

1 **Supplementary Note**

2 Cell state identification

3 To connect our gene expression signatures to cell states, we searched localized gene
4 expression data from whole mount *in situ* hybridization (e.g. using the Gene eXpression
5 Database (GXD): <http://www.informatics.jax.org/expression.shtml>), transgenic reporters or
6 single-cell expression analysis. We compared these data to a state by gene matrix that reports
7 the percent of positive cells in which transcripts of a given gene were detected
8 (**Supplementary Table 4**). Even in this context, the percent detected is sensitive to the
9 absolute expression level due to the sparse, drop-out prone nature of single-cell expression
10 data. We called a gene as specific to a state by comparing its expression estimate within that
11 state to all other states.
12

13 Our cell state annotation considers the developmental stage in which a cell state is
14 present as well as the combination of expressed genes. Its nomenclature is intended to serve
15 as an orientation, referring to cell types, anatomical structures or compartments within the
16 embryo from which a subset of the state's cells originate. This is particularly relevant for later
17 time points, where embryos are more complex and spatially resolved as well as less deeply
18 sampled. Thus, several cell states are likely to contain various related cell types and we
19 highlight these considerations below. For example, the “primitive heart tube” comprises cells
20 expressing markers for this embryonic structure, such as *Nkx2.5* and *Hcn4*, but may also
21 contain other myogenic cell types of the developing heart, such as cells of the secondary heart
22 field (PMID: 23974038, PMID: 12141429, PMID: 22521611).
23

24 We also placed our states in a lineage tree according to expected developmental
25 relationships, suggesting a hierarchical and timely ordered occurrence. However, the
26 occurrence of many states that were placed consecutively in the tree partially overlap,
27 resulting in states connected by arrows even though some of their cells may transiently
28 coexist within the same embryo (e.g. ‘23–notochord’ proceeds from ‘38–node’ even though
29 they temporally overlap in E7.5, but state 38 diminishes after this time point while 23 remains
30 stable into E8.5).
31

32 We provide the references as Pubmed IDs rather than formatting a separate reference
33 to facilitate follow up for the reader. At the end of this document we provide information on
34 the comparability between the WT single-cell reference of this study and the one by Pijuan-
35 Sala *et al.*
36

35 Epiblast

36 Cluster 17 represents the “**epiblast**,” the cell state of the embryo proper mainly composed of
37 pluripotent cells that are not part of the primitive streak. Its presence is restricted to our E6.5
38 timepoint and it displays the highest expression of *Otx2* (PMID: 15201223), *Zic2* (PMID:
39 15261827), *Nodal* (PMID: 9056778) and *Fgf5* (PMID: 24131634). *Fgf4* expression excludes
40 the possibility of cluster 17 being of extraembryonic ectodermal or endodermal nature
41 (PMID: 1618140). High *Utf1* expression argues against a mesodermal nature, as this gene
42 was reported to be expressed in pluripotent cells and extraembryonic ectoderm but not
43 expressed in primitive mesoderm (PMID: 9524124), which agrees with its comparatively
44 reduced prevalence in states of the primitive streak (see below).
45

46 Embryonic mesoderm lineage

47 The primitive streak states (37, 3, 2) were characterized by the expression of *T* (PMID:
48 1821859), *Evx1* (PMID: 1349539) and *Lhx1* (PMID: 10328927). We assigned cluster 37 to
49 “**primitive streak early**” as it is present at E7.0 and expresses *Mesp1*, which is detected in
50 ingressing mesodermal cells as they pass through the primitive streak (PMID: 10393122) as
51 well as *Dll1*, which was reported for the primitive streak and mesoderm at this stage (PMID:

52 7671806). Cluster 3 is maximal at E7.0 and characterized by a fraction of cells positive for the
53 mesendodermal markers *Foxa2* (PMID: 8375339, PMID: 12351174) and *Gsc* (PMID:
54 9671576). Importantly, it constitutes the largest fraction of primitive streak cells at E6.5 and
55 thus likely also contains cells that are committed to this state but otherwise pre-specified.
56 Consequently, we termed it **“primitive streak pre-specified and anterior.”**

57 Cluster 2 emerges around E7.5 as cluster 37 diminishes and contains cells of the
58 primitive streak and nascent mesoderm. We annotated this state **“primitive streak late,”** in
59 line with its high expression of *Hoxb1*, which extends along the primitive streak and is
60 present in the mesoderm of the posterior embryo around E7.5 (PMID: 1983472). Moreover,
61 the expression of *Eomes* is substantially reduced compared to the primitive streak states that
62 are mainly present at E7.0 (state 37 and 3).

63 Cell states of the caudal epiblast (26, 30, 31) also express *T*, *Evx1* and *Lhx1*. Their
64 more posterior position within the embryo is supported by continued *Hoxb1*, reported to be
65 expressed not only in the posterior mesoderm but also in the ectoderm anterior of and lateral
66 to the primitive streak at E7.75 (PMID: 1983472). Cluster 26 (present around E7.5-8.0) and
67 cluster 31 (present around E8.0-8.5) have a high fraction of cells that co-express *T* and *Sox2*,
68 which is characteristic of neuromesodermal progenitors (NMPs). Thus, these states were
69 annotated as **“NMPs early”** (cluster 26) and **“NMPs late”** (cluster 31) according to their
70 temporal dynamics. In addition to their mesodermal signature, both states express the
71 neuroectodermal genes *Sox3*, *Sp8*, *Nkx1-2* and *Olig3* (reported to be expressed in NMPs and
72 derivatives, PMID: 28826820). State 26 (NMPs early) appears to give rise to state 31, at least
73 partially, which might also contain more neuroectodermally-committed cells, as indicated by
74 a slightly increased fraction of cells expressing of *Pax3* (PMID: 28826820).

75 In contrast, cluster 30 emerges around E8.0-8.5, does not express *Sox2*, but instead
76 expresses genes of the **“presomitic mesoderm”** such as *Tbx6* (PMID: 8954725), *Snai1*
77 (PMID: 9671584), *Dll1* (PMID: 9671584) and *Nrarp* (PMID: 19268448). Notably, we also
78 detect the intermediate mesodermal marker *Osr1* (PMID: 10473132) in a large fraction of
79 state 30 cells, suggesting that the state may also contain cells of this tissue. Transcriptional
80 similarity (e.g. the high expression of *Hoxb1*, *Dll1*, *T*) suggests that state 30 is derived from
81 state 2, though they may also proceed directly from NMPs. Interestingly, state 31 has a
82 substantially larger fraction of cells expressing *Cyp26a1* compared to state 30, which may
83 indicate a more caudal position within the embryo (PMID: 11520679).

84 Cluster 12 is most prevalent around E7.5-8.5 and comprised of cells of the lateral plate
85 mesoderm. It expresses *Foxf1* (PMID: 11124112) and *Isl1* (PMID: 14667410). Similar to
86 state 30 (presomitic mesoderm), cluster 12 may also contain cells of the intermediate
87 mesoderm as indicated by expression of *Osr1* (PMID: 10473132). Expression of genes
88 specific to the caudal part of the embryo, such as *Cdx4* (PMID: 19906845) and *Hoxd9*
89 (PMID: 1676674), also supports a posterior position. From these data, we assigned this state
90 **“posterior lateral plate mesoderm”**, though it may contain mesoderm of other structures as
91 well.

92 Cluster 20 is present around E7.5-8.0 and resembles **“splanchnic lateral/anterior
93 paraxial mesoderm”** (PMID: 21498416). This state expresses the anterior mesoderm specific
94 *Tbx1* (PMID: 8853987), which was described to be exclusively expressed in the anterior
95 embryonic mesoderm, rostral to the node at E7.5. Furthermore, this state expresses: *Tcf21*,
96 which is expressed within the first branchial arch at E8.0 (PMID: 9733105); *Isl1*, which is
97 expressed at the earliest stages of cardiac development (PMID: 25174608); and *Prdm1*, which
98 is expressed in splanchnic mesoderm (PMID: 12204275).

99 Cluster 34 also expresses several genes characteristic for the pharyngeal mesoderm
100 (PMID: 21498416) and is present mainly around E8.5. We termed this state **“pharyngeal
101 arch mesoderm”** by the expression of *Tbx1* (PMID: 8853987, PMID: 21364285), *Six1*
102 (PMID: 21364285), *Eya1* (PMID: 21364285) and *Pax9* (PMID: 9732271), which are

103 associated with this tissue as well as other cranial mesenchymal structures. Cluster 6, present
104 at E7.5-8.5, was termed **“secondary heart field/splanchnic lateral plate mesoderm”** due to
105 the expression of *Isl1*, a marker for the secondary heart field (SHF, PMID: 25174608; PMID:
106 14667410). It also displays high *Hand2* expression, which is found in the lateral mesoderm
107 and developing heart from E7.75 onwards (PMID: 8533092). The expression of *Foxf1* and
108 *Irx3* support the notion that state 6 contains cells of the splanchnic (*Foxf1*) and also of the
109 somatic (*Irx3*) lateral plate mesoderm (PMID: 11124112). Overall, this state might be
110 composed of many cells that eventually contribute to the formation of the heart tube. Cluster
111 21, present around E8.0 and 8.5, is largely composed of cells of the **“primitive heart tube”**
112 according to the expression of *Myl7* (PMID: 11245568), *Nkx2-5* (PMID: 12141429, PMID:
113 11336496), *Hcn4* and *Tbx5* (PMID: 8853987), *Mef2c* (PMID: 8026334) and *Myocd*. *Hcn4*
114 and *Myocd* have been reported to be rather specific for the FHF (PMID: 23974038, PMID:
115 25174608). The scarcity of known markers that are distinct for the First and Second Heart
116 Fields makes it difficult to conclude with certainty whether 6 gives rise to state 21 or whether
117 21 develops from state 20 independently and in parallel. Nevertheless, higher *Hoxb1*
118 expression in state 6 compared to state 21 suggests that it is located posterior to the primary
119 heart tube.

120 Cluster 18 begins to emerge at E7.5 and continues through E8.0-8.5. We termed this
121 state **“somites”** as it expresses *Ripply2*, which is reported to be expressed at the site of somite
122 formation (Somite 0-1, PMID: 18045842), as well as *Tbx18* (PMID: 11118889) and *Pax3*
123 (PMID: 18644785), which are expressed throughout formed somites. Higher expression of
124 *Aldh1a2* (PMID: 17849458) and *Foxc1* (PMID: 8375339) in this cell state compared to
125 cluster 30 (presomitic mesoderm) indicates that this state preferentially contains cells of the
126 somatic rather than the presomitic mesoderm. Notably, in the lineage tree we connected the
127 somite state with state 20 which contains amongst others more anterior portions of paraxial
128 mesoderm that are expected to undergo somite formation within this developmental window.
129 In contrast, presomitic mesoderm of state 30 is expected to give rise to somites at later
130 developmental stages.

131 The entire lineage of hematopoietic and endothelial cells was marked by the
132 expression of *Tal1*. Cluster 13 is present at E7.0 and is thereby the first detected state
133 expressing *Tal1*, *Kdr1* and *Etv2*, characteristic for **“hematopoietic and endothelial
134 progenitors”** (PMID: 24052951). Cluster 36 likely arises from Cluster 13 and was termed
135 **“primitive blood progenitors.”** It is most prevalent around E7.5 and characterized by the
136 expression of *Hbb-bh1*, *Gata1*, *Nfe2* (PMID: 29311656), *Klf1* (PMID: 16380451) and *Runx1*
137 (PMID: 10226014), genes known to demarcate the first wave of blood production in the
138 embryo.

139 Cell states 9 and 22 appear to be **“primitive blood early and late.”** They are prevalent
140 at E8.0 and E8.5, respectively, and express marker genes mentioned for cluster 36. Within
141 this lineage, we see a progressively decreasing fraction of *Runx1+* cells from state 36 through
142 22, consistent with previous reports (PMID: 10226014). Cell state 32 also appears to arise
143 from cells of state 13 at E7.5. We characterized these cells as **“angioblasts”** by the high
144 expression of *Pecam1* (PMID: 24052951, PMID: 24550118), *Cdh5* (PMID: 24052951), *Tek*
145 (PMID: 8187650) and *Lyve1* (PMID: 27880904). This state also expresses *Gja5* and *Ephb4*,
146 an arterial and venous signature consistent with their endothelial identity (PMID: 21793101).

147 Cluster 38 and 23 represent cells of the axial mesoderm identified by the expression of
148 *Noto* (PMID: 15533813), *Shh* (PMID: 8069909) and *Foxa2* (PMID: 8375339). Cluster 38
149 emerges at E7.0 and was therefore annotated to be the **“node,”** formed at the anterior most
150 position of the primitive streak (specifically state 3). Subsequently, cluster 23 emerges around
151 E7.5 and persists into E8.5, suggesting that these are cells of the developing **“notochord”** that
152 expands anteriorly from the node.

153

154 Extraembryonic mesoderm lineage

155 Cluster 15, present around E7.5-8.5, and cluster 41, present around E8.0-8.5, were annotated
156 as the “**amnion mesoderm early**” and “**amnion mesoderm late**” respectively according to
157 the expression of Foxf1 (PMID: 11124112) and Postn, which is highly specific (PMID:
158 22966238). Foxf1 is also expressed in the “**allantois**” (cluster 5), another extraembryonic
159 mesoderm cell state that emerges around E7.0 and then persists throughout the developmental
160 window investigated here. The allantois expresses Tbx4 (PMID: 21932311, PMID: 8853987)
161 and Tbx20 (PMID: 10940636), as well as posterior Hox genes Hoxa10, Hoxa11 and Hoxa3
162 that are very specific to this tissue (PMID: 22219351).

163

164 Germline

165 Cluster 27, of which a small number of cells are already present at E6.5, is unambiguously the
166 primordial germ cell state (“**PGCs**”) according to the expression of Dppa3 and Prdm1
167 (PMID: 18583473), Prdm14 (PMID: 18622394) and Nanos3 (PMID: 20174582).

168

169 Extraembryonic and embryonic endoderm lineages

170 We identified endodermal cells according to their expression of Foxa2 (PMID: 22236333,
171 PMID: 8306889) and Sox17 (PMID: 11973269). Cluster 4 is most prevalent at E6.5 but
172 persists through E7.0, and was annotated as “**primitive and definitive endoderm,**” with two
173 distinct placements in our lineage tree (**Fig. 1c**). Cluster 4 represents a large fraction of E6.5
174 embryos (15%), suggesting an abundance of extraembryonic endoderm, though it likely also
175 contains cells of the emerging embryonic endoderm. Separating these cells further is
176 complicated by their highly similar gene expression signatures, which diverge later to become
177 more obvious (see differential analysis of gene expression for E7.5 and E8.25 in PMID:
178 22236333). Cluster 29 and 40 represent “**visceral endoderm early and late**” as these two
179 states emerge successively and both express Cubn and Amn (PMID: 20637190) as well as
180 Amot and Slc39a8 (PMID: 17576135). Cluster 14 appears to be “**parietal endoderm,**” which
181 expresses Srgn (PMID: 11369593) and Thbd (PMID: 8681807) as well as Gkn2 and Pga5
182 (PMID: 28012457). This state was captured irregularly across embryonic replicates but
183 detected consistently over development, likely due to technical variability during isolation
184 that results from its location as the outermost extraembryonic layer. Both the parietal and, to a
185 lesser extent, visceral endoderm express Sox7, which distinguishes them from cluster 7
186 (PMID: 11973269).

187 Cluster 7 is present from E7.5-8.5 and composed of embryonic endoderm cells.
188 Because the main endodermal structure present at these developmental stages is the primitive
189 gut, the state was termed “**gut endoderm.**” It is characterized by the absence or
190 comparatively reduced expression of the above mentioned visceral and parietal endoderm
191 markers, as well as by presence the definitive endoderm markers Pyy (PMID: 17683524),
192 Foxa1, Cldn8 and Sorcs2 (PMID: 22236333). Whether this cell state directly emerges from
193 state 3 or proceeds through some intermediate cells contained within state 4 is unclear.
194 Several recent works have confirmed that a subset of cells within this state is of
195 extraembryonic origin (PMID: 30787436, PMID: 30959515, and PMID: 31086336).
196 However, because the continued presence of these cells beyond E8.75 and their
197 developmental function remain unknown, we decided to place this state solely within the
198 embryonic portion of our tree.

199

200 Embryonic ectoderm lineage

201 Cluster 8 and cluster 19 are both already present at E6.5 and are generally similar in gene
202 composition to epiblast. Cluster 19 becomes less prevalent towards E7.5, while cluster 8
203 peaks in abundance at E7.0 and comprises a substantial fraction of the embryo. Both clusters
204 are enriched in Otx2, Zic2, Utf1 and Fgf5 albeit to a lesser extent than in the epiblast state 17.

205 Additionally, expression of Sox2 and Pou3f1 indicate that cells of these states originate from
206 the more anterior part of the epiblast (PMID: 24131634), which is in line with the
207 substantially lower fraction of cells expressing markers of the primitive streak. Their
208 ectodermal character is supported by the expression of Sox3 (PMID: 10446282) and a small
209 number of cells (~10%) with detectable Six3 and Hesx1, which are both neural markers
210 (PMID: 24131634). Thus, we termed both states “**ectoderm early 1**” (cluster 8) and
211 “**ectoderm early 2**” (cluster 19). We placed these states jointly within the lineage tree to
212 highlight the ambiguity of their direct developmental relationship: it is unclear whether one
213 differentiates into another or if they represent distinct regions of the differentiating pluripotent
214 field. However, state 8 displays a higher fraction of cells expressing the epiblast markers,
215 which indicates a more direct relationship to state 17. Furthermore, state 3 cells (primitive
216 streak pre-specified and anterior) are transcriptionally most similar to state 19, suggesting that
217 state 19 may be comprised of cells that are localized in a similar region within the epiblast or
218 are fated to transverse through the anterior primitive streak.

219 Cluster 1, 11, 24, and 39 represent neuroectodermal clusters characterized by the
220 expression of Sox1, Sox2 (which we consider to be a neuroectodermal marker from E7.5
221 onwards) and Sox3 (PMID: 10446282). Cluster 24 emerges at E8.0 and expresses fore- and
222 midbrain-associated Otx2, the midbrain marker Wnt1, as well as En1 and Pax2, which mark
223 the mid-hindbrain border (PMID: 11253000). This cluster also contains cells that express
224 Foxg1, which emerges around E8.5 within the anterior neural folds (PMID: 16530751) and
225 Six3, reported for the most rostral neuroectoderm (PMID: 11532921). Taken together, we
226 conclude that cluster 24 represents the developing “**fore/midbrain**” and likely emerges from
227 cluster 1, which is present at E7.5-8.0 and is Six3, Otx2, Pax2 and En1 positive.
228 Consequently, cluster 1 was termed “**neural ectoderm anterior**.” Cluster 11 is most
229 prevalent at E7.5 and annotated as “**neural ectoderm posterior**” due to the expression of the
230 hindbrain marker Gbx2 (PMID: 11253000, PMID: 11532921) and Hoxa1 (PMID: 9053316).
231 Cluster 39 is present at E8.0-8.5 and appears to be “**future spinal cord**” by its expression of
232 Hoxb8 (PMID: 8096483), Nkx6-1 and Olig2 (PMID: 15652703). We placed this cell state
233 downstream of state 11 because they both represent posterior neuroectodermal cells and
234 emerge successively. However, we include a dashed line in our lineage tree to account for a
235 possible dual origin from neuromesodermal progenitor cell states (26 and 31).

236 Cluster 33 is most prevalent at E8.5 and appears to be “**neural crest**” by the
237 expression of Sox10, Sox9, Twist, Foxd3, and Tfap2a (PMID: 24780627, PMID: 22889333,
238 PMID: 22889333). It was placed in the tree downstream of the developing brain (state 24).
239 However, more posterior structures like the spinal cord also produce neural crest cells and are
240 expected to contribute to this cell state. Cluster 35 is present around the same time and did not
241 display any enrichment of specific marker genes that would have allowed an anatomical or
242 cell type specific annotation. Its transcriptional profile appears most similar to state 33, which
243 is why it was termed “**similar to neural crest**” and placed adjacent to it in the lineage tree.

244 Cluster 10 and 16 both emerge around E7.5 and progressively increase in relative
245 abundance. The presence of Dlx5 suggests that both are non-neural ectoderm (PMID:
246 9763476). Cluster 10 expresses Six1 and Eya1, consistent with a “**preplacodal ectoderm**”
247 identity (PMID: 19027001), whereas cluster 16 was annotated as “**surface ectoderm**”
248 because it expresses Trp63 (PMID: 14757276), Tfap2c (PMID: 1989904) and Grhl3 (PMID:
249 16831572).

250

251 Extraembryonic ectoderm lineage

252 Cluster 0, 25 and 28 are Elf5 positive and belong within the extraembryonic ectoderm (Xecto)
253 lineage (PMID: 25446535). Canonical trophoblast stem cell markers like Eomes, Cdx2, Sox2
254 and Esrrb are detected in cluster 25 and 0 (PMID: 25446535, PMID: 11433360). Both are
255 present and have similar proportions within embryos at E6.5. However, cluster 0 persists as

256 development proceeds while cluster 25 rapidly diminishes. Consequently, we termed cluster
257 25 “**extraembryonic ectoderm early**” and cluster 0 “**extraembryonic ectoderm late.**” Both
258 states contain cells expressing *Spry4* as well as *Ets2* and *Ascl2*, indicating that they contain
259 cells of the proximal and distal Xecto/ectoplacental cone (PMID: 25446535). Cluster 28
260 shows substantially lower frequencies of trophoblast stem cell markers and expresses
261 differentiation markers like *Plac1* and *Prl3d1*, indicating that it is comprised of
262 “**differentiated trophoblasts,**” including cells of the ectoplacental cone and trophoblast giant
263 cells (PMID: 25446535, PMID: 18662396).

264
265
266 Comparison with the single-cell reference of WT gastrulation by Pijuan-Sala *et al.* 2019:

267 The annotation of cell states between the single-cell reference described here and the one
268 previously published by Pijuan-Sala and colleagues is overall highly congruent. However, we
269 note some differences outlined in the following:

270 Cells of our **cell states 15 and 41, amnion mesoderm early and late**, respectively,
271 correspond to a cluster termed “mesenchyme” in the reference by Pijuan-Sala *et al.* We do not
272 find any enrichment for mesenchyme markers such as *Fos*, *FN1*, *VTN* and *S100A4* in these
273 cells. However, mesenchyme encompasses many cells of the mesodermal lineage, including
274 the amnion mesenchyme as part of the extraembryonic mesenchyme, which is where the
275 annotations of both references might describe the same cells using different but overlapping
276 terminologies. Our nomenclature was based on the highly specific amnion mesoderm marker
277 Periostin (*Postn*), which is present in 35% of state 15 cells and in 88% of state 41 cells
278 (PMID: 22966238).

279 Cells of **cell state 12, posterior lateral plate mesoderm**, correspond to a cluster
280 termed “extraembryonic mesoderm” in the reference by Pijuan-Sala *et al.* The majority of
281 these cells express *Foxf1*, which is a shared marker between the extraembryonic and lateral
282 plate mesoderm (PMID: 11124112). However, we concluded that most cells are rather
283 embryonic than extraembryonic (i.e. allantois or amnion) mesoderm based on three
284 observations:

- 285 1. State 12 cells were comparatively low in the expression of allantois markers *Tbx4* (7%
286 positive cells) and *Tbx20* (9% positive cells), whereas in the allantois (state 5) these
287 markers were detected in 38% and 58% of cells, respectively.
- 288 2. The amnion marker *Postn* was not detected in state 12 cells, whereas it was detected in
289 35% and 88% of cells in amnion mesoderm early and late (state 15 and 41), respectively.
- 290 3. We detected the intermediate mesoderm marker *Osr1* in 49% of cells in state 12,
291 indicative of embryonic rather than extraembryonic mesoderm. *Osr1* and other markers
292 indicated that state 12 might be composed of posterior mesodermal cells from different
293 structures in addition to the posterior lateral plate mesoderm. Application of our cell state
294 data to a companion molecular recorder publication (Chan, Smith *et al.* 2019) also
295 suggests that posterior lateral plate clusters with the splanchnic lateral plate by shared
296 ancestry (see **Extended Data Fig. 8d** of Chan, Smith *et al.* 2019).

297 We also noted that in our reference a small subset of state 10 cells, annotated as
298 preplacodal ectoderm, is transcriptionally similar to and clusters near state 7, the gut
299 endoderm. These cells are assigned to “gut” using the Pijuan-Sala *et al.* criteria. This
300 discrepancy is apparent when cross-referencing cell state definitions using scRNA-seq data
301 from E8.5 embryos and can be seen in our Uniform Manifold Approximation and Projection
302 plot in **Fig. 1b** (see dark green state 10 cells close to yellow cells of state 7). Though this may
303 represent a minor artifact of our strategy, these cells likely represent a subtly different
304 subcluster within the gut endodermal lineage that warrants further investigation.

305