRPYFSL	na	Self	78.9362
55	FASN ₁₁₆₉₋₁₁₇₈	I PRDPSQQEL	na	Self	77.31273
56	CDCA7 ₉₄₋₁₀₃	KPRPDVTNEL	na	Self	77.95413
57	hCG1980884 ₁₈₀₇₋₁₈₁₅	KPYFPPRIL	na	Self	62.56214
58	FASN ₉₂₅₋₉₃₃	LPKTGTVSL	na	Self	74.5599
59	RACGAP1 ₄₆₅₋₄₇₃	LPQANRDTL	na	Self	100.5323
60	COPS8 ₁₆₃₋₁₇₂	LPRKPVAGAL	na	Self	99.14703

61	PACSIN2 ₃₃₇₋₃₄₅	LPSKPSSTL	na	Self	66.27471
62	FUS/BBF2H7 ₉₁₋₁₀₀	LPVSVCTPGPL	na	Self	56.58131
63	TYMS ₂₇₄₋₂₈₂	RPFPKLRIL	na	Self	64.86887
64	VEGFR3 ₁₀₅₀₋₁₀₅₈	RPGSSDRVL	na	Self	86.67953
65	UROD ₁₆₃₋₁₇₁	RPQASHQLL	na	Self	65.97249
66	HECTD1439-447	RPQVAKTLL	na	Self	105.5192
67	SIK1 ₄₃₀₋₄₄₇	RPRPVSPSSL	na	Self	78.38857
68	TBX1 ₄₄₂₋₄₅₀	RPSPPNPEL	na	Self	74.64187
69	LOC399706 ₁₃₉₋₁₄₅	SPAGSTRVL	na	Self	82.34095
70	SLC37A3 ₃₇₇₋₃₈₆	SPNDKSINAL	na	Self	79.91597
71	CA9 ₃₉₇₋₄₀₆	SPRAAEPVQL	na	Self	83.77609
72	KCDT10 ₂₈₋₃₆	SPSSKYVKL	na	Self	89.20886
73	TNKS ₁₄₂₋₁₅₀	SPSSPGSSL	na	Self	78.06154
74	MAP1S ₉₄₅₋₉₅₄	SPVYLDLAYL	na	Self	42.82756
75	HRT3 ₁₆₂₋₁₇₀	TPSNTPTGPL	na	Self	52.25194

^aOpen reading frames (ORFs)/source molecules and location of epitopes defined based on Copenhagen reference strain (VACCC, txid 10249), homo sapiens (NCBITaxon:9606)

^bamino acid sequence based on Copenhagen reference strain (VACCC, ID 10249)

^cAmino acid changes in the orthologous ECTV peptide; ●, conserved sequences with 100% homology; NSH, no significant homology; red bold residues, variation in ECTV within the VACV-derived epitope; based on Netblast (blastcl3: www.ncbi.nlm.nih.gov) using ECTV txid12643; na, not applicable

^dMS, mass spectrometry discovered VACV peptides discovered by us¹, predicted, reported as predicted VACV epitope; public, reported as positive in human or HLA class I transgenic mouse CD8⁺ T cell assay; self, naturally processed self peptides²

^eAverage rescue of pB7.2 monomer assessed in an UV-mediated peptide exchange assay¹; see Materials and Methods and Figure 5C; a threshold of $\geq 40\%$ exchange was used to select peptides for tetramer binding study

^fVARV orthologue, txid 10255 variola virus

Table S2: Homology of VACV epitopes uniquely recognised by ECTV-reactive CD8⁺ T cells

Orf: I6L^{a,b}; epitope: I6L_{272–281}

ECTV	1	MNNFVKQVASKSLKPTKKLSPSDE V ISLNECIISFNLDNFYYCNDGLFTKPINTPEDVLK	60
		MNNFVKQVASKSLKPTKKLSPSDE ISLNECIISFNLDNFYYCNDGLFTKPINTPEDVLK	
VACV	1	MNNFVKQVASKSLKPTKKLSPSDE A ISLNECIISFNLDNFYYCNDGLFTKPINTPEDVLK	60
ECTV	61	SLL I MESFAYEKMIKGLIKILISRAYINDIYFTPPFGWLTGVDD P ETHVVIKIIFNSSL	120
		SLL + MESFAYEKMIKGLIKILISRAYINDIYFTPPFGWLTGVDPETHVVIKIIFNSSL	
VACV	61	SLL V MESFAYEKMIKGLIKILISRAYINDIYFTPPFGWLTGV D YDPETHVVIKIIFNSSL	120
ECTV	121	ISIKSQVIEYLKPYNVNNSVLTTTEKELSINTFNVPSIPMSIISFFPFDTDFILVILFF	180
		ISIKSQVIEYLKPYNVNNSVLTTTEKELSINTFNVPSIPMSIISFFPFDTDFILVILFF	
VACV	121	ISIKSQVIEYLKPYNVNNSVLTTTEKELSINTFNVPSIPMSIISFFPFDTDFILVILFF	180
ECTV	181	GVYNDSYCGISYISPKERLPYIIIEILKPLVSEINMLSDEIGRTSSIRIFNSTSVKKFPTN	240
		GVYNDSYCGISYISPKERLPYIIIEILKPLVSEINMLSDEIGRTSSIRIFNSTSVKKFPTN	
VACV	181	GVYNDSYCGISYISPKERLPYIIIEILKPLVSEINMLSDEIGRTSSIRIFNSTSVKKFPTN	240
ECTV	241	TLTSICEIVYSFDES S FPTPKTFTPLNASPY IPKKIVSLL DLPSNVEIKAISRGGVDFIT	300
		TLTSICEIVYSFDES FPTPKTFTPLNASPY IPKKIVSLL DLPSNVEIKAISRGGVDFIT	
VACV	241	TLTSICEIVYSFDES F FPTPKTFTPLNASPY IPKKIVSLL DLPSNVEIKAISRGGVDFIT	300
ECTV	301	HINNKRL N TILVIAKDNFLKNSTFSGTFIKENIWKGIYTYRIKSSFPVPTIKSVTNKK	360
		HINNKRL TILVIAKDNFLKNSTFSGTFIKENIWKGIYTYRIKSSFPVPTIKSVTNKK	
VACV	301	HINNKRL T TILVIAKDNFLKNSTFSGTFIKENIWKGIYTYRIKSSFPVPTIKSVTNKK	360
ECTV	361	KICKKHCFVNSQYTTRTLSHIL	382
		KICKKHCFVNSQYTTRTLSHIL	
VACV	361	KICKKHCFVNSQYTTRTLSHIL	382

Orf: B15R^{a,b}; epitope: B15R₉₁₋₁₀₁ (C8R in ECTV)

ECTV	1	MTANFSTHVFSPQHCGCDRLTSID DV RQCLTEYIYWSSYAYRNRQCAGQLYSTLLSFRDD	75
		MTANFSTHVFSPQHCGCDRLTSID + QCLTEYIYWSSYAYRNRQCAGQLYSTLLSFRDD	
VACV	1	MTANFSTHVFSPQHCGCDRLTSID V KQCLTEYIYWSSYAYRNRQCAGQLYSTLLSFRDD	60
ECTV	76	AE S VFIDIRELVKNMPWDDVKDCA E IIRCY IPDEQKTIREISSI IIGLCAYAATYWGGEDH	135
		AE VFIDIRELVKNMPWDDVKDCEIIRCY IPDEQKTIREIS+ IIGLCAYAATYWGGEDH	
VACV	61	AEL V FIDIRELVKNMPWDDVKDCA T EIIRCY IPDEQKTIREISAI IIGLCAYAATYWGGEDH	120
ECTV	136	PTSNSLNALFVMLE L LNVDYNIIFRRMN	164
		PTSNSLNALFVMLE + LNVDYNIIFRRMN	
VACV	121	PTSNSLNALFVMLE M LNVDYNIIFRRMN	149

Note IREISS insertion at position 91 in ECTV^b but lacking in this strain of VACV^c

ECTV	1	MTANFSTHVFSPQHCGCDRLTSID DV RQCLTEYIYWSSYAYRNRQCAGQLYSTLLSFRDD	75
		MTANFSTHVFSPQHCGCDRLTSID + QCLTEYIYWSSYAYRNRQCAGQLYSTLLSFRDD	
VACV	1	MTANFSTHVFSPQHCGCDRLTSID V KQCLTEYIYWSSYAYRNRQCAGQLYSTLLSFRDD	60

ECTV	76	AE S VFIDIRELVKNMPWDDVKDCAEIIRCY IPDEQKTIREISSIIGL CAYAATYWGGEDH	135
		AE VFIDIRELVKNMPWDDVKDC EIIRCY IPDEQKT IIGL CAYAATYWGGEDH	
VACV	61	AEL V VFIDIRELVKNMPWDDVKDCVEIIRCY IPDEQKT-----IIGL CAYAATYWGGEDH	114
ECTV	136	PTSNSLNALFVMLE LL NYVDYNIIFRRMN	164
		PTSNSLNALFVMLE + LNyVDYNIIFRRMN	
VACV	115	PTSNSLNALFVMLE M LNyVDYNIIFRRMN	143

Orf: D9R^{a,b}; epitope: D9R₂₆₋₃₅

ECTV	1	MGITMDEEVIFETPRELISIKRIKD IPRSKDT HVF AACITSDGYPLIGARRTSFAFQAIL	60
		MGITMDEEVIFETPRELISIKRIKD IPRSKDT HVF AACITSDGYPLIGARRTSFAFQAIL	
VACV	1	MGITMDEEVIFETPRELISIKRIKD IPRSKDT HVF AACITSDGYPLIGARRTSFAFQAIL	60
ECTV	61	SQQNSDSIFRVSTKLLRFMYYNELREIFRRLRKGSIINNIDPHFEELILLGGKLDKKESEIK	120
		SQQNSDSIFRVSTKLLRFMYYNELREIFRRLRKGSIINNIDPHFEELILLGGKLDKKESEIK	
VACV	61	SQQNSDSIFRVSTKLLRFMYYNELREIFRRLRKGSIINNIDPHFEELILLGGKLDKKESEIK	120
		89 125	
ECTV	121	DCL K RELKEESDERITVKEFGNVILKLTTRDKLFNKVYI G YCMACFINQSLEDLSHTSIY	180
		DCL + RELKEESDERITVKEFGNVILKLTTRDKLFNKVYI YCMACFINQSLEDLSHTSIY	
VACV	121	DCL R RELKEESDERITVKEFGNVILKLTTRDKLFNKVYI S YCMACFINQSLEDLSHTSIY	180
ECTV	181	NVEIRKIKSLNDCINDDKYEYLSYIYNMLVNSK	213
		NVEIRKIKSLNDCINDDKYEYLSYIYNMLVNSK	
VACV	181	NVEIRKIKSLNDCINDDKYEYLSYIYNMLVNSK	213

Orf: A24R^b; epitope: A24R₁₀₀₂₋₁₀₁₀

ECTV	1	MKKNTDSEMDQRLGYKFLVDPKAGVFYRPLHFQYVSYSNFILHRLHEILTVKRPLLSFK	60
		MKKNTDSEMDQRLGYKFLVDPKAGVFYRPLHFQYVSYSNFILHRLHEILTVKRPLLSFK	
VACV	1	MKKNTDSEMDQRLGYKFLVDPKAGVFYRPLHFQYVSYSNFILHRLHEILTVKRPLLSFK	60
ECTV	61	NNTERIMIEISNVKVTPPDYSPIIASIKGKSYDALATFTVNIFKEVMTKEGISITKISSY	120
		NNTERIMIEISNVKVTPPDYSPIIASIKGKSYDALATFTVNIFKEVMTKEGISITKISSY	
VACV	61	NNTERIMIEISNVKVTPPDYSPIIASIKGKSYDALATFTVNIFKEVMTKEGISITKISSY	120
ECTV	121	EGKDSHLIKIPLLIIGYGNKNPLDTAKYLVPNVIGGVFINKQSVEKVGINLVEKITTWPKF	180
		EGKDSHLIKIPLLIIGYGNKNPLDTAKYLVPNVIGGVFINKQSVEKVGINLVEKITTWPKF	
VACV	121	EGKDSHLIKIPLLIIGYGNKNPLDTAKYLVPNVIGGVFINKQSVEKVGINLVEKITTWPKF	180
ECTV	181	RVVKPNSFTFSFSSVSPPNVLPTRYRHYKISLDISQLEA S NISSTKTFITVNIVLLSQYL	240
		RVVKPNSFTFSFSSVSPPNVLPTRYRHYKISLDISQLEA NISSTKTFITVNIVLLSQYL	
VACV	181	RVVKPNSFTFSFSSVSPPNVLPTRYRHYKISLDISQLEA L NISSTKTFITVNIVLLSQYL	240
ECTV	241	SRVSLEFIRRSLSYDMPPEVVYLVNAIIDS AKRITESITDFNIDTYINDLVEAEHIKQKS	300
		SRVSLEFIRRSLSYDMPPEVVYLVNAIIDS AKRITESITDFNIDTYINDLVEAEHIKQKS	
VACV	241	SRVSLEFIRRSLSYDMPPEVVYLVNAIIDS AKRITESITDFNIDTYINDLVEAEHIKQKS	300
ECTV	301	QLTINEFKYEMLHNFLPHMNYTPDQLKGFYMI S LLRKFLYCIYHTSRYPDRDSMVCHRIL	360
		QLTINEFKYEMLHNFLPHMNYTPDQLKGFYMI S LLRKFLYCIYHTSRYPDRDSMVCHRIL	
VACV	301	QLTINEFKYEMLHNFLPHMNYTPDQLKGFYMI S LLRKFLYCIYHTSRYPDRDSMVCHRIL	360

ECTV	361	TYGKYFETLAHDELENYIGNIRNDIMNNHKNRGTYAVNIHVLTTPGLNHAFFSSLLSGKFK	420
VACV	361	TYGKYFETLAHDELENYIGNIRNDIMNNHKNRGTYAVNIHVLTTPGLNHAFFSSLLSGKFK	420
ECTV	421	KSDGSYRTHPHYSWQONISIPRSVGFYDPQVKISKMFSVRKYHPSQYLYFCSSDVPERGP	480
VACV	421	KSDGSYRTHPHYSWQONISIPRSVGFYDPQVKISKMFSVRKYHPSQYLYFCSSDVPERGP	480
ECTV	481	QVGLVSQLSVLSSITNILTSEYLDLEKKICEYIRSYYKDDISYFETGFPITIENTALVASL	540
VACV	481	QVGLVSQLSVLSSITNILTSEYLDLEKKICEYIRSYYKDDISYFETGFPITIENTALVASL	540
ECTV	541	NPNMICDFVTDFFFFRRRKRMGFFGNLEVGITLVRDHMNEIRINIGAGRLVRPFLVVDNGELM	600
VACV	541	NPNMICDFVTDFFFFRRRKRMGFFGNLEVGITLVRDHMNEIRINIGAGRLVRPFLVVDNGELM	600
ECTV	601	MDVCQELESRLDDMTFSDIQKEFPHVIEVDIEQFTFSNVCESVQKFRMMSKDERKQYDL	660
VACV	601	MDVCPELESRLDDMTFSDIQKEFPHVIEVDIEQFTFSNVCESVQKFRMMSKDERKQYDL	660
ECTV	661	CDFPAEFRDGYVASSLVGINHNSGPRAILGCAQAKQAI SCLSSDIRNKIDNGIHLMPER	720
VACV	661	CDFPAEFRDGYVASSLVGINHNSGPRAILGCAQAKQAI SCLSSDIRNKIDNGIHLMPER	720
ECTV	721	PIVISKALETSKIAANCFGQHVTIALMSYKGINQEDGII IKQFIQRGGLDIVTAKKHQV	780
VACV	721	PIVISKALETSKIAANCFGQHVTIALMSYKGINQEDGII IKQFIQRGGLDIVTAKKHQV	780
ECTV	781	EIPLNFNNKERDRSNAYSKLESNGLVRLNAFLESGDAMARNISSRTLEDDFARDNQISF	840
VACV	781	EIPLNFNNKERDRSNAYSKLESNGLVRLNAFLESGDAMARNISSRTLEDDFARDNQISF	840
ECTV	841	DVSEKYTDMYKSRVERVQVELTDKVKVRVLTMKERRPILGDKFTTRTSQKGTVAYI ADET	900
VACV	841	DVSEKYTDMYKSRVERVQVELTDKVKVRVLTMKERRPILGDKFTTRTSQKGTVAYV ADET	900
ECTV	901	ELPYDENGITPDVI INSTSIFSRKTISMLIEVILTAAYSAPYNNNGENRPVCFPSSNET	960
VACV	901	ELPYDENGITPDVI INSTSIFSRKTISMLIEVILTAAYSAPYNN KGENRPVCFPSSNET	960
ECTV	961	SIDTYMQFAKQCYEHSNPQLT DDELSDKVFCEKILYDPETDKPYASKVFF GPIYYLRLRH	1020
VACV	961	SIDTYMQFAKQCYEHSNP+L+D+ELSDK+FCEKILYDPETDKPYASKVFF GPIYYLRLRH	1020
ECTV	1021	LTQDKATVRCRGKTKLIRQANEGRKRGGGIKFGEMERDCLIAHGAANTITEVLKDSEED	1080
VACV	1021	LTQDKATVRCRGKTKLIRQANEGRKRGGGIKFGEMERDCLIAHGAANTITEVLKDSEED	1080
ECTV	1081	YQDVYI CENCGDIAAQIKGINTCLRCSKLNLSPLLT KIDTTHVSKVFLTQMNARGVKVKL	1140
VACV	1081	YQDVY+ CENCGDIAAQIKGINTCLRCSKLNLSPLLT KIDTTHVSKVFLTQMNARGVKVKL	1140

ECTV	1141	DFERRPPSFYK Q LDKVDLKPSFL	1163
		DFERRPPSFYK LDKVDLKPSFL	
VACV	1141	DFERRPPSFYK P LDKVDLKPSFL	1163

^aVACV txid: 10249

^bECTV txid: 12643

^cVACV txid: 10245

Blue bold, variation outside the epitope; red bold, amino acid sequence of the epitope

TABLE S3. List of Antibodies

Antigen	Fluorochrome	Clone	Source	Catalogue number
B220	FITC	RA3-6B2	BD Biosciences	553088
B220	PerCP-Cy5.5	RA3-6B2	Biolegend	103236
B220	APC-Cy7	RA3-6B2	BD Biosciences	552094
CD69	APC	H1.2F3	Biolegend	104514
CD8a	FITC	53-6.7	BD Biosciences	553031
CD8a	PerCP-Cy5.	53-6.7	BD Biosciences	551162
CD45.2	FITC	104	Biolegend	109806
CD45.2	APC	104	Tonbo Biosciences	20-0454-U100
CXCR3	APC	CXCR3-173	BD Biosciences	562266
CD62L	PE	MEL-14	BD Biosciences	553151
KLRG1	APC	2F1/KLRG1	Biolegend	138412
CD49a	PE	Ha31/8	BD Biosciences	562115
CCR8	APC	SA214G2	Biolegend	150309
CXCR6	PE	SA051D1	Biolegend	151103
CD103	PE	2E7	Biolegend	121405
Ghost Violet 510	V510	na	Tonbo Biosciences	13-0870-T100
pB7.2 tetramer	PE	na	in house	na
pB7.2 tetramer	APC	na	in house	na
pB7.2 tetramer	BV421	na	in house	na
pB7.2 tetramer	PE-Cy7	na	in house	na
pB7.2 tetramer	APC-Cy7	na	in house	na

na, not applicable

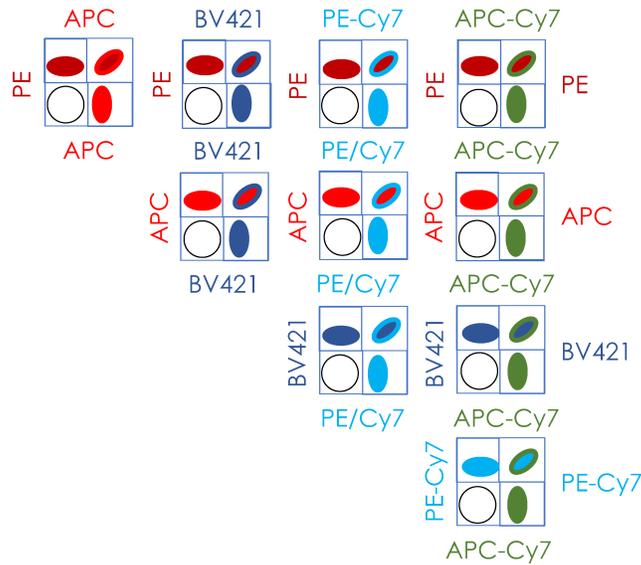
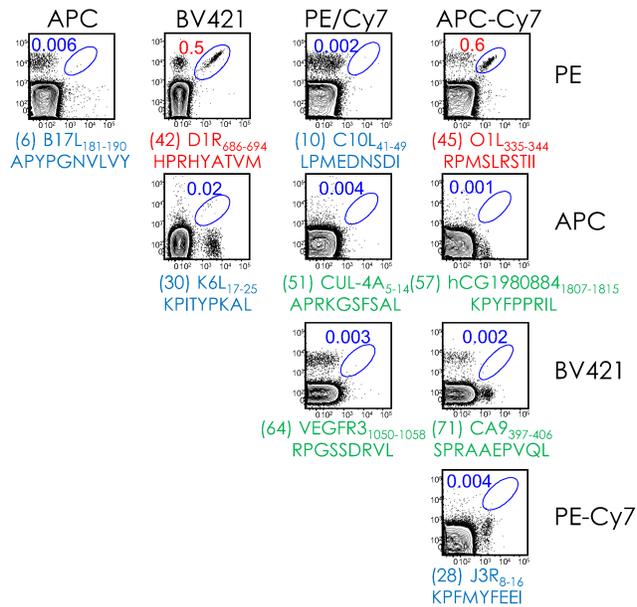


Figure S1: A schematic rendition of binary encoding of T cell epitope specificity. pMHC monomers are made to tetramers by reacting the biotinylated monomer of a single antigen specificity with two distinct fluorophore-conjugated streptavidin in two separate tubes. When a pMHC monomer of a single specificity is individually tetramerised with five different fluorophore-conjugated streptavidin molecules, and two tetramers made of two different fluorophore-conjugated streptavidin molecules mixed –and, thereby, binary encoded– one will obtain ten unique combinations as shown in the schematic. For a real example, see Figure 1A. In a variation of this binary encoding of a pMHC monomer with a single specificity, monomers of ten different specificities can be binary encoded with two unique combinations of fluorophores just with 5 different fluorophore-conjugated streptavidin molecules. For a real example, see Figure 1B. refer to refs.^{3,4} for detailed description and protocols.

Panel 1



Panel 2

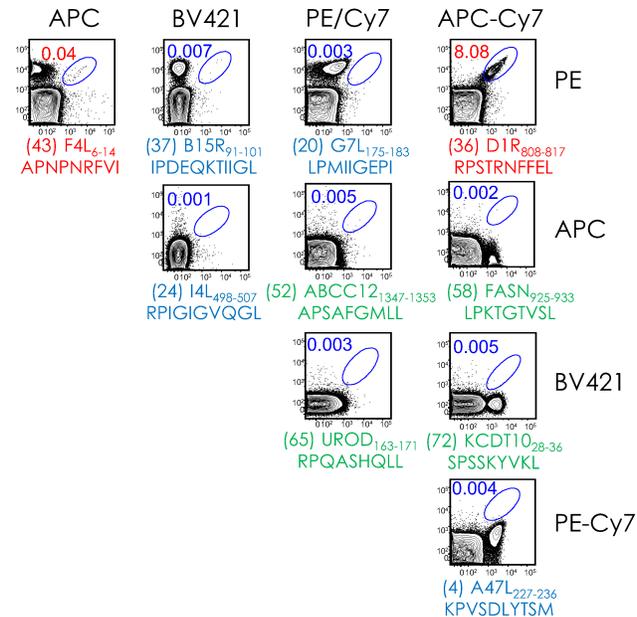
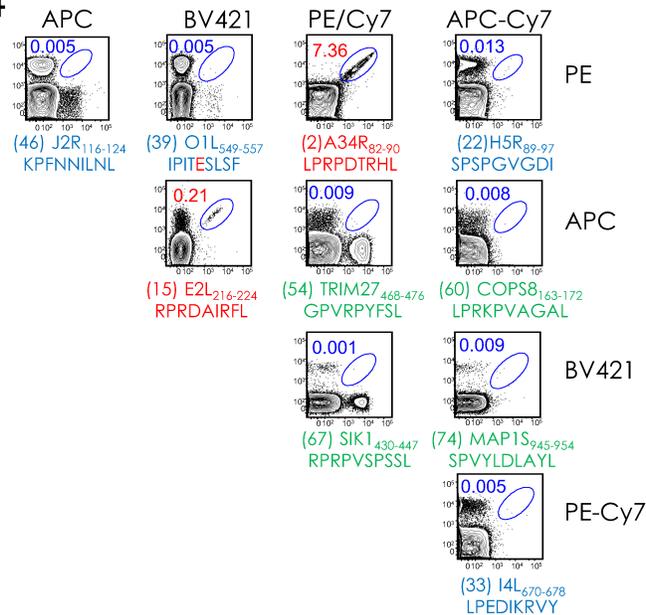


Figure S2, Panels 1&2: Tracking VACV-reactive CD8⁺ T cell responses in B7.2^{tg} mice. As in Figure 2, B7^{tg} mice were inoculated i.n. with sublethal dose of ECTV, and after 4 weeks challenged i.n. with a lethal dose of the same virus. See Materials & Methods. Spleens were harvested from mock (n=2) and ECTV (n=8) inoculated mice after 8–10 days post inoculation. CD8⁺ T cells were identified as CD8⁺ splenocytes after gating out dead cells and B220⁺ cells. Peptide-specific CD8⁺ T cells were identified using dual fluorochrome-labelled pB7.2 tetramers and binary encoding strategy as schematised in Figure S2. Red, positive VACV pB7.2 tetramer staining; blue, no staining with VACV pB7.2 tetramer; green, self p/B7.2 tetramers. *Note that Panel 3 is shown in Figure 1B.*

Panel 4



Panel 5

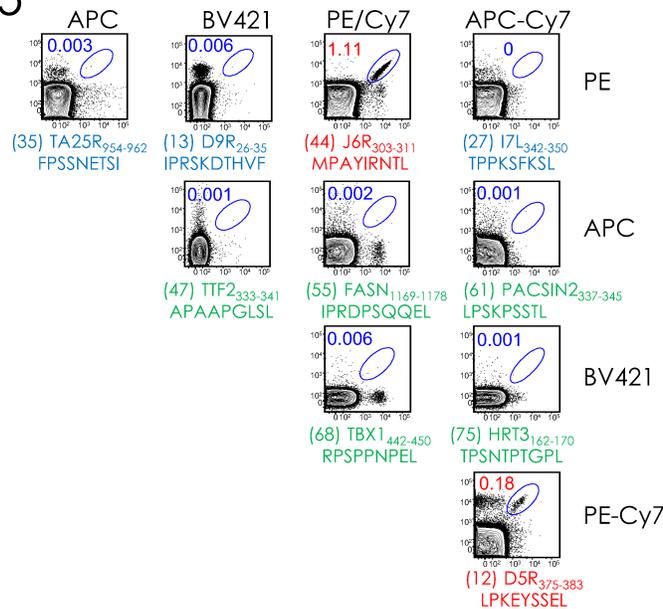
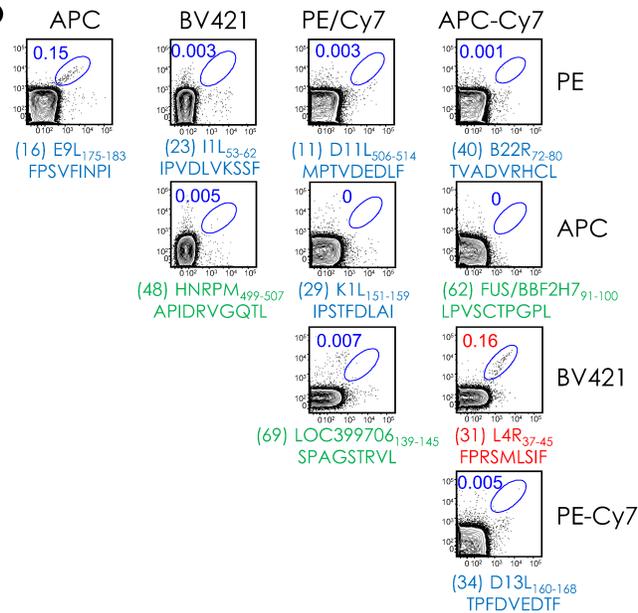


Figure S2 continued, Panels 4&5: Tracking ECTV-reactive CD8⁺ T cell responses in B7.2^{tg} mice. As in Figure 2, B7^{tg} mice were inoculated i.n. with sublethal dose of ECTV, and after 4 weeks challenged i.n. with a lethal dose of the same virus. See Materials & Methods. Splensens were harvested from mock (n=2) and ECTV (n=8) inoculated mice after 8–10 days post inoculation. CD8⁺ T cells were identified as CD8⁺ splenocytes after gating out dead cells and B220⁺ cells. Peptide-specific CD8⁺ T cells were identified using dual fluorochrome-labelled pB7.2 tetramers and binary encoding strategy as schematised in Figure S2. Red, positive VACV pB7.2 tetramer staining; blue, no staining with VACV pB7.2 tetramer; self p/B7.2 tetramers. Note that Panel 3 is shown in Figure 1B.

Panel 6



Panel 7

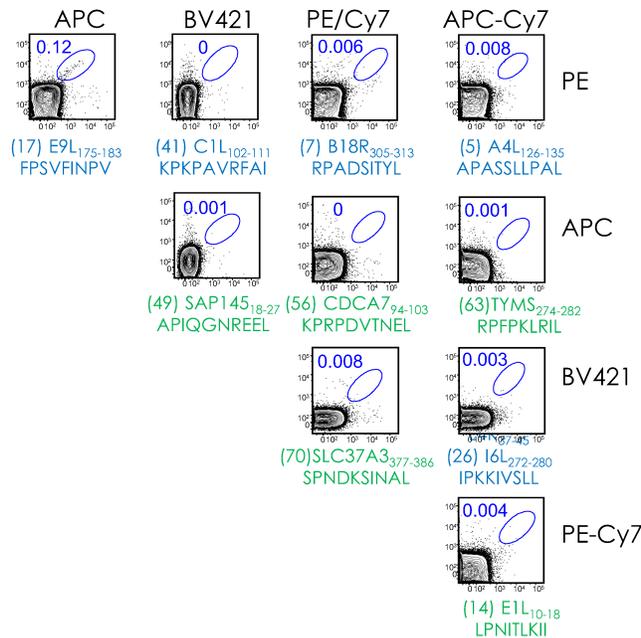


Figure S2 continued, Panels 6&7: Tracking ECTV-reactive CD8⁺ T cell responses in B7.2^{tg} mice. As in Figure 2, B7^{tg} mice were inoculated i.n. with sublethal dose of ECTV, and after 4 weeks challenged i.n. with a lethal dose of the same virus. See Materials & Methods. Spleens were harvested from mock (n=2) and ECTV (n=8) inoculated mice after 8–10 days post inoculation. CD8⁺ T cells were identified as CD8⁺ splenocytes after gating out dead cells and B220⁺ cells. Peptide-specific CD8⁺ T cells were identified using dual fluorochrome-labelled pB7.2 tetramers and binary encoding strategy as schematised in Figure S2. Red, positive VACV pB7.2 tetramer staining; blue, no staining with VACV pB7.2 tetramer; green, self p/B7.2 tetramers.

Panel 8

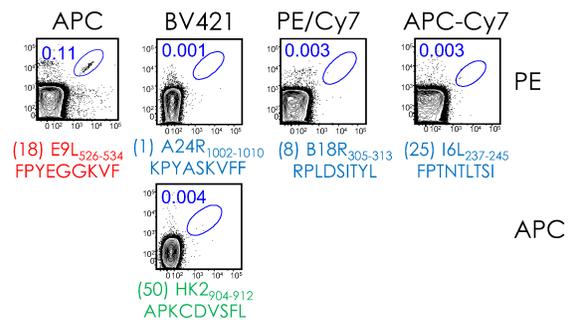


Figure S2 continued, Panel 8: Tracking ECTV-reactive CD8⁺ T cell responses in B7.2^{tg} mice. As in Figure 2, B7^{tg} mice were inoculated i.n. with sublethal dose of ECTV, and after 4 weeks challenged i.n. with a lethal dose of the same virus. See Materials & Methods. Spleens were harvested from mock (n=2) and ECTV (n=8) inoculated mice after 8–10 days post inoculation. CD8⁺ T cells were identified as CD8⁺ splenocytes after gating out dead cells and B220⁺ cells. Peptide-specific CD8⁺ T cells were identified using dual fluorochrome-labelled pB7.2 tetramers and binary encoding strategy as schematised in Figure S2. Red, positive VACV pB7.2 tetramer staining; blue, no staining with VACV pB7.2 tetramer; green, self p/B7.2 tetramers.

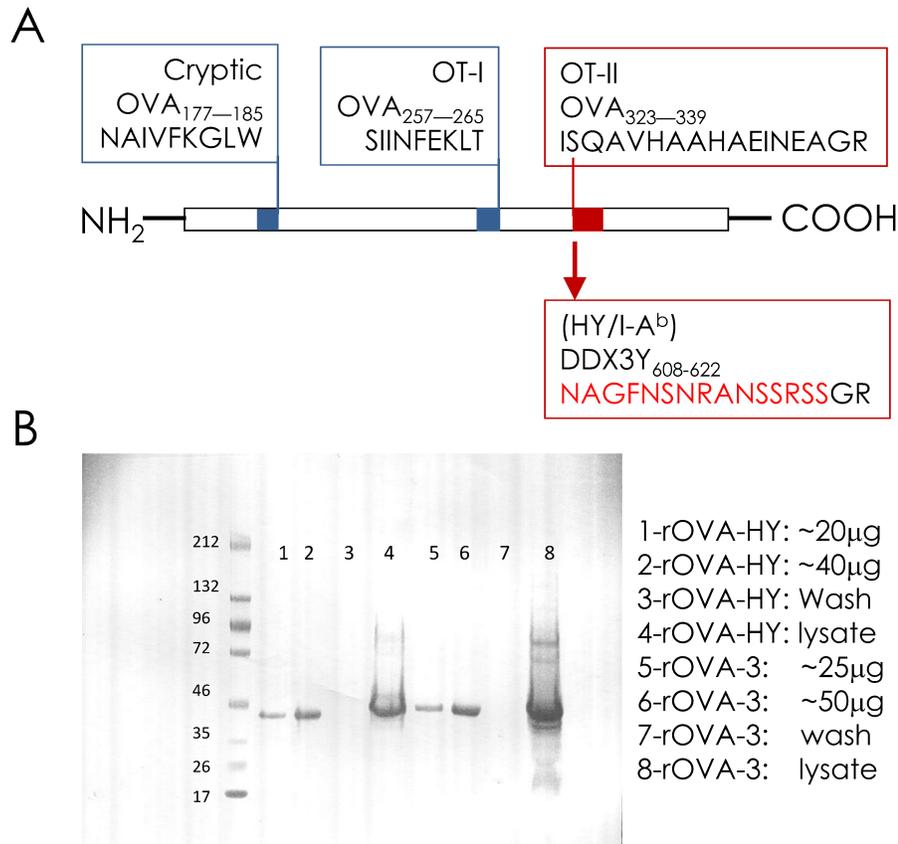


Figure S3: Design of rOVA-HY protein, and the expression and purification of rOVA-HY and rOVA-3 proteins. (A) Schematic rendition of the rOVA-HY construct in which the OT-II epitope was replaced with the H-2I-A^b-restricted HY epitope (NAGFNSNRANSSRSS) recognised by CD4⁺ T cells elicited by C57BL/6 female mice immunised with C57BL/6 male splenocytes.^{5,6} Original cryptic, and OT-I epitopes were intact. rOVA-HY containing a C-terminal hexa-histidine tag was constructed by gene synthesis and cloned into pET24a for expression in *E. coli*. (B) Representative SDS/4–12%-PAGE image showing expression of indicated rOVA protein before and after purification over metal affinity column (see Materials and Methods for details).

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