Supplemental Figures





Figure S1. Membrane and action potential properties of neurons expressing somatic vs. untargeted forms of GCaMP6f and GCaMP7f; related to Figure 3. Cultured hippocampal SomaGCaMP6f1, SomaGCaMP6f2, neurons expressing GCaMP6f, GCaMP7f and SomaGCaMP7f were patched and membrane properties recorded. (A-D) Passive membrane properties. (A) Resting potential (n = 6 cells from 2 cultures for GCaMP6f; n = 7 cells from 2 cultures for SomaGCaMP6f1; n = 6 cells from 2 cultures for SomaGCaMP6f2; n = 10 cells from 2 cultures for GCaMP7f; n = 12 cells from 2 cultures for SomaGCaMP7f). Plotted is mean plus or minus standard error throughout the figure. Throughout this figure: n.s., not significant; between GCaMP6f, SomaGCaMP6f, and SomaGCaMP6f2, Kruskal-Wallis analysis of variance followed by post-hoc test via Steel's test with GCaMP6f as control group; n.s., not significant; between GCaMP7f and SomaGCaMP7f, Wilcoxon rank sum test. See Supplemental Table 5 for full statistics for Figure S1. (B) Membrane capacitance (n = 5 cells from 2 cultures for GCaMP6f; n = 6 cells from 2 cultures for SomaGCaMP6f1; n = 6 cells from 2 cultures for SomaGCaMP6f2; n= 10 cells from 2 cultures for GCaMP7f; n = 12 cells from 2 cultures for SomaGCaMP7f). (C) Holding current while held at -65 mV (n = 5 cells from 2 cultures for GCaMP6f; n = 6 cells from 2 cultures for SomaGCaMP6f1; n = 6 cells from 2 cultures for SomaGCaMP6f2; n = 10 cells from 2 cultures for GCaMP7f; n = 12 cells from 2 cultures for SomaGCaMP7f). (D) Membrane resistance (n = 5 cells from 2 cultures for GCaMP6f; n = 6 cells from 2 cultures for SomaGCaMP6f1; n = 6 cells from 2 cultures for SomaGCaMP6f2; n = 10 cells from 2 cultures for GCaMP7f; n = 12 cells from 2 cultures for SomaGCaMP7f). (E-H) Action potential properties. (E) Averaged action potential waveforms (n = 12 cells from 2 cultures for GCaMP6f; n = 12 cells from 2 cultures for SomaGCaMP6f1; n = 12 cells from 2 cultures for SomaGCaMP6f2; n = 7 cells from 2 cultures for GCaMP7f; n = 7 cells from 2 cultures for SomaGCaMP7f). (F) Action potential width (n = 12 cells from 2 cultures for GCaMP6f; n = 12 cells from 2 cultures for SomaGCaMP6f1; n = 12 cells from 2 cultures for SomaGCaMP6f2; n = 7 cells from 2 cultures for GCaMP7f; n = 7 cells from 2 cultures for SomaGCaMP7f). (G) Action potential amplitude (n = 12 cells from 2 cultures for GCaMP6f; n = 12 cells from 2 cultures for SomaGCaMP6f1; n = 12cells from 2 cultures for SomaGCaMP6f2; n = 7 cells from 2 cultures for GCaMP7f; n = 7 cells from 2 cultures for SomaGCaMP7f). (H) Action potential threshold (n = 12 cells from 2 cultures for GCaMP6f; n = 12 cells from 2 cultures for SomaGCaMP6f1; n = 12 cells from 2 cultures for SomaGCaMP6f2; n = 7 cells from 2 cultures for GCaMP7f; n = 7 cells from 2 cultures for SomaGCaMP7f).



Figure S2. Distribution of ion channels and Ankyrin_G in neurons expressing GCaMP6f, SomaGCaMP6f1 or SomaGCaMP6f2; related to Figure 3. Cultured hippocampal neurons expressing GCaMP6f, SomaGCaMP6f1 and SomaGCaMP6f2 were immunostained using antibodies against ion channels or Ankyring epitopes. (A, left) Representative images (maximum intensity projections) of neurons expressing GCaMP6f, SomaGCaMP6f1 or SomaGCaMP6f2 (from top to bottom, respectively) and immunostained against Kv2.1 (blue). (A, right) Average fluorescent profiles down the axon of neurons with immunostained Kv2.1 in GCaMP6f (red), SomaGCaMP6f1 (blue) or SomaGCaMP6f2 (green) conditions (n = 6 GCaMP6f expressing neurons from 3 cultures; n = 6 SomaGCaMP6f1 expressing neurons from 2 cultures; n = 6 SomaGCaMP6f2 expressing neurons from 4 cultures). Normalized to the soma value. n.s., not significant, Bonferroni-corrected Kruskal-Wallis analysis of variance; see Supplemental Table 6 for full statistics for Figure S2. Plotted is mean (solid line) plus or minus standard error (shaded area) throughout the figure. (B) As in A, but for NaV1.2 (n = 6GCaMP6f expressing neurons from 3 cultures; n = 6 SomaGCaMP6f1 expressing neurons from 4 cultures; n = 6 SomaGCaMP6f2 expressing neurons from 2 cultures). (C) As in A, but for Ankyrin_G (n = 6 GCaMP6f expressing neurons from 4 cultures; n = 5 SomaGCaMP6f1 expressing neurons from 2 cultures; n = 6 SomaGCaMP6f2 expressing neurons from 2 cultures). (**D**) As in A, but for CaV2.1 (n = 5 GCaMP6f expressing neurons from 2 cultures; n = 5SomaGCaMP6f1 expressing neurons from 2 cultures; n = 5 SomaGCaMP6f2 expressing neurons from 5 cultures).



Figure S3. Fluorescent profiles of GCaMP6f, SomaGCaMP6f1 and SomaGCaMP6f2 in comparison to the fluorescent profile of the membrane in cell bodies of cultured neurons; related to Figure 3. (A, left) An image (single confocal slice) of a hippocampal neural cell body expressing GCaMP6f (green, left panel) and stained with the membrane labeling dye WGA-647 (magenta, middle panel). Merge of the left and middle panels is presented on the right panel. The yellow rectangle indicates the region of interest for fluorescent profile analysis throughout this figure. (A, right) Average fluorescent profiles of GCaMP6f (black) and the membrane (red) (n = 7 neurons from 2 cultures). Plotted is mean (solid line) plus or minus standard error (shaded area) throughout the figure. (B) As in A, but for SomaGCaMP6f1 (n = 4 neurons from 2 cultures). (C) As in A, but for SomaGCaMP6f2 (n = 6 neurons from 2 cultures).



Figure S4. Baseline fluorescence brightness of GCaMP6f and SomaGCaMP6f1 in living brain slices; related to Figure 4. Bars show average baseline brightness values for cells expressing GCaMP6f or SomaGCaMP6f1 in slice (n = 42 neurons from 4 slices from 2 GCaMP6f mice; n = 43 neurons from 8 slices from 3 SomaGCaMP6f1 mice). Error bars indicate standard error of the mean. ***P < 0.001, Kolmogorov-Smirnov test of baseline fluorescence brightness between GCaMP6f and SomaGCaMP6f1; see Supplemental Table 6 for full statistics for Figure S4.



Figure S5. Sensitivity of multiple action potentials, temporal dynamics and event rate for GCaMP6f and SomaGCaMP6f1; related to Figure 4. (A) A graph showing the df/f0 of the calcium transient elicited after a train of 1, 5, 10 and 20 current pulses (500 pA, 5 ms duration, 50 Hz) for neurons expressing GCaMP6f (dotted line) or SomaGCaMP6f1 (unbroken line, n = 7neurons from 5 slices from 2 mice for GCaMP6f; n = 5 neurons from 3 slices from 2 mice for SomaGCaMP6f1). n.s., not significant, ***P<0.001, Bonferroni-corrected Wilcoxon rank sum test of the df/f0 between GCaMP6f and SomaGCaMP6f1 expressing neurons; see Supplemental Table 6 for full statistics for Figure S5. Plotted is mean plus or minus standard error throughout the figure. (B) Bar chart showing the mean τ_{off} of calcium spikes in slice, during electrophysiological inducement of single action potentials (n = 3 neurons from 3 slices from 3 mice for GCaMP6f; n = 3 neurons from 3 slices from 3 mice for SomaGCaMP6f1). n.s., not significant, Wilcoxon rank sum test between GCaMP6f and SomaGCaMP6f1. (C) Bar chart showing the mean τ_{off} of calcium spikes in slice, during 4-aminopyridine inducement of single action potentials (n = 5 neurons from 5 slices from 4 mice for GCaMP6f; n = 5 neurons from 4 slices from 3 mice for SomaGCaMP6f1). *P < 0.05, Wilcoxon rank sum test between GCaMP6f and SomaGCaMP6f1. (D) Bar chart showing the mean event rate of calcium spikes per minute in slice (n = 8 neurons from 8 slices from 4 mice for GCaMP6f; n = 6 neurons from 6 slices from 3 mice or SomaGCaMP6f1). n.s., not significant, Wilcoxon rank sum test between GCaMP6f and SomaGCaMP6f1.



Figure S6. Temporal dynamics and calcium spike count for GCaMP6f and

SomaGCaMP6f1 expressing neurons in zebrafish larvae, driven by 4-AP; related to Figure 6. (A) A bar chart showing the mean GCaMP-spike rates for neurons in regions of the larval zebrafish forebrain expressing either GCaMP6f, SomaGCaMP6f1 or H2B-GCaMP6f (n = 101 neurons from 5 fishes for GCaMP6f; n = 146 neurons from 4 fishes for SomaGCaMP6f1; n = 513 neurons from 6 fishes for H2B-GCaMP6f). ***P<0.001, Kruskal-Wallis analysis of variance followed by post-hoc test via Steel's test; see Supplemental Table 6 for full statistics for Figure S6. Plotted is mean +/- standard error throughout this figure. (B) A bar chart showing the mean Pearson correlation coefficient between cell pairs in the larval zebrafish forebrain expressing either GCaMP6f, SomaGCaMP6f1 or H2B-GCaMP6f (n = 426 neurons from 5 fishes for GCaMP6f; n = 340 neurons from 4 fishes for SomaGCaMP6f1; n = 676 neurons from 6 fishes for H2B-GCaMP6f). White bars are for correlation coefficients calculated from raw data. Black bars are for correlation coefficieents calculated after CNMF was applied to the raw data. n.s., not significant. ***P<0.001, Kruskal-Wallis analysis of variance followed by post-hoc Tukey's HSD test;. (C) A bar chart showing the mean Pearson correlation coefficient between cell pairs in the larval zebrafish forebrain expressing either GCaMP6f (white), SomaGCaMP6f1 (gray) or H2B-GCaMP6f (black), in three distance ranges from the soma: 0-50 µm, 50-100 µm and 100-300 µm (n = 426 neurons from 5 fishes for GCaMP6f; n = 340 neurons from 4 fishes forSomaGCaMP6f1; n = 676 neurons from 6 fishes for H2B-GCaMP6f). Correlation coefficient was calculated from raw data. (D) A bar chart showing the mean Pearson correlation coefficient between cell pairs in the larval zebrafish forebrain expressing either GCaMP6f (white), SomaGCaMP6f1 (gray) or H2B-GCaMP6f (black), in three distance ranges from the soma: 0-50 μ m, 50-100 μ m and 100-300 μ m (n = 426 neurons from 5 fishes for GCaMP6f; n = 340 neurons from 4 fishes for SomaGCaMP6f1; n = 676 neurons from 6 fishes for H2B-GCaMP6f). Correlation coefficient was calculated after CNMF was applied to raw data.



Figure S7. Baseline fluorescence brightness, kinetics and pairwise correlations of GCaMP and SomaGCaMP variants in mouse striatum in vivo; related to Figure 7. (A) Bar chart showing the baseline fluorescence in vivo in the dorsal striatum for GCaMP6f, SomaGCaMP6f1 and SomaGCaMP6f2 (n = 75 neurons from 5 mice for GCaMP6f; n = 50 neurons from 2 mice for SomaGCaMP6f1; n = 80 neurons from 4 mice for SomaGCaMP6f2). ***P < 0.001, Kruskal-Wallis analysis of variance followed by post-hoc test via Steel's test with GCaMP6f as control group; see Supplemental Table 6 for full statistics for Figure S7. n.s., not significant, Kruskal-Wallis analysis of variance followed by post-hoc test via Steel's test with GCaMP6f as control group. Plotted is mean +/- standard error throughout the figure. (B) Bar chart showing the average rise time (τ_{on}) and the average decay time (τ_{off}) for neurons expressing either SomaGCaMP6f2 or GCaMP6f (n = 594 neurons from 4 mice expressing SomaGCaMP6f2, n =930 neurons from 6 GCaMP6f mice). n.s., not significant, Wilcoxon rank sum test between the rise times of SomaGCaMP6f2 and GCaMP6f expressing neurons; *** P <0.001, Wilcoxon rank sum test between the decay times of SomaGCaMP6f2 and GCaMP6f expressing neurons. (C, left) A bar chart showing the mean Pearson correlation coefficient between cell pairs in the mouse striatum expressing either GCaMP6f (white) or SomaGCaMP6f2 (gray), in three distance ranges from the soma: $0.50 \ \mu\text{m}$, $50-100 \ \mu\text{m}$ and $100-300 \ \mu\text{m}$ (n = 860 neurons from 6 mice for GCaMP6f; n = 149 neurons from 4 mice for SomaGCaMP6f2). Correlation coefficient was calculated from raw data. (C, right) A bar chart showing the mean Pearson correlation coefficient between cell pairs in the mouse striatum expressing either GCaMP6f (white) or SomaGCaMP6f2 (gray), after undergoing CNMF, in three distance ranges from the soma: 0-50 μ m, 50-100 μ m and 100-300 μ m (n = 634 cells from 4 mice for GCaMP7f; n = 1098 cells from 5 mice for SomaGCaMP7f). Correlation coefficient was calculated from raw data. (D) Bar chart showing the baseline fluorescence in vivo in the dorsal striatum for GCaMP7f and SomaGCaMP7f (n = 851 neurons from 5 mice for GCaMP7f; n = 1098 neurons from 5 mice for SomaGCaMP7f). n.s., not significant, Wilcoxon rank sum test between the baseline fluorescence of SomaGCaMP7f and GCaMP7f expressing neurons. (E) Bar chart showing the average rise time (τ_{on}) and the average decay time (τ_{off}) for neurons expressing either SomaGCaMP7f or GCaMP7f (n = 851 neurons from 5 mice for GCaMP7f; n = 1098 neurons from 5 mice for SomaGCaMP7f). n.s., not significant, Wilcoxon rank sum test between the rise or decay times of SomaGCaMP7f and GCaMP7f expressing neurons. (F, left) A bar chart showing the mean Pearson correlation coefficient between cell pairs in the mouse striatum expressing either

GCaMP7f (white) or SomaGCaMP7f (gray), in three distance ranges from the soma: 0-50 μ m, 50-100 μ m and 100-300 μ m (n = 860 neurons from 6 mice for GCaMP6f; n = 149 neurons from 4 mice for SomaGCaMP6f2). Correlation coefficient was calculated from CNMF applied to raw data. (**F**, **right**) A bar chart showing the mean Pearson correlation coefficient between cell pairs in the mouse striatum expressing either GCaMP7f (white) or SomaGCaMP7f (gray), after undergoing CNMF, in three distance ranges from the soma: 0-50 μ m, 50-100 μ m and 100-300 μ m (n = 634 cells from 4 mice for GCaMP7f; n = 1098 cells from 5 mice for SomaGCaMP7f). Correlation coefficient was calculated from CNMF applied to raw data.



Figure S8. Correlelograms based on neurons expressing GCaMP6f or SomaGCaMP6f2 imaged with a GRIN lens in the medial prefrontal cortex; related to Figure 8. (A, B) Correlelograms denoting the relationship of distance to the strength of correlated fluorescence between cell pairs from mice expressing GCaMP6f (A; n = 107 neurons from 2 mice) or SomaGCaMP6f2 (\mathbf{B} ; n = 222 neurons from 4 mice). Distance distributions are shown on the xaxis and Pearson correlation coefficients are shown on the y-axis. (top row) Analysis was performed using raw data. (bottom row) Analysis was performed using data subjected to the neuropil contamination elimination algorithm CNMF. ***P < 0.001, two-dimensional Kolmogorov-Smirnov test between GCaMP6f and SomaGCaMP6f1; see Supplemental Table 6 for full statistics for Figure S8. (C) A bar chart showing the mean Pearson correlation coefficient between cell pairs in the mouse medial prefrontal cortex expressing either GCaMP6f or SomaGCaMP6f2 (n = 107 neurons from 2 mice for GCaMP6f; n = 222 neurons from 4 mice for SomaGCaMP6f2). White bars are for correlation coefficients calculated from raw data. Black bars are for correlation coefficieents calculated from CNMF applied to the raw data. n.s., not significant. ***P<0.001, Kruskal-Wallis analysis of variance followed by post-hoc Tukey's HSD test. Plotted is mean +/- standard error throughout this figure. (D) A bar chart showing the mean Pearson correlation coefficient between cell pairs in the mouse medial prefrontal cortex expressing either GCaMP6f (white) or SomaGCaMP6f2 (gray), in three distance ranges from the soma: 0-25 μ m, 25-50 μ m and 50-100 μ m (n = 107 neurons from 2 mice for GCaMP6f; n = 222 neurons from 4 mice for SomaGCaMP6f2). Correlation coefficient was calculated from raw data. (E) A bar chart showing the mean Pearson correlation coefficient between cell pairs in the mouse medial prefrontal cortex expressing either GCaMP6f (white) or SomaGCaMP6f2 (gray), in three distance ranges from the soma: 0-25 μ m, 25-50 μ m and 50-100 μ m (n = 107 neurons from 2 mice for GCaMP6f; n = 222 neurons from 4 mice for SomaGCaMP6f2). Correlation coefficient was calculated after CNMF was applied to raw data.

Supplemental Tables

Supplemental Table 1. Related to Figures 1 and 2.

Supplemental Table 1A: Proteins that were considered in this study as potential soma targeting fragments. For amino acid sequences corresponding to the acronyms used, see **Supplemental Table 7** (successful fusions between these proteins an GCaMP6f or GCaMP7f are presented **in Figures 1, 2**).

Supplemental Table 1B: GCaMP6f fusion proteins that were screened in cultured hippocampal neurons in this project (successful candidates are presented in Figures 1, 2).

Supplemental Table 2: Statistical analysis for Figure 1, 2 and 3. Related to Figures 1, 2, and 3.

Supplemental Table 3: Statistical analysis for mouse brain slice, and for fish and mouse in vivo experiments (which include Figures 4, 6, 7, 8). Related to Figures 4, 6, 7, and 8.

Supplemental Table 4: statistical analysis for Figure 5.

<u>Figure 5G</u> Two-way analysis of variance (ANOVA) of the correlation coefficient between the ground-truth calcium dynamics and recorded calcium dynamics in the simulations for mouse, followed by post-hoc Tukey's HSD test.

Factor 1, molecules: SomaGCaMP6f2 vs GCaMP6f. Factor 2, demixing: with CMNF vs without CMNF.

n = 300 neurons from 10 simulations for SomaGCaMP6f2; n = 300 neurons from 10 simulations for GCaMP6f.

Source	SS	df	MS	F	Prob>F
Molecules	6.2098	1	6.2098	40.5516	2.72E-10
Demixing	0.0981	1	0.0981	0.6408	0.4236
Interaction	0.1592	1	0.1592	1.0394	0.3082
Error	183.1481	1196	0.1531	-	-
Total	189.6152	1199	-	-	-

Two-way ANOVA table:

Post-hoc Tukey's HSD test on Factor 1, molecules (SomaGCaMP6f2 vs GCaMP6f):

$$P = 2.9646e-10$$

<u>Figure 5H</u> Two-way analysis of variance (ANOVA) of the correlation coefficient between the ground-truth calcium dynamics and recorded calcium dynamics in the simulations for zebrafish, followed by post-hoc Tukey's HSD test.

Factor 1, molecules: SomaGCaMP6f1 vs GCaMP6f. Factor 2, demixing: with CMNF vs without CMNF.

n = 1200 neurons from 10 simulations for SomaGCaMP6f1; n = 1200 neurons from 10 simulations for GCaMP6f.

Two-way ANOVA table:

Source	SS	df	MS	F	Prob>F
Molecules	24.4007	1	24.4007	260.8343	3.76E-57
Demixing	0.2303	1	0.2303	2.4618	0.1167
Interaction	0.2002	1	0.2002	2.1397	0.1436
Error	441.1751	4716	0.0935	-	-
Total	466.0063	4719	-	-	-

Post-hoc Tukey's HSD test on Factor 1, molecules (SomaGCaMP6f1 vs GCaMP6f):

P = 1.0597e-10

Supplemental Table 5: Statistical analysis for Figure S1 – membrane and action potential properties. Related to Figure 3.

Supplemental Table 6: Statistical analysis for Figure S2, S4, S5, S6, S7, S8. Related to Figures 1-8.

Supplemental Table 7: Amino acid sequences for protein fragments used in this paper. Related to Figures 1, 2.

AnkTail-motif (Ankyrin_G (1934-2333)):

REGRIDDEEPFKIVEKVKEDLVKVSEILKKDVCVESKGPPKSPKSDKGHSPEDDWTEFSS EEIREARQAAASHAPSLPERVHGKANLTRVIDYLTNDIGSSSLTNLKYKFEEAKKDGEER QKRILKPAMALQEHKLKMPPASMRPSTSEKELCKMADSFFGADAILESPDDFSQHDQDK SPLSDSGFETRSEKTPSAPQSAESTGPKPLFHEVPIPPVITETRTEVVHVIRSYEPSSGEIPQS QPEDPVSPKPSPTFMELEPKPTTSSIKEKVKAFQMKASSEEEDHSRVLSKGMRVKEETHI TTTTRMVYHSPPGGECASERIEETMSVHDIMKAFQSGRDPSKELAGLFEHKSAMSPDVA KSAAETSAQHAEKDSQMKPKLERIIEVHIEKGPQSPCE

EE-RR:

 $\label{eq:lessel} LEIEAAFLEQENTALETEVAELEQEVQRLENIVSQYETRYGPLGSLEIRAAFLRRRNTALR TRVAELRQRVQRLRNIVSQYETRYGPL$

AcidP1-BaseP1:

AQLEKELQALEKENAQLEWELQALEKELAQGSGSAQLKKKLQALKKKNAQLKWKLQ ALKKKLAQ

nullsfGFP (mutation to abolish the fluorescence of the original sfGFP is underlined)

MSKGEELFTGVVPILVELDGDVNGHKFSVRGEGEGDATNGKLTLKFICTTGKLPVPWPT LVTTLT<u>G</u>GVQCFSRYPDHMKRHDFFKSAMPEGYVQERTISFKDDGTYKTRAEVKFEGD TLVNRIELKGIDFKEDGNILGHKLEYNFNSHNVYITADKQKNGIKANFKIRHNVEDGSVQ LADHYQQNTPIGDGPVLLPDNHYLSTQSVLSKDPNEKRDHMVLLEFVTAAGITHGMDE LYK

NLS

RKRPSDLVHVFSPPRKK

KGC

KSRITSEGEYIPLDQIDINV

ER2

FCYENEV

nullCoChR (mutation to abolish photocurrent of the original CoChR is underlined)

MLGNGSAIVPIDQCFCLAWTDSLGSDTEQLVANILQWFAFGFSILILMFYAYQTWRATC GWEEVYVCCVELTKVIIEFFHEFDDPSMLYLANGHRVQWLRYAEWLLTCPVILIHLSNL TGLKDDYSKRTMRLLVSDVGTIVWGATSAMSTGYVKVIFFVLGCIYGANTFFHAAKVYI ESYHVVPKGRPRTVVRIMAWLFFLSWGMFPVLFVVGPEGFDAISVYGSTIGHTIIDLMS<u>A</u> NCWGLLGHYLRVLIHQHIIIYGDIRKKTKINVAGEEMEVETMVDQEDEETV

KA2(1-150)

MPAELLLLLIVAFANPSCQVLSSLRMAAILDDQTVCGRGERLALALAREQINGIIEVPAK ARVEVDIFELQRDSQYETTDTMCQILPKGVVSVLGPSSSPASASTVSHICGEKEIPHIKVG PEETPRLQYLRFASVSLYPSNEDVSLAVS

KA2(1-150)-Y76A

MPAELLLLIVAFANPSCQVLSSLRMAAILDDQTVCGRGERLALALAREQINGIIEVPAK ARVEVDIFELQRDSQAETTDTMCQILPKGVVSVLGPSSSPASASTVSHICGEKEIPHIKVG PEETPRLQYLRFASVSLYPSNEDVSLAVS

KA2(1-100)

 $\label{eq:main_scale} MPAELLLLIVAFANPSCQVLSSLRMAAILDDQTVCGRGERLALALAREQINGIIEVPAK\\ ARVEVDIFELQRDSQYETTDTMCQILPKGVVSVLGPSSSP$

Ank(1-334) (Ankyrin_G (1-334))

MAHAASQLKKNRDLEINAEEETEKKKKHRKRSRDRKKKSDANASYLRAARAGHLEKA LDYIKNGVDVNICNQNGLNALHLASKEGHVEVVSELLQREANVDAATKKGNTALHIAS LAGQAEVVKVLVTNGANVNAQSQNGFTPLYMAAQENHLEVVRFLLDNGASQSLATED GFTPLAVALQQGHDQVVSLLLENDTKGKVRLPALHIAARKDDTKAAALLLQNDTNADI ESKMVVNRATESGFTSLHIAAHYGNINVATLLLNRAAAVDFTARNDITPLHVASKRGNA NMVKLLLDRGAKIDAKTRDGLTPLHCGARSGHEQVVEMLLDRAAP

AnkCT-motif (Ankyrin_G (2334-2622))

RTDIRMAIVADHLGLSWTELARELNFSVDEINQIRVENPNSLISQSFMLLKKWVTRDGKN ATTDALTSVLTKINRIDIVTLLEGPIFDYGNISGTRSFADENNVFHDPVDGWQNETPSGSL ESPAQARRLTGGLLDRLDDSSDQARDSITSYLTGEPGKIEANGNHTAEVIPEAKAKPYFP ESQNDIGKQSIKENLKPKTHGCGRTEEPVSPLTAYQKSLEETSKLVIEDAPKPCVPVGMK KMTRTTADGKARLNLQEEEGSTRSEPKQGEGYKVKTKKEIRNVEKKTH

AnkMB-motif (Ankyrin_G (1-800))

MAHAASQLKKNRDLEINAEEETEKKRKHRKRSRDRKKKSDANASYLRAARAGHLEKA LDYIKNGVDVNICNQNGLNALHLASKEGHVEVVSELLQREANVDAATKKGNTALHIAS LAGQAEVVKVLVTNGANVNAQSQNGFTPLYMAAQENHLEVVRFLLDNGASQSLATED GFTPLAVALQQGHDQVVSLLLENDTKGKVRLPALHIAARKDDTKAAALLLQNDTNAD VESKSGFTPLHIAAHYGNINVATLLLNRAAAVDFTARNDITPLHVASKRGNANMVKLLL DRGAKIDAKTRDGLTPLHCGARSGHEQVVEMLLDRSAPILSKTKNGLSPLHMATQGDH LNCVQLLLQHNVPVDDVTNDYLTALHVAAHCGHYKVAKVLLDKKASPNAKALNGFTP LHIACKKNRIRVMELLLKHGASIQAVTESGLTPIHVAAFMGHVNIVSQLMHHGASPNTT NVRGETALHMAARSGQAEVVRYLVQDGAQVEAKAKDDQTPLHISARLGKADIVQQLL QQGASPNAATTSGYTPLHLAAREGHEDVAAFLLDHGASLSITTKKGFTPLHVAAKYGKL EVASLLLQKSASPDAAGKSGLTPLHVAAHYDNQKVALLLLDQGASPHAAAKNGYTPLH IAAKKNQMDIATSLLEYGADANAVTRQGIASVHLAAQEGHVDMVSLLLSRNANVNLSN $\label{eq:construction} KSGLTPLHLAAQEDRVNVAEVLVNQGAHVDAQTKMGYTPLHVGCHYGNIKIVNFLLQ\\ HSAKVNAKTKNGYTALHQAAQQGHTHIINVLLQNNASPNELTVNGNTAL$

AnkSB-motif (Ankyrin_G (801-1521))

AIARRLGYISVVDTLKVVTEEIMTTTTITEKHKMNVPETMNEVLDMSDDEVRKASAPEK LSDGEYISDGEEGEDAITGDTDKYLGPQDLKELGDDSLPAEGYVGFSLGARSASLRSFSS DRSYTLNRSSYARDSMMIEELLVPSKEQHLTFTREFDSDSLRHYSWAADTLDNVNLVSS PVHSGFLVSFMVDARGGSMRGSRHHGMRIIIPPRKCTAPTRITCRLVKRHKLANPPPMVE GEGLASRLVEMGPAGAQFLGPVIVEIPHFGSMRGKERELIVLRSENGETWKEHQFDSKN EDLAELLNGMDEELDSPEELGTKRICRIITKDFPQYFAVVSRIKQESNQIGPEGGILSSTTV PLVQASFPEGALTKRIRVGLQAQPVPEETVKKILGNKATFSPIVTVEPRRRKFHKPITMTI PVPPPSGEGVSNGYKGDATPNLRLLCSITGGTSPAQWEDITGTTPLTFIKDCVSFTTNVSA RFWLADCHQVLETVGLASQLYRELICVPYMAKFVVFAKTNDPVESSLRCFCMTDDRVD KTLEQQENFEEVARSKDIEVLEGKPIYVDCYGNLAPLTKGGQQLVFNFYSFKENRLPFSI KIRDTSQEPCGRLSFLKEPKTTKGLPQTAVCNLNITLPAHKKETESDQDDAEKADRRQSF ASLALRKRYSYLTEPSMKTVERSSGTARSLPTTYSHKPFFSTRPYQSWTTAPITVPGPAKS GSLSSSPSNTPSA

AnkSR-motif (Ankyrin_G (1534 -1933))

SPLKSIWSVSTPSPIKSTLGASTTSSVKSISDVASPIRSFRTVSSPIKTVVSPSPYNPQVASGT LGRVPTITEATPIKGLAPNSTFSSRTSPVTTAGSLLERSSITMTPPASPKSNITMYSSSLPFK SIITSATPLISSPLKSVVSPTKSAADVISTAKATMASSLSSPLKQMSGHAEVALVNGSVSPL KYPSSSALINGCKATATLQDKISTATNAVSSVVSAASDTVEKALSTTTAMPFSPLRSYVS AAPSAFQSLRTPSASALYTSLGSSIAATTSSVTSSIITVPVYSVVNVLPEPALKKLPDSNSF TKSAAALLSPIKTLTTETRPQPHFNRTSSPVKSSLFLASSALKPSVPSSLSSSQEILKDVAE MKEDLMRMTAILQTDVPEEKPFQTDLP

$K_V 2.1$ -motif ($K_V 2.1(536-600)$)

QSQPILNTKEMAPQSKPPEELEMSSMPSPVAPLPARTEGVIDMRSMSSIDSFISCATDFPE ATRF

rSK1-tail (rSK1(351-411))

QAQKLRTVKIEQGKVNDQANTLADLAKAQSIAYEVVSELQAQQEELEARLAALESRLD VLGASLQALPSLIAQAICPLPPPWPGPSHLTTAAQSPQSHWLPTTASDCG

Nav1.6(II-III)

TVRVPIAVGESDFENLNTEDVSSESDP

Nav1.2(I-II)

YEEQNQATLEEAEQKEAEFQQMLEQLKKQQEEAQAAAAAASAESRDFSGAGGIGVFSE SSSVASKLSSKSEKELKNRRKKKKQKEQAGEEEKEDAVRKSASEDSIRKKGFQFSLEGS RLTYEKRFSSPHQSLLSIRGSLFSPRRNSRASLFNFKGRVKDIGSENDFADDEHSTFEDND SRRDSLFVPHRHGERRPSNVSQASRASRGIPTLPMNGKMHSAVDCNGVVSLVGGPSALT SPVGQLLPEGTTTETEIRKRRSSSYHVSMDLLEDPSRQRAMSMASILTNTMEELEESRQK CPPCWYKFANMCLIWDCCKPWLKVKHVVN Supplemental Table 8: Percentage of saturated pixels in images presented. Related to Figures 1, 2, 4, 6, 7, 8.

Figure 1, panels C, D, E, I, J

Variant	Saturated pixels in non- saturated image	Saturated pixels in saturated image	Total number of pixels in image	% of saturated pixels in the non-saturated image	% of saturated pixels in the saturated image
GCaMP6f	0	4825	4194304	0	0.1150369644
SomaGCaMP6f1	0	5882	4194304	0	0.1402378082
SomaGCaMP6f2	0	4819	4194304	0	0.1148939133
	0	1700	4104204	0	0.11.4170.6775
GCaMP7f	0	4789	4194304	0	0.1141/865/5
SomaGcaMP7f	0	12917	4194304	0	0.3079652786

Figure 2

Panels A, B, C

Variant	Saturated pixels in image	Total number of pixels in image	% of saturated pixels in the image
GCaMP6f	65	35680	0.180
SomaGCaMP6f1	60	35680	0.168
SomaGCaMP6f2	64	35680	0.179

Figure 4

Panel A

Variant	Saturated pixels in non- saturated image	Saturated pixels in saturated image	Total number of pixels in image	% of saturated pixels in the non-saturated image	% of saturated pixels in the saturated image
GCaMP6f	0	327	103200	0	0.3168604651
SomaGCaMP6f1	0	337	103200	0	0.3265503876
SomaGCaMP6f2	0	380	103200	0	0.3682170543

Figure 6

Panels A, B

Variant	Saturated pixels in non- saturated image	Saturated pixels in saturated image	Total number of pixels in image	% of saturated pixels in the non- saturated image	% of saturated pixels in the saturated image
GCaMP6f	11	224	28784	0.03821567538	0.7782101167
SomaGCaMP6f1	0	324	28784	0	1.125625347
GCaMP7	0	227	28784	0	0.7886325737
SomaGCaMP7f	4	351	28784	0.01389660923	1.21942746

Figure 7

Panels A, B, C, D

Variant	Saturated pixels in image	Total number of pixels in image	% of saturated pixels in the image
GCaMP6f	0	154086	0
SomaGCaMP6f1	0	154086	0
GCaMP7	0	122500	0
SomaGCaMP7f	0	122500	0

Figure 8

Panels A, B, C, D

Variant	Saturated pixels in image	Total number of pixels in image	% of saturated pixels in the image
GCaMP6f	1	64500	0.001550387597
SomaGCaMP6f12	1	64500	0.001550387597