

## Supplementary Materials for

### **Extracellular vesicle–encapsulated IL-10 as novel nanotherapeutics against ischemic AKI**

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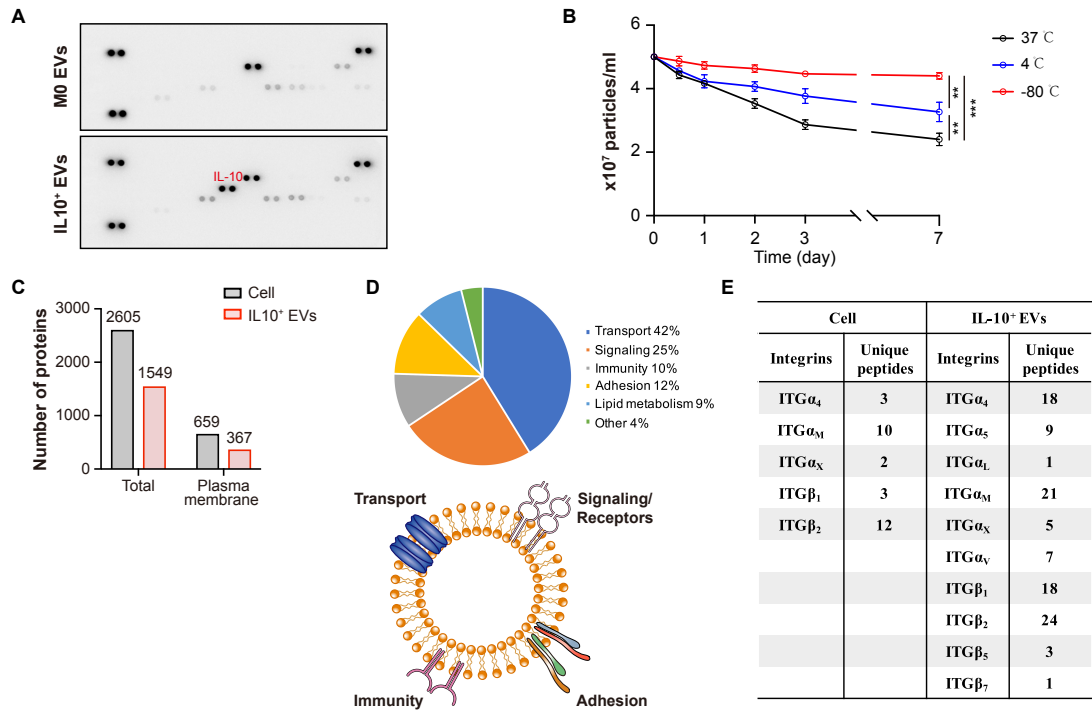
#### **The PDF file includes:**

Figs. S1 to S9  
Table S1

#### **Other Supplementary Material for this manuscript includes the following:**

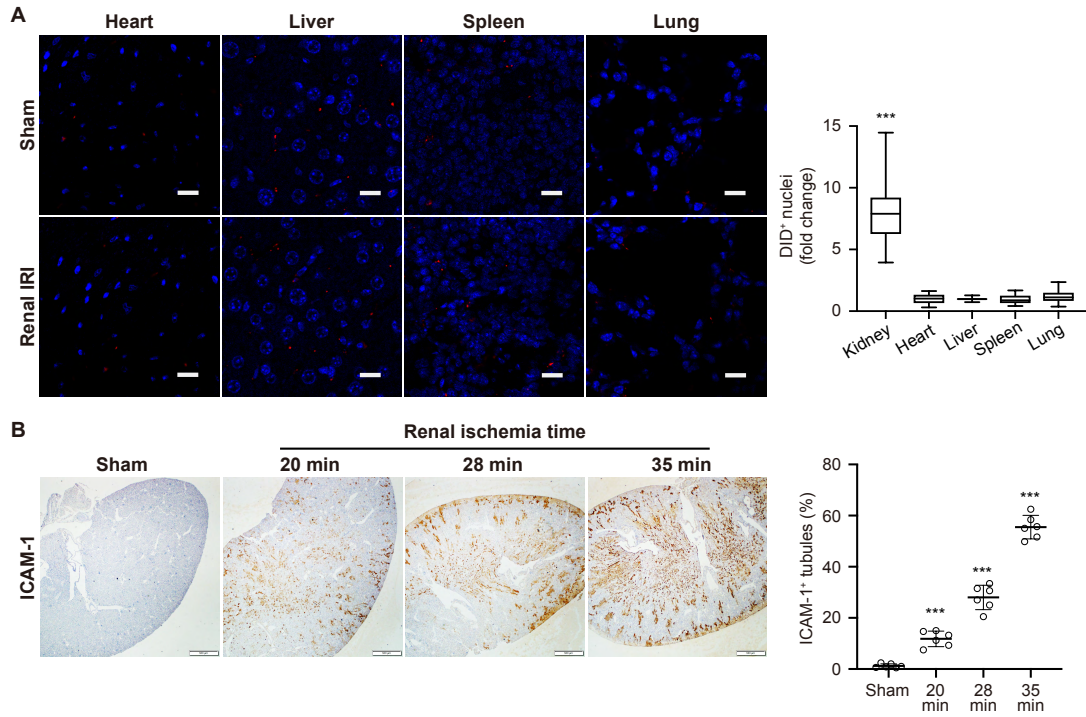
(available at [advances.sciencemag.org/cgi/content/full/6/33/eaaz0748/DC1](https://advances.sciencemag.org/cgi/content/full/6/33/eaaz0748/DC1))

Data file S1



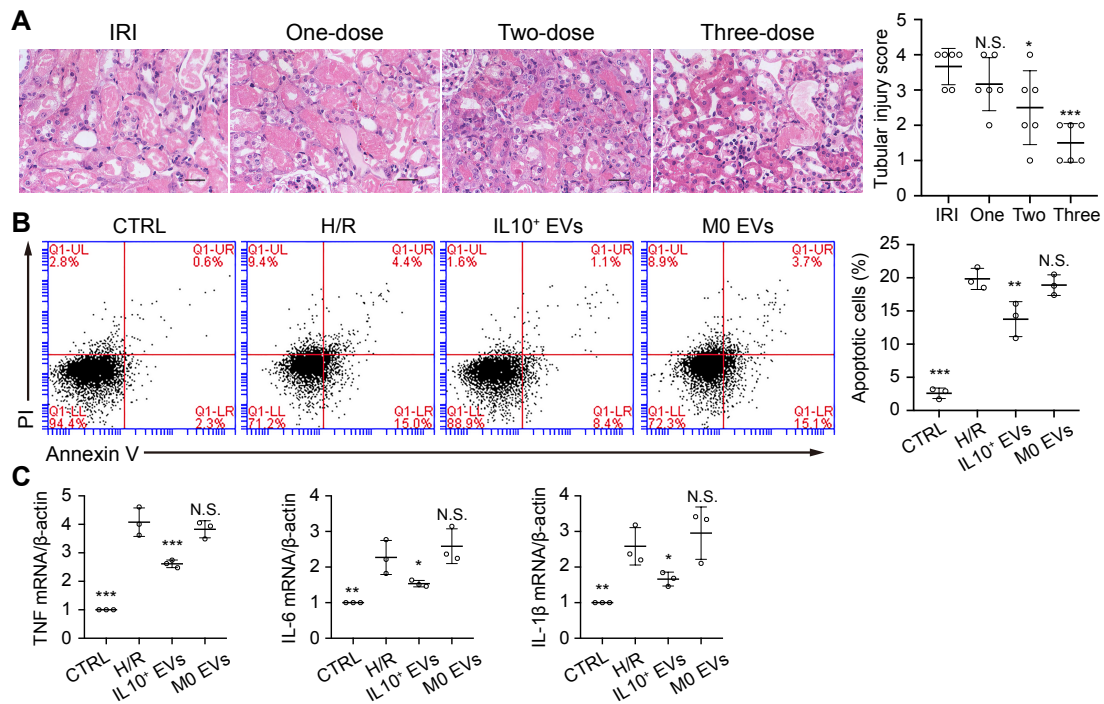
### Figure S1 Characterization of the IL-10<sup>+</sup> EVs.

(A) Cytokine antibody array analysis of IL-10<sup>+</sup> EVs and M0 EVs (EVs from untreated RAW cells). (B) Assessment of the stability of IL-10<sup>+</sup> EVs. IL-10<sup>+</sup> EVs were preserved in PBS at 37 °C, 4 °C or -80 °C for a week, and then EVs were recollected and suspended in 300  $\mu$ L PBS for NTA analysis at different time points (n=3). (C-E) Proteomic profiling of the IL-10<sup>+</sup> EVs. (C) The number of total and plasma membrane-associated proteins. (D) Functional characterization of the plasma membrane proteins identified in the IL-10<sup>+</sup> EVs. (E) Integrin expression identified in IL-10<sup>+</sup> EVs and parental RAW cells. Data are presented as mean  $\pm$  SD. \*\* p<0.01, \*\*\* p<0.001, one-way ANOVA.



**Figure S2 Tissue distribution of IL-10<sup>+</sup> EVs.**

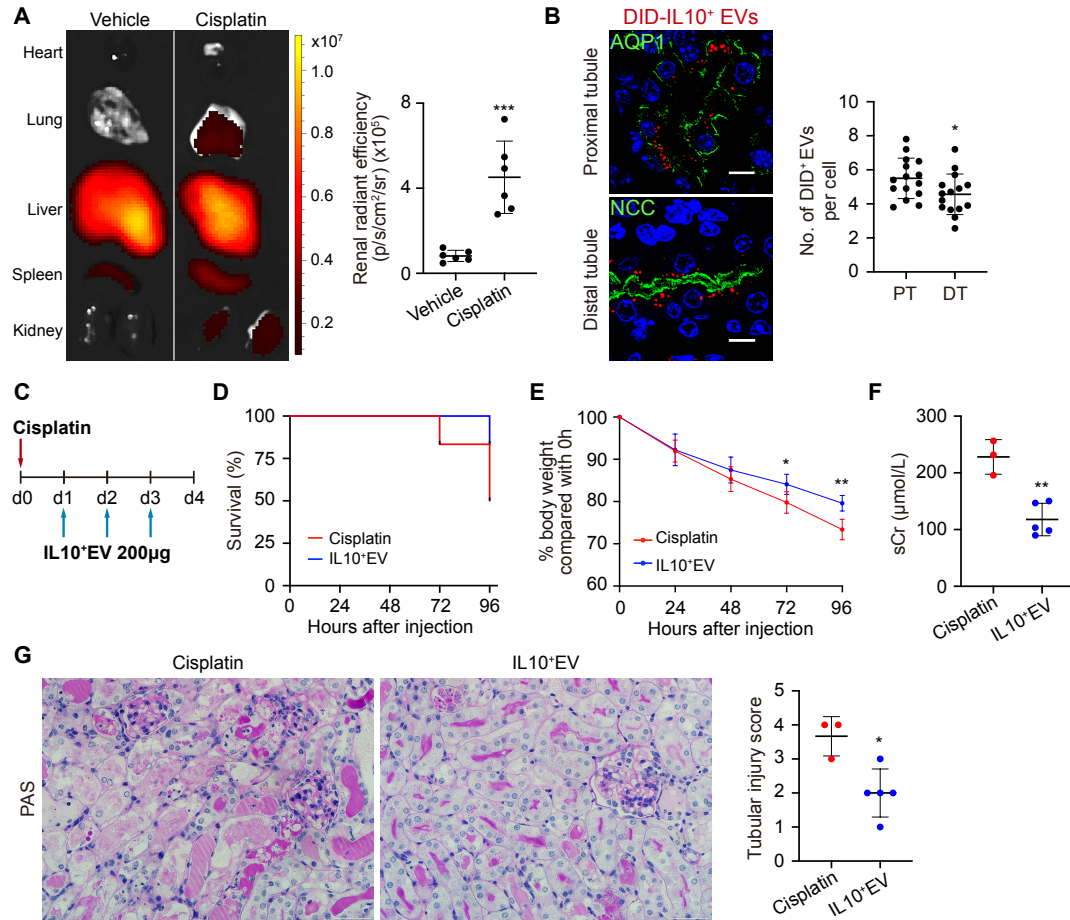
(A) Representative micrographs of the indicated organs of mice injected with DID-labelled IL-10<sup>+</sup> EVs (n=3). scale bar, 25  $\mu$ m. (B) Immunohistochemical analysis of ICAM-1<sup>+</sup> tubules (n=6). scale bar, 500  $\mu$ m. Data are presented as mean  $\pm$  SD. \*\*\* p<0.001, two-tailed t-test (A), one-way ANOVA (B).



**Figure S3 Therapeutic efficacy of IL10<sup>+</sup> EVs.**

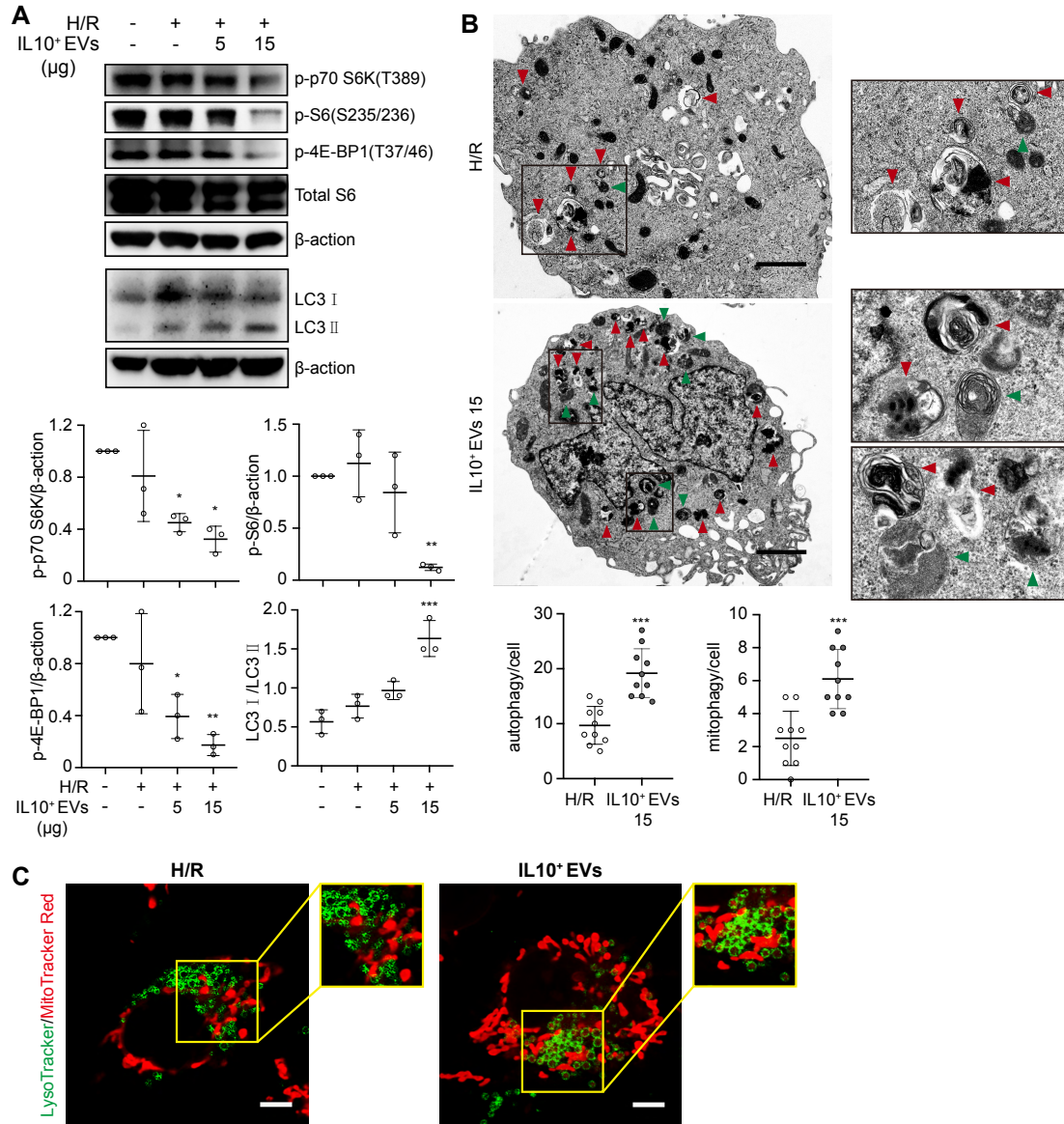
(A) IRI mice were treated with IL-10<sup>+</sup> EVs (200 μg) after reperfusion for different doses and were euthanized at 3 days post-reperfusion (n=6). scale bar, 50 μm. (B,C) Cultured TECs were stimulated with H/R and treated with IL-10<sup>+</sup> EVs (15 μg) or M0 EVs (15 μg) for 12 hours (n=3). (A) Flow cytometry analysis of the apoptosis of TECs. (B) Real-time PCR analysis of inflammatory cytokine mRNA levels in TECs. Data are presented as mean ± SD. \* p<0.05, \*\* p<0.01, \*\*\* p<0.001 vs. IRI or H/R group, one-way ANOVA.





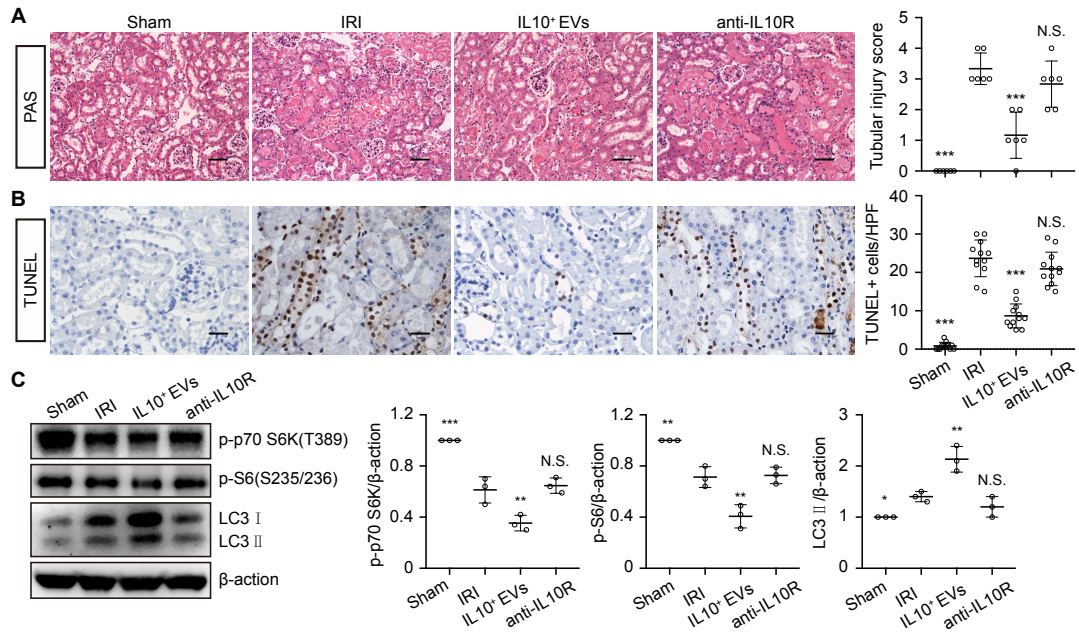
**Figure S4 IL-10<sup>+</sup> EVs protect against cisplatin-induced AKI.**

(**A and B**) Mice were injected intravenously with DID-labeled IL-10<sup>+</sup> EVs (100 µg) 24 hours after cisplatin administration (n=3). (**A**) Imaging of fluorescence intensity of indicated organs at 12 hours after injection. (**B**) Representative confocal images showed DID-labeled IL-10<sup>+</sup> EVs in proximal tubules and distal tubules. scale bar, 10 µm. (**C**) Schematic diagram of the experimental design. In brief, mice were injected intraperitoneally with cisplatin (20 mg/kg) at d0 and then were treated with IL-10<sup>+</sup> EVs (200 µg) every 24 h. (**D**) Survival and (**E**) body weight (vs. 0 hour) over time (n=6). (**F**) Serum creatinine and (**G**) representative images of PAS staining and quantification of tubular injury score at 96 h (cisplatin, n=3; IL-10<sup>+</sup> EVs, n=5). scale bar, 50 µm. Data are presented as mean ± SD. \* p<0.05, \*\* p<0.01, \*\*\* p<0.001, two-tailed t-test.



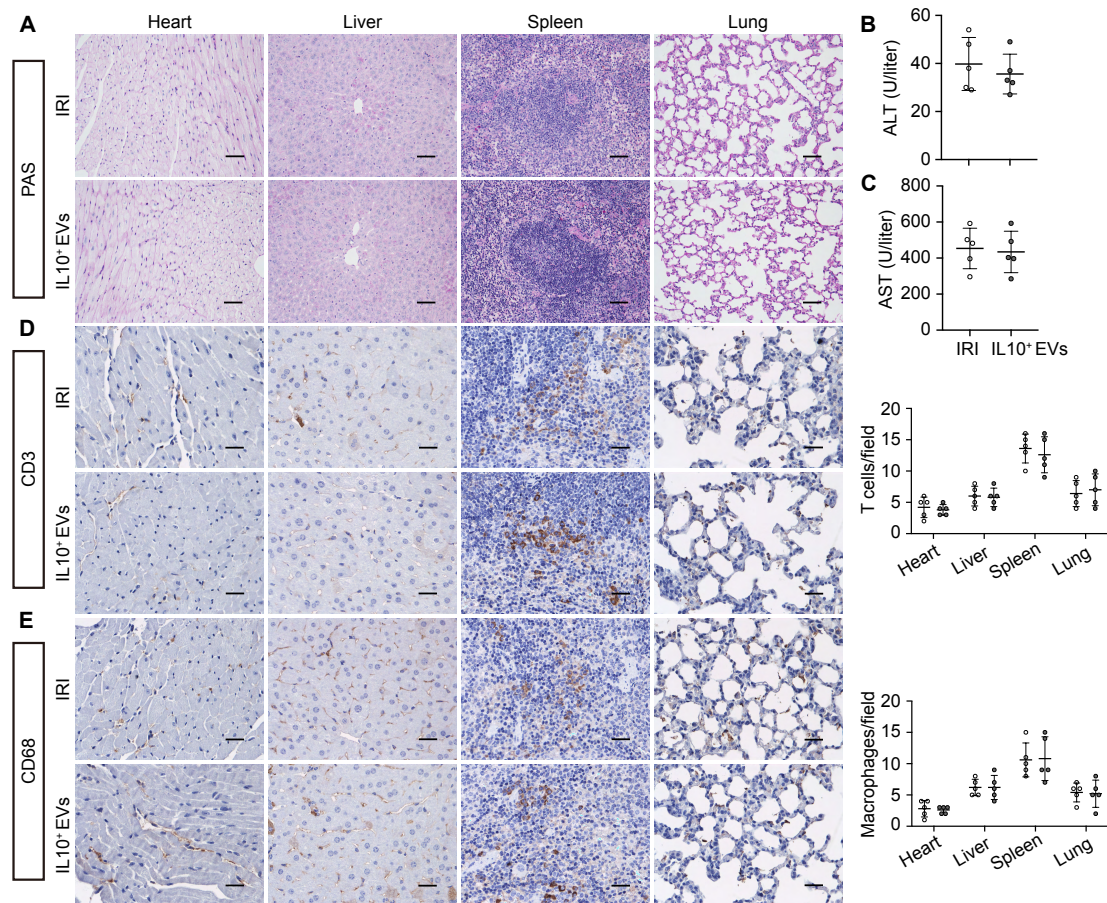
**Figure S5 Induction of mitophagy after IL10<sup>+</sup> EVs treatment in vitro.**

(A) Western blotting analysis of mTOR signaling and LC3 in TECs (n=3). (B) Representative TEM images of autophagic events in TECs. The number of autophagosomes and autolysosomes in each cell were quantified (n=3). Triangle, autophagy; Green triangle, mitophagy. Scale bar, 2 µm. Data are presented as mean ± SD. \* p<0.05, \*\* p<0.01, \*\*\* p<0.001, one-way ANOVA (A), two-tailed t-test (B). (C) Colocalization of lysosome and mitochondria in TECs was assessed using confocal microscopy. TECs were pre-stained with LysoTracker (Green) one day before H/R stimulation. Twelve hours later, TECs were stained with MitoTracker Red. Higher magnification of the indicated area (box) was shown in the right panels. Scale bar, 10 µm.



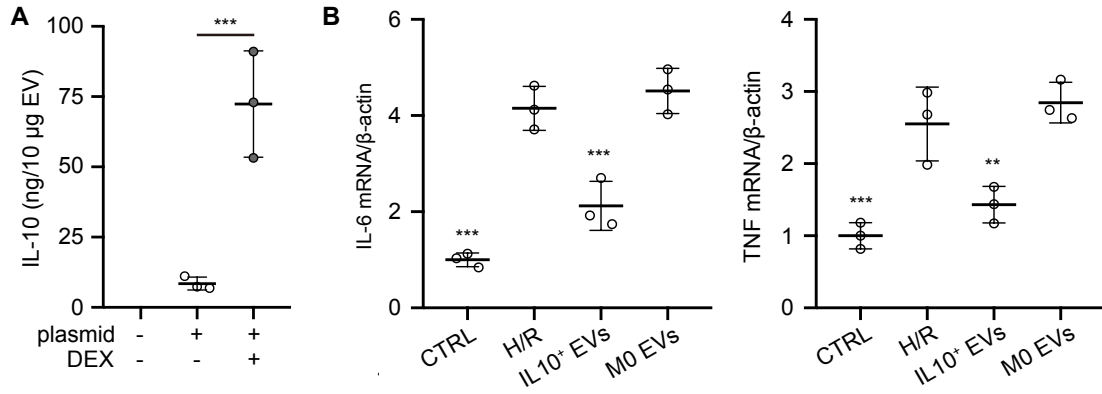
**Figure S6 Blocking IL-10 signaling impairs the therapeutic effects of IL10<sup>+</sup> EVs.** (A) Representative images of H&E staining and quantification of tubular injury (n=6). Scale bar, 100  $\mu$ m. (B) Representative images of TUNEL staining and quantification of the apoptotic cells (n=4). Scale bar, 50  $\mu$ m. (C) Western blotting analysis of mTOR signaling and LC3 in kidney tissues (n=3). Data are presented as mean  $\pm$  SD. \*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$  vs. IRI group, one-way ANOVA.





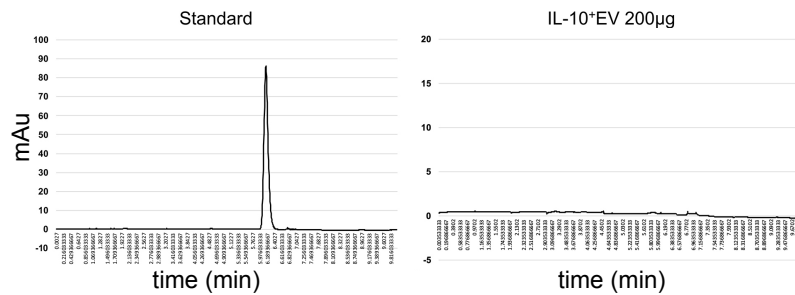
**Figure S7 Assessment of toxicity and inflammation in IL-10<sup>+</sup> EVs-treated mice.**

(A) Representative images of PAS staining of major organs from untreated IRI mice or IL-10<sup>+</sup> EVs-treated mice. Scale bar, 50  $\mu$ m. Measurement of serum alanine aminotransferase (ALT) (B) and aspartate aminotransferase (AST) (C). Immunohistochemical analysis of CD3 T cells (D) and macrophages (E) in tissue sections. Scale bar, 50  $\mu$ m. n=5. Data are presented as mean  $\pm$  SD. two-tailed t-test.



**Figure S8 Manufacturing IL-10<sup>+</sup> EVs using primary cells (BMDM).**

(A) ELISA analysis of IL-10 in BMDM-derived EVs. (B) Effects of BMDM-derived IL-10<sup>+</sup> EVs on the IL-6 and TNF mRNA levels in H/R-induced TECs. n=3. Data are presented as mean ± SD. \*\* p<0.01, \*\*\* p<0.001 vs. H/R group.



**Figure S9 HPLC analysis of DEX in IL-10<sup>+</sup> EVs.**

HPLC analysis showed there was no DEX encapsulated in IL-10<sup>+</sup> EVs.

**Table 1 Primers used in this study.**

<b>Gene</b>	<b>Forward</b>	<b>Reverse</b>
<b><math>\beta</math>-actin MUS</b>	GGGAAATCGTGCGTGAC	AGGCTGGAAAAGAGCCT
<b>CCL-2 MUS</b>	TTGAGGTGGTTGTGGAAAAGG	GTGCTGACCCCAAGAAGGAAT
<b>TNF-<math>\alpha</math> MUS</b>	AGACAGAGGCAACCTGACCAC	GCACCACCATCAAGGACTCAA
<b>IL-1<math>\beta</math> MUS</b>	GGTAAGTGGTTGCCCATCAGA	GTCGCTCAGGGTCACAAGAAA
<b>IL-6 MUS</b>	GTCACCAGCATCAGTCCCAAG	CCCACCAAGAACGATAGTCAA
<b>CCL-5 MUS</b>	CAGAATCAAGAAACCCTCTATCCTA	ACTCCCTGCTGCTTTGCCTAC
<b>iNOS MUS</b>	CAGATCGAGCCCTGGAAGAC	CTGGTCCATGCAGACAACCT
<b>CD206 MUS</b>	GTTCTGACTCTGGACACTTGC	TACTTGGACGGATAGATGGAG
<b>Arg1 MUS</b>	GCAGAGGTCCAGAAGAATGG	GGAGAAAGGACACAGGTTGC