

SUPPLEMENTAL INFORMATION

Cancer-associated substitutions in RNA recognition motifs of PUF60 and U2AF65 reveal residues required for correct folding and 3' splice site selection

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Table S1 Characterization of tested mutations

COSMIC mutation	Mutagenic primer ¹	Amino acid substitution	Domain	Secondary structure ²	FATHHM score ³	Malignancy ³	Source ³	Method ³
PUF60-419T>C	CTACTATGAGCcGGGGGAGGACA	L140P	RRM1	not assigned	0.96	Colorectal cancer	(1)	WES
PUF60-437G>A	GGACACCATCCaCCAGGCCTTTG	R146H	RRM1	α helix	0.96	Breast cancer	(2)	WGS
PUF60-436C>T	AGGACACCATCtGCCAGGCCTTT	R146C	RRM1	α helix	0.97	Colon adenocarcinoma	(3)	WES
PUF60-487G>A	ACATGTCCTGGaACTCCGTCACC	D163N	RRM1	β sheet	0.96	Hairy cell leukaemia	(4)	WES
PUF60-493G>A	CCTGGGACTCCaTCACCATGAAG	V165I	RRM1	not assigned	0.99	Colon adenocarcinoma	ICGC(COAD-US)	WES
PUF60-688G>A	ACCGCATCTAcTAGGCCTCTGTG	V230M	RRM2	β sheet	0.97	Malignant melanoma	(5)	WGS
PUF60-691G>C	GCATCTACGTGcCCTCTGTGCAC	A231P	RRM2	β sheet	0.98	Prostate cancer	(6)	WES
PUF60-719A>T	CCTCTCAGACGtTGACATCAAGA	D240V	RRM2	α helix	0.98	Prostate cancer	(7)	WES
PUF60-761_762CC>TT	GCAAGATCAAGTtTGACACTGGCC	S254F	RRM2	β sheet	N.A.	Squamous cell carcinoma	(8)	WES
PUF60-776G>A	CACACTGGCCCaGGACCCACAA	R259Q	RRM2	β sheet	0.98	Colon adenocarcinoma	(9)	WES
PUF60-802G>A	GCAAGCACAAAGaGCTACGGCTTC	G268S	RRM2	β sheet	0.96	Colon adenocarcinoma	(9)	WES
PUF60-825G>T	ATTGAGTACGAtaAAGGCCAGTCG	E275D	RRM2	bend	0.96	Endometroid carcinoma	ICGC(UCEC-US)	WGS
PUF60-892C>T	GCCAGTACTTgGGGTGGCAAGG	R298W	RRM2	β sheet	0.96	Colon adenocarcinoma	(9)	WES
U2AF65-460G>A	GCCTCTACGTGaGAACATCCCC	G154S	RRM1	β sheet	0.94	Myelodysplastic syndrome	(10)	TES
U2AF65-461G>T	CCTCTACGTGtCaACATCCCCT	G154V	RRM1	β sheet	0.96	AML	(11)	TES
U2AF65-484G>A	TTGGCCTACATaAGGAGGCCATG	E162K	RRM1	α helix	0.95	Bladder carcinoma	(12)	WES
U2AF65-485A>T	TGGCATCACTGtGGAGGCCATGA	E162V	RRM1	α helix	0.97	Melanoma	ICGC(SKCM-US)	WES
U2AF65-527G>T	GATGCGCCTGGtGGGGCTGACCC	G176V	RRM1	α helix	0.94	Liver carcinoma	TCGA-CC-A7IH-01	WES
U2AF65-527G>A	GATGCGCCTGGaGGGGCTGACCC	G176E	RRM1	α helix	0.93	Lung adenocarcinoma	ICGC(LUAD-US)	WGS
U2AF65-527G>C	GATGCGCCTGGcGGGGCTGACCC	G176A	RRM1	α helix	0.93	Breast carcinoma	27135926	WES
U2AF65-569A>T	GTTGCTGTGtGcGATTAACCAGG	Q190L	RRM1	β sheet	0.98	CLL	(14)	WES
U2AF65-584A>G	TAACCAGGACAgGAATTTGCTT	K195R	RRM1	turn	0.98	T-cell leukaemia	(15)	WES
U2AF65-588T>A	CAGGACAAGAAaTTTGCCTTTTT	N196K	RRM1	turn	0.80	AML	ICGC(LAML-KR)	WES
U2AF65-620A>G	CTCAGTGGACgGACTACCCAGG	E207G	RRM1	α helix	0.98	Melanoma	ICGC(SKCM-US)	WES
U2AF65-631G>A	AGACTACCCAGaCTATGGCCTTT	A211T	RRM1	α helix	0.98	Prostate cancer cell line	23856246	WES
U2AF65-790G>T	AGCTGTTATCtGGGGCTTACCC	G264W	RRM2	β sheet	0.99	Liver carcinoma	ICGC(LICA-CN)	WES
U2AF65-817G>A	ACCTGAACGATaACCAGGTCAAA	D273N	RRM2	α helix	0.99	Squamous cell carcinoma	(8)	WES
U2AF65-876G>T	AACCTGGTCAaGACAGTGCCAC	K292N	RRM2	β sheet	0.89	Colorectal carcinoma	(17)	WES
U2AF65-901G>A	GGCTCTCCAAGaGCTACGCCTTC	G301S	RRM2	β sheet	0.97	Renal carcinoma	ICGC(KIRP-US)	WES
U2AF65-908C>T	CAAGGGCTACGtCTTCTGTGAGT	A303V	RRM2	β sheet	0.93	Melanoma	(18)	WES
U2AF65-922G>A	TCTGTGAGTAcTAGGACATCAAC	V308M	RRM2	bend	0.96	Melanoma	(19)	WES
U2AF65-956G>T	GGCCATTGCGtGCTGAACGGCA	G319V	RRM2	α helix	0.97	Cervix adenocarcinoma	ICGC(CESC-US)	WGS
U2AF65-955G>A	AGGCCATTGCGaGGCTGAACGGC	G319R	RRM2	α helix	0.97	Squamous cell carcinoma	(20)	WES
U2AF65-977G>T	CATGCAGCTGGtGGATAAGAAGC	G326V	RRM2	turn	0.97	Glioma	TCGA-HT-A4DV-01	WES
U2AF65-976G>A	GCATGCAGCTGaGGGATAAGAAG	G326R	RRM2	turn	0.98	Lung adenocarcinoma	(21)	WES
U2AF65-977G>A	CATGCAGCTGGaGGATAAGAAGC	G326E	RRM2	turn	0.97	Pancreatic carc.	ICGC(PACA-IT)	WES
U2AF65 (PUF60 H169Y paralog)	AGATTAACCAgACAAGAATTTTG	D194Y	RRM1	turn	N.A.	Germline PUF60 deficiency	(22)	WES

Legend: ¹Mutated positions are in lower case. ² PDB accession numbers: 5KWQ (PUF60) and 2YH1 (U2AF65). ³ COSMIC database (v. 91). WES, whole exome sequencing; WGS, whole genome sequencing; TES, targeted exome sequencing.

Table S2 Cloning and RT-PCR primers

Primer	Sequence (5'-3')
Expression plasmids	
SF3B4-49F- <i>BamI</i>	ATTAGGATCCAGACGGCGGGATCTCTTT
SF3B4-49R- <i>XbaI</i>	ATTATCTAGACTGAGGGAGAGGGCCTCGAAGTG
U2AF2-R- <i>XhoI</i>	ATTACTCGAGCCAGAAGTCCCGGCGGTGAT
PUF60 and U2AF2 minigenes	
PUF60-E6-F	ACCACTCGAGCTGCCTCCTGACCATCTGTC
PUF60-E9-R	ACCATCTAGACCCTGGTCTGGCTACTCAAG
U2AF2-E5-F	ACCACTCGAGCTTCTGAGGAGCAGCAGTT
U2AF2-E5-R	ACCATCTAGAGGGTTGCTGGAGGAGGTT
U2AF2-E6-F	ACCACTCGAGTGTGTCATCATGCCCTCTG
U2AF2-E6-R	ACCATCTAGACTGGAAAAGGCCAAAGAGGT
U2AF2-E7-F	ACCACTCGAGCAGCACTTTGTCCCTCTTCC
U2AF2-E7-F	ACCATCTAGATTTGGTCTGATCCTGGCTCT
U2AF2-E8-E10-F	ACCACTCGAGTGTGAACTTTGTGCCTTTGG
U2AF2-E8-E10-F	ACCATCTAGACCAGTCTCCCTCTGCTCAAG
RT-PCR	
Vector primer PL3	GGGAGACCCAAGCTGGCTA
Vector primer PL4	AGTCGAGGCTGATCAGCGG
35F	CAGGTGCTCTCGGTTGCA
35m-amplF	GCTCGGATCCTACAGAGTCAA
U2AF2_E10R	CTCTGGACCAGCAGCTTCTT
U2AF2_E9F	AACCTGGTCAAGGACAGTGCCAC
U2AF2_E9R	GCACTGTCCTTGACCAGGTT
PUF60_E7F	GCCTTGGAGCAGATGAACTC
PUF60_E7R	AGCTTCGGGGACCTCATACT
UBE2F-F	GCATTTTCCTGATCCAAACAA
UBE2F-R	CCCTGTCTCTGTGATGTTGG
UBE2F-cr	GCTAACAGTAACCCCAGGATT

Figure S1 Sequences of wild-type splicing reporter inserts

Full reporters are schematically shown in Figures 1A, 3B and 5. Exons are in upper case and intronic sequences are in lower case. Restriction sites are underlined. Tested exons (highlighted and in bold) and flanking intronic sequences are in black, *U2AF1* exons 2 and 4 and flanking intronic sequences are in grey. Conserved dinucleotides at cryptic (*UBE2F* and *GANAB*) or alternative (*U2AF2*) splice sites are in red. Plasmid mutations are shown in Table S1.

UBE2F

ggatcc**TACAGAGTCAACTGTTTCA**TTTTATTTCAA**AATGGAGCATGTCGTCATGGAGACAGGTGCTCTCGGTTG**
CACAATAAACCGACGTTTAGCCAGgtttggttgcccttttttcatgtaaattataaaaaacttcatgttcttttca
aagacagttaatttctacatattaagcaagtcattttttctctcgtagttgtattttccatttgggttcataagt
gtgttcttttatttaataatagtgaggcaggtgatcgacttccagtgggaaggtctgagaccaccactccttggt
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gaactctgcaggcctgttttttaactcctgattctctgtatttggccatatcctaacagagtgtttgctttttgtt
tccttttttttttaatttaaagattttttgttttgttttggatag**ATGAGGGTTACTACCAGGGT**
GGAAAATTTAGTTTGA**AACTGAAGTTCCCGATGCGTACAACATGGTG**gtgagtagcctgcgttgagcctgttgt
tttacttctcggggggggtgcctgtggctgtggcccgccactggcacagggccctagcactcccctctgcgtgt
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ttcactcgggttaacatgttctaacccagcaagatttctgttgtttgcag**GTGCCGTGAGCGATGTGGAGATGCAGG**
AACACTATGATGAGTTTTTTGAGgggccc

GANAB

gaattcg**GCTTTCTGTCTCTGGTCGTGATGAGAACAGTGTGGAGTTAACCATGGCTGAGGGACCTACAAGATCA**
TCTTGACAGCACGGCCATTCCGCCTTGACCTACTAGAGGACCGAAGTCTTTTGCTTAGTGCAATGCCCGAGGAC
TCTTGGAGTTTGAGCATCAGAGGGCCCCTAGGGTCTCgtgagtacaggggttgggactgcagggaacctagtgtg
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TCAAAAGACCCAGCTGAGGGCGATGGGGCCCAGCCTGAGGAAACACCCAGGGATGGCGACAAGgcaagttctaga

OGDH

ggatccTACAGAGTCAACTGTTTCAATTTTATTTCAAATTTGGAGCATGTCGTCATGGAGACAGGTGCTCTCGGTTG
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tttaaacacaagagttcactcgggttaacatgttcttaaccagcaagatttctgttgtttgcag**GTGCCGTGAGCGA**
TGTGGAGATGCAGGAACACTATGATGAGTTTTTTGAGgggccc

U2AF2 exon 5

ggatccTACAGAGTCAACTGTTCAATTTATTTCAAATTTGGAGCATGTCGTCATGGAGACAGGTGCTCTCGGTTG
CACAATAAACCGACGTTTAGCCAGgtttggttgcccttttttcatgtaaattataaaaaacttcatgttcttttca
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GTTTTTTGAGgggccc

U2AF2 exon 6

ggatccTACAGAGTCAACTGTTCAATTTATTTCAAATTTGGAGCATGTCGTCATGGAGACAGGTGCTCTCGGTTG
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GCAGATTAACCAGGACAAGAATTTGCTTTTTGGAGgtgagctgggggagtgagtgaggtccaggaaacgtgtg
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U2AF2 exon 7

ggatccTACAGAGTCAACTGTTCAATTTATTTCAAATTTGGAGCATGTCGTCATGGAGACAGGTGCTCTCGGTTG
CACAATAAACCGACGTTTAGCCAGgtttggttgcccttttttcatgtaaattataaaaaacttcatgttcttttca
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U2AF2 exons 8-10

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CAGGAACACTATGATGAGTTTTTTGAG^{gggccc}

PUF60 exons 6-9

ggatcc**TACAGAGTCAACTGTTTCAATTTTATTTCAA**AATTGGAGCATGTCGTCATGGAGACAGGTGCTCTCGGTTG
CACAATAAACCGACGTTTAGCCAGgtttggttgcccttttttcatgtaaaattataaaaacttcatgttcttttca
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ctcccctctgtcctgctgcag**ATGGCGGCTCAGCGGCAGCGGGCGCTGGCCATCATGTGCCGCTCTACGTGGGC**
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ATGTCCTGGGACTCCGTCAACATGAAGCACAAGgtcagcaggttgggtccgccccggcacttcgggctgcctcc
caccccctgggctcgcgcagcctgacaggtgtgtccctgtgtctag**GGCTTTGCCTTCGTGGAGTATGAGGTCCCC**
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ggtctttccatctcaccgcctcttccccag**GTGGGCAGACCCAGCAACATAGGGCAGGCCAGCCCATCATAGAC**
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ATCAAGAGCGTGTGTTGAGGCCTTTGGCAAGATCAAGTCTGCACACTGGCCCGGGACCCACAAC**TGGCAAGCAA**
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tactgtgtcctcctgcccacag**AGTACGAGAAGGCCAGTCGTCCCAAGATGCTGTGTCTTCCATGAACCTCTTT**
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GGAGGCCTCCCACCTGCCGCTGCTGTGGCAGCTGCTGCAGCCACTGCCAAGATCACAGCTCAGgtgagggcccac
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agaccaggg**ctaga**taatctcgtgtgtgttactgtagcagtttgagtttaaacacaagagttcactcgggttaac
atgttctaaccagcaagatttctgttgtttgcag**GTGCCGTGAGCGATGTGGAGATGCAGGAACACTATGATGAG**
TTTTTTGAGgggccc

Figure S2 Alignment of human U2AF65, PUF60 and SF3B4 RRMs

Residues that sustained missense mutations in cancer (23) are in red; residues tested in this work are highlighted in yellow. Substitution D231V that selectively increased affinity for uridines and discriminated against purines in a short synthetic PPT-derived DNA (24) is highlighted in grey. A conserved aromatic residue that recognized a guanine in *syn* (24) is in magenta. Residues mutated in the Verheij syndrome (22,25) are highlighted in green. rU7-U2AF65, side chain contacts (s), water-mediated contacts (w) and main chain contacts (m) were compiled from the structure of the human U2AF65/rU7 complex (26).

RRM1

```

rU7-U2AF65      s s ww
PUF60-RRM1     CRVYVGSIIYYEIGEDTIRQAFAPF-----GPIKSIMSWDSVTMKKGFAFVEY177
U2AF65-RRM1    RRLYVGNIPFGITEEAMMDFFNAQMRLGGLTQAPGNPVLAV-----QINQDKNFAFLEF202
SF3B4-RRM1     ATVYVGGLDEKVSEPLLWELFLQA-----GPVVNTHMPKDRVTGQHQGYGFVEF
                :***.: : * : : * .*: :...:***:

                m
rU7-U2AF65      s m sm
PUF60-RRM1     EVPEAAQLALEQMNSVMLGGRNIKVGRPSN207
U2AF65-RRM1    RSVDEETTQAAMA-FDGIIFQGQSLKIRRPH231
SF3B4-RRM1     LSEDDADYAIKIMNMIKLYGKPIRVNKASA
                : : * : : : * : : : :

```

RRM2

```

rU7-U2AF65      w
                s s ss s
PUF60-RRM2     NRIYVASVHQDLSDDDIKSVFEAFGKIK-SCTLARDPPTTGKHKCYGFIEYEEKAQSSQDAV283
U2AF65-RRM2    HKLFIGGLPNYLNDDQVKELLSFGPLK-AFNLVKDSATGLSKGYAFCEYVDINVTQAI317
SF3B4-RRM2     ANIFIGNLDPEIDEKLLYDTFSAFGVILQTPKIMRDPDTGNSKGYAFINFASFDASDAAI
                :...: :..: : : ** : : : * ** ***. * : : : : * :

                w
rU7-U2AF65      ss s m
PUF60-RRM2     SSMNLFDLGGQYLSVGKAVT303
U2AF65-RRM2    AGLNGMQLCDKKLLVQRASV337
SF3B4-RRM2     EAMNGQYLCNRPITVSYAFK
                .:* * : : * *

```

Figure S3 Splicing pattern of PUF60-/U2AF65-dependent reporters in cells overexpressing SF3B4

A, Amino-acid substitutions in SF3B4 in cancer. Data are from the COSMIC database, v. 83 (23). Data could not be corrected for multiple database entries from the same patients or other redundancies (Dave Beare, COSMIC, the Sanger Centre, personal communication). **B**, Coexpression of the *UBE2F* (upper panel) or *OGDH* (middle panel) reporters with human SF3B4 and controls in human embryonic kidney 293 cells. RNA products (to the right) are schematically shown in Figure 1. Immunoblot from transfected cells is in the lower panel.

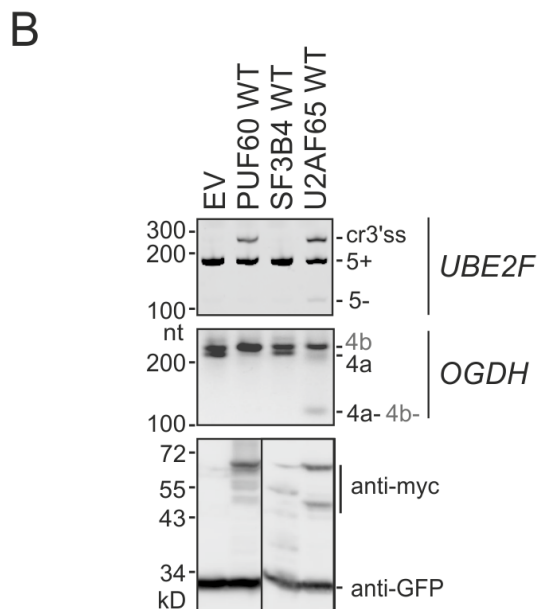
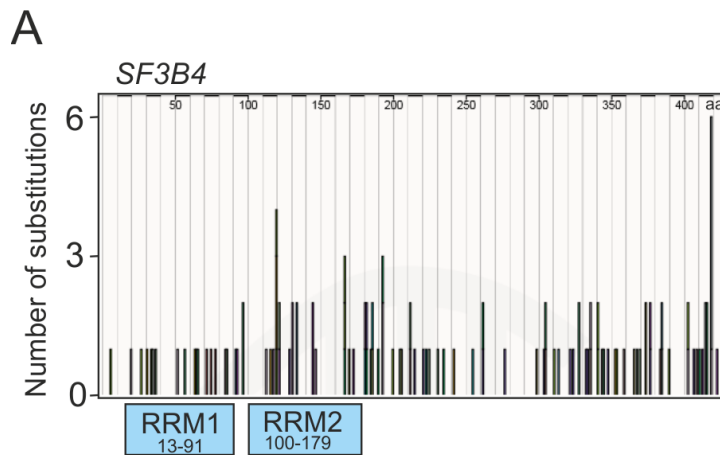


Figure S4 U2AF65 E162K binding to oligoribonucleotides derived from canonical and cryptic 3'ss of *UBE2F* exon 5

A, EMSA with purified recombinant U2AF65 E162K and RNAs indicated at the bottom. The amounts of proteins and RNAs were the same as for the WT (shown in Figure 6A,B). **B**, Fraction of bound RNA with K_d estimates. Hill coefficients were 2.23 ± 0.20 (can) and 1.84 ± 0.21 (cr).

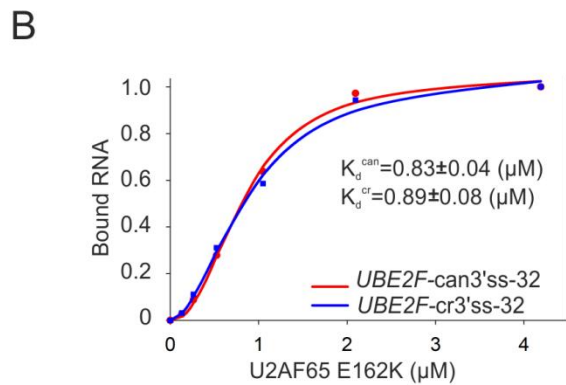
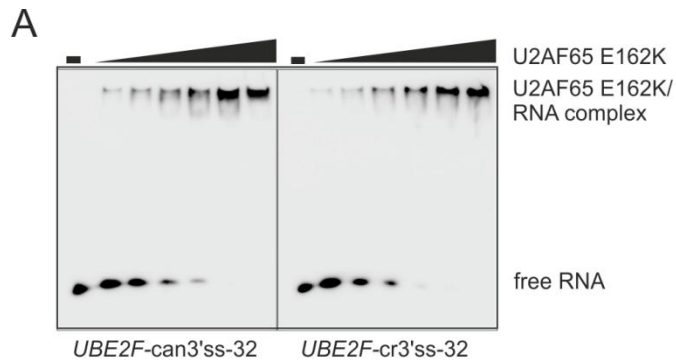


Figure S5 Non-isotopic EMSA with WT and mutated PUF60 and RNA probes derived from competing 3'ss of *UBE2F* exon 5

Substitutions are shown at the top, biotin-labelled RNA probes at the bottom. Their sequences are in Table 1. Concentration of recombinant proteins was 0.19, 0.58 and 1.76 μ M.

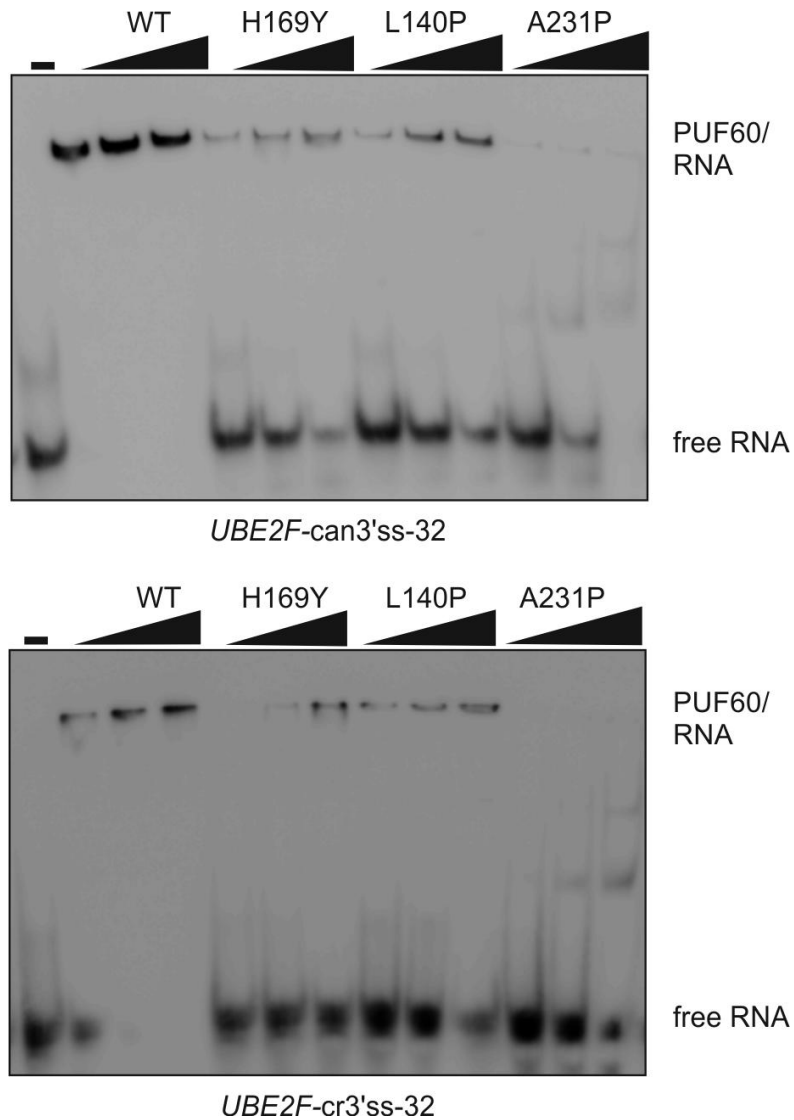


Figure S6 Binding of PUF60 H169Y to *UBE2F*-derived oligoribonucleotides

A, EMSA with purified recombinant PUF60 H169Y and 32-mer RNAs derived from canonical (left panel) and cryptic (right) 3' ss. The amount of proteins and RNAs was the same as for the WT (shown in Figure 7B). **B**, Fraction of bound RNA with approximate K_d estimates (without saturation). The Hill coefficients were 2.4 ± 0.5 (can) and 6.4 ± 2.5 (cr).

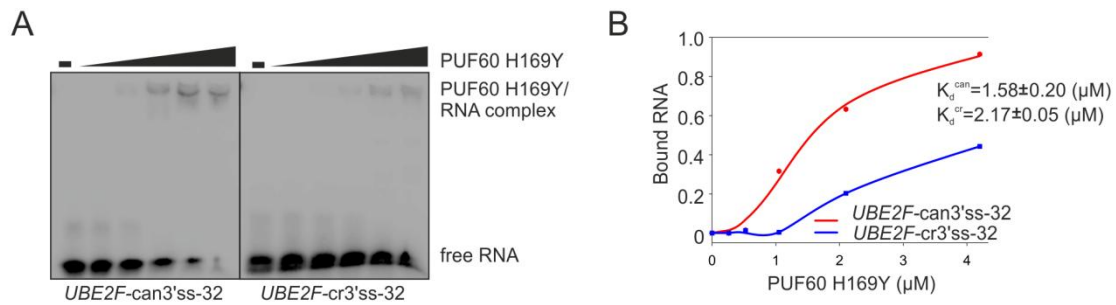


Figure S7 PUF60 L140P and A231P induce protein destabilization and misfolding

Coomassie-stained gel (upper panel) blotted with anti-His antibodies (lower panel) after overnight digestion of the indicated proteins with the TEV protease. The amount of each protein loaded on to the gel was 1.40, 0.46 and 0.14 μg .

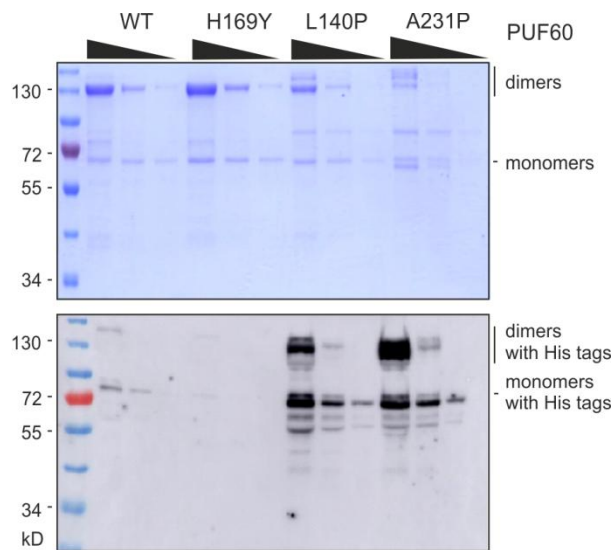


Figure S8 Solubility and stability profiles of wild-type and mutated PUF60 or U2AF65

A, PUF60. **B**, U2AF65. Scores higher than 1 denote highly soluble regions whereas scores smaller than -1 denote poorly soluble regions, as predicted by CamSol (27). Blue/red arrows denote substitutions in regions with potential increased/reduced solubility. Changes in protein stability ($\Delta\Delta G$) were predicted using a mutation cut-off scanning matrix (28).

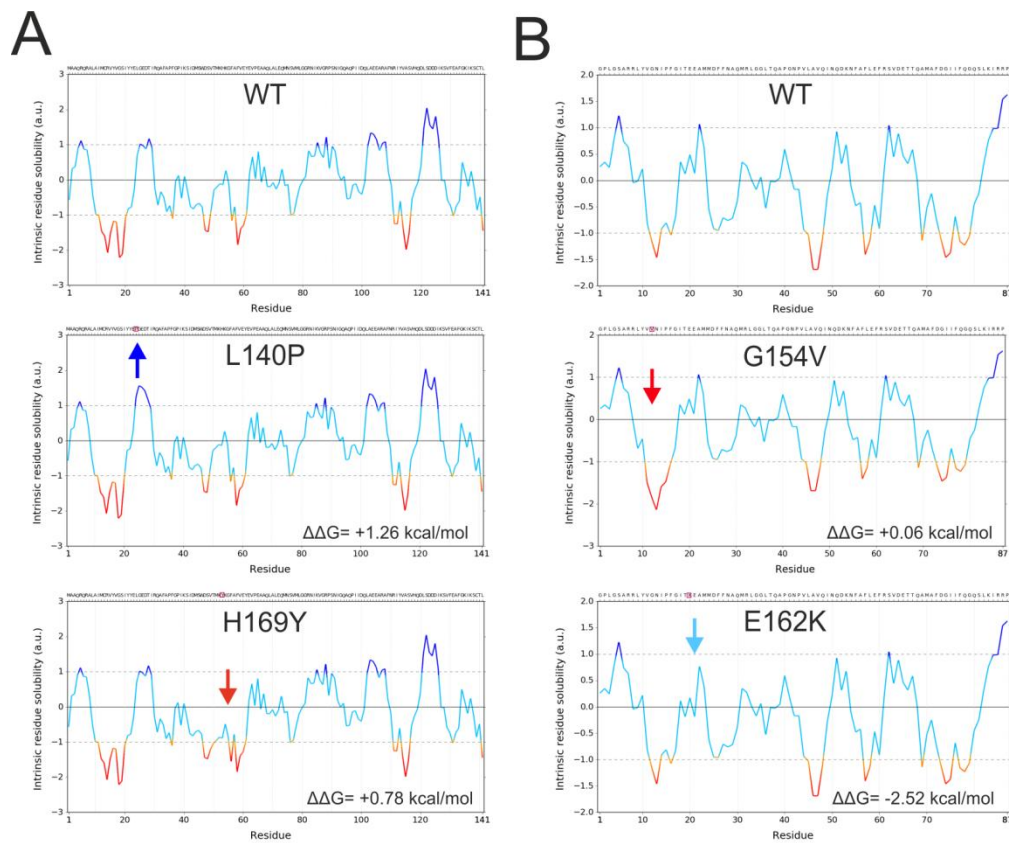


Figure S9 H169Y in the crystal structure of PUF60 RRM1

PDB: 5KW1 (Crichlow et al., unpublished). The protein is shown as cartoon in purple. H169Y and surrounding hydrophobic residues are shown in orange, nitrogen atoms in blue, and oxygen atoms in red. Dotted line denotes a hydrogen bond between H169 and L140.

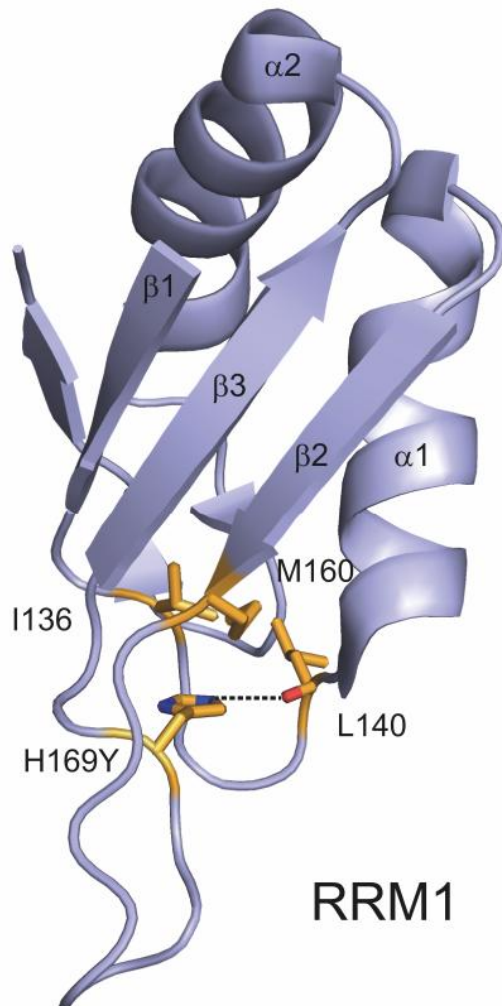
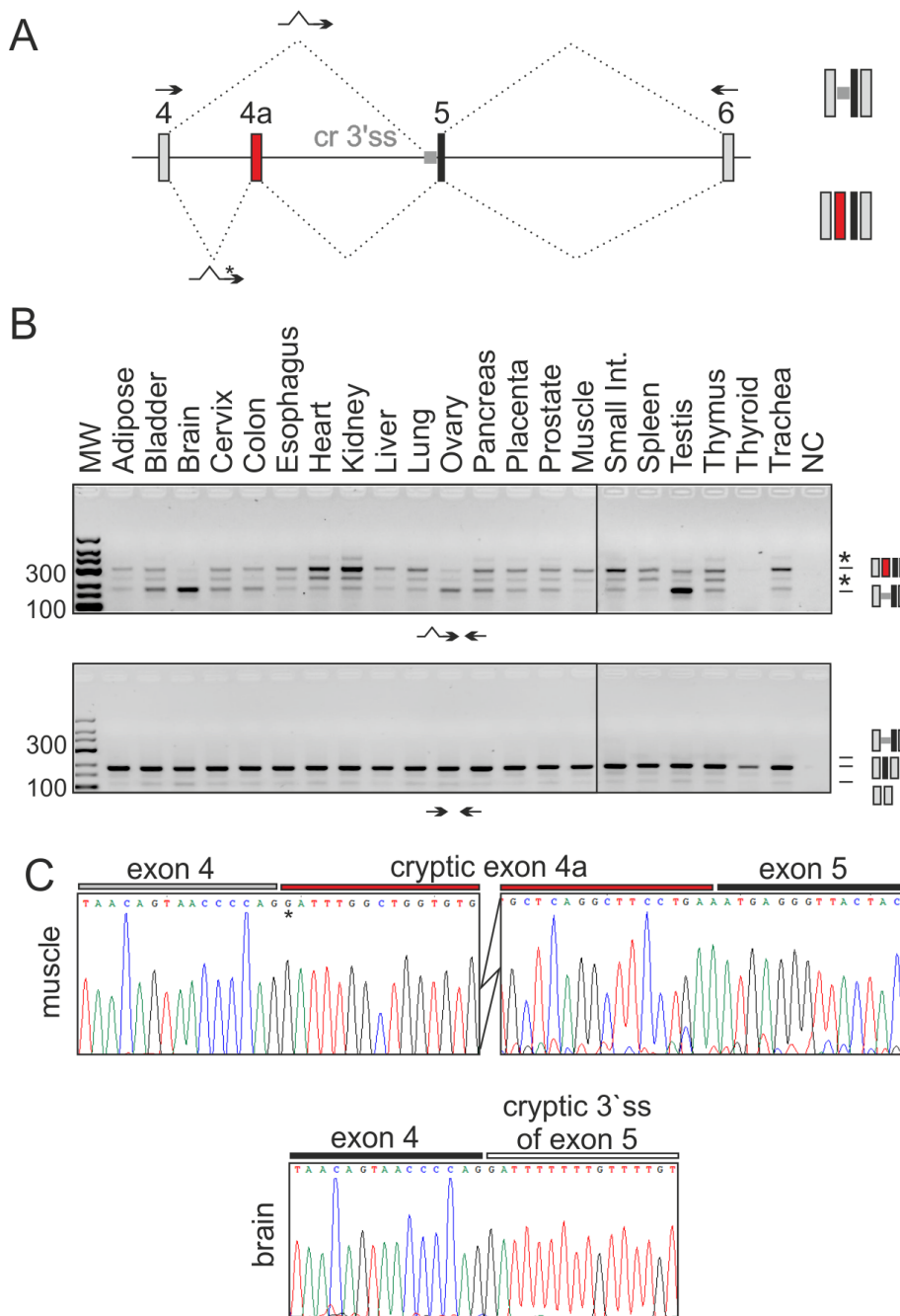


Figure S10 Utilization of cryptic 3'ss of *UBE2F* exon 5 in human tissues

A, Schematics of the *UBE2F* splicing pattern. Exons are shown as boxes, introns as lines, spliced products (shown to the right) as dotted lines. Arrows indicate locations of primers for amplifying canonical (in exon 4 and 6) and non-canonical (across cryptic 3'ss of exon 5 and in exon 6) products. Asterisk denotes a G>A mismatch between the primer and the first position of exon 4a (the sequence is in panel C). **B**, RT-PCR products amplified from cDNA samples prepared from human tissues and separated on agarose gels. Amplification primers (Table S2) are schematically shown at the bottom. The cryptic 3'ss of exon 5 could be visualized with primers in exons 4 and 6 only after >33 PCR cycles (not shown). MW, size marker (nt); NC, no target control. Minor species/heteroduplexes are denoted by asterisks. **C**, Sanger sequencing of the two non-canonical RNA products excised from panel B lanes identified to the left.



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