# **Table of Contents**

Appendix Figure S12	
Appendix Figure S25	
Appendix Figure S37	
Appendix Figure S49	
Appendix Figure S511	
Appendix Figure S613	
Appendix Figure S715	
Appendix Figure S817	

Appendix Figure S1. (a-b) Samples cluster by developmental time-points (a) and not by sequencing batches (b). First two principal components for 225 3' Tag-seq samples (75 samples at three time-points). Each dot represents a sample, samples are coloured by timepoints (a) and sequencing date (b). All genes with quantified gene expression were used for the analysis. PCA was done on the raw expression counts after applying variance stabilization transformation from DESeq2 (Love et al, 2014). (c) Residual coefficient of variation (expression variation) reflects expression heterogeneity across samples at any given expression level. Gene expression across samples (y-axis, size factor normalized read counts) for genes binned by median expression level (20 subplots for 5%-percentiles by expression level) and ordered by expression variation (x-axis). Each dot represents a gene in one of 75 samples. Top and bottom 5% of genes by expression level were removed as potential source of outliers (the first and the last sub-plots, outlined with red). Data is shown for 10-12h timepoint. (d) Gene expression variation is consistent across timepoints. Diagonal: distributions of gene expression variation values at three time-points. Upper triangular panels: Pearson correlation coefficients of gene expression variation between pairs of time-points. Lower triangular panels: scatter plots showing relationship between gene expression variation at different time-points (x and y-axis). Each dot represents a gene. Final set of 4074 genes was used at all three time-points. (e) Spearman correlation coefficients in expression variation (solid lines) and median expression levels (dashed lines) between pairs of genes located at varying quantile distances between their TSSs (actual distance intervals in kB are shown on the x-axis). Only gene pairs located on the same chromosome and with TSS-to-TSS distance <100 kB are considered. Spearman correlation coefficient (y-axis) and quantiles by distances (in kB) (x-axis) is plotted separately for gene pairs within the same TAD (blue) or split into different TADs (orange), TAD coordinates were taken from (Ramírez et al, 2018). Number of gene pairs in each group is indicated (blue and orange font for gene pairs within the same

TAD or crossing TAD border, respectively), only groups with >100 gene pairs are shown. (f) Proportion of gene expression variation explained by different components according to LIMIX variance decomposition (Methods). All three time-points (225 samples combined) were used in variance decomposition to achieve better convergence of the algorithm and more precise estimate of Cis component. Results for the final set of 4074 genes are plotted. Only Cis component (defined here as sum of Cis and (Cis x Environment) components from LIMIX) was used as a feature in subsequent random forest models.



Appendix Figure S 2. (a) Alternative measures of gene expression variation corrected for level dependence. From left to right: standard deviation after variance stabilizing transformation (sd\_vst) from DEseq2 (Anders & Huber, 2010); residual median absolute deviation (resid mad), inter-quartile range (resid iqr), and residual standard deviation (resid sd). The three later measures are LOESS residuals from regression on median expression level calculated in the same way as residual coefficient of variation (Methods). Data is shown for the final set of 4074 genes at 10-12h. Each dot represents a gene. Blue line shows LOESS regression fit, indicating no global dependence between corrected variation measures and median gene expression level. (b) Pearson correlation coefficient among different expression variation measures. Final variation measure used in the analysis is residual coefficient of variation (resid cv). Correlations were computed on the final set of 4074 genes at 10-12h. (c) Random forest performance (R^2) for predicting expression variation based only on features (left, same as Fig 1d) or on features and median expression level (right). Data is presented as mean ± SD (5-fold cross validation). (d) Random forest performance (R^2) for alternative measures of expression variation (same as in a-b). Data is presented as mean ± SD (5-fold cross validation). Random forest was run on the set of features important for predicting expression variation (resid cv, same as in Fig 1d). (e) Performance of random forest (R^2) for predicting expression variation for genes split into four quartiles by expression log2-fold change between 10-12h and 6-8h time-points. Data is presented as mean ± SD (5-fold cross validation). Numbers on the plot indicate number of genes in the corresponding quartiles. Horizontal red line indicates model performance on the full dataset (as in Fig 1d). (f) Performance of random forest for predicting expression variation when test and train sets come from different chromosome (arms). Prediction R^2 for each chromosome (arm) are shown as horizontal bars. Vertical red line indicates median performance across chromosomes.





Pearson Correlation

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0.91

0.97

0.91

0.00 0.25 0.50 0.75 1.00

Quartiles by absolute expression L2FC (10–12h vs. 6–8h)





Supplementary Figure 3. (a) GO functional enrichment (Molecular function) of genes with broad (left panel) and narrow promoters (right panel) split into four quantiles by expression variation (x-axis). Top GO terms ranked by p-value are shown (full list in Dataset EV6). P-values (Benjamini-Hochberg correction) and gene ratio from compareCluster function (R clusterProfiler package) are reported. Quartiles of expression variation (1-lowest, 4 - highest, same as in Fig. 3c) were calculated for broad and narrow promoter genes separately. Quantile intervals for broad promoter genes (1 to 4): [-1.06,-0.444]; (-0.444,-0.266]; (-0.266,-0.0754]; (-0.0754,1.89]. Quantile intervals for narrow promoter genes (1 to 4): [-0.98,-0.173]; (-0.173,0.0751]; (0.0751,0.416]; (0.416,1.99]. (b) Regulatory plasticity (standard deviation across tissues and developmental time-points) of DHSs around TSSs of genes with different expression variation and promoter shape. Each DHS was assigned to the closest gene for this analysis, only DHSs located less than 500 bp. from the annotated TSSs are considered. Median of corresponding DHS standard deviations was calculated for each gene. Left panel: comparison of regulatory plasticity of broad and narrow promoter genes. P-values = Wilcoxon test. Number of genes in each group indicated. Right panel: DHS standard deviation (x-axis) plotted against gene expression variation (y-axis). Each dot represents a gene. Colours represent types of the corresponding genes. Distributions of DHS standard deviation of each group of genes are shown above the scatter plot. (c) Expression variation of different groups of genes (same as in b) depending on whether they have orthologs in human. Only orthologs with high conservation rank were considered (Methods). P-values = Wilcoxon test. Number of genes in each group indicated. Cohen's d: 0.39, 0.1, 0.92, and 0.47 for broad, narrow TFs, narrow TATA, and other narrow groups respectively.

Fig. S3

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(a-f). Differences between broad, narrow-low and narrow-high genes by the number of different TF motifs in TSS-proximal DHSs (a, Cohen's d=0.65 for broad vs. narrow-low, d=0.28 for narrow-low vs. narrow-high), number of different TF peaks with motifs in TSS-proximal DHSs (b, Cohen's d=0.25 for broad vs. narrow-low, d=0.33 for narrow-low vs. narrow-high), polymerase II pausing index (c, Cohen's d=0.64 for broad vs. narrow-low, d=0.30 for narrowlow vs. narrow-high), number of different miRNA motifs in 3'UTR (d, Cohen's d=0.31 for broad vs. narrow-low, d=0.42 for narrow-low vs. narrow-high), number of different RNA-binding protein motifs in 3'UTRs (e, Cohen's d=0.29 for broad vs. narrow-low, d=0.36 for narrow-low vs. narrow-high), and number of TSS-distal DHSs (more than 500 bp and less than 10 kB around TSS) (f, Cohen's d=0.35 for broad vs. narrow-low, d=0.37 for narrow-low vs. narrowhigh). P-values from Wilcoxon rank test. (g) Top-50 important features for predicting expression variation within narrow promoter genes according to Boruta feature selection algorithm (with the corresponding importance for predicting expression level). Features are ordered by their importance for expression variation. Blue triangles indicate importance for variation, orange for level. Size and orientation of triangles correspond to absolute value and sign of correlation coefficient of feature with predicted variable, respectively. For binary features, point-biserial coefficient of correlation was used, otherwise Spearman coefficient of correlation. Label colours correspond to feature groups (same as in Fig. 2f). TFs enriched in TSS-proximal DHSs of broad and narrow promtoer genes (Fig 4c) are marked with red and blue start, respectively. (h) Gene scores by broad (left) and narrow (right) indices (same as Fig 4g). Colours correspond to gene types.





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(a) Expression variation (in our embryonic dataset) of genes differentially expressed (DE) upon any stress conditions in (Moskalev et al, 2015) compared to non-differentially expressed genes (non-DE). DE genes are split into 'up' (log-fold change > 0 in all stress conditions where gene is detected as DE), 'down' (log-fold change < 0 in all stress conditions where gene is detected as DE), and 'down-up' (gene is up-regulated in some experiments and down-regulated in the other) groups. Number of genes in each group is shown brackets in the figure legend. (b) Expression variation of genes differentially expressed (DE) upon different stress conditions from (Moskalev et al, 2015) compared to genes not differentially expressed in any of these experiments (non-DE). Stress conditions include: temperature (union of DE genes in three cold shock experiments at +4°C, 0°C, -4°C), radiation (union of 144 Gly, 360 Gly, and 864 Gly ionizing radiation), starvation (16 h), and fungi (union of 10 CFU and 100 CFU entomopathogenic fungus infection). P-values from Wilcoxon rank test, numbers under boxplots indicate number of genes. (b-e) Differences between DE genes (genes differentially expression in at least one stress condition from above) and non-DE genes by the number of conditions with DHS at TSS (b), and number of TSS-distal DHSs (c), polymerase II pausing index (d), number of different miRNA motifs in 3' UTR (e) for broad and narrow promoter genes separately. P-values from Wilcoxon rank test, numbers under boxplots indicate number of genes. Cohen's d: (c) 0.22 and 0.57, (d) 0.15 and 0.34, (e) 0.17 and 0.32, (f) 0.69 and 0.73 for broad and narrow promoter genes, respectively. (g-j) Same as (c-f) for comparison between genes that were found as DE in more than 10 studies with genetic perturbation (Dataset EV8) vs. genes DE in 0-10 studies. Cohen's : (g) 1.02 and 0.43, (h) 0.14 and 0.28, (i) 0.14 and 0.07, (i) 0.02 and 0.47 for broad and narrow promoter genes, respectively.

Fig. S5



Heatmap showing correlations (Spearman correlation coefficient) between expression levels and expression variations across different human tissues as well as their correlation with DE prior from (Crow *et al*, 2019). Labels contain tissue names from GTEx project; 'resid cv' stands for expression variation, 'median' indicates median expression level (log-transformed). 'Mean variation (by tissue adj)' is mean of expression variation across all tissues where a gene is expressed (corrected for expression level in each tissue separately, Methods). 'Mean variation (global adj)' is mean of coefficients of variations in all tissues where a gene was expressed, which was then corrected for mean expression level across the corresponding tissues (Methods). Colour code for the labels: red – expression levels; blue – expression variations; green – DE prior. Mean expression level and variation labels are highlighted with bold font.



#### Stomach resid cv Colon Transverse resid cv Small Intestine Terminal Ileum resid cv

Artery Coronary resid cv Artery Aorta resid cv Artery Tibial resid cv Esophagus Muscularis resid cv Esophagus Gastroesophageal Junction resid cv Prostate resid cv Prostate resid cv Nerve Tibial resid cv Nerve Tibial resid cv Nerve Tibial resid cv Adrenal Gland resid cv Brain Nucleus accumbens basal ganglia resid cv Brain Putamen basal ganglia resid cv Brain Hypothalamus resid cv Brain Caudate basal ganglia resid cv Brain Cortex resid cv Brain Cerebellar Hemisphere resid cv Brain Cerebellar Hemisphere resid cv Brain Cerebellar Hemisphere resid cv Small Intestine Terminal Ileum resid cv Colon Transverse resid cv Stomach resid cv Adipose Visceral Omentum resid cv Heart Left Ventricle resid cv Pancreas resid cv Muscle Skeletal resid cv Vagina resid cv Skin Sun Exposed Lower leg resid cv Skin Not Sun Exposed Suprapubic resid cv Spleen resid cv Uterus resid cv Artery Coronary resid cv Artery Aorta resid cv Artery Aorta resid cv Esophaguis Mucculovic control of the sid cv Spleen resid cv Artery Aorta resid cv Artery Aorta resid cv Artery Aorta resid cv Mean expression level Brain Nucleus accumbens Brain Nucleus accumbens Brain Putamen basal gang Brain Caudate basal gang Brain Caudate basal gang Brain Cortex median Brain Cortex median Brain Cortex lagulate co Brain Cerebellar Hemisph Muscle Skeletal median Hypothalamus median Putamen basal ganglia median Caudate basal ganglia median Hippocampus median Contex median Intestine Ξ Aorta Fibial ift Ventricle median rial Appendage median Visceral Omentum median Subcutaneous median idian oronary median orta median ammary Tissue median erior cingulate cortex BA24 median ntal Cortex BA9 median ebellum median ebellar Hemisphere median keletal median l median Muscularis median Gastroesophageal Junction median oid median resid cv posed Lov d median osed Lower leg median Exposed Suprapubic me ucosa median rse median Terminal lleum median ens basal ganglia median median resid cv

lestis

median

transformed lymphocytes median od median

(a-c) Distribution of promoter shape index and width by gene. (a) Histogram of genes' promoter shape index in *Drosophila* embryos (data from (Schor *et al*, 2017)). (b) Histogram of genes' promoter shape index in human lung tissue (Fantom5). (c) Histogram of genes' promoter width in human lung tissue (Fantom5). Density lines in a-c show fit of mixture distributions, vertical line indicates threshold for separating broad and narrow promoters (Methods). (d) Heatmap showing correlations (Spearman correlation coefficient) between promoter widths in different human tissues. Label names contain tissue names according to Fantom5 project.







width\_nucleus\_accumbens\_adult\_pool1

d



width\_medulla\_oblongata\_adult\_pool1 width\_lung\_adult\_pool1 width\_liver\_adult\_pool1 width\_kidney\_adult\_pool1 width\_insula\_adult\_pool1 width\_frontal\_lobe\_adult\_pool1 width\_esophagus\_adult\_pool1 width\_corpus\_callosum\_adult\_pool1 width\_colon\_adult\_pool1 width\_cervix\_adult\_pool1 width\_brain\_adult\_pool1 width\_blood\_adult\_pool1 width\_bladder\_adult\_pool1 width\_trachea\_adult\_pool1 width\_tonsil\_adult\_pool1 width\_thyroid\_adult\_pool1 width\_thymus\_adult\_pool1 width\_testis\_adult\_pool1 width\_temporal\_lobe\_adult\_pool1 width\_small\_intestine\_adult\_pool1 width\_skeletal\_muscle\_adult\_pool1 width\_retina\_adult\_pool1 width\_prostate\_adult\_pool1 width\_postcentral\_gyrus\_adult\_pool1 width\_pons\_adult\_pool1 width\_placenta\_adult\_pool1 width\_parietal\_lobe\_adult\_pool1 width\_paracentral\_gyrus\_adult\_pool1 width\_ovary\_adult\_pool1 width\_occipital\_pole\_adult\_pool1 width-ocipital pole adult pool on

(a) Random forest performance (R^2) for predicting expression variation in lung dataset using different subsets of samples (using samples metadata from GTEx website). Data is presented as mean ± SD (5-fold cross-validation). (b) Spearman correlation coefficient between mean expression variation and TF features dependent on the width of the TSS-proximal region used to associate TFs with genes. Three intervals were considered: -/+500 bp around TSS (used in the main analysis for TFs and chromatin states), -300/+200 (used for some core promoter features, e.g. promoter shape and TATA-box), and -/+2 kB. Correlations are reposted for total number of TFs in the TSS-proximal region and top-10 important TFs for predicting mean expression variation in the main model (based on Boruta feature selection). (c) DE prior of specific genes groups (GWAS hits, essential genes, drug targets) compared to the distribution of DE prior for all genes in the dataset. Cohen's d: 0.22, 0.52, and 0.57 for GWAS catalog, Drug targets, and Essential genes respectively (comparison to All genes). P-values < 2e-16 for all comparisons to All genes (Wilcoxon rank test).

