

Supplementary Figure 1: A-B:Tumor lines were transduced with lentiviral vector bearing a cDNA fusion of GFP and F-luciferase. Representative images of cells in culture showing GFP expression (A) and firefly luciferase expression detected in vitro in titrating cell number (B). Images are representative of 3 independent experiments and data represented as average  $\pm$  SE for N=3. Data representative of 3 independent experiments. C: Tumor-growth tracked in vivo by bioluminescence imaging of tumor-bearing mice shows increase in signal over time. Data as average  $\pm$  SE for N  $\geq$  5. D: GFP expression in brain sections obtained from end-stage tumor-bearing mouse. Images are representative of 3 independent experiments. E: CD31 staining was quantified using wimasis software for tube length. Data represents average  $\pm$  SE for N  $\geq$  5.. Two-sided Student's T-test was performed.\*p< 0.05. F: Ki67+ and DAPI+ cells were counted in 3 random fields in the tumor tissue section and their ratio is reported as average  $\pm$  SE for 3 images/tumor type. Two-sided Student's T-test was performed.\*p< 0.0005



Supplementary Figure 2: RNA sequencing analysis on the RNA isolated from end-stage tumors or control brains of C57bl6 mice (N=3 mice/group). A: Principal component analysis with PC1 and PC2 plotted for control sample and tumor samples. B: Heatmap of top 100 differentially expressed genes.

0.02

0.04

GeneRatio



Mut3 tumor



0.06

Count

p.adjust

3e-05

26-05

1e-05

0.08

Cation transmembrane transport Inorganic ion transmembrane transport Modulation of synaptic transmission Regulation of membrane potential Inorganic cation transmembrane transport Regulation of transmembrane transport Regulation of ion transmembrane transport Signal release Synapse organization Cognition Learning of memory Regulation of synaptic plasticity Locomotory behavior Neuron-neuron synaptic transmission Potassium ion transport Neurotransmitter transport Learning Regulation of neurotransmitter levels Positive regulation of synaptic transmission Cellular potassium ion transport Potassium ion transmembrane transport Memory Regulation of postsynaptic membrane potential Chemical synaptic transmission, postsynaptic

Supplementary Figure 3: Differential expression analysis was performed for each tumor sample using control as a reference. Genes that were significantly different were used for pathway enrichment analysis. Top 25 differentially expressed pathways as compared to naïve brain have been shown for CT2A and Mut3. The statistical test was a hypergeometric test as implemented in the ClusterProfiler R/Bioconductor package (Yu, G. et al, 2012).





005 tumor Innate immune response Immune effector response Response to cytokine Defense response to other organism Regulation of immune response Cellular response to cytokine stimulus Response to bacterium T cell activation Adaptive immune response (AIR) Leukocyte mediated immunity Leukocyte migration Lymphocyte mediated immunity Lymphocyte proliferation AIR based on somatic recombination of immune receptors built from Ig superfamily domains Defense response to bacterium Response to interferon-gamma Leukocyte chemotaxis Antigen processing and presentation (APP) Cellular response to interferon-gamma Response to interferon-beta APP of peptide antigen Cellular response to interferon-beta APP of exogenous peptide antigen APP of exogenous peptide antigen via MHC class II

#### GL261 tumor Innate immune response Response to cytokine Cellular response to cytokine stimulus Interspecies interactions between organisms Symbiosis, encompassing mutualism through parasitism Defense response to other organism Positive regulation of immune response Multi-organism cellular process Viral process Adaptive immune response (AIR) Response to interferon-beta Response to virus AIR based on somatic recombination of immune receptors built from Ig superfamily domains Response to interferon-gamma Lymphocyte mediated immunity Defense response to virus Antigen processing and presentation (APP) Cellular response to interferon-beta APP of peptide antigen Cellular response to interferon-gamma APP of exogenous antigen APP of exogenous peptide antigen APP of peptide or polysaccharide antigen via MHC class II APP of peptide antigen via MHC class II APP of exogenous peptide antigen via MHC class II

Supplementary Figure 4: Differential expression analysis was performed for each tumor sample using control as a reference. Genes that were significantly different were used for pathway enrichment analysis. Top 25 differentially expressed pathways as compared to naïve brain have been shown for 005 and GL261. The statistical test was a hypergeometric test as implemented in the ClusterProfiler R/Bioconductor package (Yu, G. et al, 2012).



Supplementary Figure 5: Mass cytometry and flow Cytometry comparison. Mixed mouse tumor samples were stained for fluorescent-tagged or metal-tagged antibodies and run through flow cytometer or CyTOF, respectively. Cells were pre-gated for live cells. Gates in each plot show frequency of positive cells. n=4 independent biological replicates. Paired two-sided T-test showed no significant difference. NS= not significant for p<0.05.





Supplementary Figure 7: Percent population of activated and resting microglia in different tumor types were compared to naïve brain. Data represented as average  $\pm$  SE for n=3 mice/group. Two-sided student's t-test with Holm-Sidak corrections was applied \*\*\*p≤ 0.0005.



Supplementary Figure 8: A: Markers used for broad population characterization of immune cells in GL261 tumor model were overlaid on a SPADE diagram. Color represents the arc-sinh transformed median expression of each marker. Size of the node represents abundance of population. Flow plots depicting those populations are plotted underneath each SPADE diagram.



S3A overlaid on viSNE plots for GL261 tumor model. Each dot represents a single cell and color gradient correlates with expression level of the defined marker.

-18.27

5.904 0

tSNE2



Supplementary Figure 10: A: CD39 expression overlaid on viSNE of concatenated samples of individual tumor types. Each dot represents a single cell and color gradient correlates with expression level of CD39. B: Histogram overlays of CD39 expression in total ungated sample, CD4 and CD8 subsets for the four tumor types. C: Data represented in (B) is plotted as average  $\pm$  SE for n=3 mice/group. Two-sided Student's t-test was applied. \*p≤ 0.05, \*\*p≤ 0.005.



Supplementary Figure 11: Relative percentage of cell populations expressing various T cell function markers in CD4 (A) and CD8 (B) T cell populations in 4 tumor types represented as average  $\pm$  SE for n=3 mice/ group. Two-sided Student's t-test with Holm-Sidak corrections for multiple comparisons was applied. \*p≤ 0.05; \*\*p≤ 0.005.



Supplementary Figure 12: Tumors harvested at end stage were sectioned and stained with CD3, CD4, CD8, Foxp3, CD68 and Iba-1. Number of brown cells were counted as positive for each marker in each tumor type and plotted as number of cells identified/field. Data represented as average  $\pm$  SE for N  $\geq$  5. Student's t-test with Holm-Sidak corrections for multiple comparisons was applied \*p< 0.05; \*\*p< 0.005.



Supplementary Figure 13: Tumors were classified into immunologically active (GL261 and 005) and immunologically silent (CT2A and Mut3) based on RNAseq analysis. FlowSOM analysis was performed followed by abundance analysis. Populations that were not significantly different were plotted. Data is represented as average  $\pm$  SE for each cluster characterized. B-C: Relative percentage of cell populations expressing various T cell function markers in CD4 (A) and CD8 (B) T cell populations in immunologically inert and active tumor types represented as average  $\pm$  SE for n=6 mice/group, representative plots from 2 experiments. Two-sided Student's t-test with Holm-Sidak corrections for multiple comparisons was applied \*p ≤ 0.05.



Supplementary Figure 14: Percentage of cells for populations not showing significant differences were plotted as average  $\pm$  SE (n=6 mice/group, representative plot from 2 independent experiments).

Cell- lineage		Activation
CD3 CD4 CD8 CD11b CD11c NKp30 CD19 CD20 IgM CD172ab CD45 CD64 CD127 CD33 CD169 Ki67 Basophil	CX3CR1 CD14 CD16 CD115 CD103 CD86 CD15 CCR2 TMEM119 Siglec8 CD68 HLA-DR CD180 CD163 CD304 STING CCR3	CD279/PD1 CD39 CD69 Tim3 CTLA4 CD25 PD-L1 CD66b

Supplementary Figure 15: Antibody panel for anti-human CyTOF analysis separated into lineage-identifying and activation status markers.



Supplementary Figure 16: CyTOF on isolated lymphocytes from tumor tissue and matched blood from 5 GBM patients and 5 healthy donors was performed. A: FlowSOM analysis for the three tissue types. B: Frequency of T cell subsets were calculated from total T cells GBM tumor tissue, blood from GBM patients and control blood. Data represented as average  $\pm$  SE for n=5 independent biological repeats. Two-sided Student's t-test with Holm-Sidak corrections for multiple comparisons was applied. \*p $\leq$  0.05.





Supplementary Figure 17: Uncropped western blot images shown in Figure 1b