

## Additional File 1

### **Removal of *N*-linked glycans in cellobiohydrolase Cel7A from *Trichoderma reesei* reveals higher activity and binding affinity on crystalline cellulose**

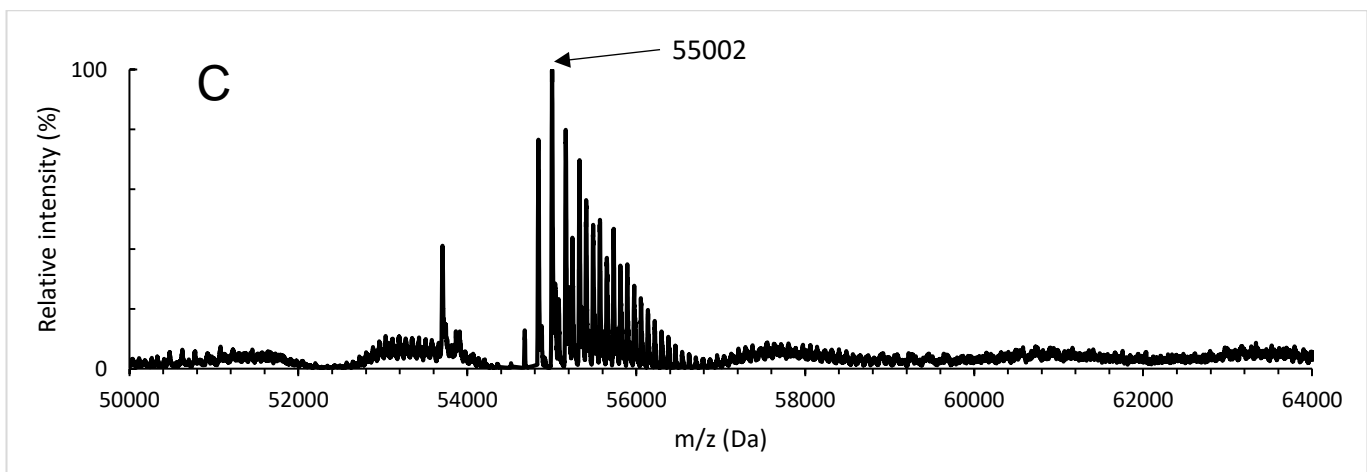
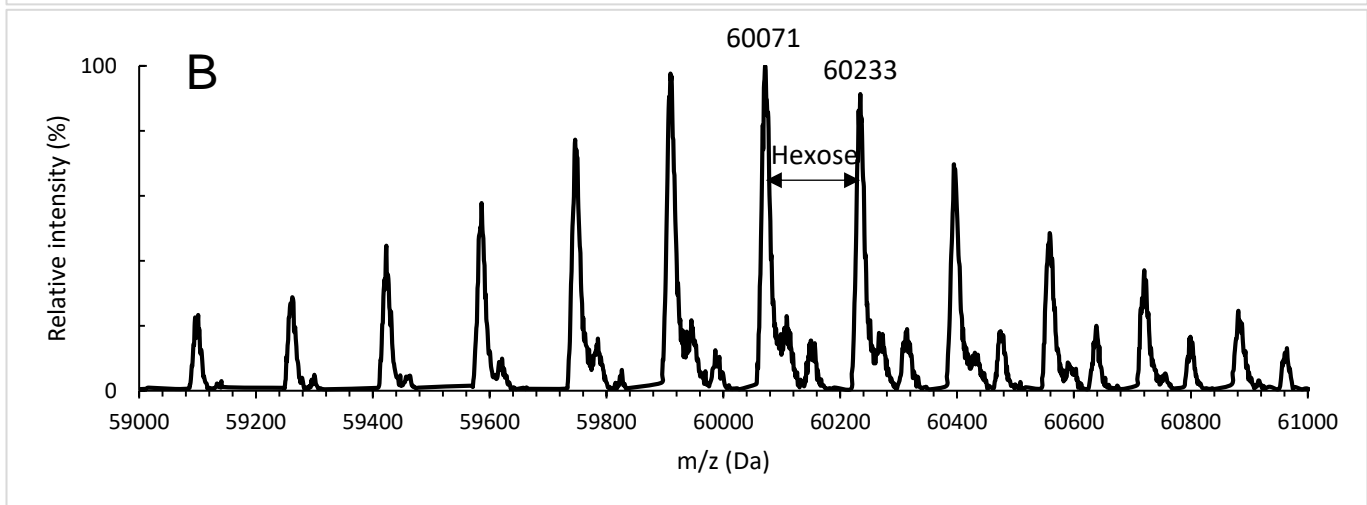
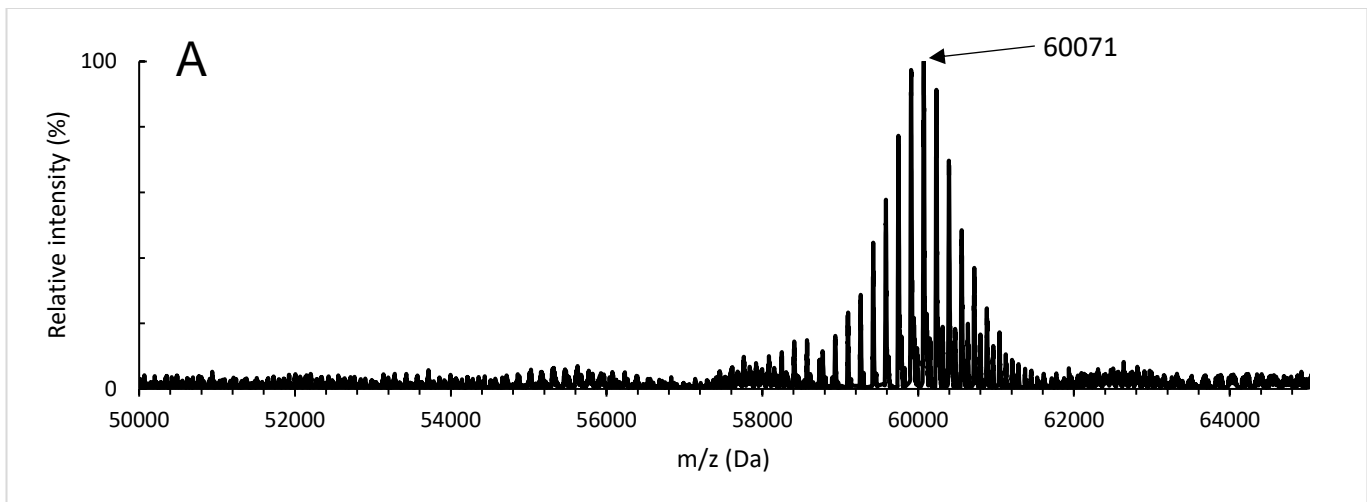
Bartłomiej M. Kołaczkowski<sup>1</sup>, Kay S. Schaller<sup>2</sup>, Trine Holst Sørensen<sup>3</sup>, Günther H. J. Peters<sup>4</sup>, Kenneth Jensen<sup>3</sup>, Kristian B. R. M. Krogh<sup>3</sup>, Peter Westh<sup>2\*</sup>

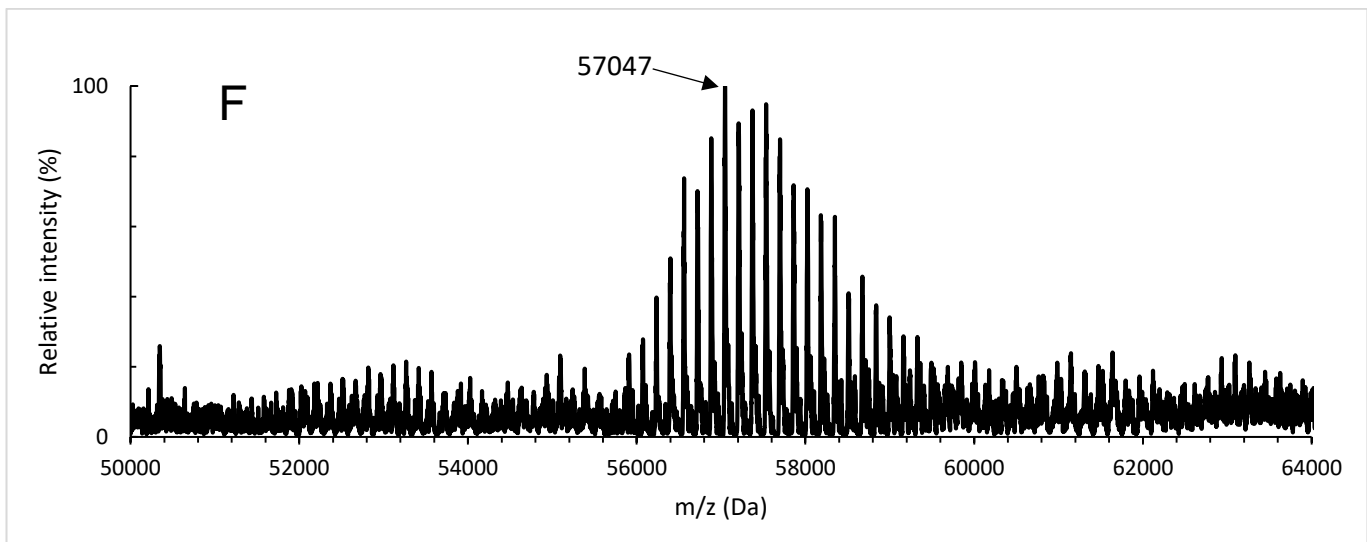
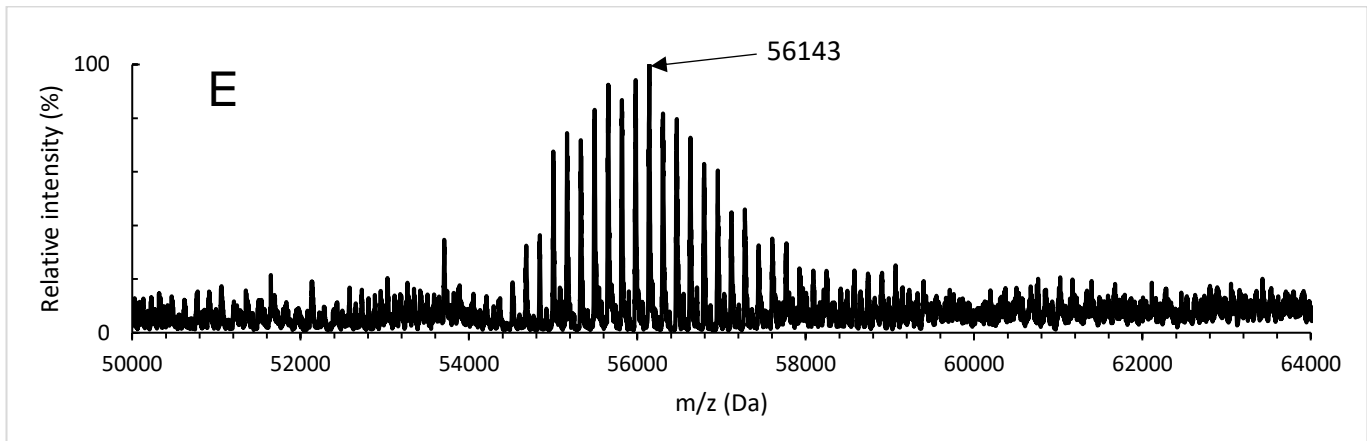
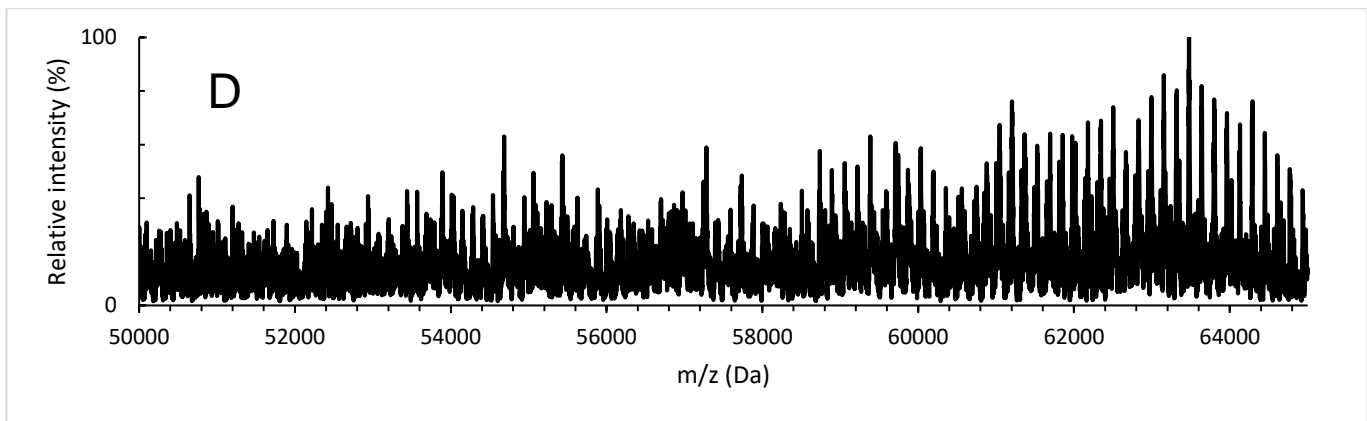
<sup>1</sup>Roskilde University, INM, Universitetsvej 1, Building 28, DK-4000, Roskilde, Denmark

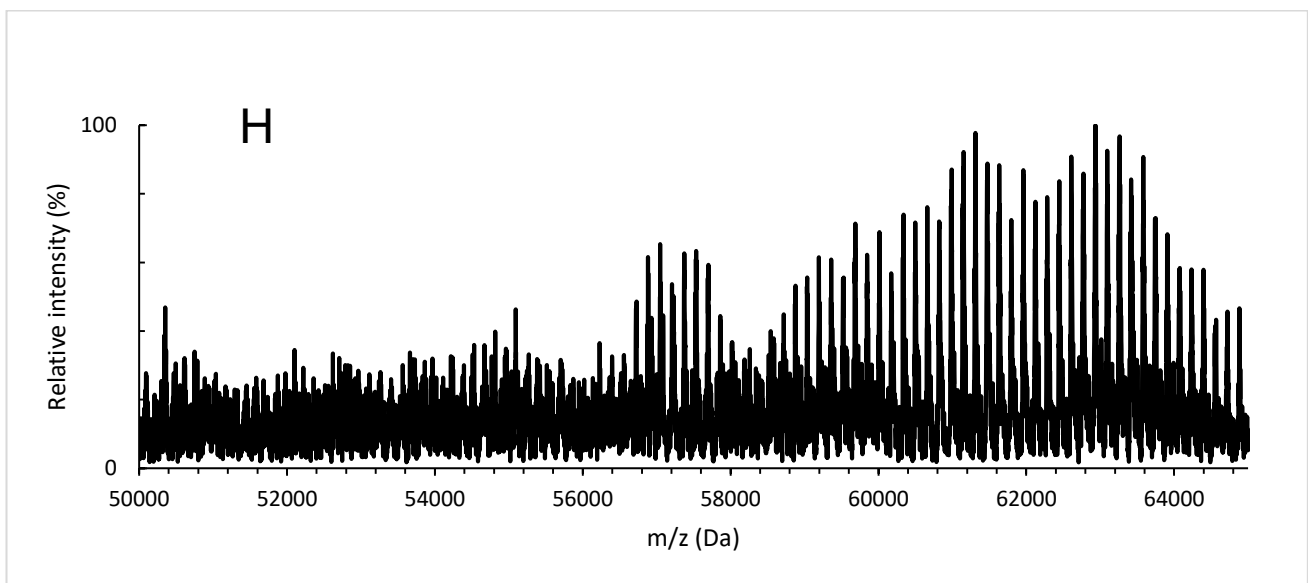
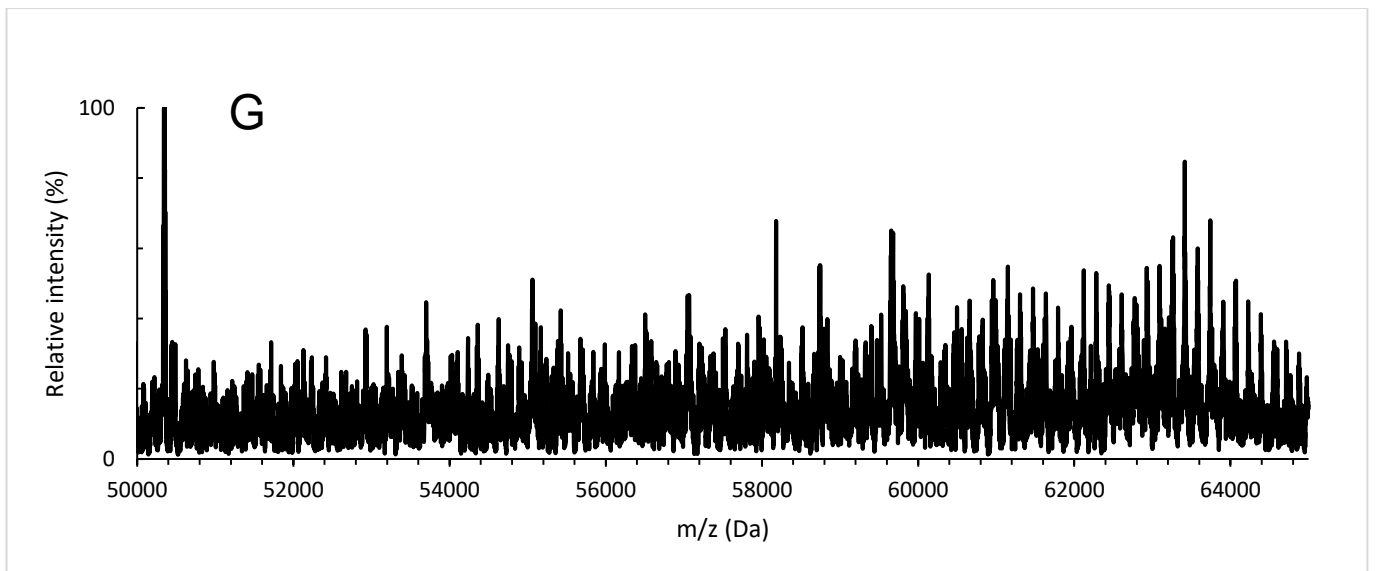
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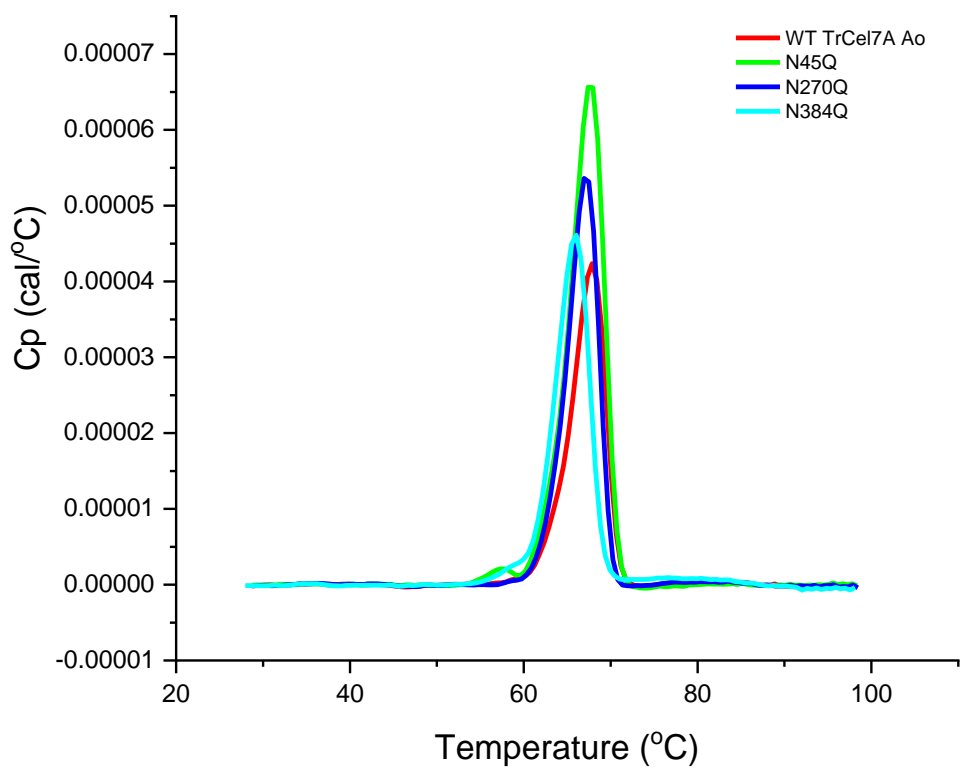
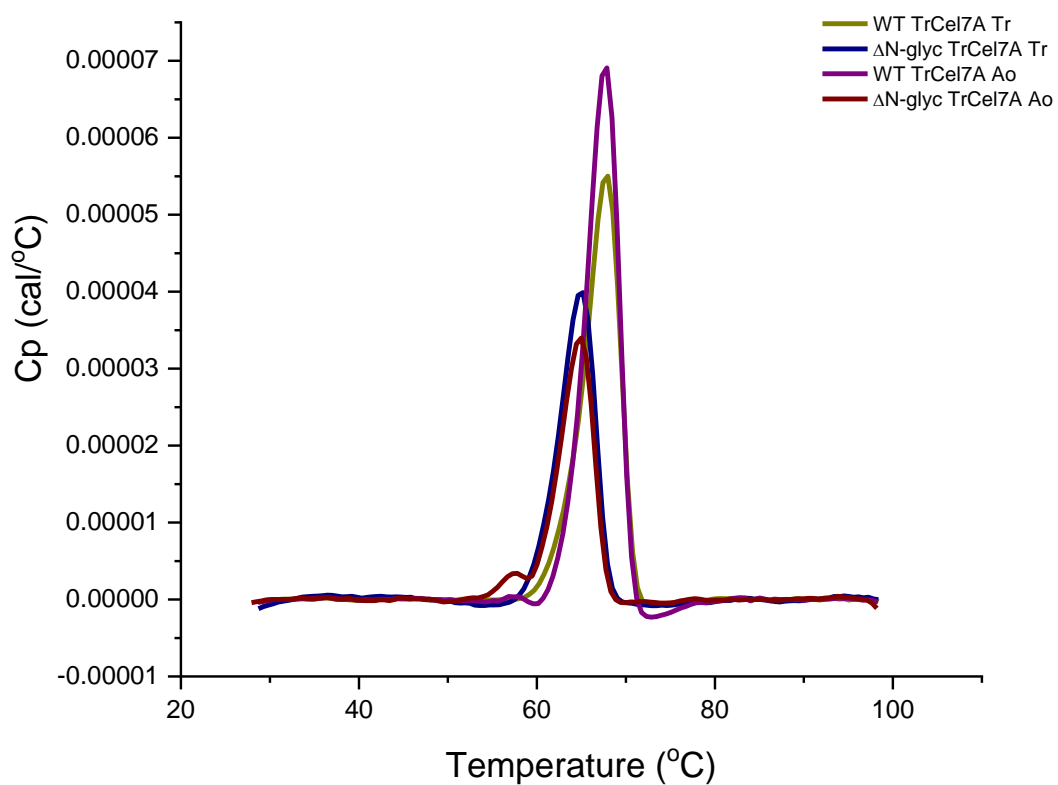
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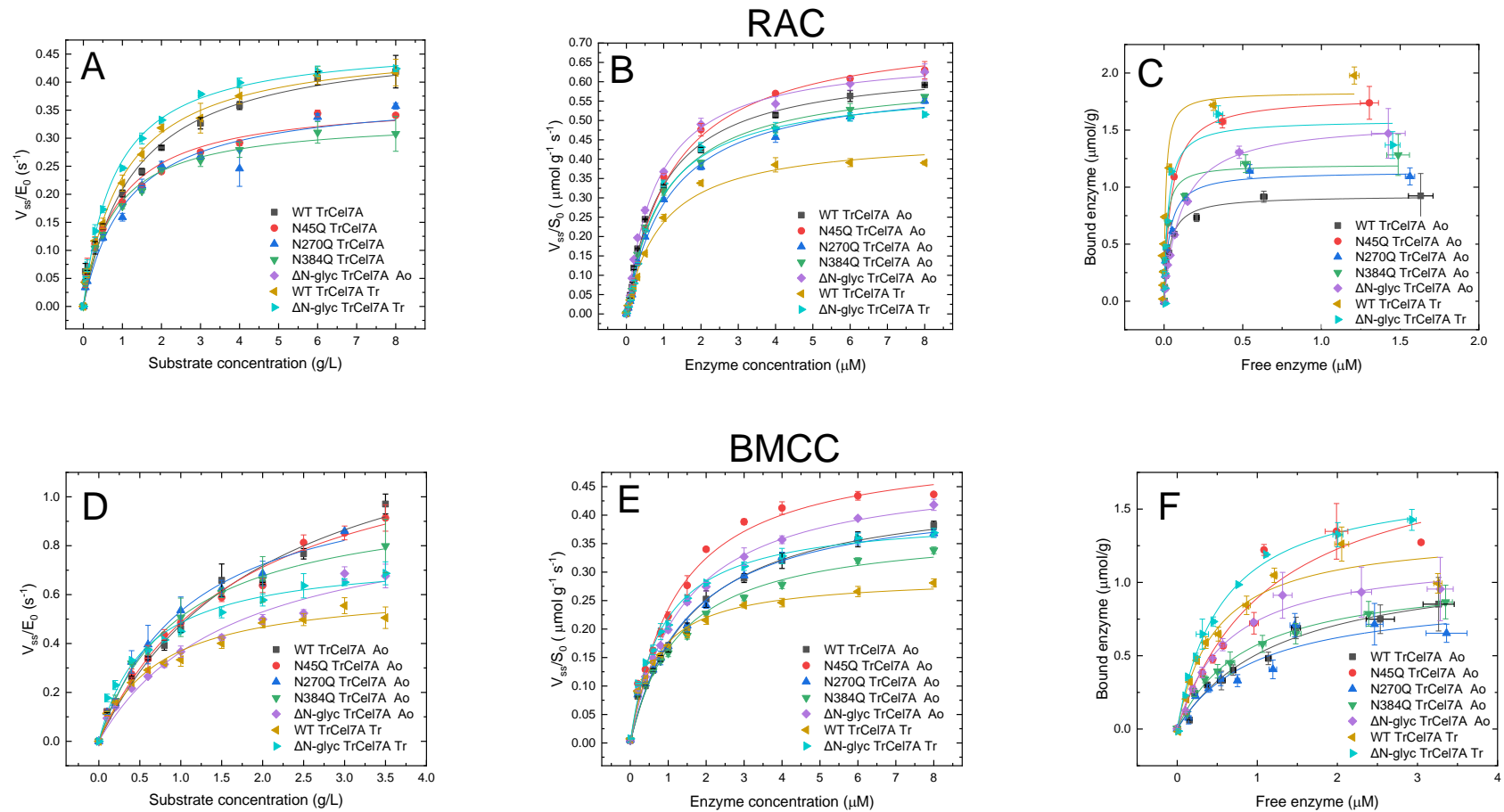
**Figure S1.** Mass distribution profile of full length TrCel7A enzymes **A)** WT expressed in *T. reesei*, **B)** the zoom of mass spectrum shown in A); the spacing between peaks corresponds to the mass difference of hexose (162 Da) **C)**  $\Delta$ N-glyc variant expressed in *T. reesei*, **D)** WT expressed in *A. oryzae*, **E)**  $\Delta$ N-glyc variant expressed in *A. oryzae*, **F)** N45Q variant expressed in *A. oryzae* **G)** N270Q variant expressed in *A. oryzae* and **H)** N384Q variant expressed in *A. oryzae*.



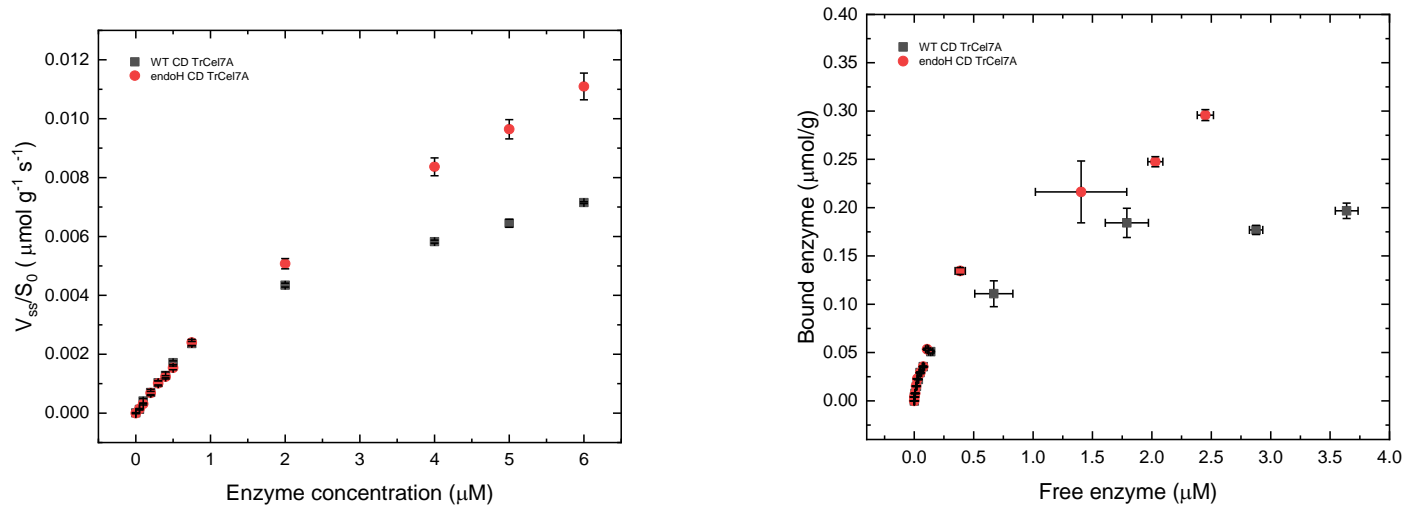
**Figure S2.** DSC scans of all WT TrCel7A and TrCel7A variants used in this work.

**Table S1.** Steady-state kinetic and binding affinity parameters (25°C) of *Tr*Cel7A WT and the variants with modified *N*-glycosylation pattern. The parameters were derived from convMM, invMM and binding isotherm using Avicel, RAC and BMCC. The  $\pm$  values correspond to the error of non-linear fit of Michaelis-Menten curves and binding isotherm curves. The parameters statistically different from the others at the 0.05 level of significance are indicated with letter 'a' (Table S3).

Avicel								
Enzyme	convMM			invMM		Kinetic substrate accesability $\Gamma_{\text{attack}}$ ( $\mu\text{mol/g}$ )	Adsorption isotherms	
	$\text{conv}V_{\text{max}}/E_0$ ( $\text{s}^{-1}$ )	$\text{conv}K_M$ ( $\text{g L}^{-1}$ )	$\eta$ ( $\text{Lg}^{-1}\text{s}^{-1}$ )	$\text{inv}V_{\text{max}}/S_0$ ( $\mu\text{mol g}^{-1}\text{s}^{-1}$ )	$\text{inv}K_M$ ( $\mu\text{M}$ )		$\Gamma_{\text{max}}$ ( $\mu\text{mol/g}$ )	$K_d$ ( $\mu\text{M}$ )
WT <sub>Ao</sub>	0.14 $\pm$ 0.00 <sup>a</sup>	4.2 $\pm$ 0.5	0.034	0.021 $\pm$ 0.001 <sup>a</sup>	4.5 $\pm$ 0.3 <sup>a</sup>	0.15 $\pm$ 0.01	0.20 $\pm$ 0.01	0.36 $\pm$ 0.06
N45Q <sub>Ao</sub>	0.12 $\pm$ 0.00	3.7 $\pm$ 0.6	0.032	0.028 $\pm$ 0.001 <sup>a</sup>	5.8 $\pm$ 0.3 <sup>a</sup>	0.23 $\pm$ 0.01	0.29 $\pm$ 0.01 <sup>a</sup>	0.3 $\pm$ 0.03
N270Q <sub>Ao</sub>	0.13 $\pm$ 0.00	4.6 $\pm$ 0.4	0.027	0.018 $\pm$ 0.001 <sup>a</sup>	4 $\pm$ 0.3 <sup>a</sup>	0.14 $\pm$ 0.01	0.24 $\pm$ 0 <sup>a</sup>	0.53 $\pm$ 0.04 <sup>a</sup>
N384Q <sub>Ao</sub>	0.14 $\pm$ 0.00 <sup>a</sup>	4.2 $\pm$ 0.6	0.034	0.018 $\pm$ 0 <sup>a</sup>	3.4 $\pm$ 0.2	0.13 $\pm$ 0.01	0.20 $\pm$ 0.01	0.25 $\pm$ 0.03
$\Delta N$ -gly <sub>CAo</sub>	0.10 $\pm$ 0.00 <sup>a</sup>	3 $\pm$ 0.6	0.032	0.025 $\pm$ 0 <sup>a</sup>	6.1 $\pm$ 0.2 <sup>a</sup>	0.25 $\pm$ 0.01	0.40 $\pm$ 0.01 <sup>a</sup>	1 $\pm$ 0.07 <sup>a</sup>
WT <sub>Tr</sub>	0.15 $\pm$ 0.01 <sup>a</sup>	5 $\pm$ 0.8	0.029	0.008 $\pm$ 0 <sup>a</sup>	1.7 $\pm$ 0.1 <sup>a</sup>	0.05 $\pm$ 0.01	0.15 $\pm$ 0.01 <sup>a</sup>	0.13 $\pm$ 0.03 <sup>a</sup>
$\Delta N$ -gly <sub>CTr</sub>	0.13 $\pm$ 0.01	3.3 $\pm$ 0.7	0.040	0.012 $\pm$ 0 <sup>a</sup>	2.8 $\pm$ 0.1	0.09 $\pm$ 0.01	0.17 $\pm$ 0.01 <sup>a</sup>	0.16 $\pm$ 0.04
RAC								
Enzyme	convMM			invMM		Kinetic substrate accesability $\Gamma_{\text{attack}}$ ( $\mu\text{mol/g}$ )	Adsorption isotherms	
	$\text{conv}V_{\text{max}}/E_0$ ( $\text{s}^{-1}$ )	$\text{conv}K_M$ ( $\text{g L}^{-1}$ )	$\eta$ ( $\text{Lg}^{-1}\text{s}^{-1}$ )	$\text{inv}V_{\text{max}}/S_0$ ( $\mu\text{mol g}^{-1}\text{s}^{-1}$ )	$\text{inv}K_M$ ( $\mu\text{M}$ )		$\Gamma_{\text{max}}$ ( $\mu\text{mol/g}$ )	$K_d$ ( $\mu\text{M}$ )
WT <sub>Ao</sub>	0.11 $\pm$ 0.01	0.32 $\pm$ 0.11	0.35	0.44 $\pm$ 0.01 <sup>a</sup>	2.1 $\pm$ 0.1	3.9 $\pm$ 0.3	3.3 $\pm$ 0.1 <sup>a</sup>	0.11 $\pm$ 0.01
N45Q <sub>Ao</sub>	0.09 $\pm$ 0.00	0.14 $\pm$ 0.03	0.64	0.43 $\pm$ 0.02 <sup>a</sup>	2.6 $\pm$ 0.3	4.7 $\pm$ 0.3	5.6 $\pm$ 0.2	0.13 $\pm$ 0.02
N270Q <sub>Ao</sub>	0.10 $\pm$ 0.00	0.15 $\pm$ 0.02	0.66	0.37 $\pm$ 0.01	2.2 $\pm$ 0.1	3.6 $\pm$ 0.1	4.8 $\pm$ 0.1	0.2 $\pm$ 0.03
N384Q <sub>Ao</sub>	0.10 $\pm$ 0.00	0.2 $\pm$ 0.04	0.49	0.37 $\pm$ 0.01	2.4 $\pm$ 0.2	3.8 $\pm$ 0.2	4.3 $\pm$ 0.2 <sup>a</sup>	0.07 $\pm$ 0.02
$\Delta N$ -gly <sub>CAo</sub>	0.12 $\pm$ 0.00 <sup>a</sup>	0.29 $\pm$ 0.04	0.41	nd	nd	nd	nd	nd
WT <sub>Tr</sub>	0.11 $\pm$ 0.00	0.23 $\pm$ 0.04	0.47	0.23 $\pm$ 0.01 <sup>a</sup>	1.6 $\pm$ 0.1 <sup>a</sup>	2.1 $\pm$ 0.1	6.3 $\pm$ 0.3 <sup>a</sup>	0.11 $\pm$ 0.02
$\Delta N$ -gly <sub>CTr</sub>	0.10 $\pm$ 0.01	0.25 $\pm$ 0.07	0.39	0.32 $\pm$ 0.01 <sup>a</sup>	2.1 $\pm$ 0.2	3.3 $\pm$ 0.2	7.1 $\pm$ 0.2 <sup>a</sup>	0.1 $\pm$ 0.02
BMCC								
Enzyme	convMM			invMM		Kinetic substrate accesability $\Gamma_{\text{attack}}$ ( $\mu\text{mol/g}$ )	Adsorption isotherms	
	$\text{conv}V_{\text{max}}/E_0$ ( $\text{s}^{-1}$ )	$\text{conv}K_M$ ( $\text{g L}^{-1}$ )	$\eta$ ( $\text{Lg}^{-1}\text{s}^{-1}$ )	$\text{inv}V_{\text{max}}/S_0$ ( $\mu\text{mol g}^{-1}\text{s}^{-1}$ )	$\text{inv}K_M$ ( $\mu\text{M}$ )		$\Gamma_{\text{max}}$ ( $\mu\text{mol/g}$ )	$K_d$ ( $\mu\text{M}$ )
WT <sub>Ao</sub>	0.35 $\pm$ 0.02	0.69 $\pm$ 0.09	0.52	0.23 $\pm$ 0.01 <sup>a</sup>	2.7 $\pm$ 0.2 <sup>a</sup>	0.64 $\pm$ 0.04	3.9 $\pm$ 0.2 <sup>a</sup>	1.24 $\pm$ 0.12 <sup>a</sup>
N45Q <sub>Ao</sub>	0.31 $\pm$ 0.01	0.53 $\pm$ 0.07	0.59	0.30 $\pm$ 0.01 <sup>a</sup>	2.9 $\pm$ 0.2 <sup>a</sup>	0.96 $\pm$ 0.04	4.0 $\pm$ 0.2 <sup>a</sup>	0.74 $\pm$ 0.09
N270Q <sub>Ao</sub>	0.35 $\pm$ 0.02	0.52 $\pm$ 0.09	0.67	0.19 $\pm$ 0.01	1.1 $\pm$ 0.1	0.55 $\pm$ 0.04	3.0 $\pm$ 0.2	1.27 $\pm$ 0.2 <sup>a</sup>
N384Q <sub>Ao</sub>	0.38 $\pm$ 0.02	0.7 $\pm$ 0.13	0.54	0.20 $\pm$ 0.01	1.9 $\pm$ 0.2 <sup>a</sup>	0.52 $\pm$ 0.04	4.5 $\pm$ 0.3 <sup>a</sup>	1.09 $\pm$ 0.18 <sup>a</sup>
$\Delta N$ -gly <sub>CAo</sub>	0.34 $\pm$ 0.02	0.43 $\pm$ 0.07	0.79	0.28 $\pm$ 0.01 <sup>a</sup>	1.8 $\pm$ 0.2 <sup>a</sup>	0.82 $\pm$ 0.05	4.4 $\pm$ 0.4 <sup>a</sup>	1.88 $\pm$ 0.42 <sup>a</sup>
WT <sub>Tr</sub>	0.36 $\pm$ 0.02	0.49 $\pm$ 0.07	0.73	0.14 $\pm$ 0 <sup>a</sup>	0.6 $\pm$ 0.1 <sup>a</sup>	0.40 $\pm$ 0.02	2.8 $\pm$ 0.2	0.47 $\pm$ 0.1
$\Delta N$ -gly <sub>CTr</sub>	0.38 $\pm$ 0.02	0.41 $\pm$ 0.07	0.91	0.20 $\pm$ 0	0.9 $\pm$ 0.1	0.52 $\pm$ 0.03	2.4 $\pm$ 0.1 <sup>a</sup>	0.33 $\pm$ 0.05 <sup>a</sup>

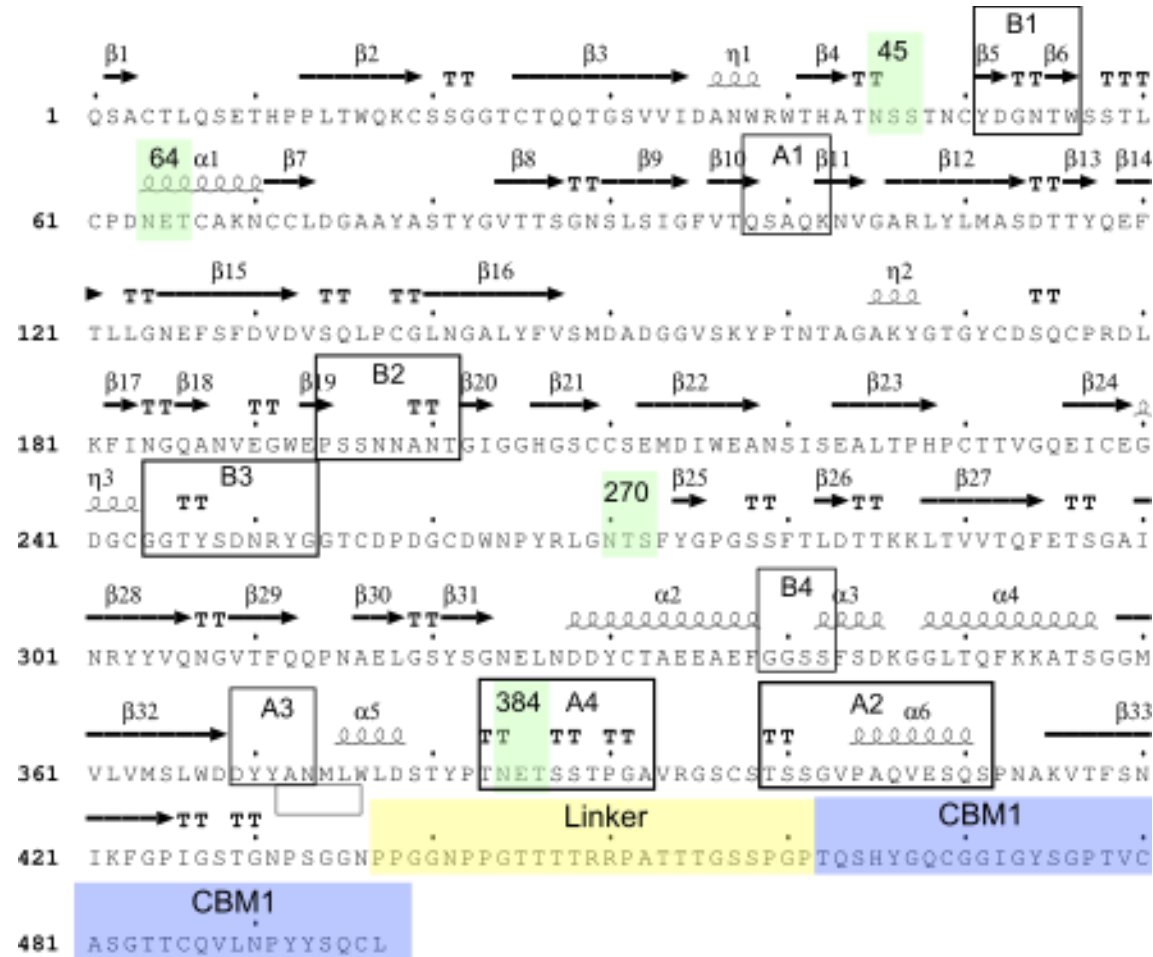


**Figure S3.** Steady-state kinetic analysis and binding isotherm for *TrCel7A* enzymes on RAC and BMCC at 50°C. A) C) In the <sup>conv</sup>MM analysis, low enzyme concentration of 50 nM is saturated with increasing RAC concentration (0-8 g/L) and BMCC concentration (0-3.5 g/L), respectively. B) E) In the <sup>inv</sup>MM approach, low substrate concentration of 0.4 g/L RAC and 0.75 g/L BMCC, respectively, is saturated with enzyme concentration. The lines indicate a non-linear fit from equation (1) and (2). C) F) Binding isotherm of *TrCel7A* wt and variants on 0.4 g/L RAC and 0.75 g/L BMCC, respectively, at 50°C. The lines show the fitted Langmuir equation (see equation 4). Error bars represent the standard deviations from a triplicate measurement. Error bars represent standard deviations from triplicate measurements.

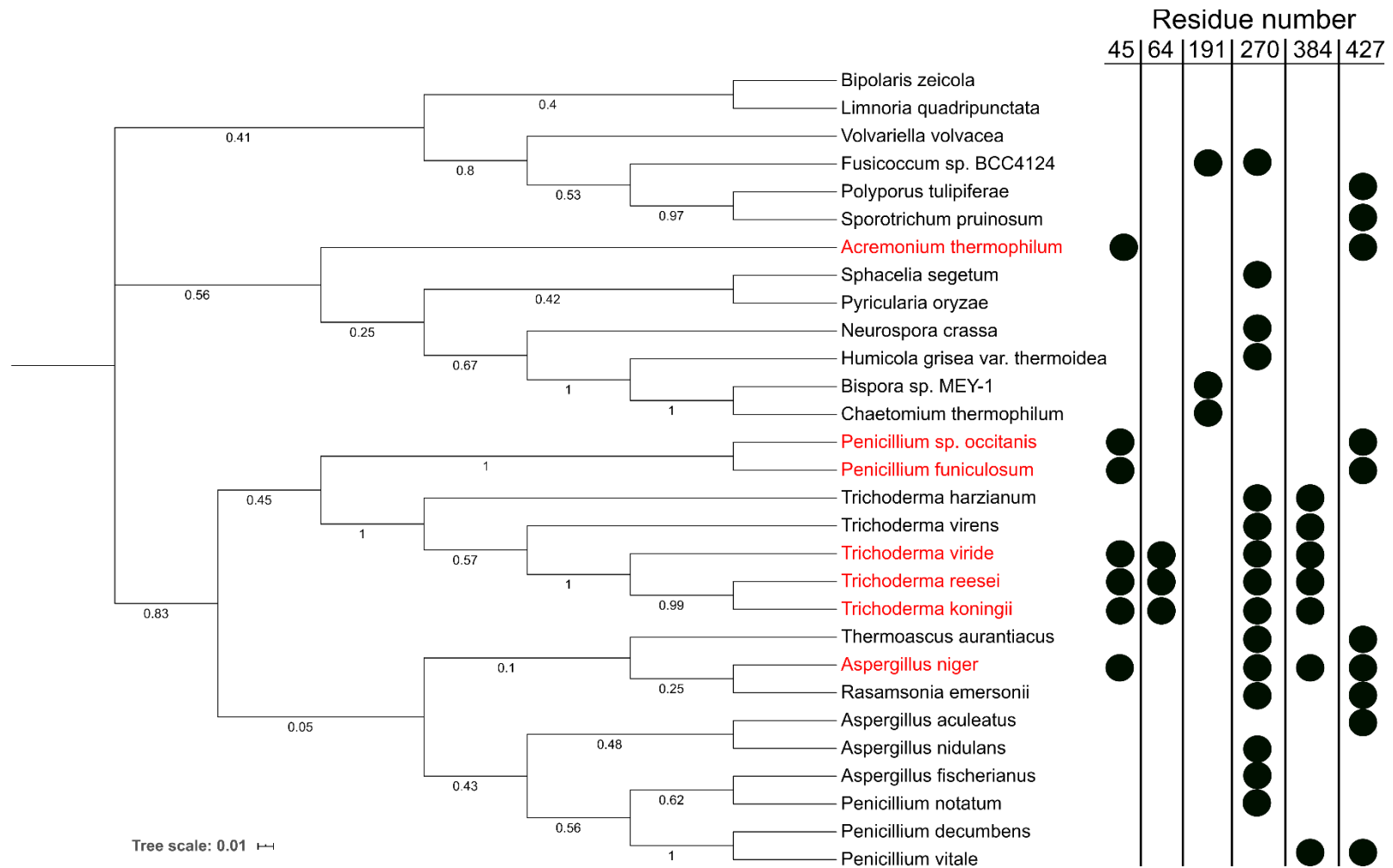


**Figure S4.**  $\text{InvMM}$  analysis (left) and binding isotherm (right) of TrCel7A CD and endoH treated CD TrCel7A on 12 g/L Avicel. The results show increased activity and binding upon reducing content of *N*-glycosylation. The enzyme was expressed in *A. oryzae*. Error bars represent the standard deviations from triplicate measurements.





**Figure S5.** Architecture-based amino acid sequence of the *Trichoderma reesei* Cel7A. Linker and CBM are indicated with yellow and blue color. Experimentally determined secondary structure elements of the catalytic domain are shown on top of the sequence with the following figures and symbols: alpha helices (helices,  $\alpha$  and  $\eta$  for a 310-helix), beta strands (arrows,  $\beta$ ), turns (T, strict  $\beta$ -turns as TT and strict  $\alpha$ -turns as TTT). Green frames indicate all *TrCel7A* N-glycosylation motifs with the consensus motif (N-X-S/T). Black frames indicate loops characterized by Momeni *et al.* (2013) [1]. The secondary structure annotation were added using ESPript 3.0 web server with default parameters [2].



**Figure S6.** Phylogenetic tree of the characterized Cel7A sequences from GH7. In total 29 sequences were extracted from the CAZy database[3] and used for the multiple sequence alignment, using TrCel7A as the reference sequence. The most common N-glycosylation motifs were annotated in the separate columns and their presence were indicated by black circle. The red colored organisms indicate presence of N-glycosylation site at N45.

**Table S2.** Statistical comparison by analysis of means of the parameters derived from non-linear fit of Michaelis-Menten and binding isotherms curves (50 °C). The parameters that exceeded upper or lower limit are statistically different from the others at the 0.05 level of significance.

convMM					invMM								Adsorption isotherms											
convV <sub>max</sub>					convK <sub>M</sub>				invV <sub>max</sub>				invK <sub>M</sub>				Γ <sub>max</sub>				K <sub>d</sub>			
Avicel																								
Level	Lower Limit	Estimate	Upper Limit	Limit Exceeded	Lower Limit	Estimate	Upper Limit	Limit Exceeded	Lower Limit	Estimate	Upper Limit	Limit Exceeded	Lower Limit	Estimate	Upper Limit	Limit Exceeded	Lower Limit	Estimate	Upper Limit	Limit Exceeded	Lower Limit	Estimate	Upper Limit	Limit Exceeded
N45Q <sub>AO</sub>	0.27	0.34	0.36		4.2	11.6	13.7		0.028	0.048	0.034	Upper	1.51	2.92	2.46	Upper	0.08	0.23	0.16	Upper	0.00	0.69	0.81	
N270Q <sub>AO</sub>	0.27	0.35	0.36		4.2	11.6	13.7		0.029	0.031	0.034		1.51	2.22	2.46		0.08	0.13	0.17		-0.60	1.00	1.41	
N384Q <sub>AO</sub>	0.27	0.35	0.36		4.8	10.6	13.1		0.028	0.037	0.034	Upper	1.46	2.54	2.52	Upper	0.09	0.10	0.15		0.03	0.30	0.78	
WT <sub>AO</sub>	0.25	0.40	0.38	Upper	1.4	18.1	16.5	Upper	0.029	0.034	0.034		1.47	2.45	2.50		0.10	0.10	0.15		0.14	0.26	0.67	
WT <sub>Tr</sub>	0.28	0.32	0.36		5.3	9.0	12.6		0.030	0.020	0.033	Lower	1.70	1.16	2.28	Lower	0.10	0.08	0.15	Lower	0.08	0.27	0.73	
ΔN-glyc <sub>AO</sub>	0.28	0.24	0.35	Lower	5.9	6.2	12.0		0.027	0.048	0.035	Upper	1.33	3.54	2.64	Upper	0.07	0.27	0.17	Upper	-0.18	1.00	0.99	Upper
ΔN-glyc <sub>Tr</sub>	0.28	0.32	0.35		6.1	7.4	11.8		0.029	0.032	0.033		1.67	1.71	2.30		0.09	0.11	0.15		-0.01	0.40	0.82	
RAC																								
N45Q <sub>AO</sub>	0.35	0.37	0.42		0.5	0.9	1.1		0.57	0.74	0.66	Upper	0.73	1.23	1.21	Upper	1.16	1.79	1.61	Upper	0.00	0.05	0.04	Upper
N270Q <sub>AO</sub>	0.34	0.38	0.43		0.4	1.1	1.3		0.57	0.61	0.66		0.70	1.15	1.23		1.20	1.14	1.58	Lower	0.00	0.03	0.04	
N384Q <sub>AO</sub>	0.35	0.34	0.42	Lower	0.5	0.8	1.1		0.58	0.62	0.66		0.74	1.03	1.19		1.21	1.20	1.56	Lower	0.01	0.02	0.03	
WT <sub>AO</sub>	0.34	0.48	0.43	Upper	0.4	1.3	1.2	Upper	0.58	0.65	0.66		0.77	0.95	1.16		1.21	0.92	1.57	Lower	-0.01	0.03	0.04	
WT <sub>Tr</sub>	0.34	0.47	0.43	Upper	0.5	1.0	1.1		0.58	0.46	0.66	Lower	0.70	0.94	1.24		1.20	1.83	1.58	Upper	0.01	0.01	0.02	Lower
ΔN-glyc <sub>AO</sub>	0.36	0.30	0.42	Lower	0.6	0.5	1.0	Lower	0.58	0.68	0.65	Upper	0.81	0.82	1.12		1.11	1.58	1.66		-0.04	0.11	0.08	Upper
ΔN-glyc <sub>Tr</sub>	0.35	0.47	0.42	Upper	0.6	0.8	1.1		0.58	0.59	0.66		0.76	0.93	1.17		1.19	1.56	1.59		0.01	0.02	0.03	
BMCC																								
N45Q <sub>AO</sub>	0.63	1.33	1.07	Upper	0.3	1.7	1.5	Upper	0.37	0.52	0.44	Upper	0.81	1.23	1.30		0.82	1.96	1.84	Upper	-0.05	1.20	1.26	
N270Q <sub>AO</sub>	0.65	1.13	1.05	Upper	0.4	1.1	1.3		0.37	0.44	0.45		0.68	1.49	1.43	Upper	0.95	0.93	1.71	Lower	-0.37	0.99	1.59	
N384Q <sub>AO</sub>	0.68	1.01	1.01		0.5	1.0	1.3		0.37	0.38	0.44		0.70	1.29	1.41		0.99	1.06	1.67		-0.08	0.85	1.30	
WT <sub>AO</sub>	0.55	1.46	1.14	Upper	0.1	2.1	1.7	Upper	0.36	0.45	0.45	Upper	0.66	1.59	1.45	Upper	0.85	1.17	1.81		-0.56	1.28	1.77	
WT <sub>Tr</sub>	0.75	0.63	0.95	Lower	0.5	0.7	1.2		0.38	0.29	0.43	Lower	0.84	0.66	1.27	Lower	1.08	1.34	1.58		0.35	0.48	0.86	
ΔN-glyc <sub>AO</sub>	0.64	0.96	1.05		0.1	1.6	1.6	Upper	0.37	0.48	0.44	Upper	0.75	1.38	1.36	Upper	1.05	1.20	1.61		0.21	0.62	1.01	
ΔN-glyc <sub>Tr</sub>	0.76	0.76	0.93		0.7	0.6	1.1	Lower	0.38	0.40	0.43		0.86	0.82	1.25	Lower	1.05	1.71	1.61	Upper	0.36	0.55	0.85	

**Table S3.** Statistical comparison by analysis of means of the parameters derived from non-linear fit of Michaelis-Menten and binding isotherms curves (25 °C). The parameters that exceeded upper or lower limit are statistically different from the others at the 0.05 level of significance.

convMM					invMM								Adsorption isotherms											
convV <sub>max</sub>					convK <sub>M</sub>				invV <sub>max</sub>				invK <sub>M</sub>				Γ <sub>max</sub>				K <sub>d</sub>			
Avicel																								
Level	Lower Limit	Estimate	Upper Limit	Limit Exceeded	Lower Limit	Estimate	Upper Limit	Limit Exceeded	Lower Limit	Estimate	Upper Limit	Limit Exceeded	Lower Limit	Estimate	Upper Limit	Limit Exceeded	Lower Limit	Estimate	Upper Limit	Limit Exceeded	Lower Limit	Estimate	Upper Limit	Limit Exceeded
N45Q <sub>A0</sub>	0.12	0.12	0.14		2.2	3.7	5.6		0.012	0.028	0.015	Upper	2.5	5.8	3.9	Upper	0.19	0.29	0.23	Upper	0.12	0.30	0.32	
N270Q <sub>A0</sub>	0.12	0.13	0.14		1.8	4.6	6.0		0.012	0.018	0.015	Upper	2.6	4.0	3.8	Upper	0.18	0.24	0.23	Upper	0.01	0.53	0.44	Upper
N384Q <sub>A0</sub>	0.12	0.14	0.14	Upper	2.2	4.2	5.5		0.012	0.018	0.014	Upper	2.8	3.4	3.6		0.19	0.20	0.23		0.12	0.25	0.33	
WT <sub>A0</sub>	0.12	0.14	0.14	Upper	2.2	4.2	5.5		0.012	0.021	0.015	Upper	2.6	4.5	3.8	Upper	0.19	0.20	0.23		0.07	0.36	0.38	
WT <sub>Tr</sub>	0.12	0.15	0.14	Upper	1.9	5.0	5.8		0.013	0.008	0.014	Lower	2.9	1.7	3.6	Lower	0.19	0.15	0.22	Lower	0.15	0.13	0.29	Lower
ΔN-glyC <sub>A0</sub>	0.12	0.10	0.14	Lower	2.2	3.0	5.6		0.012	0.025	0.015	Upper	2.3	6.1	4.1	Upper	0.17	0.40	0.24	Upper	-0.07	1.00	0.51	Upper
ΔN-glyC <sub>Tr</sub>	0.12	0.13	0.14		2.5	3.3	5.2		0.013	0.012	0.014	Lower	2.8	2.8	3.7		0.19	0.17	0.22	Lower	0.14	0.16	0.30	
RAC																								
N45Q <sub>A0</sub>	0.09	0.09	0.11		0.10	0.16	0.30		0.31	0.43	0.37	Upper	1.7	2.6	2.6		4.6	5.6	5.7		0.04	0.13	0.16	
N270Q <sub>A0</sub>	0.09	0.10	0.11		0.12	0.15	0.29		0.32	0.37	0.37		1.7	2.2	2.6		4.6	4.8	5.7		-0.01	0.20	0.21	
N384Q <sub>A0</sub>	0.09	0.10	0.11		0.09	0.20	0.32		0.32	0.37	0.37		1.7	2.3	2.6		4.7	4.3	5.6	Lower	0.06	0.07	0.14	
WT <sub>A0</sub>	0.09	0.11	0.12		0.02	0.32	0.38		0.32	0.44	0.37	Upper	1.8	2.1	2.5		4.7	3.3	5.6	Lower	0.02	0.11	0.18	
WT <sub>Tr</sub>	0.09	0.11	0.11		0.08	0.23	0.33		0.32	0.23	0.37	Lower	1.7	1.6	2.6	Lower	4.6	6.3	5.6	Upper	0.05	0.11	0.15	
ΔN-glyC <sub>A0</sub>	0.09	0.12	0.12	Upper	0.06	0.29	0.35		nd	nd	nd		nd	nd	nd		nd	nd	nd		nd	nd	nd	
ΔN-glyC <sub>Tr</sub>	0.09	0.10	0.11		0.05	0.25	0.35		0.32	0.32	0.37	Lower	1.7	2.1	2.6		4.6	7.0	5.6	Upper	0.07	0.09	0.13	
BMCC																								
N45Q <sub>A0</sub>	0.31	0.31	0.39		0.24	0.53	0.75		0.16	0.30	0.23	Upper	0.4	2.9	1.8	Upper	2.2	4.0	3.7	Upper	0.18	0.74	0.86	
N270Q <sub>A0</sub>	0.31	0.35	0.39		0.28	0.52	0.71		0.18	0.19	0.21		0.8	1.1	1.4		2.3	3.0	3.5		-0.14	1.27	1.19	Upper
N384Q <sub>A0</sub>	0.30	0.38	0.40		0.20	0.70	0.79		0.17	0.20	0.22		0.5	1.9	1.7	Upper	1.9	4.5	3.9	Upper	-0.01	1.09	1.06	Upper
WT <sub>A0</sub>	0.30	0.35	0.40		0.19	0.69	0.80		0.16	0.23	0.22	Upper	0.3	2.7	1.9	Upper	2.0	3.9	3.8	Upper	-0.15	1.24	1.19	Upper
WT <sub>Tr</sub>	0.31	0.36	0.39		0.29	0.49	0.70		0.18	0.14	0.21	Lower	0.9	0.6	1.3	Lower	2.6	2.8	3.3		0.31	0.47	0.73	
ΔN-glyC <sub>A0</sub>	0.31	0.34	0.39		0.31	0.43	0.67		0.17	0.28	0.21	Upper	0.7	1.8	1.4	Upper	2.1	4.4	3.8	Upper	-0.32	1.88	1.36	Upper
ΔN-glyC <sub>Tr</sub>	0.31	0.38	0.39		0.34	0.41	0.65		0.18	0.20	0.21		0.9	0.9	1.3		2.6	2.4	3.2	Lower	0.37	0.33	0.68	Lower

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